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A MANUAL
OF
BACTERIOLOGY

BY
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Epidemiological Society of London, of the Royal Academy of Medicine of
Rome, of the Academy of Medicine of Rio de Janeiro, of the
American Academy of Medicine, etc., etc.

ILLUSTRATED BY HELIOTYPE AND CHROMO-LITHOGRAPHIC PLATES

AND

TWO HUNDRED AND SIXTY-EIGHT ENGRAVINGS

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PREFACE.

THE progress of our knowledge relating to the bacteria has been so rapid and the literature of the subject is now so extensive that it is no small task to keep pace with this progress, even when one has the literature at hand and devotes a large share of his time to bacteriological studies. Fortunately, recent researches in this department of science have been largely made by exact methods and by trained investigators, and the results can be accepted as well established. A manual of bacteriology, therefore, which fairly represents the present state of knowledge, will consist largely of a statement of facts established by experimental data, and cannot fail to be of value to physicians and to advanced students of bacteriology as a work of reference. The present volume is an attempt to supply such a manual, and at the same time a *text book of bacteriology for students* and guide for laboratory work. That portion of the book which is printed in large type will, it is hoped, be found to give an accurate and sufficiently extended account of the most important pathogenic bacteria, and of bacteriological technology, to serve as a text book for medical students and others interested in this department of science. The descriptions of non-pathogenic bacteria, and of the less important or imperfectly described species of pathogenic bacteria, are given in smaller type. In the preparation of this manual various text books in foreign languages have been consulted, and I am especially indebted to the works of Flügge, of Baumgarten, and of Eisenberg. But the descriptions of species and experimental data have been very largely taken from original memoirs. The illustrations also are to a considerable extent reproductions from the original papers of those engaged in research work.

NEW YORK, April 1st, 1892.

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PART FIRST.

CLASSIFICATION, MORPHOLOGY, AND GENERAL BACTERIOLOGICAL TECHNOLOGY.

- I. HISTORICAL. II. CLASSIFICATION. III. MORPHOLOGY. IV. STAINING
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TERIA. X. INCUBATING OVENS AND THERMO REGU-
LATORS. XI. EXPERIMENTS UPON ANIMALS.
XII. PHOTOGRAPHING BACTERIA.

PART FIRST.

I.

HISTORICAL.

It is probable that Leeuwenhoeck, "the father of microscopy," observed some of the larger species of bacteria in fæces, putrid infusions, etc., which he examined with his magnifying glasses (1675), but it was nearly a century later before an attempt was made to define the characters of these minute organisms and to classify them (O. F. Müller, 1773).

In the absence of any reliable methods for obtaining pure cultures, it is not surprising that the earlier botanists, in their efforts to classify microorganisms, fell into serious errors, one of which was to include under the name of infusoria various motile bacteria. These are now generally recognized as vegetable organisms, while the *Infusoria* are unicellular animal organisms.

Ehrenberg (1838), under the general name of *Vibrioniens*, describes four genera of filamentous bacteria, as follows :

1. *Bacterium*—filaments linear and inflexible ; three species.
2. *Vibrio*—filaments linear, snake-like, flexible ; nine species.
3. *Spirillum*—filaments spiral, inflexible ; three species.
4. *Spirochæte*—filaments spiral, flexible ; one species.

These vibrioniens were described by Ehrenberg as "filiform animals, distinctly or apparently polygastric, naked, without external organs, with the body uniform and united in chains or in filiform series as a result of incomplete division."

Dujardin (1841) also placed the vibrioniens of Ehrenberg among the infusoria, describing them as "filiform animals, extremely slender, without appreciable organization, and without visible locomotive organs."

Charles Robin (1853) suggested the relationship of Ehrenberg's vibrioniens with the genus *Leptothrix*, which belongs to the algæ ; and Davaine (1859) insisted that the vibrioniens are vegetable organ-

isms, nearly allied to the algæ. His classification will be found in the "Dictionnaire Encyclop. des Sciences Médicales," art. "Bactéries" (1868). This view is also sustained by the German botanist Cohn and is now generally accepted.

Spallanzani, in 1776, endeavored to show by experiment that the generally received theory of the spontaneous generation of microorganisms in organic liquids was not true. This he did by boiling putrescible liquids in carefully sealed flasks. The experiment was not always successful, but in a certain number of instances the liquids were sterilized and remained unchanged for an indefinite period. The objection was raised to these experiments that the oxygen of the air was excluded by hermetically sealing the flasks, and it was claimed, in accordance with the views of Gay-Lussac, that free admission of this gas was essential for the development of fermentation.

This objection was met by Franz Schulze (1836), who admitted air to boiled putrescible liquids by drawing it through strong sulphuric acid, in which suspended microorganisms were destroyed. He thus demonstrated that boiled solutions, which, when exposed to the air without any precautions, quickly fell into putrefaction, remained unchanged when freely supplied with air which had been passed through an agent capable of quickly destroying all living organisms contained in it.

Schwann (1839) demonstrated the same fact by another method. Air was freely admitted to his boiled liquids through a tube which was heated to a point, which insured the destruction of suspended microorganisms. The same author is entitled to the credit of having first clearly stated the essential relation of the yeast plant—*Saccharomyces cerevisiæ*—to the process of fermentation in saccharine liquids, which results in the formation of alcohol and carbonic acid.

Helmholtz, in 1843, repeated the experiments of Schwann with calcined air, and arrived at similar results—i.e., he found that the free admission of calcined air to boiled organic infusions did not produce fermentation of any kind.

It was objected to these experiments that the air, having been subjected to a high temperature, had perhaps undergone some chemical change which prevented it from inaugurating processes of fermentation.

This objection was met by Schröder and Von Dusch (1854) by a very simple device which has since proved to be of inestimable value in bacteriological researches. These observers showed that a loose plug of cotton, through which free communication with the external air is maintained, excludes all suspended microorganisms, and that

air passed through such a filter does not cause the fermentation of boiled organic liquids.

The experiments of Pasteur and of Hoffman, made a few years later, showed that even without a cotton filter, when the neck of the flask containing the boiled liquid is long drawn out and turned downward, the contents may be preserved indefinitely without change. In this case suspended particles do not reach the interior of the flask, as there is no current of air to carry them upward through its long-drawn-out neck, and they are prevented by the force of gravity from ascending.

Tyndall showed at a later date that in a closed chamber, in which the air is not disturbed by currents, all suspended particles settle to the floor of the chamber, leaving the air optically pure, as is proved by passing a beam of light through such a chamber.

Notwithstanding the fact that the experimenters mentioned had succeeded in keeping boiled organic liquids sterile in flasks to which the oxygen of the air had free access, the question of the possibility of spontaneous generation—*heterogenesis*—still remained unsettled, inasmuch as occasionally a development of bacterial organisms did occur in such boiled liquids.

This fact was explained by Pasteur (1860), who showed that the generally received idea that the temperature of boiling water must destroy all living organisms was a mistaken one, and that, especially in alkaline liquids, a higher temperature was required to insure sterilization. His experiments showed that a temperature of 110° to 112° C. (230° to 233.6° F.), which he obtained by boiling under a pressure of one and a half atmospheres, was sufficient in every case. These experiments, which have been repeated by numerous investigators since, settled the spontaneous-generation controversy; and it is now generally admitted that no development of microorganisms occurs in organic liquids, and no processes of putrefaction or fermentation occur in such liquids, when they are completely sterilized and guarded against the entrance of living germs from without.

Pasteur at a later date (1865) showed that the atmospheric organisms which resist the boiling temperature are in fact reproductive bodies, or *spores*, which he described under the name of “*corpuscles ovoides*” or “*corpuscles brillants*.” Spores had been previously seen by Perty (1852) and Robin (1853), but it was not until 1876 that the development of these reproductive bodies was studied with care by Cohn and by Koch. The last-named observer determined the conditions under which spores are formed by the anthrax bacillus. Five years later (1881) Koch published his valuable researches relating to the resisting power of anthrax spores to heat and to chemical agents.

The development of our knowledge relating to the bacteria, stimulated by the controversy relating to spontaneous generation and by the demonstration that various processes of fermentation and putrefaction are due to microorganisms of this class, has depended largely upon improvements in methods of research. Among the most important points in the development of bacteriological technique we may mention, *first*, the use of a cotton air filter (Schröder and Von Dusch, 1854); *second*, the sterilization of culture fluids by heat (methods perfected by Pasteur, Koch, and others); *third*, the use of the aniline dyes as staining agents (first recommended by Weigert in 1877); *fourth*, the introduction of solid culture media, and the "plate method" for obtaining pure cultures, by Koch in 1881.

The various improvements in methods of research, and especially the introduction of solid culture media and Koch's "plate method" for isolating bacteria from mixed cultures, have placed bacteriology upon a scientific basis, and have shown that many of the observations and inferences of the earlier investigators were erroneous owing to the imperfection of the methods employed.

Since it has been demonstrated that certain infectious diseases of man and the lower animals are due to organisms of this class, physicians have been especially interested in bacteriological researches, and the progress made during the past fifteen years has been largely due to their investigations. It was a distinguished French physician, Davaine, who first demonstrated the etiological relation of a microorganism of this class to a specific infectious disease. The anthrax bacillus had been seen in the blood of animals dying from this disease by Pollender in 1849 and by Davaine in 1850, but it was several years later (1863) before the last-named observer claimed to have demonstrated by inoculation experiments the causal relation of the bacillus to the disease in question.

The experiments of Davaine were not generally accepted as conclusive, because in inoculating an animal with blood containing the bacillus, from an infected animal which had succumbed to the disease, the living microorganism was associated with material from the body of the diseased animal. This objection was subsequently removed by the experiments of Pasteur, Koch, and many others with pure cultures of the bacillus, which were shown to have the same pathogenic effects as had been obtained in inoculation experiments with the blood of an infected animal.

The next demonstration of the causal relation of a parasitic microorganism to an infectious malady was made by Pasteur, who devoted several years to the study of an infectious disease of silkworms which threatened to destroy the silk industry of France—*pébrine*.

In 1873 Obermeier, a German physician, announced the discovery, in the blood of patients suffering from relapsing fever, of a minute, spiral, actively motile microorganism—the *Spirochæte Obermeieri*—which is now generally recognized as the specific infectious agent in this disease.

The very important work of Koch upon traumatic infectious diseases was published in 1878.

In 1879 Hansen reported the discovery of bacilli in the cells of leprous tubercles, and subsequent researches have shown that this bacillus is constantly associated with leprosy and presumably bears an etiological relation to the disease.

In the same year (1879) Neisser discovered the “gonococcus” in gonorrhœal pus.

The bacillus of typhoid fever was first observed by Eberth, and independently by Koch, in 1880, but it was not until 1884 that Gaffky’s important researches relating to this bacillus were published.

In 1880 Pasteur published his memoir upon fowl cholera, and the same year appeared several important communications from this pioneer in bacteriological research upon the “attenuation” of the virus of anthrax and of fowl cholera and upon protective inoculations in these diseases.

In 1880 the present writer discovered a pathogenic micrococcus, which he subsequently named *Micrococcus Pasteuri*, and which is now generally recognized as the usual agent in the production of acute croupous pneumonia—commonly spoken of as the “diplococcus pneumoniae,” but described in the present volume under the name of *Micrococcus pneumoniae crouposa*.

In 1881 several important papers by Koch and his colleagues appeared in the first volume of the “Mittheilungen” published by the Imperial Board of Health of Germany.

The following year (1882) Koch published his discovery of the tubercle bacillus.

The same year Pasteur published his researches upon the disease of swine, known in France as *rouget*.

The same investigator (Pasteur) also published in 1882 his first communication upon the subject of rabies.

Another important discovery was made in 1882 by the German physicians Löffler and Schütz, viz., that of the bacillus of glanders.

Koch published his discovery of the cholera spirillum—“comma bacillus”—in 1884.

The same year (1884) Löffler discovered the diphtheria bacillus.

Another important publication during the same year was that of Rosenbach, who, by the application of Koch’s methods, fixed defi-

nately the characters of the various microorganisms found in pus from acute abscesses, etc.

The tetanus bacillus was discovered in 1884 by Nicolaier, a student in the laboratory of Prof. Flügge, of Göttingen. That this bacillus is the cause of tetanus in man has been demonstrated by the subsequent researches of numerous investigators. For an exact knowledge of its biological characters we are especially indebted to Kitasato.

So far as human pathology is concerned, no important pathogenic microorganism has been discovered since the year 1884 until the present year (1892). After numerous unsuccessful researches by competent bacteriologists, a bacillus has finally been discovered by Pfeiffer, of Berlin, and independently by Canon, which is believed to be the specific cause of influenza.

Having briefly passed in review some of the principal events in the progress of our knowledge in this department of scientific investigation, it will be of interest to students to know something more of the literature of bacteriology. Important papers have appeared in medical and scientific journals in all countries, and research work of value has been done by enthusiastic investigators of nearly every nation. The brilliant pioneer work done by Pasteur and by Koch has attracted to them many pupils and has made France and Germany the leading countries in this line of investigation. The very great advantages of Koch's methods of research, introduced at the commencement of the last decade, have attracted many students from various parts of the world to Berlin, and to other cities of Germany where instruction was to be obtained from some of Koch's earlier pupils. But to-day bacteriological laboratories have been established in all parts of the world, and it is no longer necessary to go to Germany to obtain such instruction. The literature of the subject is, however, largely in the German and French languages. We can only refer here to such periodicals as are principally devoted to bacteriological research work.

The *Zeitschrift für Hygiene* has been published since 1886, and contains numerous valuable papers, contributed for the most part by the pupils of Koch and of Flügge, who are the editors of the journal. Nine volumes, each containing three numbers, have thus far been published (1891).

The *Annales de l'Institut Pasteur* is a monthly journal which has been published since 1888. It is edited by Duclaux, and contains many important papers and reviews, as well as the statistics of the Pasteur Institute relating to preventive inoculations against hydrophobia.

The *Annales de Micrographie* is a monthly journal, published in Paris. It is now (1892) in its fourth year. The principal editor is Miquel.

The *Centralblatt für Bakteriologie und Parasitenkunde* is a weekly journal which has been published by Gustav Fischer, of Jena, since 1887. The editors are Uhlworm, of Cassel; Löffler, at present professor at Greifswald; and Leuckart, professor at Leipzig.

A most important work for students of bacteriology is the *Jahresbericht* of Baumgarten, which has been published since 1885 by Harald Bruhn, Braunschweig, Germany. This gives a brief abstract of nearly every paper of importance relating to the subject which has been published during the year.

II.

CLASSIFICATION.

THE earlier naturalists—Ehrenberg (1838), Dujardin (1841)—placed the bacteria among the infusoria; but they are *now recognized as vegetable microorganisms*, differing essentially from the infusoria, which are unicellular animal organisms. One of the principal points in differentiating animal from vegetable organisms among the lowest orders of living things is the fact that animal organisms receive food particles into the interior of the body, assimilating the nutritious portion and subsequently extruding the non-nutritious residue; vegetable organisms, on the other hand, are nourished through the cell wall which encloses their protoplasm, by organic or inorganic substances held in solution.

Ehrenberg (1838), under the name of vibrioniens, established four genera, as follows:

1. *Bacterium*—filaments linear and inflexible.
2. *Vibrio*—filaments linear, snake-like, flexible.
3. *Spirillum*—filaments spiral, inflexible.
4. *Spirochæte*—filaments spiral, flexible.

Dujardin (1841) united the two genera *Spirillum* and *Spirochæte* of Ehrenberg, and added to the description of the generic characters as follows:

1. *Bacterium*—filaments rigid, with a vacillating movement.
2. *Vibrio*—filaments flexible, with an undulatory movement.
3. *Spirillum*—filaments spiral, movement rotatory.

It will be seen that this classification leaves no place for the motionless bacilli, such as the anthrax bacillus and many others, and does not include the spherical bacteria, now called micrococci.

The classification of Davaine (1863) provides for the motionless, filamentous bacteria, but does not include the micrococci. This author first insisted that the vibrioniens of Ehrenberg are truly vegetable organisms, allied to the algæ. He makes four genera, as follows:

Filaments straight or bent,	{	Moving spontaneously,	{	Rigid <i>Bacterium</i> .
but not in a spiral,		Motionless,		Flexible <i>Vibrio</i> .
				<i>Bacteridium</i> .
Filaments spiral,				<i>Spirillum</i> .

Following Davaine, the French bacteriologists frequently speak of the motionless anthrax bacillus as *la bactérie*.

Hoffman in 1869 included in his classification the spherical bacteria, and pointed out the fact that motility could not be taken as a generic character, as it was not constant in the same species and depended to some extent upon temperature conditions, etc.

Having determined that the bacteria are truly vegetable organisms, the attention of botanists has been given to the question as to what class of vegetable organisms they are most nearly related to. There are decided differences of opinion in this regard. While Davaine, Rabenhorst, and Cohn insist upon their affinities with the algæ, Robin, Nägeli, and others consider them fungi. One of the principal characters which distinguish the algæ from the fungi is the presence of chlorophyll in the former and its absence in the latter. Now, the bacteria are destitute of chlorophyll, and in this regard resemble the fungi; yet in others their affinities with the *Palmellaceæ* and *Oscillatoriaceæ* are unmistakable. It is not necessary, however, that we should consider them as belonging to either of these classes of the vegetable kingdom. By considering them a distinct class of unicellular vegetable organisms, under the general name of bacteria, we may avoid the difficulties into which the botanists have fallen.

We must refer briefly, however, to the classification proposed by some of the leading German botanists.

Nägeli, placing the bacteria among the lower fungi, which give rise to the decomposition of organic substances, divides these into three groups:

1. The *Mucorini*, or mould fungi.
2. The *Saccharomycetes*, or budding fungi, which produce alcoholic fermentation in saccharine liquids.
3. The *Schizomycetes*, or fission fungi, which produce putrefactive processes, etc.

Cohn, under the name of *Schizophytes*, has grouped these low vegetable organisms, whether provided or not with chlorophyll, into two tribes having the following characters:

1. GLÆOGENES—cells free or united into glairy families by an intercellular substance.
2. NEMATOGENES—cells disposed in filaments.

In the first tribe he has placed the genera *Micrococcus* (Hallier), *Bacterium* (Dujardin), *Merismopedia* (Meyer), *Sarcina* (Goodsir), and *Ascococcus* (Billroth), with various genera of unicellular algæ containing chlorophyll.

In the second tribe we have the genera *Bacillus* (Cohn), *Leptothrix* (Kg.), *Vibrio* (Ehr.), *Spirillum* (Ehr.), *Spirochaete* (Ehr.), *Streptococcus* (Billr.), *Cladothrix* (Cohn), and *Streptothrix* (Cohn), associated with genera of green filamentous algæ.

The German botanist Sachs unites the fungi and the algæ into a single group, the *Thallophytes*, in which he establishes two parallel series, one including those containing chlorophyll, and the other without, as follows:

THALLOPHYTES.

Forms with chlorophyll.

Forms without chlorophyll.

Class I.—Protophytes.

A. Cyanophyceæ (Oscillatoriaceæ, etc.).

B. Palmellaceæ.

A. Schizomycetes (Bacteria).

B. Saccharomycetes.

Zopf, who insists upon the polymorphism of these low organisms, divides the bacteria into four groups:

Genera.	
1. COCCACEÆ.—Up to the present time, only known in the form of cocci.	<i>Streptococcus</i> , <i>Merismopedia</i> , <i>Sarcina</i> , <i>Micrococcus</i> , <i>Ascococcus</i> .
2. BACTERIACEÆ.—Have for the most part spherical, rod-like, and filamentous forms; the first (cocci) may be wanting; the last are not different at the two extremities; filaments straight or spiral.	<i>Bacterium</i> , <i>Spirillum</i> , <i>Vibrio</i> , <i>Leuconostoc</i> , <i>Bacillus</i> , <i>Clostridium</i> .
3. LEPTOTRICHEÆ. — Spherical, rod-shaped, and filamentous forms; the last show a difference between the two extremities; filaments straight or spiral; spore formation not known.	<i>Crenothrix</i> , <i>Beggiatoa</i> , <i>Phragmidiothrix</i> , <i>Leptothrix</i> .
4. CLADOTRICHEÆ. — Spherical, rod-shaped, filamentous, and spiral forms; the filamentous form presents pseudo-branches; spore formation not known.	<i>Cladothrix</i> .

The main objection to this classification is that it assumes a pleomorphism for the bacteria of the second group—Bacteriaceæ—which has only been established for a few species, and which appears not to be general among the rod-shaped and spiral bacteria.

De Bary divides the bacteria into two principal groups, one including those which form endospores, and the other those which are reproduced by arthrospores. But our knowledge is yet too imperfect to make this classification of value, and the same may be said of Hueppe's recent attempt at classification, in which the mode of reproduction is a principal feature.

The classification of Baumgarten (1890) appears to us to have more practical value, and, with slight modifications, we shall adopt it in the present volume. This author divides the bacteria into two principal groups, as follows:

GROUP I. Species relatively monomorphous.

GROUP II. Species pleomorphous.

The first group includes the *micrococci*, the *bacilli*, and the *spirilla*; the second group the *spirulina* of Hueppe, *leptotricheæ* (Zopf), and *cladotricheæ*.

The pleomorphous species described by Hauser under the generic name *Proteus* are included in the second group among the *spirulina*. In the present volume we have described these pleomorphous species among the *bacilli*.

The COCCI, in the classification of Baumgarten, constitute a single genus with the following subgenera: 1, *Diplococcus*; 2, *Streptococcus*; 3, *Merismopedia* (Zopf)—“*Merista*” (Hueppe); 4, *Sarcina* (Goodsir); 5, *Micrococcus* (“*staphylococci*”).

The BACILLI are included in a single genus embracing all of

those species which only form rod-shaped cells, and filaments composed of red-like segments; or straight filaments not distinctly segmented, which may be rigid or flexible.

The SPIRILLA are also included in a single genus, embracing all of those species in which the filaments are spiral in form and the segments more or less spiral or "comma-shaped"—filaments either rigid or flexible.

This simple morphological classification of the monomorphous group of Baumgarten corresponds with the nomenclature now generally in use among bacteriologists. We speak of the spherical bacteria as *cocci* or as *micrococci*, of the rod-shaped bacteria as *bacilli*, and of the spiral bacteria as *spirilla*.

It is true, however, that we are sometimes embarrassed to decide whether a particular microorganism belongs to one or the other of these morphological groups or so-called genera. Among the bacilli, for example, we may have, in the same pure culture, rods of very different lengths, some being so short that if alone they might be taken for cocci, while others may have grown out into long filaments. But if we are assured that the culture is pure the presence of rod forms establishes the diagnosis, and usually the cocci-like elements, when carefully observed, will be seen to be somewhat longer in one diameter than in the other. The German bacteriologists generally insist upon placing among the bacilli all straight bacteria in which, as a rule, one diameter is perceptibly greater than that transverse to it; and several species of well-known bacteria which were formerly classed as micrococci are now called bacilli—e.g., Friedländer's bacillus ("pneumococcus"), *Bacillus prodigiosus*.

The distinction made by Cohn and others between the genus *Bacterium* (Duj.) and the genus *Bacillus* (Cohn) cannot be maintained, inasmuch as we may have short rods and quite long filaments in the same pure culture of a single species; and, again, the character upon which the genus *Vibrio* (Ehr.) was established—viz., the fact that the filaments are flexible and the movements sinuous—is not a sufficient generic or even specific character, for in a pure culture there may be short rods which are rigid, and long filaments which are flexible and have a sinuous movement. We therefore to-day speak of all the elongated forms as bacilli, unless they are spiral and have a corkscrew-like motion, in which case they are known as spirilla.

The bacteria are also classified according to their biological characters, and it will be necessary to consider the various modes of grouping them from different points of view other than that which relates to their form. This is the more important inasmuch as we are not able to differentiate species by morphological characters

alone. Thus, for example, there are among the spherical bacteria, or micrococci, numerous well-established species which the most expert microscopist could not differentiate by the use of the microscope alone; the same is true of the rod-shaped bacteria. The assumption often made by investigators who are not sufficiently impressed with this fact, that two microorganisms from different sources, or even from the same source, are the same because stained preparations examined under the microscope look alike, has led to serious errors and to much confusion. As an example of what is meant we may refer to the pus organisms. Before the introduction of Koch's "plate method" micrococci had been observed in the pus of acute abscesses. Some of these were grouped in chains—streptococci—and some were single, or in pairs, or in groups of four; but whether these were simply different modes of grouping in a single species, or whether the chain micrococci represented a distinct species, was not determined with certainty. That there were in fact four or more distinct species to be found in the pus of acute abscesses was not suspected until Rosenbach and Passet demonstrated that this is the case, and showed that not only is the streptococcus a distinct species, but that among the cocci not associated in chains there are three species which are to be distinguished from each other by their color when grown on the surface of a solid culture medium. One of these has a milk-white color, one is of a lemon-yellow color, while the third is a golden-yellow.

Those microorganisms which form pigment are called *chromogenes*, or chromogenic; those which produce fermentations are spoken of as *zymogenes*, or zymogenic; those which give rise to disease processes in man or the lower animals are denominated *pathogenes*, or pathogenic. We cannot, however, classify bacteria under the three headings chromogenes, zymogenes, and pathogenes, for some of the chromogenic species are also pathogenic, as are some of the zymogenes. These characters must therefore be considered separately as regards each species, and in studying its life history and distinguishing characters we determine whether it is chromogenic or non-chromogenic; whether it produces special fermentations; and whether it is or is not pathogenic when inoculated into the lower animals. In making the distinction between pathogenic and non-pathogenic microorganisms we must remember that a certain species may be pathogenic for one animal and not for another. Thus the anthrax bacillus, which is fatal to cattle, sheep, rabbits, guinea-pigs, and mice, does not kill white rats; the bacillus of mouse septicæmia kills house mice, but field mice are fully immune from its pathogenic effects; on the other hand, the bacillus of glanders is fatal to field mice but not to house mice.

Again, it must be remembered that pathogenic power also depends, to a greater or less extent, upon the dose injected into an animal as compared to its body weight. Some pathogenic organisms in very minute doses give rise to a fatal infectious malady; others are only able to overcome the vital resisting power of the tissues and fluids of the body when introduced into the circulation, or into the subcutaneous tissue or abdominal cavity, in considerable amounts. Some pathogenic bacteria invade the blood; others multiply only in certain tissues of the body; and others again multiply in the intestine and by the formation of poisonous products which are absorbed show their pathogenic power.

Another classification of the bacteria relates to the environment favorable to their development. Thus we speak of saprophytic and parasitic bacteria, or of SAPROPHYTES and PARASITES.

The *saprophytes* are such as exist independently of a living host, obtaining their supply of nutriment from dead animal or vegetable material and from water containing organic and inorganic matters in solution. The *strict parasites*, on the other hand, depend upon a living host, in the body of which they multiply, sometimes without injury to the animal upon which they depend for their existence, but frequently as harmful invaders giving rise to acute or chronic infectious diseases. Microorganisms which ordinarily lead a saprophytic existence, but which can also thrive within the body of a living animal, are called *facultative parasites*. Thus the leprosy bacillus, which is only found in leprosy tissues, is a strict parasite; while the typhoid bacillus, the cholera spirillum, etc., are facultative parasites, inasmuch as they are capable of maintaining an independent existence, for a time at least, external to the bodies of living animals.

It seems probable that the pathogenic organisms which are only known to us to-day as strict parasites were, at some time in the past, saprophytes, which gradually became accustomed to a parasitic mode of existence, and, under the changed conditions of their environment, finally lost the power of living in association with other saprophytes exposed to variations of temperature, etc. The tubercle bacillus, for example, is known to us only as a parasite which has its habitat in the lungs, lymphatic glands, etc., of man and of certain of the lower animals. But we are able to cultivate it in artificial media external to the body; and it is in accord with modern views relating to the development of species to suppose that at some time in the past it was able to lead a saprophytic existence. Not to admit this forces us to the conclusion that, at some time subsequent to the appearance of man and the lower animals in which it is now found as a parasite, it was created with its present biological characters, which restrict it to a parasitic existence in the bodies of these ani-

mals, and that, consequently, the immense destruction of human life which has resulted from its parasitic invasion of successive generations was designed when it was created. The opposite view is supported by numerous facts which show that these low organisms, like those higher in the scale, are subject to modifications as a result of changed conditions of environment, and that such modifications, in the course of time, may become well-established specific characters.

Again, the bacteria may be grouped into *aërobic* and *anaërobic* species. This is a very important distinction, which was first established by Pasteur, who found that certain bacteria will only grow when freely supplied with oxygen, while others absolutely decline to grow in the presence of this gas. The latter, which are spoken of as *strict anaërobics*, may be cultivated in a vacuum or in an atmosphere of hydrogen. Those species which grow either in the presence of oxygen or when it is excluded are called *facultative anaërobics*.

Certain bacteria produce a peptonizing ferment which has the power of liquefying gelatin. This has led to the classification of those microorganisms of this class which grow in Koch's flesh-peptone-gelatin as *liquefying* and *non-liquefying* bacteria.

Again, we speak of them as *motile* or *non-motile*.

It is evident that these biological characters, although all-important in the definition of species, cannot serve us in an attempt to establish natural genera; for the lines are not sharply drawn between the saprophytes and the parasites, the *aërobics* and the *anaërobics*, etc., inasmuch as we have facultative parasites and facultative *anaërobics* which we cannot include in either class, and which yet do not form a distinct class by themselves. We therefore adhere to the morphological classification, although this is open to criticism. For example, among the rod-shaped organisms which we call bacilli and describe under the generic name *Bacillus* there are some which multiply by binary division only, while others form endogenous reproductive bodies known as *spores*. Certainly so important a difference in the mode of reproduction should be sufficient to separate these rod-shaped organisms into two natural groups or genera.

As heretofore stated, the German bacteriologist Hueppe has attempted a classification based upon the mode of reproduction, in which he makes two groups, or "tribes," one in which reproduction occurs by the formation of endogenous spores—"endospores"—the other in which it occurs by the formation of "*arthrospores*."¹ The latter group includes all of those bacteria in which no other mode of multiplication is known than that by binary division, which is common to all. In the present state of our knowledge this classification

¹ An account of this mode of reproduction is given on page 19.

is scarcely to be considered of practical value, inasmuch as the question of spore formation is still undetermined for a large number of species.

In the following table we shall give the characters of the different genera which have been described by recent botanists and bacteriologists, arranged under the three headings, MICROCOCCI, BACILLI, SPIRILLA. Where we doubt the propriety of maintaining a distinct generic name upon the supposed distinguishing characters, the description will be printed in small type.

MICROCOCCI.

General Characters.—Spherical bacteria which are reproduced by binary division; usually without spontaneous movements; do not form endogenous spores. (According to some authors, certain cells, known as arthrospores, may be distinguished by their greater size and refractive power, and these are supposed to have greater resistance to desiccation than the ordinary cocci resulting from binary division, and to serve as reproductive bodies.) Some micrococci are not precisely round, but are somewhat oval in form; and when in process of division the cocci, necessarily, are more or less elongated in one diameter before a complete separation into two spherical elements has occurred.

MICROCOCCUS.—Division in one direction; cocci single, in pairs, or accidentally associated in irregular groups; sometimes held together in irregular masses by a transparent, glutinous, intercellular substance. (Micrococci belonging to this genus are frequently described as “staphylococci,” and *Staphylococcus* is used by Rosenbach as a generic name for the pus cocci described by him, which are solitary or associated in irregular groups, as above described.)

ASCOCOCCUS.—Cocci associated in globular or lobulated, zoöglœa masses by a rather firm intercellular substance.

LEUCONOSTOC.—Cocci, solitary or in chains, surrounded by a thick, gelatinous envelope and forming zoöglœa of cartilaginous consistence.

STREPTOCOCCUS.—Division in one direction only; cocci associated in chains.

Diplococcus.—Division in one direction only; cocci associated in pairs.

Association in pairs is common to all of the micrococci, inasmuch as they multiply by binary division. When such association has rather a permanent character, it is customary to speak of the microorganism as a diplococcus, but we doubt the propriety of recognizing this mode of association as a generic character.

MERISMOPEDIA.—Division in two directions, forming groups of four, which remain associated in a single plane—“tetrads.”

SARCINA.—Division in three directions, forming packets of eight

or more elements, which remain associated in more or less regular cubical masses.

BACILLI.

General Characters.—Rod-shaped and filamentous (not spiral) bacteria in which there is no differentiation between the extremities of the rods; reproduction by binary division in a direction transverse to the long axis of the rods, or by binary division and the formation of endogenous spores; rigid or flexible; motile or non-motile.

BACILLUS.—Characters as given above.

Bacterium.—This genus, established by Dujardin, is now generally abandoned, the species formerly included in it being transferred to the genus *Bacillus*. As defined by Cohn, the generic characters were: Cells cylindrical or elliptical, free or united in pairs during their division, rarely in fours, never in chains, sometimes in zoöglœa (differing from the zoöglœa of spherical bacteria by a more abundant and firmer intercellular substance), having spontaneous movements, oscillatory and very active, especially in media rich in alimentary material and in presence of oxygen.

Clostridium.—Rod-shaped bacteria which form large, endogenous, and usually oval spores; these are centrally located, and during the stage of spore formation the rods become fusiform.

SPIRILLA.

General Characters.—Curved rods or spiral filaments; rigid or flexible; reproduction by binary division, or by binary division and the formation of endogenous spores (or by arthrospores?); movements rotatory in the direction of the long axis of the filaments.

SPIRILLUM.—Characters as above.

Spirochæte.—Flexible, spiral filaments; movements rotatory.

Vibrio.—Filaments flexible, straight or sinuous; movements sinuous.

A considerable number of bacteria which are usually seen as short, curved rods, but which may grow out into long, spiral filaments, are described by some authors under the generic name *Vibrio*, e.g., the so-called "comma bacillus" of Koch—"Spirillum cholerae Asiaticæ"; the spirillum of Finkler and Prior—"Vibrio proteus"; the spirillum described by Gameléia—"Vibrio Metschnikovi," etc. These microorganisms have not the characters which distinguished the genus *Vibrio* as established by Ehrenberg, and we prefer to follow Flügge in describing them under the generic name *Spirillum*.

The pathogenic bacteria now known belong to one or the other of the above-described genera, and the attention of bacteriologists has been given chiefly to the study of micrococci, bacilli, and spirilla. But the botanists place among the bacteria certain other forms which are found in water, and which, in a systematic account of this class of microorganisms, demand brief attention at least. These are included in Baumgarten's second group, which includes the pleomorphic bacteria.

SPIRULINA (Hueppe).—The vegetative cells are sometimes rod-shaped and sometimes spiral; in suitable media they may grow out

into long, straight, wavy, or spiral filaments. These filaments may break up into cocci-like reproductive elements—"arthrospores."

LEPTOTRICHEÆ (Zopf).—The vegetative cells present rod-shaped and spiral forms, and grow out into straight, wavy, or spiral filaments; these may show a difference between the two extremities, of base and apex. Cocci-like reproductive bodies are formed by segmentation of the rod-shaped elements in these filaments. In some of the species the segments are enclosed in a common sheath. *Subgenera*: **LEPTOTHRIX**, **BEGGIATOA**, **CRENOTHRIX**, **PHRAGMIDIOTHRIX** (for generic characters see page 12).

CLADOTRICHEÆ (Zopf).—The vegetative cells are rod-shaped or spiral, and grow out into straight or spiral filaments, which may present pseudo-ramifications. A single genus, **CLADOTHRIX** (see page 12).

III.

MORPHOLOGY.

IN the present chapter we shall give a general account of the morphology, modes of grouping, and dimensions of the bacteria.

The standard of measurement used by bacteriologists is the micromillimetre, or the one-thousandth part of a millimetre. This is represented by the Greek letter μ . One μ (micromillimetre) is equal to about one-twenty-five-thousandth of an English inch.

The spherical bacteria, or micrococci, differ greatly in size, and also in the mode of grouping when, as a result of binary division, they remain associated one with another. The smallest may measure no more than 0.1μ , while some of the larger species are from one to two μ in diameter. The enormous number of these minute organisms which may be contained in a small drop of a pure culture may be easily estimated in a rough way. Compare a single micrococcus, for example, with a sphere having a diameter of one-twenty-fifth of an inch. If our micrococcus is one of the larger sort, having a diameter of one μ , it would take a chain of one thousand to reach across the diameter of such a sphere, and its mass, as compared to the larger sphere, would be as 1 to 523,600,000.

The number of cocci in a milligramme of a pure culture of *Staphylococcus pyogenes aureus* has been estimated by Bujwid, by counting, at 8,000,000,000.

Not only do different species differ in dimensions, but considerable differences in size may be recognized in the individual cocci in a pure culture of the same species. On the other hand, there are numerous species which so closely resemble each other in size and mode of association that they cannot be differentiated by a microscopic examination alone, and we must depend upon other characters, such as color, growth in various culture media, pathogenic power, etc., to decide the question of identity or non-identity.

When in active growth the micrococci necessarily depart from a typical spherical form just before dividing, and under these circumstances may be of a short or long oval. When division has taken place, if the two members of a pair remain associated they are often more or less flattened at the point of contact (Fig. 1, a).

The *staphylococci* are characterized by the fact that, for the most part, the individual cocci in a culture are solitary (Fig. 1, *b*). But, inasmuch as multiplication occurs by binary division, we also have pairs and occasionally a group of four—probably from the accidental apposition of two pairs (Fig. 1, *c*). When in a culture the cocci are for the most part associated in pairs (Fig. 1, *d*), we speak of the organism as a *diplococcus*. Frequently after staining

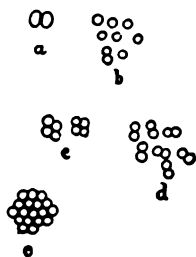


FIG. 1.

and mounting a preparation we find that the cocci are associated in irregular groups, although we may have endeavored to distribute them in a drop of distilled water. This results from the fact that they are surrounded by a glutinous material which causes them to

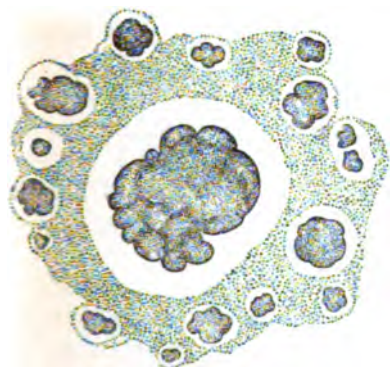


FIG. 3.



FIG. 2.

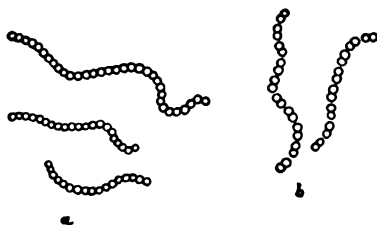


FIG. 4.

adhere to each other (Fig. 1, *e*). A mass of cocci held together in this way by a transparent, glutinous, intercellular substance is spoken of as a *zoöglæa* (Fig. 2). In the genus *Ascococcus* the intercellular substance is quite firm and the *zoöglæa* are in the form of spherical or irregularly lobulated masses surrounded by a resistant envelope of jelly-like material (Fig. 3).

When, as a result of division in one direction only, the cocci

remain united in chains (Fig. 4, *a*), they are described as *streptococci*, and are sometimes spoken of as in *chaplets* or in *torula chains*. In such chains we frequently find the evidence of recent division of the cocci, as shown by the grouping of the elements of the chain into pairs (Fig. 4, *b*).

When division occurs habitually in two directions, groups of four result, which are spoken of as *tetrads*. This is the distinguishing character of the genus *Merismopedia*. In these groups of four the individual cocci are often flattened at the points of contact, as in Fig. 5, *b*. We also find pairs and groups of three in pure cultures of species belonging to this genus, as shown in Fig. 5, *c*. In these, transverse division has not yet occurred in one or in both elements of a pair. This association of micrococci in tetrads seems to be maintained, in some species at least, by the fact that each group of four is enclosed in a jelly-like *capsule*. The extent of this capsule differs in the same species under different circumstances; as a rule, it is most apparent when a culture has been made in a liquid medium. Some of

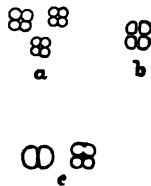


FIG. 5.

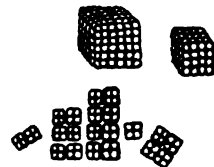


FIG. 6.

the diplococci have a similar capsule. The jelly-like substance does not stain well with the aniline colors and is seen as a transparent halo around the stained cocci. Some authors (Fränkel and Pfeiffer) believe that this capsule is formed by the swelling up of the cell membrane as a result of the imbibition of water.

When division occurs in three directions packets of eight or more elements are formed. This mode of association characterizes the genus *Sarcina*. The "packet form" is best seen in an unstained preparation from a fresh culture, in which a little material suspended in water is examined under a comparatively low-power objective—one-sixth (Fig. 6).

Among the bacilli there is room for a wider range of morphological characters. They differ not only in dimensions and in modes of grouping, but in form. The relation of the transverse to the longitudinal diameters affords a great variety of forms, varying from a short oval element to a slender rod or elongated filament. But it must be remembered that we may have short rods and long filaments in a pure culture of the same bacillus—the typhoid bacillus, for

example. There are also considerable differences in the transverse diameter of bacilli belonging to the same species when cultivated in different media, or even in the same medium, although, as a rule, the transverse diameter is tolerably uniform in pure cultures.

Again, the form of the extremities of the rods is to be observed (Fig. 7). This may be square, or the corners may be slightly rounded, or the extremities may be quite round or lance-oval, or the outlines of the rod may be spindle-shaped from the formation of

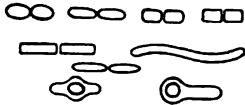


FIG. 7.



FIG. 8.

a large central spore—"clostridium"—or one end may be dilated from the formation of a large terminal spore.

In old cultures we frequently find irregular forms due to swellings and constrictions, which probably occur in bacilli which have but little vitality or are already dead. These are spoken of as *involution forms* (Fig. 8).

The bacilli multiply by binary division in a direction transverse to the longitudinal axis, and, as a result of such binary division, long



FIG. 9.

chains in which the elements remain associated may be formed (Fig. 9); or the rods may be for the most part solitary or united in pairs. Like the micrococci, the bacilli are sometimes surrounded by a gelatinous envelope or capsule. They may also be united by a glutinous material into zoöglœa masses.

Bacilli which under certain conditions are seen as short rods may, under other circumstances, grow out into long filaments, and these may be associated in bundles or in tangled masses.

The spirilla differ from the bacilli in the form of the rods and fila-

ments, which are curved or spiral. The shorter elements in a pure culture may be simply curved, as in *a*, Fig. 10, while the spiral form becomes apparent in those which are longer, and we may have one or several turns of the spiral (Fig. 10, *b*). The spiral form may be but slightly marked (Fig. 10, *c*), or the turns may be close and deep as in a corkscrew (Fig. 10, *d*). Again, the curved filaments may be short and rigid, or long and flexible (Fig. 10, *e*).

In the genus *Cladothrix*, which is placed by botanists among the bacteria, the filaments appear to branch; but this branching is only apparent, and there is no true dichotomous branching in this class of microorganisms. The false branching of *Cladothrix dichotoma*, Cohn, is shown in Fig. 11. The fact that some of the larger species of bacilli and spirilla are provided with slender, whip-like appendages called *flagella* has been known for many years, and it has for some time been suspected that all of the motile organisms

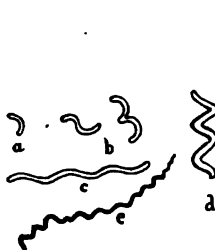


FIG. 10.

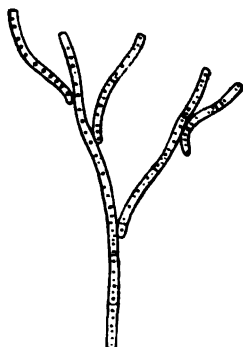


FIG. 11.

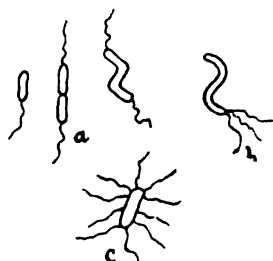


FIG. 12.

of this class are provided with similar appendages and that these are organs of locomotion. Recently, by improvements in methods of staining, Löffler has demonstrated the presence of flagella in many species in which they had heretofore escaped observation. They are sometimes single, at the ends of the rods (Fig. 12, *a*); or there may be several at the extremity of a single rod (Fig. 12, *b*); again, they are seen in considerable numbers around the periphery of the rod (Fig. 12, *c*).

The bacilli and spirilla sometimes contain in the interior of the cells granules of different kinds. These may appear like little oil drops or they may be more opaque. In the genus *Beggiatoa* grains of sulphur are found in the interior of the cells. Again, we may find vacuoles in the protoplasm; or, in stained preparations, deeply stained granules, which are not spores, may be seen at the extremities of the rods—end-staining. The morphological characters depending upon the formation of endogenous spores will be referred to hereafter.

IV.

STAINING METHODS.

THE rapid development of our knowledge with reference to the minute microorganisms under consideration depends very largely upon the discovery that they may be stained by various dyes, and especially by the aniline colors. Weigert (1876) was the first to employ these colors in studying the bacteria, and Koch at once recognized the value of the method and made use of it in his researches.

The basic aniline colors are those employed, and among these the most useful are fuchsin, methylene blue, gentian violet, Bismarck brown, and vesuvin.

Staining upon the Cover Glass or Slide.—By a “cover-glass preparation” we mean that material supposed to contain bacteria has been spread out upon a thin glass cover, dried, and stained for microscopical examination. A small drop of a liquid culture may, for



FIG. 13.

example, be spread upon a perfectly clean cover glass by means of a platinum wire held in a glass handle (Fig. 13). Or we may place a drop of water in the centre of the thin glass cover, and by means of the same instrument take a little material from a culture made upon the surface of a solid medium and distribute it through the drop. In this case we must be careful to take very little of the material, as the smallest quantity will contain an immense number of bacteria, and for a satisfactory view of the individual cells it is necessary that they be well separated from each other, in some parts of the preparation at least, and not massed together.

Where the object is to make a cabinet preparation for permanent preservation, special care should be taken to distribute the bacteria uniformly through the drop of water. The next step consists in evaporating the liquid so that the bacteria may remain attached to the surface of the glass cover. This may be done by simple exposure to the air or by the application of gentle heat. When the bacteria are

suspended in an albuminous medium it will be necessary, after the film is dry, to heat the preparation sufficiently to coagulate the albumen, in order that it may not be washed off in the subsequent staining process. This is best done, in accordance with Koch's directions for the preparation of tuberculous sputum, by passing the cover glass, held in slender forceps, rather quickly through the flame of an alcohol lamp three times in succession. In this operation it must be remembered that too much heat will destroy the preparation, while too little will fail to accomplish the object in view—coagulation of the albumen. In passing the cover glass through the flame the smeared side is to be held upward. The time required will be about three seconds for passing it three times as directed; but this will vary according to the intensity of the flame, and some little experience is necessary in order to obtain the best results.

The operation of "fixing," or coagulating the albumen, may also be effected by exposure in a dry-air oven, heated to 120° to 130° C., for a few minutes (two to ten minutes), as directed by Ehrlich.

Bacteria simply suspended in distilled water adhere very well to the cover glass when treated as directed, but if they have been taken from a liquefied gelatin culture the film is very apt to be washed away during the staining process. This is best avoided by taking as little as possible of the gelatin medium and suspending the bacteria to be examined in a drop of water, which dilutes the gelatin and washes it away from the surface of the cells.

Smear Preparations.—In various infectious diseases bacteria are found in the blood and tissues of the body, and their presence may be demonstrated by making what is called a smear preparation. A little drop of blood may be spread upon the thin glass cover, or it may be brought in contact with the freshly cut surface of one of the vascular organs, as the liver or spleen. It is especially desirable that the material used for such a preparation be small in amount and distributed evenly in a very thin layer. In Germany it is the custom, in making smear preparations, to press the material between two glass covers, which are then separated by sliding them apart, thus leaving a thin layer upon each. This answers very well, but the writer prefers to spread the material by drawing across the face of the cover glass the end of a well-ground and polished glass slide. This method is especially useful for spreading blood in a uniform layer, in which the corpuscles are evenly distributed and retain their normal form. A very small drop of blood is placed near one edge of the cover glass, which is placed upon a smooth surface; the glass slide is held at a very acute angle and is gently drawn across the cover glass, without any pressure.

Most bacteriologists make their preparations upon the cover glass,

as above described, but the writer has for a number of years made his mounts of bacteria upon the glass slide, and believes that this method has some advantages for every-day work. The thin glass covers required when a preparation is to be examined with an immersion objective of high power, are easily broken and often dropped from the fingers or forceps. When the material to be examined is spread and dried directly upon the glass slide, the operation is attended with less difficulty and fewer accidents and the results are quite as good. In this case the slide is held in the fingers during the various steps in the operation of distributing, drying, and staining, while the thin glass cover must be held in delicate forceps.

Contact Preparations.—When a dry and clean cover glass is brought in contact with a colony or surface culture we may often obtain a very pretty preparation, showing the bacteria in a single layer, and preserving the arrangement, as regards growth, which characterizes the species. Similar preparations may sometimes be obtained from the surface of liquid cultures, when the bacteria grow upon the surface as a thin film. The cover glass is to be gently brought into contact with this surface growth, which adheres to it and is dried and stained by the usual methods.

Staining of the dried film is quickly effected by using an aqueous solution of one of the aniline colors above mentioned. For general use the writer prefers a solution of fuchsin, on account of the promptness of its staining action, and because, in preparations for permanent preservation, it is not as likely to fade as methylene blue or gentian violet. It is also a better color than blue or violet in case a photograph is to be made from the preparation.

It is best to keep on hand saturated alcoholic solutions of the staining agents named, and to make an aqueous solution whenever required by the addition of a few drops to a little water in a watch glass or test tube; for the aqueous solutions do not keep well on account of the precipitation of the dye as a fine powder, which renders the solution opaque. The addition of ten per cent of alcohol to the aqueous solution will, however, prevent this precipitation; but, as a rule, freshly prepared solutions are the best. These should be filtered before use. We may place a few drops of the filtered solution upon the dried film on the slide or cover glass, or the thin cover may be floated upon a little of the solution in a watch glass. In some cases it is best to use heat to expedite the staining, and this may be done by holding the slide or the watch glass over the flame of an alcohol lamp until steam commences to be given off. If the heating is carried too far the preparation is likely to be spoiled by the precipitation of the staining agent. As a rule, heating will not be necessary, and when an aqueous solution of fuchsin (one part to

one hundred of water) is used most bacteria are stained within a few seconds to a minute. At the end of this time the staining solution is to be washed away by means of a gentle stream of water, or by moving the cover glass about in a vessel containing distilled water.

Decolorization.—It often happens that the albuminous material associated with the bacteria which we propose to examine is stained so deeply as to obscure the view of these; and, generally, we will obtain more satisfactory preparations by the use of a decolorizing agent, by which the background is cleared up and the outlines of the cells more clearly defined. The agents chiefly used for this purpose are alcohol, diluted acids, and solution of iodine with potassium iodide (Gram's solution).

Koch recommends a solution containing sixty parts of alcohol to forty parts of water. The cover glass is to be quickly passed through this solution two or three times. Some bacteriologists prefer to use absolute alcohol.

Or we may use dilute acetic acid (one-half to one per cent) or very dilute hydrochloric acid (ten drops to half a litre of water).

For decolorizing preparations containing the tubercle bacillus strong solutions of the mineral acids are employed (one part of nitric or of sulphuric acid to three parts of water).

Gram's solution contains one part of iodine and two parts of potassic iodide in three hundred parts of water. Special directions will be given for the use of these agents when we give an account of the staining methods most useful for the various pathogenic organisms.

Double Staining.—After decolorizing the background of albuminous material we may again stain this with a contrast stain, such as eosin or vesuvin. In mounts made from pure cultures, either liquid or solid, a single stain, for the bacteria only, is all that we require, and our aim is to have the background as free as possible from any material which would obscure the view.

After staining, decolorizing, and washing the preparation the cover glass or slide is again dried by exposure to the air or gentle heat, and is then ready for the permanent mounting in Canada balsam. If the bacteria have been stained upon the slide, a small drop of balsam dissolved in xylol is placed in the middle of the preparation and a clean, thin glass cover applied.

If it is the intention to make the microscopical examination with an immersion objective of high power, or to make photomicrographs from it, only the thinnest glass covers should be used—one-two-hundredths of an inch or less.

If the preparation is not intended for permanent preservation,

the examination may be made without drying the surface upon which the stained bacteria are spread, the water taking the place of balsam in a permanent mount; or we may dry the film and use a drop of cedar oil between the slide and cover.

While simple aqueous solutions of the aniline colors, when freshly prepared, will promptly stain most bacteria, certain agents may be added to these which aid in the preservation of the solution, or which act as mordants, and are useful in special cases.

We shall only give here a few of the standard solutions which are most frequently employed by experienced bacteriologists:

1. *Aniline-Gentian-Violet* (Ehrlich).

Saturated alcoholic solution of gentian violet,	5 cc.
Aniline water,	100 cc.

2. *Aniline-Methyl-Violet* (Ehrlich-Weigert).

Saturated alcoholic solution of methyl violet,	11 cc.
Absolute alcohol,	19 cc.
Aniline water,	100 cc.

Aniline water for the above solutions is prepared by shaking in a test tube one part of aniline oil with twenty parts of distilled water, and, after allowing it to stand for a short time, filtering the saturated aqueous solution through a moistened filter. If the solution is not perfectly transparent it should be filtered a second time.

3. *Carbol-Fuchsin* (Ziehl's solution).

Fuchsin,	1 cc.
Alcohol,	10 cc.

Dissolve and add 100 cc. of a five-per-cent solution of carbolic acid.

4. *Alkaline Blue Solution* (Löffler's solution).

Saturated solution of methylene blue,	30 cc.
Solution of caustic potash of 1:10,000,	100 cc.

These solutions keep better than the simple aqueous solutions, but after having been kept for a time they are likely to lose their staining power as a result of the precipitation of the aniline color.

The following special methods of staining cover-glass preparations will be found useful in certain cases:

Gram's Method.—The dried film upon a slide or cover glass is stained with an aqueous solution of methyl violet or with aniline-gentian-violet solution (No. 1); it is then placed in the iodine solution for a minute or two (iodine one part, potassic iodide two parts, water

three hundred parts); then washed in alcohol, dried, and, if for permanent preservation, mounted in balsam.

METHODS OF STAINING THE TUBERCLE BACILLUS.—Numerous methods of staining the tubercle bacillus in sputum dried upon a cover glass have been proposed, but we shall only give here two or three of the most approved methods, either one of which may be relied upon for satisfactory results if carefully followed.

1. *The Ehrlich-Weigert Method.*—Place in a watch glass a little of the aniline-methyl-violet solution (No. 2); float upon the surface of this the cover glass with the dried film downward; heat over a small flame until it begins to steam, then allow it to stand for from two to five minutes; decolorize in a tray containing one part of nitric acid to three parts of water—the cover glass, held in forceps, is gently moved about in the decolorizing solution for a few seconds. It is then washed off in sixty-per-cent alcohol to remove the remaining blue color—this usually takes but a second or two—and then in water. For a contrast stain a saturated aqueous solution of vesuvin may be used, a few drops being left upon the cover glass for five minutes. The stained preparation is then washed, dried, and mounted in balsam.

2. *The Ziehl-Neelson Method.*—Float the cover glass upon the carbol-fuchsin solution (No. 3); heat gently until steam commences to rise—from three to five minutes' time will usually be sufficient; wash off in water, and decolorize in nitric or sulphuric acid, twenty-five-per-cent solution, then in sixty-per-cent alcohol for a very short time to remove remaining color from albuminous background; wash well in water and mount in Canada balsam.

3. *Friedländer's Method.*—Spread and dry the sputum upon the slide; fix by passing the slide three times through the flame of an alcohol lamp or Bunsen burner; place upon the dried film three or four drops of carbol-fuchsin (No. 3); heat gently over a flame until steam is given off; wash in a dish of distilled water; drain off excess of water, and add a few drops of the following decolorizing solution:

Acid, nitric, pure,	5 cc.
Alcohol (eighty per cent),	to 100 cc.

—usually the preparation will be decolorized in about half a minute; wash in water; add a few drops of an aqueous solution of methylene blue as a contrast stain; allow the stain to act for about five minutes, without heating; wash again in water, dry, and mount in balsam, or for a temporary mount use a drop of cedar oil.

4. *Gabbett's Method.*—This is a slight modification only of a very useful method recommended by B. Fränkel in 1884. The contrast stain is added to the decolorizing solution. After staining with

carbol-fuchsin solution (No. 3) the cover glass is placed for one or two minutes in a solution containing:

Sulphuric acid (twenty-five-per-cent solution),	100 cc.
Methylene blue,	2 cc.

Wash, dry, and mount in cedar oil or balsam.

METHODS OF STAINING SPORES.—When preparations containing the spores of bacilli are stained by any of the methods above given, these remain unstained and appear as highly refractive bodies in the interior of the rods or filaments in which they have been formed, or scattered about in the field if they have been set free. Owing to the contrast with the stained protoplasm of the rod or spore-bearing filament, they are especially well seen in recent cultures; while in older cultures the bacilli often do not stain well, or are entirely disintegrated and spores only are to be seen. The discovery was made at about the same time by Buchner (1884) and by Hueppe that spores may be stained if they are first exposed to an elevated temperature for some time. This may be accomplished by placing the slide or cover glass, upon which the spore-containing culture has been dried, in a hot-air oven at a temperature of 120° C. for an hour; or a higher temperature (180° C.) may be employed for a shorter time (fifteen minutes); or the cover glass may be passed through the flame of an alcohol lamp or Bunsen burner eight or ten times, instead of three times as is customary when the object in view is simply to coagulate the albumen and fix the film upon the cover glass. After such treatment the spores may be stained with an aqueous solution of one of the basic aniline colors—fuchsin, methyl violet, etc.—but the bacilli no longer take the stain so well.

To obtain satisfactory *double-stained preparations*, showing both spores and bacilli, a different method is employed.

The film upon the cover glass is passed through the flame three times, as heretofore directed; it is then floated upon aniline-fuchsin solution in a watch glass, and this is heated to near the boiling point for an hour—*Neisser's method*. The aniline-fuchsin solution is prepared by shaking an excess of aniline oil in a test tube with distilled water, filtering the saturated solution into a watch glass, and then adding a few drops of a saturated alcoholic solution of fuchsin. After this prolonged action of the hot staining fluid the spores of some bacilli are deeply stained, while others do not take the stain so well. The cover glass is next washed in water and then placed in a decolorizing solution containing twenty-five parts of hydrochloric acid to seventy-five parts of alcohol. This removes the stain from the bacilli, but, if not allowed to act too long, leaves the spores still stained. The preparation is next stained in a saturated aqueous

solution of methylene blue ; and if the operation has been successfully carried out the spores will be stained red and the protoplasm of the bacilli in which they are present will be blue.

Möller has recently (1891) published the following *new method of staining spores* :

The cover-glass preparation, dried in the air, is passed three times through a flame or placed for two minutes in absolute alcohol ; it is then placed in chloroform for two minutes and washed in water ; it is now immersed in a five-per-cent solution of chromic acid for from half a minute to two minutes and again thoroughly washed in water ; next a solution of carbol-fuchsin is poured upon it and it is heated over a flame until it commences to boil, for sixty seconds ; the carbol-fuchsin solution is then poured off and the cover glass is immersed in a five-per-cent solution of sulphuric acid until the film is decolorized, after which it is again thoroughly washed in water. It is then placed for thirty seconds in an aqueous solution of methylene blue or of malachite green, and again washed in water, after which the preparation should be dried and mounted in balsam. As a result of this procedure the spores are stained dark red and the protoplasm of the bacilli blue or green.

METHODS OF STAINING FLAGELLA.—Koch first succeeded in demonstrating the flagella of certain bacilli and spirilla by staining them with an aqueous solution of hæmatoxylin, and dilute chromic acid as a mordant. Recently Löffler (1889) has succeeded in demonstrating, by an improved staining method, the presence of flagella in a considerable number of species in which they had not previously been seen, although generally suspected to be present. His method is as follows :

Löffler's Method.—The following solution is used as a mordant :

No. 1.

Solution of tannin of twenty per cent.	10 cc.
Saturated (cold) solution of ferrous sulphate,	5 cc.
Aqueous or alcoholic solution of fuchsin,	1 cc.
(Or one cubic centimetre alcoholic solution of methyl violet.)	

No. 2.

A one per-cent solution of caustic soda.

No. 3.

A solution of sulphuric acid of such strength that one cubic centimetre is exactly neutralized by one cubic centimetre of the soda solution.

According to Löffler, solution No. 1 is just right for staining the flagellum of *Spirillum concentricum*, but for certain other bacteria it is necessary to add to this some of No. 2 or of No. 3. Thus, for the cholera spirillum from half a drop to a drop of the acid solution is

added to sixteen cubic centimetres of No. 1. For the bacillus of typhoid fever one cubic centimetre of No. 2 is added to sixteen cubic centimetres of No. 1. *Bacillus subtilis* requires twenty-eight to thirty drops of No. 2; the bacillus of malignant oedema thirty-six to thirty-seven drops, etc.

By carefully conducted experiments Löffler has found that success in staining the flagella depends upon adding just the right quantity of acid or alkali, a very slight variation from the proper quantity being sufficient to give an imperfect or negative result. In general, those bacilli which produce an acid reaction in a neutral culture medium require the addition of the alkaline solution, and those which cause an alkaline reaction require the addition of an acid—acetic or sulphuric.

Löffler gives the following detailed account of his method :

A small quantity of a pure culture is suspended in a few drops of distilled water. Upon perfectly clean glass covers small drops of water are distributed by means of a platinum wire loop; these are sowed with bacilli from the first drop. These little drops are spread out upon the cover with a platinum wire and allowed to dry in the air, after which they are passed through the flame in the usual way to fix the bacteria to the cover glass. Great care must be taken not to heat the preparation too much, as this prevents the flagella from taking the stain. Löffler recommends that the cover glass be held between the thumb and forefinger in passing it through the flame, instead of in forceps, as this insures it from being overheated.

The mordant (solution No. 1) is now placed upon the cover glass so as to completely cover it as an arched drop. The cover glass is then carefully heated over a flame until steam commences to be given off; too much heat causes a precipitate which cannot be washed away. The mordant is left upon the cover glass for from half a minute to a minute, and during this time it is gently moved to and fro. The cover glass is then washed by means of a stream of distilled water. All remnants of the mordant attached to the margins of the cover glass should then be washed away with absolute alcohol. The staining solution is now dropped upon the surface of the glass cover so as to completely cover it, and heat is applied as before for about a minute until steam commences to be given off. The staining solution recommended is a *neutral*, saturated aniline-water-fuchsin solution.

METHODS OF STAINING BACTERIA IN TISSUES.—The solutions recommended for staining cover-glass preparations are also used in staining bacteria in thin sections of the various organs, in which they are found in certain infectious diseases; but, in general, a longer time is required to stain sections, and it is best not to hasten

the process by the use of heat. To obtain good thin sections, the material, cut in small cubes, must be very thoroughly hardened in absolute alcohol. The piece selected for cutting may be attached to a cork by the use of melted glycerin jelly, which is hardened by placing the cork and attached piece of tissue in alcohol. This answers for well-hardened pieces of liver, kidney, etc., but the hollow viscera and tissues of loose structure will require embedding in paraffin or celloidin. Any well-made sledge microtome will answer for cutting the sections, if the knife is properly sharpened. The sections should, of course, be cut under alcohol, and they can scarcely be too thin when the object is to demonstrate the presence or absence of bacteria. Very thin sections may be cut dry by embedding in paraffin having a melting point of 50° C. In this case the knife is set at a right angle to the material to be cut, and the sections are spread out upon and attached to the glass slide for staining.

One of the most useful solutions for staining tissues is Löffler's alkaline solution of methylene blue (No. 4). A freshly-prepared solution will stain sections in four or five minutes. Superfluous color is removed by immersing the sections in diluted alcohol or in a one-half-per-cent solution of acetic acid for a few seconds. The sections are dehydrated in absolute alcohol, cleared up with oil of cedar, and mounted in a drop of cedar oil for examination, or in balsam if they are to be preserved.

Gram's method may be used as directed for cover-glass preparations, the sections being first stained in aniline-gentian-violet solution (No. 1), then washed in water, or in aniline water as recently (1892) recommended by Botkin, then decolorized in the iodine solution (see page 29). The sections when decolorized are again washed in water, dehydrated in absolute alcohol, cleared in cedar oil, and mounted in balsam.

Weigert's Method.—This is a modification of Gram's method in which the sections are dehydrated by the use of aniline oil. The stained section, after having been washed, is transferred to a clean glass slide, the excess of water is removed by the use of filtering paper, and the iodine solution is placed upon it in sufficient quantity to cover the entire section. When sufficiently decolorized this is removed in the same way. The section is then dehydrated by placing a few drops of aniline oil upon it, removing this with filtering paper, and repeating the operation once or twice. The aniline oil must then be completely removed by the use of xylol, after which the section is mounted in balsam.

Kühne's Method.—The object of this method is to prevent the removal of the color from stained bacteria in sections during the treatment which such sections usually receive before they are ready

for mounting—i.e., during the washing and dehydrating processes usually employed. For staining, Kühne prefers a methylene-blue solution prepared as follows: Methylene blue, 1.5 parts; absolute alcohol, ten parts; triturate in a watch glass and add gradually one hundred parts of a solution of carbolic acid containing five parts in one hundred of water. The section is placed in this solution for about half an hour, then washed in water and decolorized in a weak solution of hydrochloric acid—ten drops to five hundred grammes of water. This part of the operation must be conducted very carefully, and usually thin sections will only require to be dipped in the acid solution for an instant, after which they must be at once immersed in a solution of lithium—eight drops of a saturated solution of carbonate of lithium in ten grammes of water. They are then allowed to remain in a bath of distilled water for a few minutes, after which they are dipped into absolute alcohol, which Kühne colors by the addition of methylene blue. The sections are then placed in aniline oil which contains a little methylene blue in solution, where they are dehydrated without the color being extracted from the stained bacteria present. The aniline-oil blue solution is prepared by adding an excess of dry methylene blue to a small quantity of clarified aniline oil. The undissolved pigment settles to the bottom, and a few drops of the colored solution are added to a little aniline oil in a watch glass to make the colored dehydrating bath. The section is next washed out in pure aniline oil—not colored—after which every trace of aniline oil is to be removed by the use of xylol. The section is cleared up in turpentine and mounted in balsam.

Ziehl-Neelson Method, for the tubercle bacillus in tissues.—Leave the sections for fifteen minutes in carbol-fuchsin solution (No. 3); decolorize in sulphuric or nitric acid, twenty-five-per-cent solution; wash in sixty-per-cent alcohol; place in a saturated aqueous solution of methylene blue for contrast stain; wash, dehydrate, and mount in balsam.

The following method of staining sections for the purpose of demonstrating bacteria present in the tissues is recommended by Pregl (1891) as a *substitute for the method of Kühne*. The results are said to be excellent, and it is much simpler and more expeditious.

The sections are made from tissues embedded in paraffin, and are attached to clean glass slides with albumen-glycerin. Or they may be attached to a cover glass by the following method when not embedded in paraffin: The sections, completely dehydrated, are taken out of absolute alcohol on a thin glass cover, upon which they are extended; a piece of filter paper is applied to the side of the cover glass to absorb the alcohol, and before the section is completely dry a drop of acetone-celloidin solution is placed upon it by means of a

glass rod. The cover glass is now moved about in the air to promote rapid evaporation of the alcohol, and is then placed in water. The section now remains attached to the cover glass during subsequent manipulations. The *acetone-celloidin solution* referred to is prepared by adding celloidin in small, dry pieces to acetone until a concentrated solution is obtained. A large drop of this added to five cubic centimetres of absolute alcohol makes a suitable solution for use. This must be kept in a glass-stoppered bottle, and will require to be frequently renewed, as it is not suitable for use after having absorbed moisture from the air. The acetone as obtained from dealers contains considerable water and must be dehydrated by adding to it red-hot sulphate of copper.

The sections, attached to a slide or cover glass by one of the methods mentioned, are stained with Kühne's carbol-methylene-blue solution, which is dropped upon them from a pipette. Usually they will be sufficiently stained at the end of half a minute to a minute, but in some cases a longer time and the application of heat will be desirable. They are then washed in water and immediately placed in fifty-per-cent alcohol, where they remain until the sections have a pale-blue color with a greenish tinge. They are now completely dehydrated in absolute alcohol and subsequently cleared up in xylol.

STAINING SECTIONS OF GELATIN STICK CULTURES.—Fischl, Weigert, and Neisser have given an account of methods for staining stick cultures in gelatin of non-liquefying bacteria. The object of this is to show the mode of growth and the association of individual cells in undisturbed cultures. Neisser gives the following directions: The gelatin cultures are inoculated, by several punctures, with the microörganism to be studied. When the development is deemed sufficient the cylinder of gelatin is removed from the test tube by gently warming its walls. It is then placed for several days—one to eight, according to its size and thickness—in a one-per-cent solution of bichromate of potassium. While in this solution it must be exposed to the light, which causes a change in the gelatin, rendering it insoluble. The gelatin cylinder is thoroughly washed and then hardened in alcohol, first of seventy per cent and then of ninety-six per cent. It is then cut into suitable pieces, and these are attached to a cork in the usual manner and placed for twenty-four hours in absolute alcohol. Thin sections may now be made with a microtome, and these are attached to a glass slide and stained by Gram's or Weigert's method or by the use of Löffler's solution (No. 4). The decolorization should be effected by the use of alcohol and not with an acid solution. When Gram's method is used decolorize by the alternate use of alcohol and oil of cloves. Clear the preparation with oil of bergamot.

V.

CULTURE MEDIA.

To obtain a satisfactory knowledge of the biological characters of the different species of bacteria, it is necessary to isolate them in "pure cultures" and to study their growth in various culture media. By a pure culture we mean a cultivation containing a single species only; and to be absolutely sure that we have a pure culture it is desirable that all of the bacteria in a culture shall be the progeny of a single cell. The methods of obtaining pure cultures will be given later. At present we propose to give an account of the various culture media commonly employed by bacteriologists, and the methods of preparing them for use.

By a natural culture medium we mean one which, as obtained in nature, contains the necessary pabulum for the development of one or more species of bacteria. An artificial culture medium is one which is prepared artificially by adding nutritive material to water. A sterile medium is one which does not contain any living micro-organisms. We may obtain natural media in a sterile condition, but artificial media require sterilization, as they are infallibly contaminated with living "germs" from the atmosphere during the process of preparing them. Sterilization is usually effected by heat. Forceps, glass tubes, etc., may be sterilized by passing them through the flame of an alcohol lamp or Bunsen burner.

NATURAL CULTURE MEDIA.—The most important natural culture medium is *blood serum*, which may be obtained from one of the lower animals—preferably from oxen or calves. This is to be collected in a sterilized jar, with every precaution to insure cleanliness, at the moment of slaughtering the animal. Or the blood of a calf, sheep, or dog may be collected at the laboratory by a carefully conducted operation, in which the femoral or carotid artery is connected with a sterilized glass tube leading into a sterilized receptacle, such as a Woulf's bottle, into one neck of which a cotton plug has been placed to permit the air to escape as the bottle fills with blood through a tube which is secured in the other neck. When blood is passed directly from an artery into a sterilized receptacle the serum will not subsequently require sterilization. The writer is in

the habit of collecting it in this way, and, after the serum has separated, of drawing it off in little flasks having a long neck, as shown in Fig. 14. The neck of the flask, previously sterilized by heat, is slipped into the Woulf's bottle beside the cotton plug, the bulb (α) having been previously gently heated to expand the contained air. As the heated air cools a partial vacuum is formed and the clear serum mounts into the little flask. One after another is filled in this way, and each one is hermetically sealed in the flame of a lamp.



FIG. 14.

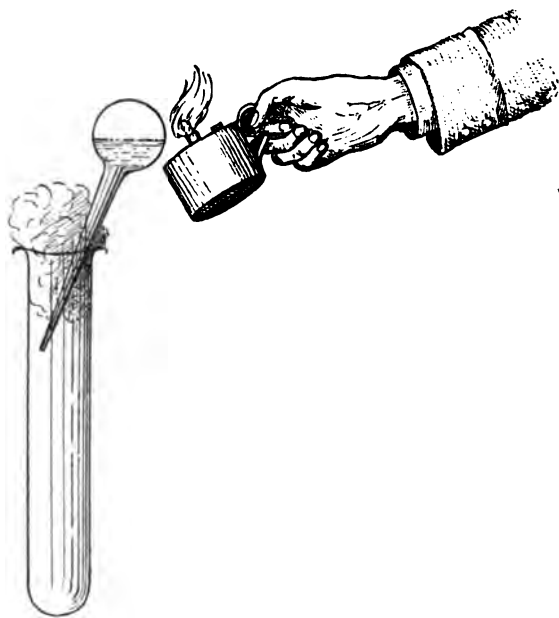


FIG. 15.



FIG. 16.

as soon as it is withdrawn. The sterile blood serum may be preserved indefinitely in this way, and may be used as a liquid culture medium in the little flask, or it may be transferred to a test tube and solidified by heat whenever a solid blood-serum medium is required. The advantage of preserving blood serum and other liquid media in these little flasks is in the fact that they may be preserved indefinitely without becoming contaminated or drying up, and that they are easily transported, while a liquid medium in a test tube must be kept upright. The contents of one of these flasks are readily

transferred to a test tube by breaking off the sealed extremity with sterile forceps and slipping it past the cotton plug, which must be partly withdrawn for the purpose. Upon applying gentle heat to the bulb its contents are forced out into the test tube (Fig. 15). Blood serum which is collected without these special precautions will require sterilization by heat, for which directions will be given later.

To obtain the clear serum from blood collected as above directed, the jars containing it are set aside in a cool place in order that a firm clot may form, care being taken not to shake them. After the clot has formed they may be transported to the laboratory, where they are placed in an ice box or in a cool cellar for from twenty-four to forty-eight hours. By this time the serum has separated from the clot, and it may be transferred to sterilized test tubes by means of a suction pipette (Fig. 16), or may be distributed in little flasks as above directed.

Milk is largely used as a culture medium, and is especially useful in studying the biological characters of various microorganisms, as shown by their causing coagulation of the casein, or otherwise; or an acid or alkaline reaction of the liquid; or peptonization of the precipitated casein, etc. In the udder of healthy cows milk is quite sterile, and by proper precautions it may be drawn into sterilized flasks without any contamination and kept indefinitely without undergoing coagulation or any other change. But in practice it is easier to sterilize it in test tubes or small flasks by the use of heat than to obtain it in a sterile condition from the udder of the cow.

Urine has been used to some extent as a culture medium, and many bacteria multiply in it abundantly, although, on account of its acid reaction, other species fail to grow in it. As contained in the healthy bladder it is sterile, but the mucous membrane of the *meatus urinarius* always contains numerous bacteria upon its surface, and some of these are sure to be carried away with the current when urine is passed.

A culture fluid which the writer has found extremely useful, in tropical countries where it is to be obtained, is the transparent fluid contained in the interior of unripe cocoanuts—called *agua coco* by the Spaniards. In countries where the cocoanut is indigenous this cocoanut water is largely used as a refreshing drink. It contains about four per cent of glucose in solution, together with some vegetable albumen and salts. Some microorganisms multiply in it without appropriating the glucose, while others split this up, producing an abundant evolution of carbon dioxide and giving to the fluid a very acid reaction. The following are the results of an analysis

made for me by Dr. L. L. Van Slyke in the chemical laboratory of Johns Hopkins University : The weight of the fluid obtained from six nuts averaged 339.1 grammes. The specific gravity averaged 1.02285. The amount of water averaged 95 per cent ; the amount of inorganic ash, 0.618 per cent ; the amount of glucose, 3.97 per cent ; the amount of fat, 0.119 per cent ; the amount of albuminoids, 0.133 per cent.

As this fluid is contained in a germ-proof receptacle, no sterilization is required when it is drawn off with proper precautions in the little flasks heretofore described.

Hydrocele fluid has been used as a culture medium, and many bacteria multiply in it abundantly.

Other natural culture media are found in the animal and vegetable kingdoms, which are used, either cooked or raw, as solid substrata upon which bacteria may be cultivated. One of the most useful of these is the *potato*, which is a favorable medium for the development of numerous species, and upon which (cooked) many of them present characters of growth which are so distinctive as to aid greatly in the differentiation of species.

Other *tubers*, *roots*, or *fruits* may also be used as solid media, or their juices extracted and employed as liquid media. Cooked fish and meats of various kinds are also suitable media for certain species—*e.g.*, the phosphorescent bacteria grow very well upon the surface of boiled fish, and in a dark room give off a bright, phosphorescent light.

Eggs, sterilized by boiling, have been used by some bacteriologists, especially for the cultivation of anaërobic species.

ARTIFICIAL CULTURE MEDIA.—A great variety of liquid media have been employed by bacteriologists, the most useful of which are infusions of beef or mutton, with the addition of a little peptone. But Pasteur has shown that some species of bacteria will grow in a medium which does not contain any albuminous material, nitrogen being obtained from salts containing ammonia.

Pasteur's solution, which is rarely used at present, contains : Distilled water, one hundred parts ; cane sugar, ten parts ; tartrate of ammonia, one part, with the addition of the ashes from one gramme of yeast.

Cohn modified this by leaving out the cane sugar, which favors the development of moulds. These fluids are not, however, intended for general use in the cultivation of bacteria, but to demonstrate certain facts relating to their physiology.

Infusions of meat, or “flesh water,” are made by chopping fine lean beef or mutton (one pound) and covering it with water (one litre). This is placed in an ice chest for twenty-four hours, and the

aqueous extract is then obtained by filtration through muslin by pressure. This extract is cooked, filtered, and carefully neutralized by the addition of a solution of carbonate of sodium, which is added drop by drop. Usually we add to this one-half per cent of chloride of sodium. The addition of ten grammes of peptone to a litre of this meat infusion constitutes the *flesh-peptone solution* which is largely used in the preparation of solid culture media, to be described hereafter.

The addition of five per cent of glycerin to the above infusion makes a useful liquid medium for the cultivation of the tubercle bacillus (Roux and Nocard). The liquid should be again neutralized after adding the glycerin, which commonly has an acid reaction.

Bouillon is made by cooking the chopped meat—one pound in a litre of water—for about half an hour in a large glass flask or an enamelled iron kettle. The filtered bouillon is then carefully neutralized with sodium carbonate, and again boiled for an hour to precipitate all coagulable albuminoids. It is again filtered and distributed in test tubes or small flasks, in which it is subsequently sterilized. For certain pathogenic bacteria a bouillon made from the flesh of a fowl or of a rabbit is preferable to beef bouillon.

Flesh infusion may also be made from one of the standard beef extracts, such as Liebig's (five grammes to a litre of water).

Various *vegetable infusions* may also be used as culture media, such as yeast water, potato water, infusion of hay, of barley, or of wheat, of dried fruits, beer wort, etc.

SOLID CULTURE MEDIA.—The introduction of solid culture media, and especially the use of gelatin and agar-agar, as first recommended by Koch (1881), for the isolation and differentiation of species, was a most important advance in bacteriological technology. We are concerned here only with the composition and preparation of these media.

Flesh-Peptone-Gelatin.—This is made by adding ten per cent of the best French gelatin to the flesh-peptone solution above described. This is the standard gelatin medium, but more or less gelatin may be added to serve a special purpose. Thus, in Havana during the summer months the writer used a medium containing twenty per cent of gelatin, because when but ten per cent was used the gelatin was liquefied by the normal temperature of the atmosphere. Ten-per-cent gelatin, of good quality and carefully prepared, will stand a temperature of 20° to 22° C. (68° to 71.6° F.) without melting. When twenty per cent of gelatin is used the melting point is about 8° C. higher. It must be remembered that exposure to a boiling temperature reduces the melting point of gelatin. It is therefore desirable to accomplish the operations of cook-

ing and sterilizing in as short a time as is practicable. The French gelatin used comes in thin sheets ; this is broken up and added to the flesh-peptone solution.

Usually we prepare a litre of nutrient gelatin at one time, and for this quantity one hundred grammes of gelatin will be required for the standard preparation (ten per cent). It is well to allow it to soak for a time in the liquid before applying heat for the purpose of dissolving it. Then apply gentle heat until it is completely dissolved. The gelatin of commerce usually has an acid reaction, and it will be necessary to carefully neutralize the medium after it has been added. A slightly alkaline reaction is usually no disadvantage, but certain pathogenic bacteria will not grow when there is a trace of acid present. The

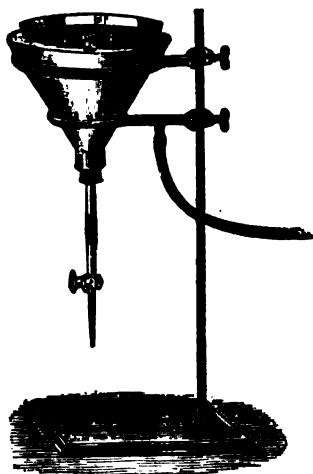


FIG. 17.

next step consists in *clarifying* the nutrient medium. It is allowed to cool to about 50° C., and an egg, previously broken into one hundred grammes of water, is gradually added while stirring the liquid with a glass rod. A whole egg is used for a litre of the solution. Heat is again applied and the solution is kept at the boiling point for about ten minutes, during which time the egg albumen is precipitated and carries down with it all insoluble particles, which without this clarifying process would have interfered with the transparency of the medium, even when carefully filtered. The hot solution is then *filtered*. A hot-water funnel (Fig. 17) is usually employed, as the gelatin solution does not pass through filtering paper very rapidly, and when cooled to near the point of solidifying ceases to pass.

The advantages of the gelatin medium are that it is perfectly transparent, that it is easily melted for making "plates," and that many bacteria exhibit in it special characters of growth by which they may be differentiated from others which resemble them in form. The principal disadvantage is the low melting point, which prevents us from making use of this medium for cultivating bacteria in an incubating oven at a higher temperature than about 22° C. for ten-per-cent gelatin.

This disadvantage is overcome by using *agar-agar* instead of gelatin. This is prepared in Japan and other Eastern countries from certain species of gelatinous algæ. It comes to us in the form of bundles of dried strips, which form a stiff jelly when dissolved in water in the proportion of one to two per cent. This jelly remains solid at a temperature of 40° C. and above. It was first employed by Hesse, one of Koch's collaborators in the office of the imperial board of health of Berlin. Koch, who was in search of a transparent jelly which would stand the temperature required for the cultivation of certain pathogenic bacteria (37° to 38° C.), quickly recognized its value and introduced it into general use.

The *agar-agar jelly* is more difficult to filter than the gelatin medium, and some skill is required in order to obtain a transparent solution. It will bear long boiling without losing its quality of forming a stiff jelly. From ten to twenty grammes are added to a litre of flesh infusion, or we may make a *peptonized agar* in accordance with the following formula which is given by Salomonson: Add to one litre of distilled water five grammes Liebig's extract, thirty grammes peptone, five grammes cane sugar, fifteen grammes agar. Cook for an hour, render slightly alkaline, and cool to below 60° C. Clarify and cook again for an hour or more.

Glycerin-agar is made by adding five per cent of glycerin to the peptonized agar made by the above formula or by the use of the flesh-peptone infusion. This is a very favorable medium for the cultivation of the tubercle bacillus—first used by Roux and Nocard.

Agar-gelatin, a medium which has recently come into favor and is said to be very useful, as it resembles gelatin in transparency and has a considerably higher melting point than ten-per-cent gelatin, is made by adding fifty grammes of gelatin and 7.5 grammes of agar to a litre of flesh-peptone solution. Care should be taken not to cook this longer than is necessary.

In making all of these agar culture media the main difficulties encountered result from the difficulty of dissolving the agar and the slowness with which the solution passes through filtering paper. These difficulties are best met as follows: Break up the sticks of agar into small fragments and allow them to soak in cold water for twenty-

four hours. Pour off the water and add the flesh-peptone solution. Boil for several hours until the agar is completely dissolved. Neutralize by adding gradually a solution of carbonate of soda (or render slightly alkaline). Filter.

The last operation is the most troublesome, and various plans have been proposed to avoid the tedious filtration through filtering paper in a hot-water filter. A method which gives satisfactory results is to place the filter containing the hot agar solution, and the flask which is to receive the filtrate, in a steam sterilizing apparatus, where it is left in an atmosphere of streaming steam until the filtra-

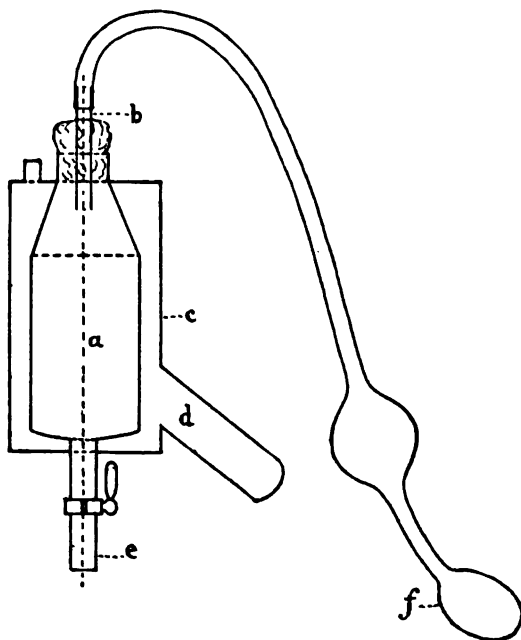


FIG. 18.

tion is completed. Or the solution may be put in a tall jar and left in the steam sterilizer for several hours until it is clear as a result of sedimentation. The clear solution is then obtained by decantation. Or by conducting the operation in a tall cylindrical vessel, and allowing sedimentation to occur in the steam sterilizer and the agar subsequently to solidify by cooling, the cylinder of jelly may be removed from the jar and the part containing the sediment can be cut away. The transparent portion is then melted again and distributed in test tubes for use.

In the present volume we frequently refer to the nutrient medium made by adding one to two per cent of agar-agar to the standard flesh-peptone solution as "nutrient agar" or simply as "agar."

The following method of filtering agar has recently (1890) been proposed by Karlinsky. It is a modification of the method previously described by Jakobi and depends upon the use of pressure.

In Fig. 18, *a* is a cylindrical vessel of tin, which is closed above by a perforated rubber cork, through which is passed a glass tube, *b*. This is enclosed in a larger tin cylinder, *c*, which contains water, which may be kept hot by placing an alcohol lamp under the projecting arm *d*. The central cylinder has a tube, *e*, passing through the bottom of the hot-water cylinder, and which is provided with a



FIG. 19.

stopcock for drawing off the filtered solution. Before pouring the hot agar solution into the cylinder *a*, a cotton filter about ten centimetres thick is placed at the bottom of this cylinder and hot water is poured upon it while the stopcock of the outlet tube is open. This washes out the cotton and prepares the filter for the agar solution. The apparatus is supported upon a tripod, not shown in the figure. Filtration is said to occur rapidly when the air in the central cylinder is compressed by means of the hand bellows attached to the tube *b*.

Unna (1891) has devised a filtering apparatus for agar which is shown in Fig. 19. In this the pressure of steam is utilized. A hollow

sphere of copper, supported upon a tripod, is so constructed that an upper hemispherical segment can be removed to give access to the interior. An opening at the bottom contains a perforated rubber cork, through which the stem of an enamelled iron funnel passes. A simple filter of filtering paper is used in this funnel, and this is filled to a depth of two centimetres with well-burned *kieselgur* (diatomaceous earth in which the organic matter has been destroyed by heat). The hot solution of agar is poured into the funnel, and hot water into the space between it and the copper vessel; this must not come too near the top of the funnel—not nearer than three centimetres. The hemispherical cover is then secured in its place by means of a clamp screw shown in the figure. By placing a Bunsen burner under the projecting arm the water is made to boil and a sufficient steam pressure secured. A small stopcock attached to the cover of the copper vessel permits the escape of steam if the pressure is too great. According to Unna, solutions containing as much as three per cent of agar can be filtered by means of this apparatus, and a litre of two-per-cent agar will pass through it in about two hours.

For special purposes various substances are added to the above-described solid and liquid media. A favorable addition for the growth of a considerable number of bacteria is from one to three per cent of *glucose*. The phosphorescent bacteria grow best in a medium containing two to three per cent of *sodium chloride*. The addition of three to four per cent of *potassium nitrate* is made in conducting experiments designed to test the reducing power of certain bacteria, by which this salt is decomposed with the production of nitrites. *Acids* are also added in various proportion to test the ability of bacteria under investigation to grow in an acid medium. From 1:2,000 to 1:500 of hydrochloric acid may be used for this purpose. The addition of *litmus* to milk or other culture media is frequently resorted to for the purpose of ascertaining whether acids or alkalies are developed during the growth of bacteria under investigation. The addition of *aniline colors* which are variously changed by the products of growth of certain species has also been resorted to in the differentiation of species. Various disinfecting agents, such as *carbolic acid*, etc., have also been used for the same purpose, and it has been shown by experiment that some bacteria will grow in a medium containing such agents in a proportion which would entirely restrain the development of others.

Quite recently the *soluble silicates* which form a jelly-like mass have been proposed as a culture medium for certain bacteria which do not grow in the usual media. Kühne (1890), Winogradsky (1891), and Sleskin (1891) have made experiments which indicate that this medium has considerable value.

Winogradsky uses in the preparation of his silicate jelly the following salts :

Ammonium sulphate,	0.4 gramme.
Magnesium sulphate,	0.05 "
Potassium phosphate,	0.1 "
Calcium chloride,	a trace.
Sodium carbonate,	0.6 to 0.9 gramme.
Distilled water,	100 grammes.

To this he adds a solution of silicic acid. According to Kühne, a solution containing 3.4 per cent of silicic acid and having a specific gravity of 1.02 may be preserved in a liquid condition. To this the salts are added in greater or less amount, according to the consistence desired.

Sleskin states that a suitable jelly is formed by the addition of 1.15 to 1.45 per cent of the salts, and recommends that concentrated, sterilized solutions be added to the acid. He dissolves separately, in as little water as possible, the sulphates, the potassium phosphate and sodium carbonate, and the calcium chloride.

The use of a culture medium containing an extract from the jequirity seeds has recently been recommended by Kaufmann (1891), who has found, by experimenting upon various bacteria, that such a medium is useful in differentiating species.

The *jequirity solution*, which may be used as a liquid medium or may be employed in the preparation of nutrient gelatin or agar, is prepared as follows : Ten grammes of jequirity seeds are bruised in a mortar and the shells removed ; they are then placed in one hundred cubic centimetres of water and cooked for two hours in the steam sterilizer ; after allowing the infusion to cool it is filtered. The filtered liquid has a pale-yellow color and a neutral or slightly alkaline reaction. Certain bacteria grow in this solution without producing any change in its color ; others, which produce an acid reaction, cause it to be decolorized ; others, which produce an alkaline reaction of the medium, change the color to green.

Cooked Potato.—Schröter first used cooked potato as a culture medium for certain chromogenic bacteria (1872), and Koch subsequently called attention to the great value of potato cultures for differentiating species. His plan of preparing potatoes is as follows: Sound potatoes are chosen in which the epidermis is intact. These are thoroughly washed and scrubbed with a brush to remove all dirt. The "eyes" and any bruised or discolored spots are removed with a sharp-pointed knife. They are again thoroughly washed in water, and are then placed for an hour in a bath containing mercuric chloride in the proportion of 1:500, to thoroughly disinfect the surface. They are then placed in a steam sterilizer for about three-quarters of an hour, and after an interval of twenty-four hours

are again steamed for fifteen minutes. It is well to wrap each potato in tissue paper before placing it in the bichloride bath, and to leave it in this protecting envelope until it is placed in the glass dish in which it is preserved from contamination by atmospheric germs after being inoculated with some particular microörganism. Just before such inoculation the potato is cut in halves with a sterilized (by heat) table knife. The bacteria to be cultivated are placed upon the cut surface and the potato is preserved in a glass dish (Fig. 20).

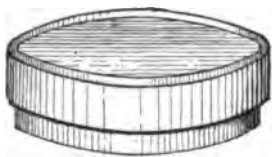


FIG. 20.

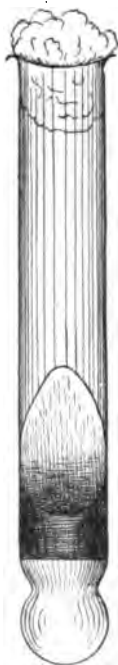


FIG. 21.



FIG. 22.

A more convenient method, and one which secures the potato more effectually from atmospheric organisms, is to cut a cylinder, about an inch in diameter, from a sound potato, by means of a tin instrument resembling a cork borer or apple corer. This cylinder is cut obliquely into two pieces having the form shown in Fig. 22, and each piece is placed in a large test tube having a cotton air filter, in which it is sterilized. This method, first employed by Bolton, has been slightly modified by Roux, who recommends that a receptacle for catching the water which separates during the sterilizing process be formed by making a constriction around the test tube an inch above its lower extremity. This is done by the use of a blowpipe.

The cylinder of potato rests upon the constricted portion of the tube, as shown in Fig. 21.

Sometimes a *potato paste* is employed. The potatoes are boiled for an hour and the skins removed, after which they are mashed with a little sterilized water, placed in suitable plates, and sterilized by exposure for half an hour on three successive days in the steam sterilizer. *Bread paste* may be made in the same way, and is a very favorable medium for the growth of certain bacteria and also for the common moulds.

VI.

STERILIZATION OF CULTURE MEDIA.

A MOST important part of bacteriological technology consists in the sterilization of the various culture media employed. A sterile medium is essential for maintaining a pure culture, and we can only obtain an exact knowledge of the biological characters of a species by studying its growth in various media, its physiological reactions, its pathogenic power, etc., independently of all other microorganisms—*i.e.*, in pure cultures.

We may sterilize a culture medium either by heat or by filtration through a substance which does not permit bacteria to pass. The last-mentioned method is useful for certain special purposes; but, in general, sterilization of culture media, and of the vessels in which they are preserved, is effected by heat.

The scientific use of heat as an agent for sterilizing our culture media depends upon a knowledge of the thermal death-point of the various microorganisms which are liable to be present in them, and upon various facts relating to the manner in which heat is applied. All this has been determined by experiment, and before giving practical directions for sterilization it will be well to consider the experimental data upon which our methods are based.

As a rule, bacteria which do not form spores are killed at a comparatively low temperature. Thus, in a series of experiments made by the writer upon the thermal death-point of various pathogenic organisms, the pus cocci were found to be the most resistant, and all of these were killed by exposure for ten minutes to a temperature of 62° C. (143.6° F.). There are several species of bacteria known, however, which not only are not killed by this temperature, but are able to grow and multiply at a temperature of 65° to 70° C. (Miquel, Van Tieghem, Globig). But it is safe to say that exposure to a boiling temperature for a minute or two will infallibly destroy all microorganisms in the absence of spores, *when they are in a moist condition or moist heat is used—i.e.*, when they are directly exposed to the action of boiling water or of steam. The power of dry heat to destroy microorganisms in a desiccated condition is a different matter and will require special consideration.

The spores of bacilli have a much greater resisting power, and the vitality of some of these reproductive bodies, from known species, is not destroyed by a boiling temperature maintained for several hours. Thus Globig found that the spores of a certain bacillus from the soil—his “red potato bacillus”—required six hours' exposure to streaming steam in order to destroy it. Steam under pressure, at a temperature of 115° C., killed it in half an hour; at 125° C. in five minutes. This extreme resisting power is exceptional, however, and many spores are destroyed in a few minutes by the boiling temperature of water.

In practice we assume that some of the more resistant spores, which are frequently present in the atmosphere, may have fallen into our culture material, and to insure its sterilization we subject it to a temperature which can be depended upon to destroy these; or we resort to the method of *discontinuous heating*. This method was first employed by Tyndall (1877), and is now in general use in the bacteriological laboratories of Germany, having been adopted by Koch and his pupils; while in France a single sterilization by means of steam under pressure, securing a higher temperature, is still the favorite method with many.

In the method by discontinuous heating we subject the culture material for a short time to the temperature of boiling water, thus destroying all bacteria in the vegetative stage. After an interval, usually of twenty-four hours, we repeat the operation for the purpose of destroying those which in the meantime have developed from spores which may have been present. Again the material is put aside, and after twenty-four hours it is again heated to the boiling point. This is usually repeated from three to five times. The object in view is to kill the growing bacteria which are developed from spores which were present; and, as a matter of experience, we find that this method of sterilization is more reliable than a single prolonged boiling, unless this be effected at a higher temperature than that of boiling water at the ordinary pressure of the atmosphere. Discontinuous heating is especially useful for the sterilization of liquids which would be injured by prolonged boiling—as is the case with solutions of gelatin—or which are coagulated by the boiling temperature. By means of a water bath, the temperature of which is regulated automatically, we may conduct the operation at any desired degree. Thus in sterilizing blood serum we use a temperature a little below that at which coagulation occurs (about 70° C.).

Test tubes, flasks, and apparatus of various kinds are commonly sterilized by dry heat in a *hot-air oven*. This is usually made of sheet iron, with double walls, and shelves for supporting the articles

to be sterilized. The form shown in Fig. 23 is commonly used in bacteriological laboratories.

It must be remembered that a much higher temperature is required for the destruction of microorganisms when dry heat is employed than is the case with moist heat. The experiments of Koch and Wolffhügel (1881) show that a temperature of 120° to 128° C. (248° to 262° F.) is required to destroy the spores of mould fungi, and micrococci or bacilli in the absence of spores. For the spores of bacilli a temperature of 140° C. (284° F.), maintained for three hours, was required.

In practice we usually maintain a temperature of about 150° C.

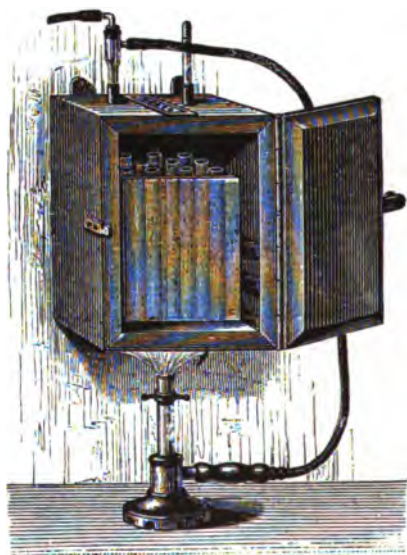


FIG. 23.

(302° F.) for an hour or more; and it is customary to sterilize all test tubes and flasks, which are to be used as receptacles for culture media, in the hot-air sterilizer. This procedure could no doubt, however, be dispensed with in many cases and reliance be placed upon the sterilization of the flask, together with its contents, in the steam sterilizer, especially with such culture media as are not injured by long exposure to a boiling temperature—*e.g.*, bouillon and agar-agar.

When we propose to cultivate aërobic bacteria, or such as require oxygen for their development, a cotton air filter is placed in the mouth of each test tube and flask before it is sterilized in the hot-air oven. This is a loose plug of cotton, pushed into the neck of the flask for an inch or more, and projecting from its mouth for a short distance. These cotton filters should fill the tube completely and

uniformly, but should not be packed so closely that there is difficulty in removing them.

Steam Sterilizers.—Steam at the ordinary pressure of the atmosphere has the same temperature as boiling water, and in practice is preferable to a water bath for several reasons. The form of steam sterilizer adopted by Koch, after extensive experiments made in collaboration with Löffler and Gaffky, is now generally used in bacteriological laboratories. This is shown in Fig. 24. It consists of a cylindrical vessel of zinc which is covered with a jacket of felt. The cover, also covered with non-conducting material, has an aperture at the top for the escape of steam. A glass tube, which is in communication with the interior of the vessel, serves to show the



FIG. 24.

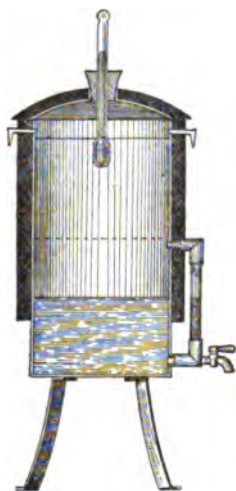


FIG. 25

height of the water when the apparatus is in use. The bottom of the cylindrical vessel should be of copper. A Bunsen burner having three jets will commonly be required to keep the water in ebullition and the upper part of the steam sterilizer filled with "live steam," which should escape freely from the aperture in the cover to insure a temperature of 100° C. in the steam chamber. A perforated zinc or copper shelf in the interior of the cylinder serves to support the flasks, etc., which are to be sterilized. Usually they are lowered into the cylinder in a light wire basket, or tin pail with perforated bottom, of proper diameter to slip easily into the sterilizer.

Fig. 25 is a sectional view of this sterilizer.

The steam sterilizer shown in Fig. 26¹ is an American invention,

¹ The Arnold steam sterilizer, manufactured at Rochester, N. Y.

which answers the purpose admirably, and which has the advantage of getting up steam very quickly and also of using comparatively little gas.

The use of steam under pressure, by which higher temperatures are obtained, requires a more expensive apparatus, made on the principle of Papin's digester. The form manufactured by Müncke is one of the best. This is shown in Fig. 27. It is provided with a pressure gauge and a safety valve. A single sterilization in this apparatus, at a temperature of 115° C., for half an hour, will usually

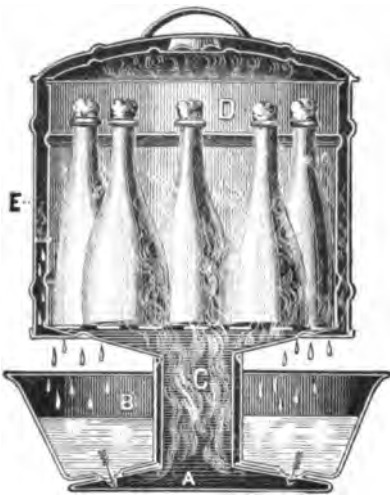


FIG. 26.



FIG. 27.

suffice, and for liquid culture media or for agar-agar this method is entirely satisfactory; but a gelatin medium which is exposed to this temperature loses its property of forming a jelly at 20° to 22° C., and consequently its value as a solid culture medium. In practice the simpler form of apparatus in which streaming steam is used will be found to answer every requirement. To insure sterilization with this it is customary to resort to discontinuous heating, as heretofore described. The standard flesh-peptone-gelatin medium should, as a rule, be subjected to a temperature of 100° C. for ten minutes, at intervals of twenty-four hours, four days in succession. Bouillon, flesh infusions, and agar-agar jelly may be steamed for an hour at a time two or three days in succession.

It is always advisable to test the sterilization of culture material before making use of it. This is done by placing it for a few days in an incubating oven at 30° to 35° C. If a considerable quantity of material in test tubes has been prepared at one time, it will be sufficient to put a few tubes in the incubating oven to test sterilization.

Failure to make this test often leads to serious complications in experimental investigations. A laboratory sometimes becomes infected with resistant spores, which are not all destroyed by the usual methods of sterilization, and these may not develop until some time has elapsed after the supposed sterilization.

Sterilization of Blood Serum.—Blood serum which has been collected in test tubes or small flasks, as heretofore directed, is

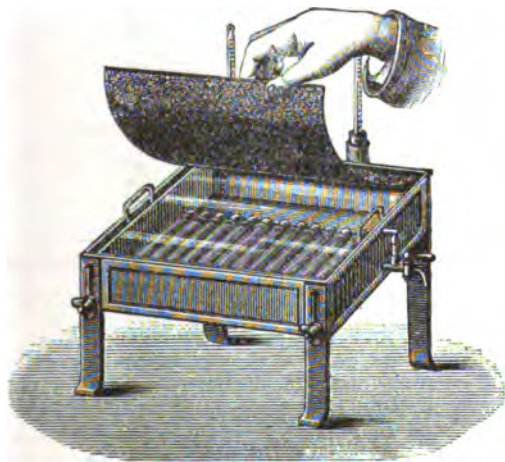


FIG. 28.

sterilized in a water bath at 60° C. (140° F.) by the method of discontinuous heating. It is usually left in the hot-water bath for about an hour, and this is repeated, at intervals of twenty-four hours, for five to seven days. This rather tedious process may be avoided by collecting the serum in the first instance with proper precautions to prevent it from becoming contaminated with atmospheric organisms. A special apparatus was devised by Koch for sterilizing blood serum, but an improvised hot-water bath which is regulated to a temperature of 60° C. by an automatic thermo-regulator will answer the purpose. After being sterilized the serum is solidified by careful exposure to a temperature of about 68° C., which causes it to coagulate, forming a transparent, jelly-like mass. When coagulated at a higher temperature it becomes opaque. The time required for this operation varies from half an hour to an hour, and it is best to remove the tubes from the receptacle in which they are exposed to

heat as soon as the serum is solidified. Koch's apparatus for coagulating blood serum is shown in Fig. 28. It is customary to place the test tubes in an oblique position, so that a large surface may be exposed upon which to cultivate the tubercle bacillus or whatever microörganism may be under investigation. A form of apparatus designed for both sterilizing and coagulating blood serum is shown in Fig. 29. It is manufactured by Müncke in accordance with the directions of Hueppe, and special precautions have been taken to secure a uniform temperature in all parts of the air chamber. We

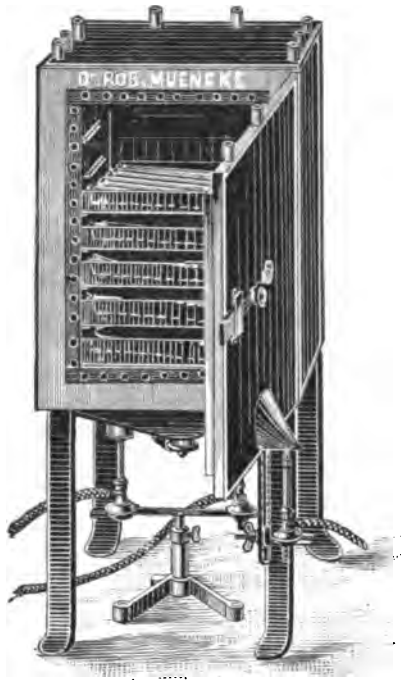


FIG. 29.

may remark that since it has been shown by Roux and Nocard that the tubercle bacillus grows very well in agar-agar jelly to which five per cent of glycerin has been added, blood serum is not so largely used as a culture medium in bacteriological laboratories.

Sterilization by Filtration.—This method is especially useful for separating the soluble substances contained in a liquid culture of bacteria from the living cells. It has been demonstrated that several of the most important pathogenic bacteria produce toxic substances during their growth which may cause the death of susceptible animals independently of the living bacteria; and this demonstration

has been made either by sterilizing a pure culture by means of heat, or by separating the bacteria from the culture liquid by filtration. Some of these toxic products of bacterial growth are destroyed by a comparatively low temperature; the method of sterilization by filtration is therefore very important in researches relating to the composition and pathogenic power of these soluble products. Pasteur, in his earlier experiments, used plaster of Paris as a filter, and

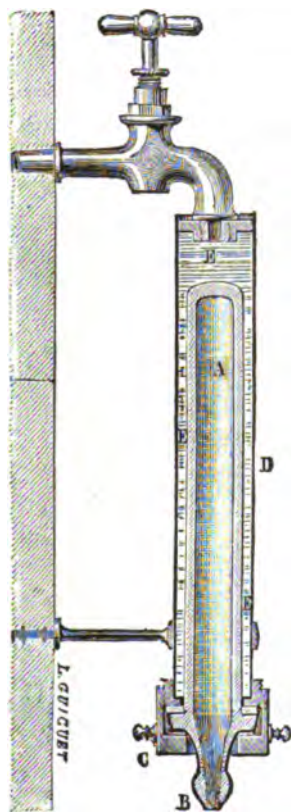


Fig. 30.

subsequently resorted to the use of unglazed porcelain, through which a liquid may be forced by pressure, but which does not permit of the passage of suspended particles, however small.

As the porcelain filter is the most reliable and convenient for accomplishing the object in view, we shall not describe other methods of filtration which have been proposed and successfully used. The porcelain used is a very fine paste, manufactured at Sèvres, which is moulded into cylinders (*bougies*) of the form proposed by Chamberlain and baked at a high temperature.

In Fig. 30 the Pasteur-Chamberlain filter is shown as arranged for the filtration of water. A is the hollow porcelain cylinder, which is enclosed in a metal case, D. The metal case is tightly clamped against a projecting shoulder at the lower part of the porcelain filter, a ring of rubber being interposed to secure a tight joint. When water under pressure is admitted to the space E, between the cylinder of porcelain and the metal case, it slowly filters through, and, running down the inner wall of the filter, escapes at B into a receptacle placed to receive it. If we fill the space E with a liquid culture of bacteria and apply sufficient pressure (one or two atmospheres), a clear filtrate is obtained which is entirely sterile if the porcelain filter is sound and made of proper material. After the



FIG. 31.

filter has been in use for some time, however, it may permit the passage of bacteria, and it will be necessary to subject it to a high temperature for the purpose of destroying all organic matter contained in the porous porcelain.

We may use the Chamberlain filter without a metal case by immersing it in a cylindrical glass vessel containing the liquid to be filtered, as shown in Fig. 31. The porcelain cylinder is connected with an aspirator bottle, *a*, and a small Erlenmeyer flask, *b*, is interposed to catch the filtrate when it overflows from the interior of the filter. Of course all the necessary precautions must be taken with reference to the sterilization of the interior of the *bougie*, of the flask *b*, and of the rubber tube connecting the two.

Another arrangement of the Pasteur-Chamberlain filter for laboratory purposes is shown in Fig. 32. In this form of apparatus a

receptacle, R, is provided for the liquid to be filtered, and a pump for compressing air is attached to it by a rubber tube. Instead of this pump, water pressure may be used indirectly by attaching a strong bottle to the water supply and allowing it to fill slowly with water, and at the same time to force out the air through a tube connected with the filtering apparatus. For this purpose the bottle, having a capacity of a quart or more, should be provided with a rubber stopper through which two short tubes are passed. One of these is connected with the water supply and the other with the filter. Of course this is only practicable when a water supply with sufficient pressure is available.



FIG. 32.

As a rule, filtration cannot be substituted with advantage for sterilization by heat in the preparation of culture media. Albuminous liquids pass through the filter with difficulty, and the process of sterilization by discontinued heating will usually prove more satisfactory than filtration, which requires extreme precautions to prevent accidental contamination of the filtered liquid. Moreover, the filter may change the composition of the medium passed through it by preventing the passage of colloid and albuminous material in solution. Thus, in an attempt to separate blood corpuscles from the serum by filtration through a Chamberlain filter, the writer obtained a transparent liquid which did not coagulate by heat—*i.e.*, the albuminous constituents of the serum did not pass through the filter.

VII.

CULTURES IN LIQUID MEDIA.

PRIOR to the introduction of gelatinous media by Koch in 1881, cultures were made in various organic liquids, and these are still largely used, being for certain purposes preferable to solid media. The method of preparing and sterilizing the flesh infusions and other organic liquids commonly used has already been given. We are here concerned with the various modes of using these nutritive liquids in cultivating bacteria.

Flasks and tubes of various forms have been employed by different investigators, but the most useful receptacle for liquid as well as for solid culture media is *the ordinary test tube*. These are carefully cleaned, plugged with a cotton air filter, sterilized in the hot-air oven at 150° C., and are then ready to receive the filtered liquid. Usually the tube should not be filled to more than one-third to one-half of its capacity. Sterilization of the culture liquid is then effected by placing the tubes in the steam sterilizer for half an hour on three successive days. Before using, the tubes should be placed for a few days in an incubating oven at 30° to 35° C. to test the sterilization. This is especially important with liquid media, for if a single living spore is present it may give rise to an abundant progeny, which will be distributed through the liquid in association with the species which has been planted. In solid cultures, on the contrary, such a spore would give rise to a colony, which by its locality and characters of growth would probably be recognized as different from the species planted, and consequently accidental. This is the great danger in the use of liquid media; imperfect sterilization, or accidental contamination by atmospheric germs, may lead the inexperienced student into serious errors resulting from the assumption that the micro-organisms present in his cultures are all derived from the seed he planted.

On the other hand, liquid media are more convenient than solid when it is the intention to isolate by filtration the soluble products of bacterial growth; for injection into animals to test pathogenic power; for experiments on the germicidal or antiseptic power of chemical agents, etc.

For larger quantities of liquid than can be held in an ordinary test tube the small flasks with a flat bottom, known as Erlenmeyer flasks, are very convenient (Fig. 33).

In his earlier researches Pasteur used flasks and tubes of various forms, which served a useful purpose, but have been displaced in his laboratory by the simpler form of apparatus shown in Fig. 34. This is a little flask having a cover which is ground to fit the neck. This cover is drawn out above into a narrow tube which admits oxygen to the flask through a cotton air filter. To obtain access to the interior of the flask for the purpose of introducing bacteria to start a culture, or to obtain material for microscopical examination, the cover is detached at the ground joint by a gentle twisting motion.

There is much less danger that a sterile culture liquid will become



FIG. 33.



FIG. 34.

contaminated during the momentary removal of the cover from one of these little flasks, or of the cotton plug from a test tube, than is usually supposed. Abundant laboratory experience demonstrates that such contamination by bacteria floating in the atmosphere rarely occurs. The spores of mould fungi are commonly more abundant in the air, but even these do not very frequently fall into the culture liquid when the tube is opened to inoculate it with the bacteria it is proposed to cultivate. This inoculation is best made with a platinum wire, bent into a loop at the free extremity, and sealed fast into the end of a glass rod (Fig. 35). This is sterilized in the flame of a Bunsen burner or alcohol lamp by bringing the platinum wire to a red heat and passing the end of the glass rod which carries it through the flame several times. With this instrument we may transfer a little drop from a culture to the sterile fluid in another

tube for the purpose of starting a new culture. Or we may start a pure culture from a drop of blood taken from the veins of an animal which has been inoculated with anthrax, or any similar infectious disease in which the blood is invaded by a bacterial parasite.

But if we have not a pure culture to start with our liquid media do not afford us the means of obtaining one; and if two or more bacteria which resemble each other in their morphology are associated in such a culture we cannot differentiate them, and are likely to infer that we have a pure culture of a single microorganism when this, is not really the case.

But if we have pure stock to start with we may maintain pure cultures in liquid media without any special difficulty.

Various *characters of growth*, etc., are to be observed in cultivating different microorganisms in liquid media. Thus some grow at the surface in the form of a thin film or membranous layer—"mycoderma"—while others are distributed uniformly through the liquid, rendering it opalescent or more or less milky and opaque; others, again, form little flocculi which are suspended in the transparent



FIG. 35.

fluid. Usually, when active growth has ceased, the bacteria fall to the bottom of the tube as a more or less abundant, white or colored, pulverulent or glutinous deposit. In some cases the liquid is colored with a soluble pigment formed during the growth of the bacteria, and usually this is formed most abundantly at the surface, where there is free access of oxygen. The reaction of the medium is often changed as a result of the growth of bacteria in it. From being neutral it may become decidedly alkaline or acid in its reaction. These changes may be observed by adding a litmus solution before sterilization of the culture medium, and observing the change of color when an acid-producing bacterium is under cultivation. The reducing power of bacteria upon various aniline colors may also be studied; also their power to break up various organic substances, as shown by the evolution of gas or other volatile products which may be collected, or by substances which remain in solution and can be studied by ordinary chemical methods.

Drop Cultures.—When we desire to study the life history of a microorganism and to witness its development from spores, for example, its motions, etc., the method of cultivation in a hanging drop

of culture fluid, attached to a thin glass cover and suspended over a circular excavation ground out of a glass slide, is very useful. Such a drop culture may be left under the microscope and kept under observation for hours or days.

In making these drop cultures it is necessary to sterilize the glass slides and thin glass covers by heat, and to take every precaution to prevent the inoculation of the drop of culture liquid with any other bacteria than those which are to be studied.

The simplest form of moist chamber for drop cultures consists of an ordinary glass slide having a concave depression, about fifteen millimetres in diameter, ground out in its centre. This and the thin glass cover, having been sterilized by exposure in the hot-air oven at 150° C. for an hour or more, or by passing them through the flame of an alcohol lamp, are ready for use. The cover glass is held in sterile forceps, and a little drop of the culture fluid containing the bacterium to be studied is transferred to its centre by means of the platinum loop heretofore described. It is best to spread the drop out as thin as possible, and it may be inoculated, from a pure cul-



FIG. 26.

ture, with a platinum needle (Fig. 36) after it has been placed upon the cover. This is then inverted over the hollow place in the glass slide, and it is customary to prevent the entrance of air and attach the cover by spreading a little vaseline around the margin of the excavation.

Another form of moist chamber is made by attaching a glass ring, having parallel, ground surfaces, to the centre of a glass slide by a suitable cement.

In Ranvier's moist chamber there is a central eminence surrounded by a groove ground into the glass slide, and the drop of culture fluid is in contact with a polished glass surface below as well as above. This affords a more satisfactory view under the microscope.

The Author's Culture Method.—In a paper read at the meeting of the American Association for the Advancement of Science, in August, 1881, the writer described a method of conducting culture experiments which he has since used extensively and with very satisfactory results. The liquid culture medium is preserved in little flasks having a long neck which is hermetically sealed. The principal advantages connected with the use of these little flasks, or "Stern-

berg's bulbs," as they are sometimes called, are that a culture medium may be preserved in them indefinitely and that they are easily transported from place to place; whereas test tubes, Pasteur's flasks, and similar receptacles must be kept upright, and after a time the culture liquid in them is changed in its composition by evaporation. They are also liable to be contaminated by the entrance of mould fungi when kept in a damp place. The spores of these fungi, falling upon the surface of the cotton air filter, germinate, and the mycelium grows down through the cotton into the interior of the tube, where a new crop of spores is quickly formed. It is, therefore, a convenience to have sterile culture liquids always ready for use in a receptacle which can be packed in a box and transported from place to place; but for every-day use in the laboratory the ordinary

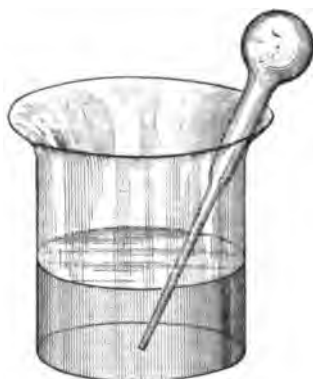


FIG. 37.

test tube, with its cotton air filter, is the most economical and convenient receptacle for culture liquids as well as for solid media. With reference to the method of making and using these little flasks, I quote from a paper published in the *American Journal of the Medical Sciences* in 1883:¹

The culture flasks employed contain from one to four fluidrachms. They are made from glass tubing of three- or four-tenths inch diameter, and those which the writer has used in his numerous experiments have all been "home-made." It is easier to make new flasks than to clean old ones, and they are thrown away after being once used. Bellows operated by foot, and a flame of considerable size—gas is preferable—will be required by one who proposes to construct these little flasks for himself.² After a little practice they are made rapidly; but as a large number are required, the time and labor expended in their preparation are no slight matter. After blowing a bulb at the extremity of a long glass tube, of the diameter mentioned, this is provided with a slender neck, drawn out in the flame, and the end of this

¹ "The Germicide Value of Certain Therapeutic Agents," op. cit., vol. clxx.

² A glass-blower ought to make them for two or three dollars per hundred.

is hermetically sealed. Thus one little flask after another is made from the same piece of tubing until this becomes too short for further use. To introduce a culture liquid into one of these little flasks, heat the bulb slightly, break off the sealed extremity of the tube and plunge it beneath the surface of the liquid (Fig. 37). The quantity which enters will of course depend upon the heat employed and the consequent rarefaction of the enclosed air. Ordinarily the bulb is filled to about one-third of its capacity with the culture liquid, leaving it two-thirds full of air for the use of the microscopic plants which are to be cultivated in it. . . . Sterilization is effected by heat after the liquid has been introduced and the neck of the flask hermetically sealed in the flame of an alcohol lamp.

Sterilization may be effected by boiling for an hour in a bath of paraffin or of concentrated salt solution, by which a temperature considerably above that of boiling water is secured. The writer is in the habit of preparing a considerable number of these flasks at one time, and leaving them, in a suitable vessel filled with water, for twenty-four hours or longer on the kitchen stove.¹

To inoculate the liquid contained in one of these little flasks with microorganisms from any source, the end of the tube is first heated to destroy germs attached to the exterior; the extremity is then broken off with sterilized (by heat) forceps; the bulb is very gently heated, so as to force out a little air, and the open end is plunged into the liquid containing the organism to be cultivated (or into a vein, or one of the solid viscera of an animal dead from an infectious germ disease, such as anthrax).

Inoculation from one tube to another may also be effected by means of the ordinary platinum wire needle.

Before the introduction of Koch's plate method for isolating bacteria in pure cultures, certain methods had been proposed, and employed to some extent, which at present have a historical value only.

Thus Klebs (1873) proposed to take from a first culture in which two or more species were associated a minute quantity, by means of a capillary tube, and with this to inoculate a second culture. By repeating this procedure several times he expected to exclude all except the species which was present in the greatest abundance and which multiplied most rapidly in the medium employed.

The *method by dilution*, first employed with precision by Brefeld (1872) in obtaining pure cultures of mould fungi, and subsequently by Lister for the isolation of bacteria, consists in so diluting a minute quantity of the mixed culture that the number of bacteria in the dilution may be less than one for each drop of the liquid. If now a single drop be added to each of a series of tubes containing a small quantity of sterile bouillon, some of the inoculations made may give a pure culture, as the drop may have contained but a single vegetative cell.

Another method of obtaining a pure culture in liquid media, when several microorganisms are associated which have a different ther-

¹ Where a steam sterilizer is at hand they will be most conveniently sterilized in the usual way, by subjecting them to the boiling temperature for an hour at a time on three successive days.

mal death-point, consists in the application of heat and thus destroying all except the most resistant species. This method is especially applicable when one of the species, only, forms spores. By subjecting the mixed culture to a temperature which is sufficient to destroy all the vegetative cells in it, the more resistant spores are left and, under favorable conditions, may subsequently vegetate and give us a pure culture of the species to which they belong.

VIII.

CULTURES IN SOLID MEDIA.

THE introduction of solid culture media in 1881 by the famous German bacteriologist, Robert Koch, inaugurated a new era in the progress of our knowledge relating to the bacteria. His methods enable us to obtain pure cultures with ease and certainty, and to study the morphological and biological characters of each species free from the complications which led to so much error and confusion before these methods were introduced. We have already given an account of the method of preparing and sterilizing the various solid culture media, and are here concerned with the manner in which they are used and the special advantages which they afford.

Koch's flesh-peptone-gelatin, which contains ten per cent of gelatin, is a transparent jelly which liquefies at from 22° to 24° C. It is a favorable culture medium for a great number of bacteria, and many species show definite characters of growth in this medium which serve to differentiate them. One of the most prominent of these characters depends upon the fact that some bacteria liquefy gelatin and others do not. This is made apparent when we make *stick cultures*—also called “stab cultures.” This is the usual manner of inoculating a solid culture medium, and is illustrated in Fig. 38. A platinum needle, consisting of a piece of platinum wire inserted into a glass rod which serves as a handle, is passed through the flame of an alcohol lamp to sterilize it. When cooled, which occurs very quickly, the point is introduced into the material containing the bacteria to be planted in the gelatin medium. We may obtain our seed for a pure culture from a single colony, from another stick culture, from the blood of an infected animal, etc. The point of the needle is then carried into the sterilized jelly, as shown in the figure, care being taken to introduce it in the central line and in a direction parallel



FIG. 38.

with the sides of the tube. It is best always to hold the tube inverted during the inoculation, and not to remove the cotton air filter until we are ready to make it. The cotton plug is then returned to its place and the platinum needle again brought to a red heat to destroy any bacteria which remain attached to it.

Sometimes it is an advantage to have the culture medium with a



FIG. 39.

sloping surface, as shown in Fig. 39. We may then draw the needle over the surface in a longitudinal direction, and by this means distribute the seed in a line along which development will take place.

The characters of growth in these stick cultures in gelatin are

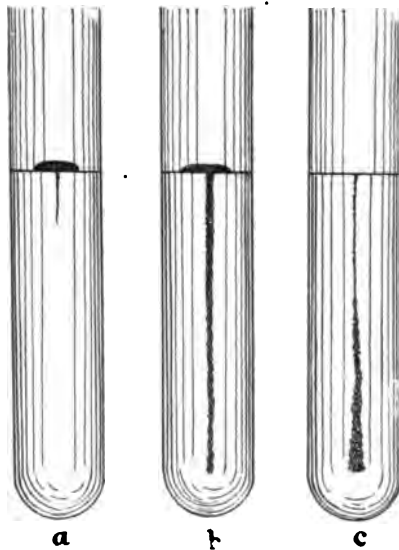


FIG. 40.

very various. Non-liquefying bacteria may grow only on the surface, as at *a*, Fig. 40; or both on the surface and along the line of puncture, as at *b*; or only at the bottom, as at *c*. In the first case the microorganism is *aërobic*—that is, it requires oxygen, and grows only in the presence of this gas. In the second case it is not strictly *aërobic*, but may grow either in the presence of oxygen

or in its absence—a *facultative anaërobic*. In the third case the microorganism is an *anaërobic*, which cannot grow in the presence of oxygen, and consequently does not grow upon the surface of the culture medium or along the upper portion of the line of puncture.

Again, we have differences as to the character of growth upon the surface or along the line of puncture. The surface growth may be a little mass piled up at the point where the needle entered the gelatin; or it may form a layer over the entire surface, and this may be thin or thick, dry or moist, viscid or cream-like, and of various colors—green, blue, red, or yellow, of different shades—or more frequently of a milk-white color.

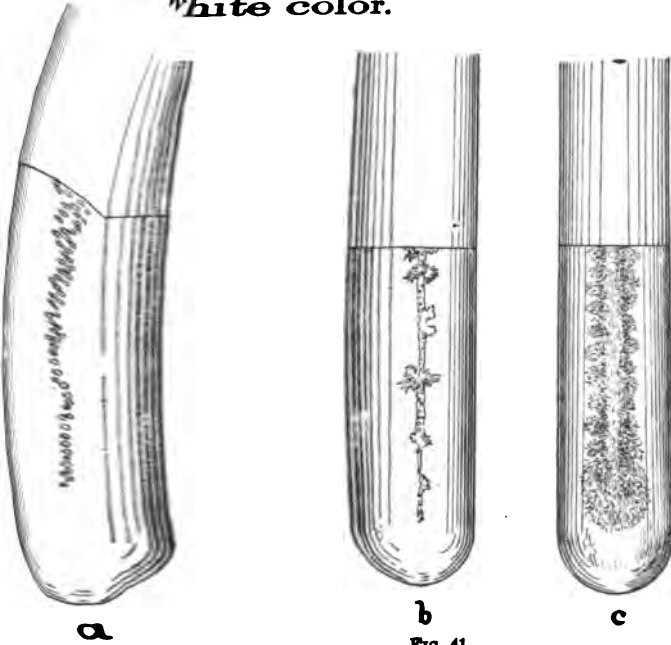


FIG. 41.

The growth along the line of puncture also differs greatly with different species. We may have a number of scattered spherical colonies (a, Fig. 41), and these may be translucent or opaque; or we may have little tufts, like moss, projecting from the line of puncture (b, Fig. 41); or slender, filamentous branches may grow out into the gelatin (c, Fig. 41).

The liquefying bacilli also present different characters of growth. Thus liquefaction may take place all along the line of puncture, forming a long and narrow funnel of liquefied gelatin (a, Fig. 42); or we may have a broad funnel, as at b; or a cup-shaped cavity, as at c; or the upper liquefied portion may be separated from that which is not liquefied by a horizontal plane surface, as at d.

The characters of growth in agar-agar jelly are not so varied, but this medium possesses the advantage of not liquefying at a temperature of 35° to 38° C., which is required for the development of certain pathogenic bacteria. Variations in mode of growth are also manifested in nutrient agar similar to those referred to as produced by non-liquefying bacteria in flesh-peptone-gelatin. These relate to the surface growth and to growth along the line of puncture. One character not heretofore mentioned consists in the formation of gas bubbles in stick cultures either in gelatin or agar.

Colonies.—If we melt the gelatin or agar in a test tube, pour the liquid medium into a shallow glass dish previously sterilized,

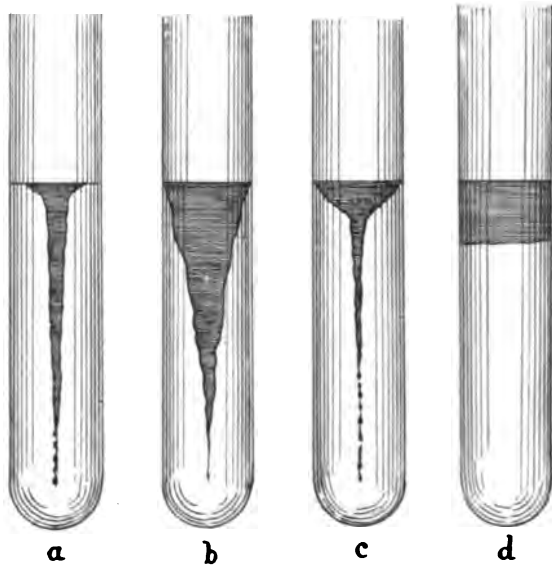


FIG. 42.

and allow it to cool while properly protected by a glass cover, we will have a broad surface of sterile nutrient material. If now we expose it to the air for ten or fifteen minutes, and again cover it and put it aside for two or three days at a favorable temperature, we can scarcely fail to have a number of colonies upon the surface of the culture medium, which have been developed from atmospheric germs which were deposited upon it during the exposure. Each of these colonies, as a rule, is developed from a single bacterium or spore, and consequently the little mass, visible to the naked eye, which we call a colony, is a pure culture of a particular species. In this experiment we are more apt to have colonies of mould fungi than of bacteria, but the principle is the same, viz., that a colony developed from a single germ is a pure culture. By touching our platinum

needle, then, to such a colony, which is quite independent of, and well separated from, all others, we may make a stick culture in gelatin or agar, and preserve the pure culture for further study. This is a most important advantage which pertains to the use of solid culture media. It is a singular fact that, as a rule, colonies of bacteria which lie near each other do not grow together, but each remains distinct. If there are but few colonies, each one, having plenty of room, may grow to considerable size; if there are many and they are crowded, they remain small, but are still independent colonies.

Now, these colonies differ greatly in their appearance and characters of growth, according to the species (Fig. 43). Some are spherical, and these may be translucent or opaque, or they may have an opaque nucleus surrounded by a transparent zone. Again, the

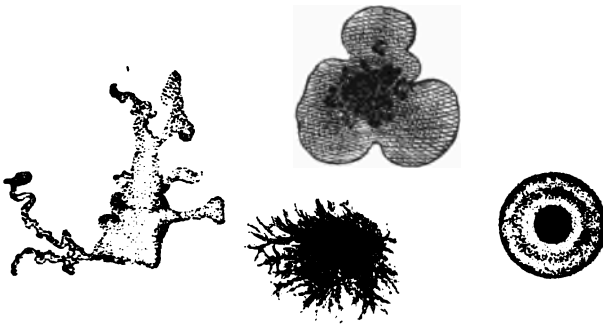


FIG. 43.—Colonies of Bacteria.

outlines may be irregular, giving rise to amoeba-like forms, or to a fringed or plaited margin, or the form may be that of a rosette, etc.; or the colony may appear to be made up of overlapping scales or masses, or of tangled filaments; or it may present a branching growth. In the case of liquefying bacteria, when the colonies have developed in a gelatin medium they commonly do not at once cause liquefaction of the gelatin, but at the end of twenty-four hours or more the gelatin about them commences to liquefy and they are seen in a little funnel of transparent liquefied gelatin; or in other cases little opaque drops of liquefied gelatin are seen, which, as the liquefaction extends, run together. All of these characters are best studied under a low-power lens, with an amplification of five to twenty diameters; and by a careful observation of the differences in the form and development of colonies we are greatly assisted in the differentiation of species.

Single, isolated colonies do not always contain a single species, for they are not always developed from a single cell. We may have

deposited upon our plate, exposed as above described, a little mass of organic material containing two or more different bacteria, and this would serve as the nucleus of a colony from which we could not obtain a pure culture.

Koch's Plate Method.—In the experiment above described, colonies were obtained from air-borne germs which were deposited upon the surface of our gelatin medium. By Koch's famous "plate method" we obtain colonies of any particular microorganism which we desire to study, or of two or more associated bacteria which we desire to study separately in pure cultures. Evidently, when we have obtained separate colonies of different bacteria upon the surface of a solid culture medium, we can easily obtain a pure culture of each by inoculating stick cultures from single colonies.

To obtain separate colonies we resort to the ingenious method of Koch. Three test tubes containing a small quantity of nutrient gelatin (or of agar) are commonly employed. The tubes are numbered 1, 2, and 3. The first step consists in liquefying the nutrient jelly by heat, and it will be well for beginners to place the tubes in a water bath having a temperature of about 40° C. (104° F.) for the purpose of keeping the culture material liquid, and at the same time at a temperature which is not high enough to destroy the vitality of the bacteria which are to be planted. We next, by means of a platinum-wire loop or the platinum needle used for stick cultures, introduce into tube No. 1 a small amount of the culture, or material from any source, containing the bacteria under investigation. Care must be taken not to introduce too much of this material, and it must be remembered that the smallest visible amount may contain many millions of bacteria. The reason for using three tubes will now be apparent. It is usually impossible to introduce a few bacteria into tube No. 1, but we effect our object by dilution, as follows: With the platinum-wire loop we take up a minute drop of the fluid in tube No. 1, through which the bacteria have been distributed by stirring, and carry it over to tube No. 2. Washing off the drop by stirring, we may repeat this a second or third time—this is a matter of judgment and experience; often it will suffice to carry over a single *öse* (the German name for the platinum-wire loop). Next we carry over one, or two, or three *öse* from tube No. 2 to tube No. 3. By this procedure we commonly succeed in so reducing the number of bacteria in tube No. 3 that only a few colonies will develop upon the plate which we subsequently make from it; or it may happen that the dilution has been carried too far and that no colonies develop upon the plate made from this tube, in which case we are likely to get what we want from tube No. 2. The next step is to pour the liquid gelatin upon sterilized glass plates, which are num-

bered to correspond with the tubes. The plates used by Koch are from eight to ten centimetres wide and ten to twelve centimetres long. They must be carefully cleaned and sterilized in the hot-air oven, at 150°C. , for two hours. They may be wrapped in paper before sterilization, or placed in a metal box especially made for the purpose. In order that the liquid gelatin may be evenly distributed upon the plate the apparatus shown in Fig. 44 is used. This consists of a glass plate, *g*, supported by a tripod having adjustable feet. By means of the spirit level *l* the glass plate is adjusted to a horizontal position. The sterilized glass plate is placed in a glass tray, shown in the figure, and the gelatin from one of the tubes is carefully poured upon it and distributed upon its surface with a sterilized glass rod, care being taken not to bring it too near the edge of the plate. The glass tray is then covered until the gelatin has cooled sufficiently to become solid, after which plate No. 1 is removed and plates Nos. 2 and 3 are made in the same way. In



FIG. 44.

order to save time it is customary to fill the glass tray shown in the figure with ice water, to place a second glass support upon it, and upon this the sterilized glass plate upon which the liquid gelatin is poured. This is protected by a glass cover, as before, until the gelatin becomes solid.

The three plates, prepared as directed, are put aside in a glass jar of the form shown in Fig. 44, one being supported above the other by a bench of sheet zinc or glass.

Petri's Dishes.—A modification of the plate method of Koch, which has some advantages, consists in the use of three small glass dishes of the same form as the larger one used by Koch to contain the plates. These dishes of Petri are about ten to twelve centimetres in diameter and one to 1.5 centimetres high, the cover being of the same form as the dish into which the gelatin is poured. These dishes take less room in the incubating oven than the larger glass jar used in the plate method, and they do not require the use of a levelling apparatus. The colonies also may be examined and counted, if desired, without removing the cover, and consequently

without the exposure which occurs when a plate prepared by Koch's method is under examination.

In agar-agar cultures or in gelatin cultures of non-liquefying bacteria made in Petri's dishes, we may examine and count colonies, without removing the cover, by inverting the dish.

In pouring the liquefied gelatin from the test tubes in which the dilution has been made into sterilized Petri's dishes, care must be taken to first sterilize the lip of the test tube by passing it through the flame of a lamp. We may at the same time burn off the top of the cotton plug, then remove the remaining portion with forceps, when the lip has cooled, for the purpose of pouring the liquid into the shallow dish.

Von Esmarch's Roll Tubes.—Another very useful modification of Koch's plate method is that of von Esmarch. Instead of pouring the liquefied gelatin or agar medium upon plates or in shallow

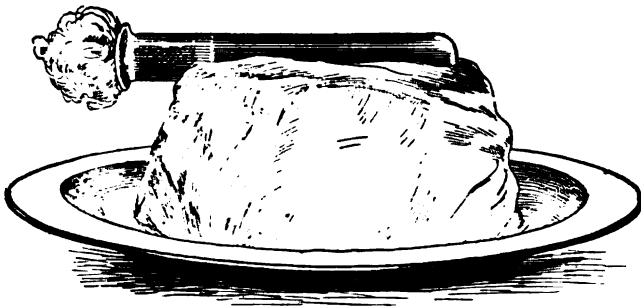


FIG. 45.

dishes, it is distributed in a thin layer upon the walls of the test tube containing it. This is done by rotating the tube upon a block of ice or in iced water. Esmarch first used a tray containing iced water, and to prevent the wetting of the cotton filter a cap of thin rubber was placed over the end of the tube. It is more convenient to turn the tubes upon a block of ice having a horizontal flat surface, in which a shallow groove is first made by means of a test tube containing hot water (Fig. 45). Or, in the winter, we may turn the tube under a stream of cold water from the city supply—*i.e.*, from a faucet in the laboratory. A little practice will enable the student to distribute the culture medium in a uniform layer on the walls of the test tube, and as soon as it is quite solidified these may be placed aside for the development of colonies from the bacteria which had been introduced. When roll tubes are made from the agar jelly it is best to place the tubes in a nearly horizontal position, for if placed upright at once the film of jelly is likely to slip from the walls of the

tube. This is due to the fact that a little fluid is pressed out of the jelly, probably by a slight contraction while cooling. If the tubes are slightly inclined from the horizontal the film does not slip and the fluid accumulates at the bottom. After a day or two they may be placed in an upright position.

These roll tubes possess several advantages. They are quickly made and take but little space in the incubating oven, and the film of jelly is protected from contamination by atmospheric germs. When colonies have formed we may examine them through the thin walls of the tube, either with a pocket lens or a low-power objective. In making a stick culture from a single colony in one of these roll tubes, we invert the tube, remove the cotton air filter, and pass the point of a sterilized platinum needle up to the selected colony. In the same way we obtain material for microscopical examination.

Streak Cultures.—In his earlier experiments with solid culture media Koch made "streak cultures" by drawing the point of a platinum needle, charged with bacteria, over the surface of a gelatin or agar plate; and this method is still useful in certain cases. If we draw the needle over the moist surface several times in succession the greater number of bacteria will be deposited in the first streak, and in the second or third single cells are likely to be left at such intervals from each other that each will develop an independent colony. If the streaks were made with impure stock we may thus succeed in getting separate colonies of the several bacteria contained in it, so that this method may be employed for obtaining pure cultures. But for this purpose it is much inferior to the plate method, and it is chiefly used for observing the growth of bacteria on the surface of solid culture media. Thus we commonly make a streak upon the surface of cooked potato or solidified blood serum in studying the development of various bacteria on these culture media.

Cultures upon Blood Serum.—The use of blood serum as a solid medium is practically restricted to stick cultures and streak cultures, for we cannot substitute it for the gelatin and agar media in making plates and roll tubes. This is because it only becomes solid at a temperature which would be fatal to most bacteria (70°C.), and when once made solid by heat cannot again be liquefied. Its use is, therefore, restricted mainly to the cultivation of bacteria for which it is an especially favorable medium. It may be used, however, in combination with a gelatin or agar medium. For this purpose it is most conveniently kept in a fluid condition in the little flasks heretofore described ("Sternberg's bulbs").

The gelatin or agar jelly in test tubes is liquefied by heat and cooled in a water bath to about 40°C. The desired amount of sterile blood serum is then forced into each tube by passing the slender

neck of the little flask along the side of the cotton filter (see Fig. 46) and applying gentle heat to the bulb. The slender neck is first sterilized by passing it through a flame, and the point is broken off with sterile forceps. After inoculating the liquefied medium in the test tubes in the usual manner we may make plates or roll tubes.

Cultures on Cooked Potato.—The method of preparing potatoes for surface cultures has already been given (page 48). It was in using them that Koch first got his idea of the importance of solid media, which led to his introduction of the use of gelatin and agar-agar and the invention of the plate method. By means of streak

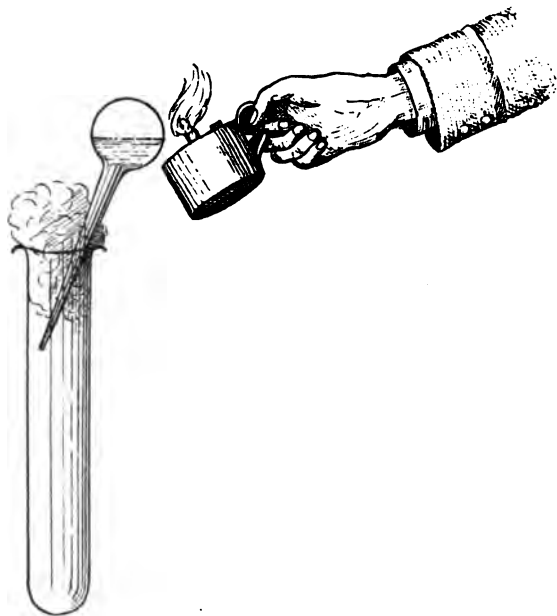


FIG. 46.

cultures upon potato he had succeeded in obtaining isolated colonies and pure cultures. We now use the potato chiefly for the purpose of differentiating species. Some bacteria grow on the surface of cooked potato and some do not. Those which do present various characters of growth. Thus we have differences as to color, as to rapidity of growth, as to the character of the mass formed—thick or thin, viscid, moist or dry, restricted to line of inoculation or extending over the entire surface, etc.

Instead of using a cut section of the potato in the manner heretofore described, we may make a *purée* by mashing the peeled and cooked tubers and distributing the mass in Erlenmeyer flasks. After

thorough sterilization by steam the culture medium is ready for use. In the same way other vegetables, or bread, etc., may be used for special purposes, and especially for cultures of the mould fungi.

Potatoes usually have a slightly acid reaction, and on this account certain bacteria will not grow upon them. This acid reaction is not constant and differs in degree, and as a result we may have decided differences in the growth of the same species upon different potatoes. To overcome this objection the writer has sometimes neutralized the cones of potato in test tubes (see Fig. 21, page 48) by first boiling them in water containing a little carbonate of soda. The liquid is poured off after they have been in the steam sterilizer for half an hour, and they are returned for sterilization.

Salomonson's Method of cultivation in capillary tubes has a historical value only since the introduction of Koch's plate method.

IX.

CULTIVATION OF ANAËROBIC BACTERIA.

PASTEUR (1861) first pointed out the fact that certain species of bacteria not only grow in the entire absence of oxygen, but that for some no growth can occur in the presence of this gas. Such bacteria are found in the soil, and in the intestines of man and the lower animals. The cultivation of "strict anaërobics" calls for methods by which oxygen is excluded. The "facultative anaërobics" grow

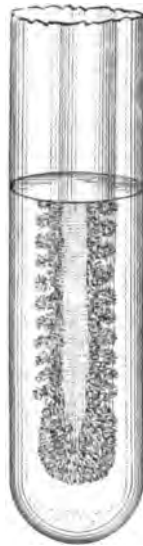


FIG. 47.



FIG. 48.

either in the presence or absence of oxygen. There are various gradations in this regard, from the strictly aërobic species which require an abundance of oxygen and will not grow in its absence, to the strictly anaërobic species which will not grow if there is a trace of oxygen in the medium in which we propose to cultivate them. Among the most interesting pathogenic bacteria which are strictly anaërobic are the bacillus of tetanus, the bacillus of malignant œdema, and the bacillus of symptomatic anthrax.

If we make an inoculation of one of the species which is not strictly anaërobic into a test tube containing nutrient gelatin or agar-agar, we may have a development all along the line of puncture, and this may be more abundant below, as in Fig. 47. But when we make a long stick culture with a strict anaërobic the development occurs only near the bottom of the line of puncture (Fig. 48).

We may then, if we have a pure culture to start with, propagate these anaërobic bacilli in *long stick cultures*. It is best to use tubes which have been recently sterilized, as boiling expels the air from the culture medium; and a very slender needle should be used in making the inoculation. To prevent the absorption of oxygen a layer of sterilized olive oil may be poured into the tube after the inoculating puncture has been made, or it may be filled up with agar jelly which has been cooled to about 40° C. Roux has proposed to prevent the absorption of oxygen by the culture medium by planting an aërobic bacterium—*Bacillus subtilis*—upon the surface, after making a long stick culture with the anaërobic species. The agar jelly is first boiled and quickly cooled; the inoculation is then made with a slender glass needle; some sterile agar cooled to 40° C. is poured into the tube, and when this is solid the aërobic species is planted upon the surface. The top of the test tube is then closed hermetically and it is placed in the incubating oven. The aërobic species exhausts the oxygen in the upper part of the tube by its growth on the surface of the culture medium, and the anaërobic species grows at the bottom of the tube. To obtain material for a new culture or for microscopical examination the test tube is broken near its bottom.

Cultures in liquid media may be made by exhausting the air in a suitable receptacle or by displacing it with hydrogen gas. The first-mentioned method has been largely used in Pasteur's laboratory, but methods in which hydrogen gas takes the place of atmospheric air in the culture tube are more easily applied and require simpler apparatus. The flask shown in Fig. 49 may be used in connection with an air pump. The sterile culture liquid is first introduced into a long-necked flask and inoculated with the anaërobic bacillus to be cultivated. The neck of the flask is then drawn out in a flame at *c*. The open end is then connected with a Sprengle's pump or some other apparatus for exhausting the air. The flask is placed in a water bath at 40° C., which causes ebullition at the diminished pressure, and the exhaustion is continued for about half an hour. The narrow neck is then sealed at *c* by the use of a blowpipe flame.

The flask shown in Fig. 49, which can be made from a test tube, may also be used in connection with a hydrogen apparatus. In this case a slender glass tube is passed into the flask, as shown in Fig.

50, and this is connected with a hydrogen apparatus by a rubber tube. The hydrogen is allowed to bubble through the culture liquid in a full stream for ten to fifteen minutes, in order that all of the oxygen in the flask may be removed by displacement. Then, while the gas is still flowing, the flask is sealed at *a* with a blow-pipe flame, the hydrogen tube being left in position and melted fast to the flask. Some little skill is required in the successful performance of the last step in this procedure, and it will be easier for those

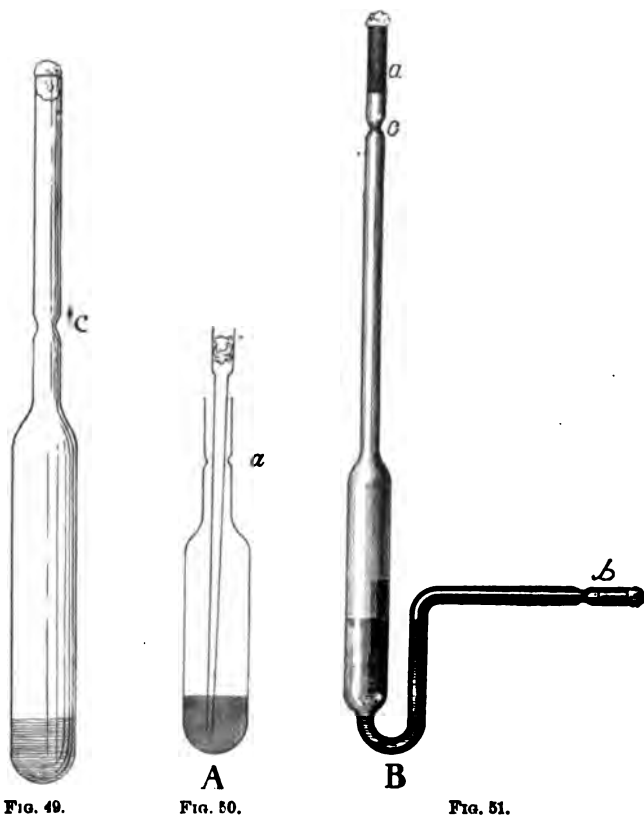


FIG. 49.

FIG. 50.

FIG. 51.

who are not skilful in the use of the blowpipe to use Salomonson's tube, shown in Fig. 51. In this, hydrogen is admitted through the arm *b*, and escapes through the cotton plug *a*. The vertical tube is sealed at *c* while the gas is flowing, and then the horizontal tube at *b*.

Fränkel's Method.—Instead of these tubes specially made for the purpose, an ordinary test tube may be used, as recommended by Fränkel. This is closed by a soft rubber cork through which two glass tubes pass—one, reaching nearly to the bottom of the test tube,

for the admission of hydrogen, which passes through the liquefied culture medium ; and the other a short tube for the escape of the gas. The outlet tube is sealed in the flame of a lamp while the gas is freely flowing, and after sufficient time has elapsed to insure the complete expulsion of atmospheric oxygen—which, when the hydrogen flows freely, requires about four minutes (Fränkel)—melted paraffin is applied freely to the rubber stopper to prevent leakage of the hydrogen and entrance of oxygen. A roll tube may then be made after the manner of Esmarch, and, after colonies have developed, the anaerobic culture will appear as shown in Fig. 52.

To isolate anaerobic bacteria in pure cultures it is well to make a

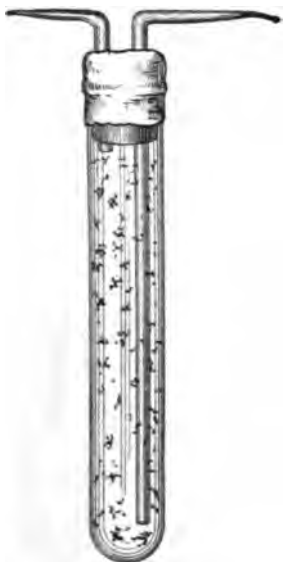


FIG. 52.

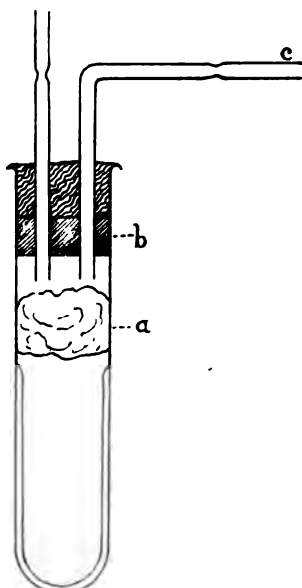


FIG. 53.

series of dilutions as heretofore described for aerobic cultures ; we will then usually obtain isolated colonies in tube No. 2 or No. 3 of a series, and by removing the rubber stopper we may transplant bacteria from these colonies to deep stick cultures in nutrient gelatin or agar.

The Writer's Method.—The following simple method has been successfully employed by the writer:

Three Esmarch roll tubes are prepared as is usual for aerobic cultures. The cotton air filter, or a portion of it, is then pushed down the tubes for a short distance, as shown at *a*, Fig. 53. A section of a soft rubber stopper carrying two glass tubes is then pushed into the

gelatin, previously cooled upon its walls, which contains the bacteria under investigation. When the anaerobic colonies have developed the test tube must be broken to get at them, or the cylinder of gelatin may be removed by first warming the walls of the tube.

Another *method, recommended by Liborius*, consists in distributing the bacteria in test tubes nearly filled with nutrient gelatin or agar which has been recently boiled to expel air. Colonies of anaerobic bacteria will develop near the bottom of such a tube, while the aerobic species will only grow near the surface. The cylinder of jelly is removed by heating the walls of the tube, and sections are

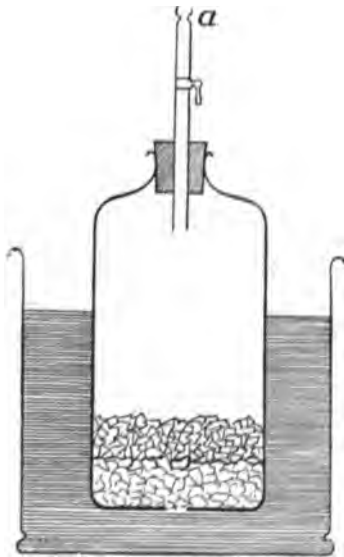


FIG. 56.

made with a sterilized knife for the purpose of obtaining material from individual colonies for further cultures, etc.

Koch and his pupils are in the habit of testing the aerobic character of bacteria in plate cultures by covering the recently made plates with a thin sheet of mica which has been sterilized by heat. The strictly aerobic species do not grow under such a plate; but, according to Liborius, the exclusion of oxygen is not sufficiently complete for the growth of strict anaerobics.

Buchner's Method consists in the removal of oxygen by means of pyrogallic acid. The anaerobic species under investigation is planted in recently boiled agar jelly in a small test tube. This is placed in a larger tube having a tightly fitting rubber stopper, as shown in Fig. 55. The small tube is supported by a bent-wire

stand, and in the lower part of the large tube are placed ten cubic centimetres of a ten-per-cent solution of caustic potash, to which one gramme of pyrogalllic acid is added. The absorption of the oxygen takes some time, but, according to Buchner, it is finally so complete that strict anaerobics grow in the small tube.

In practice, cultivation in an atmosphere of hydrogen will be found the most convenient method, and for this any form of hydrogen generator may be used. The writer is in the habit of using the form shown in Fig. 56. A perforation a quarter of an inch in diameter is drilled through the bottom of a wide-mouthed bottle. Some fragments of broken glass are then put into the bottle, form-

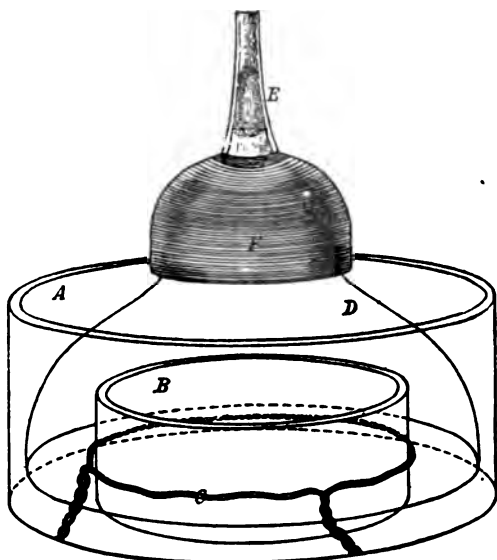


FIG. 57.

ing a layer two or three inches thick. Upon this is placed a quantity of granulated zinc. This bottle has a tightly fitting cork, through which passes a metal tube having a stopcock. The bottle is placed in a glass jar containing diluted sulphuric acid (one part by weight of sulphuric acid to eight parts of water). The acid, rising through the perforation in the bottom of the bottle, when it comes in contact with the zinc gives rise to an abundant evolution of hydrogen, which escapes by the tube *a* when the stopcock is open. When this is closed the gas forces the acid back from contact with the zinc. To remove any trace of oxygen present the gas may be passed through a solution of pyrogalllic acid in caustic potash.

Evidently plates prepared by Koch's method, or Esmarch roll

tubes, may be placed in a suitable receiver and the air exhausted, or hydrogen substituted for atmospheric air. Such an apparatus for hydrogen has been devised by Blücher and is shown in Fig. 57. A glass dish, A, contains a smaller dish, B, which has a diameter of about seven centimetres. The small dish is kept in its position in the centre of the larger one by the wire ring, having three projecting arms, which is shown in the figure. The culture medium containing the anaërobic bacteria to be cultivated is poured into the small dish and the glass funnel D is put in position. This is held in its place by a weight of lead which encircles the neck of the funnel at F. A mixture of glycerin and water (twenty to twenty-five per cent) is poured into the dish A to serve as a valve to shut off the atmospheric air from the interior of the funnel D. Hydrogen gas is introduced through the tube E, which is connected by a rubber tube with a hydrogen apparatus.

A somewhat similar apparatus has been devised by Botkin, in which the hydrogen is admitted beneath a bell jar covering small glass dishes containing the culture medium. We believe that in practice the writer's method (page 81), in which Esmarch roll tubes are first made, will be found more convenient than either of the last-mentioned methods of preserving plates in an atmosphere of hydrogen; or roll tubes may be prepared in the way usually practised in cultivating aërobic bacteria, and these may be placed in a suitable receptacle which can be filled with hydrogen.

X.

INCUBATING OVENS AND THERMO-REGULATORS.

THE saprophytic bacteria generally, and many of the pathogenic species, grow at the ordinary temperature of occupied apartments (20° to 25° C.); but some pathogenic species can only be cultivated at a higher temperature, and many of those which grow at the "room temperature" develop more rapidly and vigorously when kept in an incubating oven at a temperature of 35° to 38° C. Every bacteriological laboratory should therefore be provided with one or more brood ovens provided with thermo-regulators to maintain a constant temperature. These incubating ovens are made with double walls surrounding an air chamber. The space between the double walls is filled with water, which is usually heated by a small gas flame. The gas passes through the thermo-regulator, and its flow is automatically controlled for any temperature to which this is adjusted. The exterior of the incubating oven is covered with felt or asbestos to prevent the loss of heat by radiation. A simple and cheap form which answers every purpose is shown in Fig. 58. The quadrangular box with double walls should be made of zinc or copper. An outer metal door covered with non-conducting material, and an inner door of glass, give access to the interior space; and a thermometer introduced through an aperture in the top (Fig. 58, *b*) shows the temperature of this space when the door is closed. The stopcock *e* permits the drawing off of the water from the space between the double walls, and the glass tube *d* shows the height of the water, as it is connected with the space containing it. The thermo-regulator passes through an aperture at one side of the oven into the water, the temperature of which controls the flow of gas.

The ordinary thermo-regulator is shown in Fig. 59 as manufactured by Rohrbeck. A glass receptacle, shaped like an ordinary test tube, has an arm, *c*, for the escape of the gas, which enters by the bent tube *a*, which passes through a perforated cork and is adjustable up and down. Tube *a* is connected with the gas supply and tube *c* with the burner by means of rubber tubing. A glass partition extending downward as a tube, *g*, makes an enclosed space in

the lower part of the instrument, and this, when immersed in water, acts as a thermometer bulb. This space contains mercury below and air or the vapor of ether above. When the air is expanded by heat the mercury is forced up the tube *g* until it meets the end of the inlet tube for gas at *h*, and by shutting off the flow of gas prevents the temperature from going any higher. A small opening in the inlet tube at *e* permits a small amount of gas to flow, so that the flame under the brood oven (Fig. 58, *f*) may not be entirely extinguished. The lower end of the bent tube *a* is bevelled, so that a triangular opening is formed, which is closed gradually by the rising

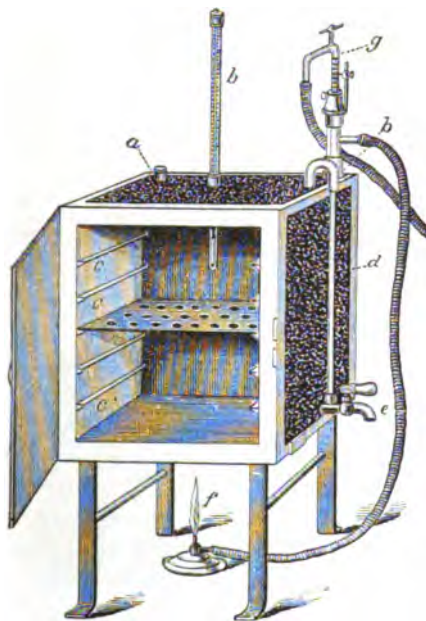


FIG. 58.

mercury, instead of abruptly as would be the case if the lower end of the tube *a* were cut off square. To adjust the temperature in the air space of the incubating oven when the thermo-regulator is in position, a full flow of gas is admitted to the burner until the thermometer (Fig. 58, *b*) shows the desired temperature; then the bent tube *a* is pushed down through the cork until its lower extremity meets the mercury and the flame *f* is somewhat reduced. The apparatus is then left for a time, to see whether the flame runs too high or too low, and a further adjustment is made. When the changes in the exterior temperature are slight and the gas pressure regular the temperature in the air chamber is controlled with great precision. But this is not the case under the reverse conditions. Changes in

the pressure of gas, especially, interfere with the maintenance of a constant temperature, and for this reason a pressure regulator will be required when great precision is desired. That of Moitessier is commonly used in bacteriological laboratories (Fig. 60). But for most purposes variations of temperature of 1° to 2° C. are not of great importance. For ordinary use a brood oven should be regulated to about 35° to 37° C. It is best to have a little cylindrical screen of mica around the gas jet beneath the incubating oven, for the purpose of preventing the flame from being extinguished by currents of air (Fig. 61).

Koch's ingenious automatic device for shutting off the gas if the flame is accidentally extinguished is shown in Fig. 62.

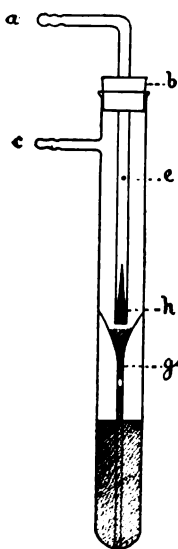


FIG. 59.

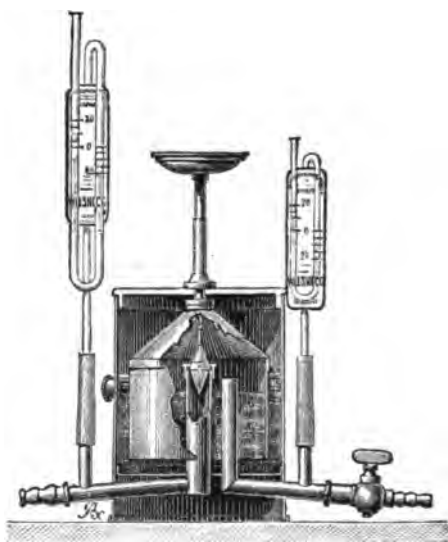


FIG. 60.

Another form of thermo-regulator, which answers very well, is that of Reichert (Fig. 63). In this the gas enters at *a* and escapes at *c*. The mercury, which fills the bulb, shuts off the gas at the point for which the instrument is regulated. By means of the screw *d* the height of the mercury in the tube may be very accurately adjusted for any desired temperature.

The regulator of Bohr, shown in Fig. 64, is more sensitive than that of Reichert, and rather simpler in construction than the usual form shown in Fig. 59. The thermometer bulb *a* contains only air, and the gas which passes through the tube *f* is shut off at the proper temperature by the mercury in the U-shaped tube *c*. The stopcock *b* is left open when the bulb *a* is immersed in the water

bath, and when the proper temperature is reached is closed so as to confine the air in the bulb. An increase of temperature now causes



FIG. 61.

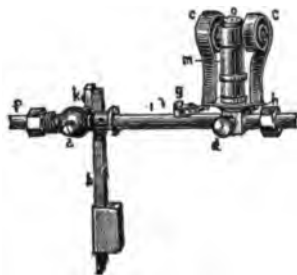


FIG. 62.

the air in the bulb to expand, the mercury in the U-tube is forced up

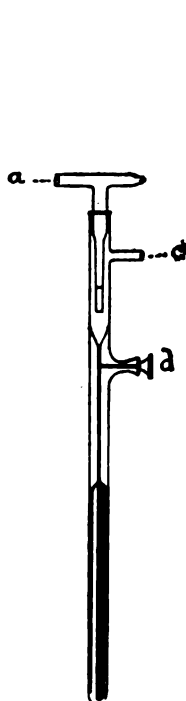


FIG. 63.

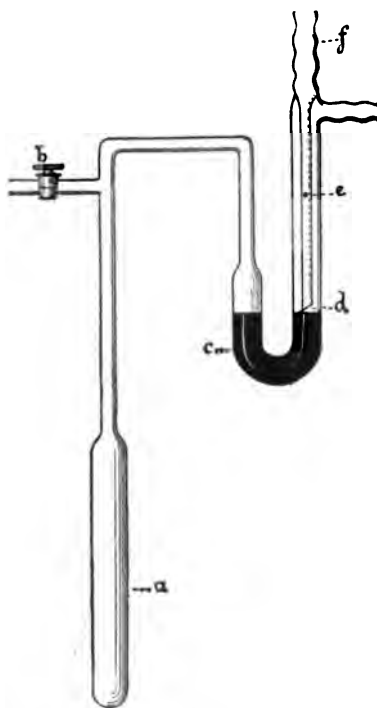


FIG. 64.

and shuts off the gas flowing through the tube *f* at its lower extremity, *d*. A small opening, *e*, permits sufficient gas to pass to

maintain a small flame which must not be sufficient by itself to keep up the desired temperature in the water bath.

Altmann has recently (1891) described a thermo-regulator which is made by Müncke, of Berlin, and which is shown in Fig. 65. This is said to act with great precision. It is a modification of Reichert's

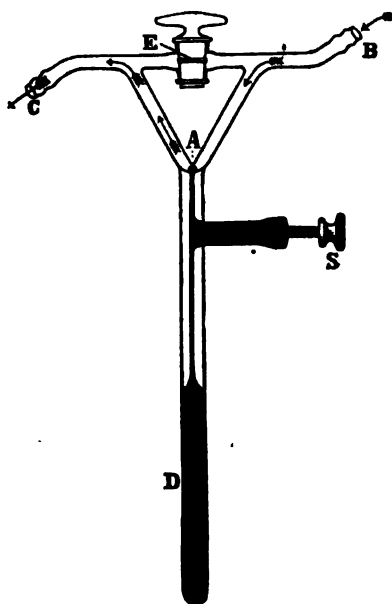


FIG. 65.



FIG. 66.

regulator. Its mode of action will be readily understood by a reference to the figure.

A thermo-regulator which gives very accurate results, which are not influenced by differences in pressure, is that invented by the

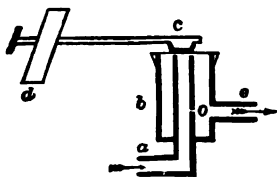


FIG. 67.

writer over twenty years ago. The regulating thermometer may contain mercury only, or air and mercury, as shown in the thermo-regulator for gas (Fig. 59). In the simplest form a large bulb containing mercury is used, and a platinum wire is hermetically sealed in the glass so as to have contact with the mercury (Fig. 66, *a*).

Another platinum wire passes down the tube of the thermometer, *b*, and is adjustable for any desired temperature. The gas passes through a valve which is controlled by an electro-magnet. A simple form of valve is shown in Fig. 67. The bent tube *a* is connected with the gas supply by a piece of rubber tubing. The upright arm of this tube is enclosed in a larger tube, *b*, having an out-

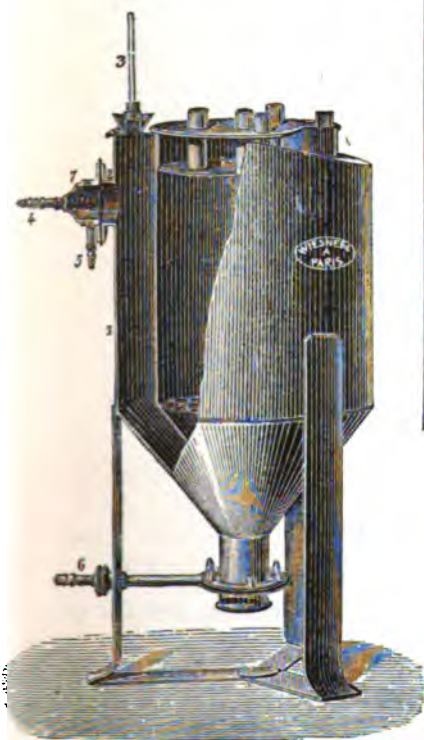


FIG. 68.

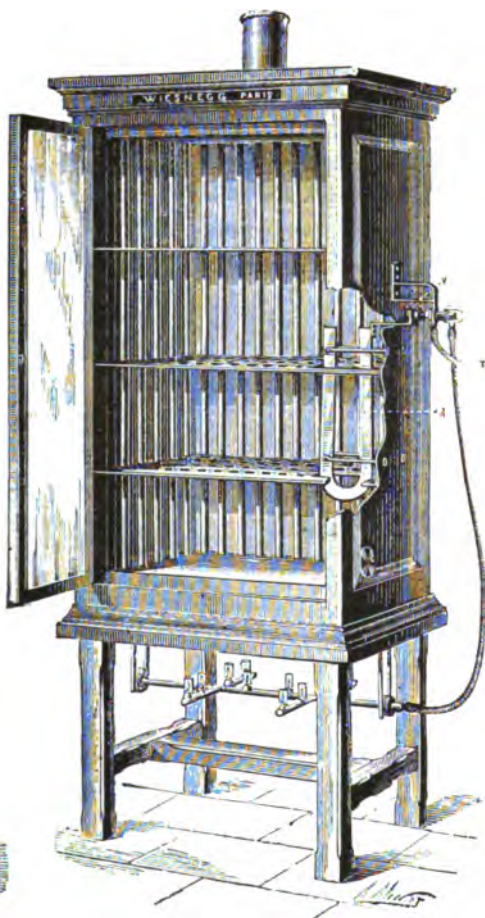


FIG. 69.

let, *e*, which is connected with the burner under the incubating oven. The upper end of this larger tube is closed by means of a piece of sheet rubber, which prevents the escape of gas. When this is depressed by means of the lever *c*, the flow of gas through the valve is arrested. The lever *c* has attached to it the armature *d*, and is operated by an electro-magnet under the control of the regulating thermometer.

When the thermometer is immersed in a water bath the temperature of which it is desired to regulate, and the proper electric connections are made, it acts as a circuit breaker. When the desired temperature is reached the mercury in the tube of the thermometer touches the wire *b* (Fig. 66), an electric circuit is completed, and the valve is closed, shutting off the gas supply and preventing the temperature from going any higher. When contact is broken in the thermometer tube the valve opens and permits the gas to flow again. A small opening, *o* (Fig. 67), permits the constant flow of a sufficient amount of gas to prevent the flame from being extinguished. In practice, however, it is better to have a small side jet of gas, quite independent of that which passes through the valve, which burns constantly and relights the principal jet when

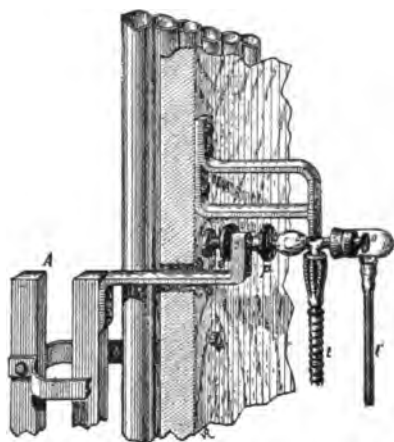


FIG. 70.

the valve is opened. This apparatus is very well adapted for regulating the temperature of a water bath with precision, but for general use in connection with incubating ovens the ordinary gas regulator is preferable, on account of the trouble connected with keeping a galvanic battery in order when it is required to act at frequent intervals "on a closed circuit," for weeks and months together.

The incubating apparatus of D'Arsonval is shown in Fig. 68. It is a cylindrical vessel of copper having double walls, and is provided with the thermo-regulator of D'Arsonval, by which very accurate regulation is maintained at any desired temperature. In its form this apparatus is not as convenient as are the brood ovens made in the form shown in Fig. 58, with a swinging door which gives easy access to the interior, which is provided with one or more shelves upon which the cultures are placed. Various modifications

of this simple and convenient incubating oven are manufactured by Rohrbeck and by Müncke, of Berlin. The apparatus of D'Arsonval, and other forms in favor at the French capital, may be obtained from Wiesnegg, of Paris. The last-named manufacturer also supplies the incubating oven and thermo-regulator recently described by Roux (1891). This is shown in Fig. 69. The regulator is formed of two metallic bars, one of steel and the other of zinc; these are soldered together in the shape of a letter **U**; the regulator is seen in position in the cut (Fig. 69). The most dilatable metal (zinc) is on the outside. When the temperature is raised the arms of the **U** approach each other, and the reverse when it falls. The method by which regulation is effected is shown in Fig. 70. The **U**-shaped regulator is placed vertically, and one of its branches, *A*, is firmly fixed to the wall of the incubating oven; the other, free arm carries a horizontal bar which projects through the wall of the incubator in an opening which permits it to move freely under the influence of a change in the temperature within. The end of this projecting bar is turned up at a right angle and the screw *p* passes through it; this can be fixed at any desired point by means of the nut *e*. The end of the screw *p* rests against the stem of a conical brass valve which controls the flow of gas. The valve is closed by a spiral spring and opened by the screw *p* under the control of the thermo-regulator.

XI.

EXPERIMENTS UPON ANIMALS.

THE pathogenic power of various bacteria has been demonstrated by injecting pure cultures into susceptible animals. As a rule, the herbivora are more susceptible than the carnivora, and this is perhaps to be explained in accordance with the theory of natural selection. Carnivorous animals often feed upon the bodies of animals which have succumbed to infectious diseases, and upon dead animals in which putrefactive changes have commenced. In their struggles with each other they are wounded by teeth and claws soiled with infectious material which would cause a fatal disease if inoculated into the more susceptible herbivorous animals. As this has been going on for ages, we may suppose that, by survival of the fittest, a race tolerance has been acquired. The lower animals have their own infectious diseases, some of which are peculiar to certain species and some common to several. As a rule, the specific infectious diseases of man cannot be transmitted to lower animals, and man is not subject to the diseases of the same class which prevail among animals. But certain diseases furnish an exception to this general rule. Thus tuberculosis is common to man and several of the lower animals; relapsing fever may by inoculation be transmitted to monkeys; diphtheria may be transmitted to pigeons and guinea-pigs. On the other hand, anthrax and glanders may be contracted by man as a result of accidental inoculation or contact with an infected animal.

Nearly allied species sometimes present very remarkable differences as to susceptibility. Thus the bacillus of mouse septicæmia is fatal to house mice but not to field mice, while, on the other hand, field mice are killed by the bacillus of glanders and house mice are immune from this pathogenic bacillus.

The animals most commonly used for testing the pathogenic power of bacteria are the mouse, the guinea-pig, and the rabbit. Domestic fowls and pigeons are also useful for certain experiments. The dog and the rat are of comparatively little use on account of their slight susceptibility.

Inoculations are made directly into the circulation through a vein, into the subcutaneous connective tissue, or into one of the serous cavities—usually the peritoneal.

The ordinary hypodermic syringe may be used in making injections, but this is difficult to sterilize on account of the leather piston, and complications are liable to arise from its use which it is best to avoid. The best way to sterilize a piston syringe is to wash it thoroughly with a solution of bichloride of mercury of 1 : 1,000, and then to remove every trace of bichloride by washing in alcohol. But one never feels quite sure that the most careful washing will insure sterilization, and it is best to use a syringe which may be sterilized by

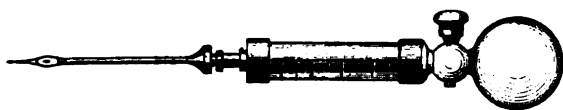


FIG. 71.

heat, such as that of Koch, shown in Fig. 71. In this the metal point and glass tube are easily sterilized in a hot-air oven. Fluid is drawn into the syringe and forced out of it by a rubber ball which has a perforation to be covered by the finger.

The writer has for some years been in the habit of making injections in animals with an improvised glass syringe. This is made from a piece of glass tubing in the same form as the collecting tubes heretofore described. A bulb is blown at one end of the tube, and the other end is drawn out to form a slender tube which serves as the

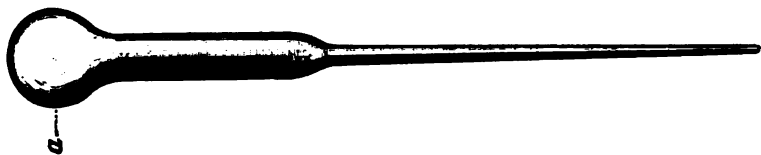


FIG. 72.

needle of the syringe (Fig. 72). By gently heating the bulb in an alcohol lamp and immersing the open end of the capillary tube in the fluid to be injected, this rises into the syringe as the expanded air cools. Having introduced the glass point beneath the skin or into the cavity of the abdomen of the animal to be injected, the contents of the tube are forced out by again heating the bulb by means of a small alcohol lamp. The glass point is easily forced through the thin skin of a mouse or of a young rabbit; but for animals with a thicker skin it is necessary to cut through, or nearly through, the skin with some other instrument. A small pair of curved scissors answers very well for this purpose.

Generally, in making injections into animals, it is customary to remove the hair for some distance around the point of inoculation with scissors and razor, and then to sterilize the surface by careful washing with a solution of bichloride of mercury. This precaution is necessary in researches in which pathogenic bacteria are being tested, in order to remove any possibility of accidental inoculation with germs other than those under investigation, and, as a consequence, a mistaken inference as to the pathogenic action of the species under investigation. But when we know the specific pathogenic power of a certain microorganism it is hardly necessary to take this precaution, as a few drops of culture will contain millions of the bacteria, while contamination, if it occurs from the surface of the body, must be by a comparatively small number of bacteria, which are likely to be of a harmless kind which will have no influence on the result of the experiment.

Instead of sterilizing the surface, the writer usually clips away a small portion of skin with curved scissors, not cutting deep enough to draw blood, but leaving a bare surface through which the point of the syringe can be introduced with very little danger of carrying bacteria into the connective tissue other than those contained in the syringe.

In making injections into the peritoneal cavity care must be taken not to wound the liver or the distended stomach. The intestine is not very likely to be wounded, as it slips out of the way. By seizing a longitudinal fold of the abdominal wall and pushing the point of the syringe quite through it, and then releasing the fold and carefully withdrawing the instrument until the point remains in the cavity, the danger of wounding the intestine will be reduced to a minimum.

Injections into the circulation are made by exposing a vein and carefully introducing the needle of the syringe in the direction of the blood current. Care must of course be taken not to inject air. In the rabbit one of the large veins of the ear may be conveniently penetrated by the point of a hypodermic syringe without any previous dissection. The ear is first washed with a solution of bichloride of mercury or simply with warm water. The animal had better be carefully wrapped in a towel to control its movements. The veins are distended by compressing them near the base of the ear. When the point of the needle has not been properly introduced, and the fluid to be injected escapes in the surrounding connective tissue, it will commonly be best to withdraw the syringe and make the attempt upon another vein. As pointed out by Abbott, the needle of the syringe should be ground flat at the point, and not curved as is commonly the case.

Large quantities of fluid may be injected into the cavity of the abdomen or into the circulation by slowly forcing the fluid through a slender canula, properly introduced, which is coupled with a large syringe by means of rubber tubing, or with a glass receptacle from which the fluid is forced by the pressure of air pumped in with a rubber hand ball.

Mice are usually injected subcutaneously near the tail. The little animal is first seized by a long pair of forceps, or "mouse tongs, and the hair is clipped away on the back just above the tail. If solid material is to be introduced a little pocket is made with scissors or with a lancet, into which the infectious material is carried by means of a platinum needle or slender forceps. Liquids may be injected by the little glass syringe heretofore described, the point of which is easily forced through the skin.

Pasteur's method of inoculating rabbits with the virus of hydrophobia consists in trephining the skull and injecting the material beneath the dura mater. An incision through the skin is first made to one side of the median line a short distance back of the eyes. The edges of the wound are separated, and a small trephine (five or six millimetres in diameter) is used to remove a button of bone. The emulsion of spinal cord from a hydrophobic animal is then carefully injected beneath the dura mater—two or three drops will be sufficient. The wound is washed out with a two-per-cent solution of carbolic acid and closed with a couple of sutures.

Injections into the intestine are made by carefully opening the abdomen with antiseptic precautions, gently seizing a loop of the intestine, and passing the point of the syringe through its walls; the loop is then returned and the incision in the walls of the abdomen carefully closed with sutures and dressed antiseptically.

Inoculations into the anterior chamber of the eye of rabbits and other animals have frequently been practised, and offer certain advantages in the study of the local effects of pathogenic microorganisms. The animal should be fastened to an operating board, belly down, and its head held by an assistant, who at the same time holds the eyelids apart. The conjunctiva is seized with forceps to steady the eye, and an incision about two millimetres long is made through the cornea with a cataract knife. Through this opening a small quantity of a liquid culture may be injected, or a bit of solid material introduced with slender curved forceps.

Ordinary injections give but little pain and do not call for the use of an anæsthetic. When anæsthesia is required ether will usually be preferable to chloroform. Rabbits, especially, are very apt to die from chloroform, no matter how carefully it may be administered. Dogs, rats, and mice stand ether very well. The smaller animals

may be brought under the anæsthetic by placing them in a covered jar into which a pledget of cotton wet with ether has been dropped. Before making injections into the anterior chamber of the eye it is well to use a two-per-cent solution of cocaine as a local anæsthetic.

Mice which have been inoculated are usually kept in a glass jar having a wire-gauze cover. A quantity of cotton is put into the jar to serve as a shelter for the little animal, and it is well to partly fill the jar with dry sawdust. Larger animals are kept in suitable cages of wire or wood, and, as a rule, each one should be kept in a separate cage while under observation after an inoculation experiment.

In experimenting upon animals the following points should be kept in view and noted :

(a) *The age and weight of the animal.* Young animals are, as a rule, more susceptible than older ones, and with many pathogenic bacteria the lethal dose of a culture bears some relation to the size of the animal.

(b) *The point of inoculation.* Injections into the circulation are generally more promptly fatal and require a smaller dose than those into a serous cavity or into the connective tissue. Pathogenic bacteria introduced into the abdominal cavity reach the circulation more promptly than those injected subcutaneously. But certain microorganisms owe their pathogenic power to the local effect about the point of inoculation and the absorption of toxic products formed in the limited area invaded, and do not enter the general circulation, or at least do not multiply in the circulating fluid, and quickly disappear from it.

(c) *The age of the culture injected.* Old cultures sometimes have greater and sometimes less pathogenic potency than recent cultures. Some kinds of virus become "attenuated" when kept. But when the pathogenic power depends chiefly upon toxic products formed during the growth of the bacteria, old cultures are, as a rule, more potent than those recently made.

(d) *The medium in which the pathogenic bacteria are suspended.* Cultures in albuminous media, like blood serum, are in some cases more potent than bouillon cultures ; and the virulence of several pathogenic bacteria is greatly intensified by successive cultures—by inoculation—in the bodies of susceptible animals. Ogston found that pus cocci cultivated in the interior of eggs had an increased virulence. According to Arloing, Cornevin, and Thomas, the activity of a culture of the bacillus of symptomatic anthrax is doubled by adding one-five-hundredth part of lactic acid to the culture fluid.

(e) *The quantity injected* is evidently an essential point when the result depends largely upon the toxic products formed in the cul-

ture medium. It is also an essential point when pathogenic bacteria are injected which kill susceptible animals in very minute doses, for it has been shown by the experiments of Watson Cheyne and others that in the case of some of these, at least, there is a limit below which infection does not occur.

Inoculated animals should be carefully observed, and a note made of every symptom indicating a departure from the usual condition of health, such as fever, loss of activity, loss of appetite, weakness, emaciation, diarrhoea, convulsions, dilated pupils, the formation of an abscess or a diffuse cellulitis extending from the point of inoculation, etc. The temperature is usually taken in the rectum. The temperature of small animals, like rabbits and guinea-pigs, varies considerably as a result of external conditions. In the rabbit the normal temperature may be given as about 102° to 103° F. ; in the guinea-pig it is a little lower.

In making a post-mortem examination of an inoculated animal it is best to stretch it out on a board, belly up, by tying its legs to nails or screws fastened in the margin of the board. When the abdomen is dirty, as is usually the case, it should be carefully washed with a disinfecting solution. An incision through the skin is then made in the median line the full length of the body, and the skin is dissected back so as to expose the anterior walls of the abdomen and thorax. These cavities are then carefully opened with a sterilized knife or scissors, and the various organs and viscera examined. Attention should also be given to the appearances at the point of inoculation. To ascertain whether the microorganism injected has invaded the blood, smear preparations should be made with blood obtained from a vein or from one of the cavities of the heart. It will be well also to make a smear preparation from a cut surface of the liver and spleen. In the various forms of acute septicæmia the spleen is usually found to be enlarged. If but few microorganisms are present in the blood and tissues they may escape observation in stained smear preparations, and it will be necessary to make cultures to demonstrate their presence. A little blood from a vein or from one of the cavities of the heart is transferred, by means of a platinum loop (*öse*) or a sterilized collecting tube (see page 38), to a test tube containing liquefied nutrient gelatin or agar-agar, and an Esmarch roll tube is made. This is put aside for the development of colonies from any scattered bacteria which may be present. As a rule, it will be best to make agar cultures, as these can be placed in the incubating oven at 35° to 38° C. Stick cultures may also be made and will serve to show the presence of microorganisms, but will not give information as to how numerous they may be. The roll tube also has the advantage of showing whether there is a

mixed infection or whether a pure culture of a single microorganism is obtained from the blood. In the same way cultures may be made from material obtained from the liver or spleen, and it may happen that one or both of these organs contain bacteria when none are found in the blood. Before passing the platinum needle or collecting tube into the organ, the surface, which has been more or less exposed to contamination, should be sterilized by applying to it a hot spatula; then at the moment of lifting the spatula the sterilized needle is introduced into the interior of the organ, and the blood and crushed tissue adhering to it at once carried over to the culture medium. Or blood obtained with proper precautions from a vein, a cavity of the heart, or the interior of the spleen or liver, may be used to inoculate another animal.

Animals are also sometimes inoculated by excoriating the cutis as in vaccination. They may also, in rare cases, be infected by introducing cultures into the stomach, either mixed with the food ingested or by injection through a tube. Infection by inhalation is accomplished by causing the animal to respire an atmosphere, in a properly enclosed space, in which the pathogenic organism is suspended, by the use of a spray apparatus for liquid cultures, or some form of powder blower for powders containing the bacteria in a desiccated condition.

One method of obtaining a pure culture of pathogenic bacteria consists in the inoculation of susceptible animals with material containing a pathogenic species in association with others which are not. When the blood is invaded by the pathogenic species and the animal dies from an acute septicæmia, we may usually obtain a pure culture by inoculating a suitable culture medium with a minute drop of blood taken from a vein or from one of the cavities of the heart. Sometimes, however, a mixed infection occurs and some other microorganism is associated in the blood with that one which was the immediate cause of the death of the animal.

XII.

PHOTOGRAPHING BACTERIA.

WELL-MADE photomicrographs are unquestionably superior to drawings made by hand as a permanent record of morphological characters. This being the case, bacteriologists would no doubt resort to this method more generally but for the technical difficulties and the time and patience required in overcoming these. Koch, in his earlier studies, gave much time to photographing bacteria, and with very remarkable success. In his work on "Traumatic Infective Diseases" (1878) he says :

"With respect to the illustrations accompanying this work, I must here make a remark. In a former paper¹ on the examination and photographing of bacteria I expressed the wish that observers would photograph pathogenic bacteria in order that their representations of them might be as true to nature as possible. I thus felt bound to photograph the bacteria discovered in the animal tissues in traumatic infective diseases, and I have not spared trouble in the attempt. The smallest, and in fact the most interesting bacteria, however, can only be made visible in animal tissues by staining them and by thus gaining the advantage of color. But in this case the photographer has to deal with the same difficulties as are experienced in photographing colored objects—*e.g.*, colored tapestry. These have, as is well known, been overcome by the use of colored collodion. This led me to use the same method for photographing stained bacteria, and I have, in fact, succeeded, by the use of eosin-collodion, and by shutting off portions of the spectrum by colored glasses, in obtaining photographs of bacteria which had been stained with blue and red aniline dyes. Nevertheless, from the long exposure required and the unavoidable vibrations of the apparatus, the picture does not have sharpness of outline sufficient to enable it to be of use as a substitute for a drawing, or, indeed, even as evidence of what one sees. For the present, therefore, I must abstain from publishing photographic representations ; but I hope, at a subsequent period when improved methods allow a shorter exposure, to be able to remedy this defect."

¹ The paper referred to is published in Cohn's "Beiträge zur Biologie d. Pflanzen."

Since the above was written considerable progress has been made in removing the technical difficulties, and a few bacteriologists have succeeded in making very satisfactory photomicrographs. As specimens of what may be done with the best apparatus and the highest degree of skill, we may call attention to the photomicrographs in the *Atlas der Bakterienkunde* of Fränkel and Pfeiffer, and those of Roux in the *Annales of the Pasteur Institute*. The writer, also, has devoted much time to making photomicrographs which have served as illustrations for several of his published works.

Those who have had no practical experience in making photomicrographs are apt to expect too much and to underestimate the technical difficulties. Objects which under the microscope give a beautiful picture, which we desire to reproduce by photography, may be entirely unsuited for the purpose. In photographing with high powers it is necessary that the objects to be photographed be in a single plane and not crowded together or overlying each other. For this reason photographing bacteria in sections presents special difficulties, and satisfactory results can only be obtained when the sections are extremely thin and the bacteria well stained. Even with the best preparations of this kind much care must be taken in selecting a field for photography. It must be remembered that the expert microscopist, in examining a section with high powers, has his finger on the fine adjustment screw and focuses up and down to bring different planes into view. He is in the habit of fixing his attention on that part of the field which is in the focus and disregarding the rest. But in a photograph the part of the field not in focus appears in a prominent way which mars the beauty of the picture. In a cover-glass preparation made from a pure culture, when the bacteria are well distributed, this difficulty does not present itself, as the bacteria are all lying in a single plane; but the portion of the field which can be shown at one time is limited by the spherical aberration of the objective, which the makers do not seem able to overcome in high-power lenses of wide angle, at least not without loss of defining power.

Usually preparations of bacteria are stained for photography, but with some of the larger forms, such as the anthrax bacillus, very satisfactory photomicrographs may be made from unstained preparations. In this case a small quantity of a recent culture is put upon a slide, covered with a thin cover glass, and placed at once upon the stage of the microscope. The main difficulty to be encountered results from the change of location of the suspended bacteria resulting from the pressure of the objective in focussing. Motile bacteria, of course, cannot be photographed in this way without first arresting their movements by means of some germicidal agent ;

and in general it will be found more satisfactory to fix the micro-organisms to be photographed to a slide or cover glass by desiccation and heat, and to stain them with one of the aniline colors.

Objects which are opaque cannot be photographed by transmitted light, and objects which have a deep orange or red color are practically opaque for the actinic rays which are at the violet end of the spectrum. Such objects simply intercept the light, but this gives the outlines, and, where there are no details of structure, is all that is required to illustrate the form and mode of grouping. Softer and more satisfactory photomicrographs of bacteria are made when the staining is not such as to entirely arrest the actinic rays. Among the aniline colors Bismarck brown and vesuvin are the most suitable, care being taken, with the larger bacteria especially, not to make the staining too intense. Objects which are transparent for the actinic rays, or nearly so, give a very feeble photographic image, or none at all, on account of the want of contrast in the impression made upon the sensitive plate. This is the case when we attempt to photograph, by ordinary white light, objects which are stained violet or blue. But this want of contrast in the negative can be overcome by the use of specially prepared plates and colored screens of glass interposed between the object and the source of light. The so-called orthochromatic plates are more sensitive to the rays toward the red end of the spectrum than ordinary plates. They are prepared by treating the plates with a solution of eosin, of erythrosin, or of rose bengal (Vogel), and may now be purchased in this country from dealers in dry plates. If we shut off the violet rays by the use of a yellow screen, objects having a yellow or orange color may be photographed upon orthochromatic plates, although the time of exposure will be quite long owing to the comparatively feeble actinic power of the yellow rays.

We may also make photomicrographs of objects stained with methylene blue or with fuchsin, because objects stained with these colors are opaque for the rays from the red end of the spectrum, and sufficiently so with yellow light to give a good photographic contrast. Fränkel and Pfeiffer recommend the use of a green light-filter (green glass screen) for all preparations stained with methyl violet, fuchsin, or methylene blue; and for brown-stained preparations a pure blue light. The writer has been in the habit of using a yellow glass screen for fuchsin-stained preparations, and has had excellent results, but the time of exposure is necessarily long. A yellow glass screen may be prepared by dissolving tropæolin in negative varnish, and pouring this upon a clean glass slide, where it is permitted to dry.

To show bacteria in photographs in a satisfactory manner we

require an *amplification* of five hundred to one thousand diameters : and as it is often desirable to make comparisons as to the dimensions of microorganisms which resemble each other in form, it is best to adopt a standard amplification. The writer has himself adopted, and would recommend to others, a standard amplification of one thousand diameters. This is about as high a magnifying power as we can get with satisfactory definition, or as we require, and it is a convenient number when measurements are made from the photograph. The beginner, after having put his apparatus in position, should focus the lines of a stage micrometer upon the screen with the optical apparatus which he proposes to use ; then by moving the screen forward or back as required, and carefully focusing the lines, he will ascertain what is the position of the screen for exactly one thousand diameters. If the stage micrometer is ruled with lines which are one one-thousandth of an inch apart, it is evident that when projected upon the screen they should be one inch apart to make the amplification one thousand diameters. But it must be remembered that any change in the position of the optical combination will change the amplification. If, therefore, the cover correction of the objective is changed, or the position of the eyepiece—if one is used—it will be necessary to again adjust the distance of the screen.

Apparatus required.—A first-class immersion objective of one-twelfth of an inch or higher power, a substantial stand which can be placed in a horizontal position, and a camera which can be coupled with the microscope tube, are the essential pieces of apparatus. If sunlight is to be used a heliostat will also be required.

The oil-immersion objectives of any good maker may be used, but the apochromatic objectives and projection eyepieces of Carl Zeiss, of Jena, are especially to be recommended. Indeed, those who can afford it will do well to get Zeiss' complete apparatus, which includes a stand having a mechanical stage and a camera mounted upon a metal frame conveniently provided with focussing appliances, etc. However, good work may be done with less expensive apparatus.

The stand should be substantial and well made, with a delicate, fine adjustment. A mechanical stage is not essential, but is a great convenience in finding and adjusting to the centre of the screen a satisfactory field to photograph. The substage should be provided with a good apochromatic condenser, and with appliances for moving the condensing lens forward and back and for centring it, with diaphragms, etc.

By the use of a high-power objective, like the one-eighteenth-inch oil-immersion of Zeiss, the desired amplification may be obtained

without the use of an eyepiece ; and, as a rule, it is best not to use an ordinary eyepiece to secure increased amplification, as this is obtained at the expense of definition. But an amplifier may be used in the tube of the microscope, as first recommended by Woodward. In this case the amplifier must be carefully adjusted with reference to the distance of the screen, to secure the best possible definition.

The projection eyepieces of Zeiss are constructed especially for photography and possess a decided advantage. By the use of his three-millimetre apochromatic oil-immersion objective and projection eyepiece No. 3 we may obtain an amplification of one thousand diameters with excellent definition.

Light.—Sunlight is in many respects the most satisfactory for photography, but has the disadvantage that it is not always available. In some sections of the country weeks may pass without a single clear day suitable for making photomicrographs. In addition to the uncertainty arising from cloudy weather, we have to contend with the fact that the sun is only available for use with a heliostat for a limited time during each day, and that this time is greatly restricted in Northern latitudes during the winter months. When sunlight is to be employed the microscope and camera must be set up in a room having a southern exposure on a line corresponding with the true meridian of the place. The heliostat is placed outside the window in such a position that when properly adjusted the light of the sun will fall upon the condenser attached to the substage of the microscope. The condenser must be carefully centred, so that the circle of light falling upon the screen shall be uniform in intensity and outline.

The calcium, magnesium, or electric light may be used as a substitute for sunlight, but they are all rather expensive, unless, in the case of the electric light, a suitable current is available without the expense of generating it for the special purpose in view. The writer has obtained very good results with the calcium light, but has no experience in the use of the electric light. Woodward, as a result of extended experiments, arrived at the conclusion that "the electric light is by far the best of all artificial lights for the production of photomicrographs." He used a Grove battery of fifty elements to generate the current, and a Duboscq lamp. The current from a dynamo would no doubt be much cheaper and more conveniently used, if an electric-lighting plant was in the vicinity.

The apparatus shown in Fig. 73 was designed by Mr. Pringle for the use of the calcium light. It will serve to illustrate the arrangement of the microscope and camera in connection with any other light as well. An oil lamp may be placed in the position of the oxy-hydrogen burner ; or, if sunlight is to be employed, a heliostat will be placed in the same position.

When a colored screen is used this may be placed either before or behind the condensing lens—we prefer to place it behind, although

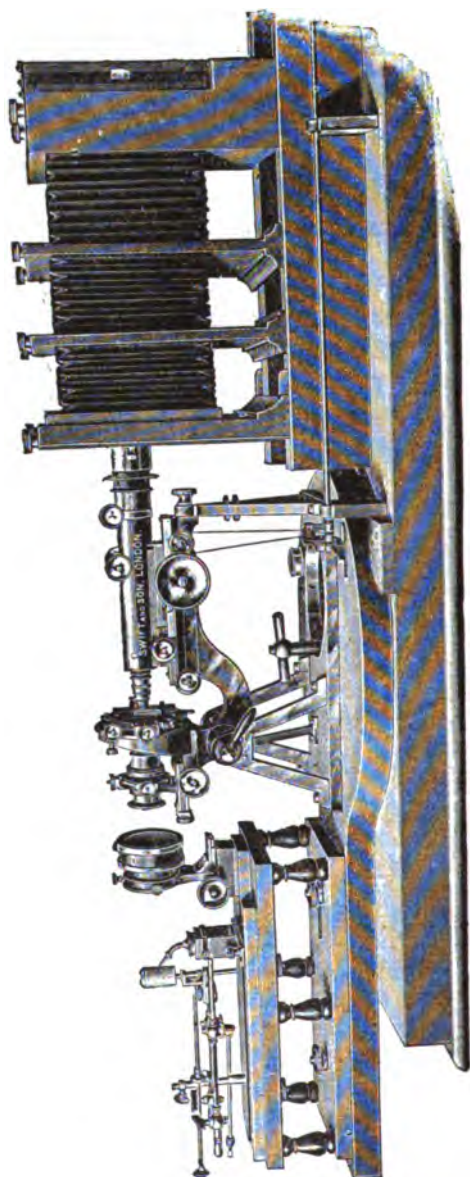


FIG. 73.

Neuhauss has shown that it makes no difference in the length of the exposure.

We cannot in the present volume give full details with reference

to the technique of making photomicrographs, but append an account of a form of apparatus which we have used with great satisfaction :

"Photomicrography by Gaslight.—Those who have had much experience in making photomicrographs will agree with me that one of the most essential elements of success is the use of a suitable source of illumination.

"Without question the direct light of the sun, reflected in a right line by the mirror of a heliostat, is the most economical and, in some respects, the most satisfactory light that can be used. But we cannot command this light at all times and places, and it often happens that, when we are ready to devote a day to making photomicrographs, the sun is obscured by clouds or the atmosphere is hazy. Indeed, in some latitudes and at certain seasons of the year a suitable day for the purpose is extremely rare. The use of sunlight also requires a room having a southern exposure and elevated above all surrounding buildings or other objects by which the direct rays of the sun would be intercepted. For these reasons a satisfactory artificial light is extremely desirable.

"The oxyhydrogen lime light, the magnesium light, and the electric arc light have all been employed as a substitute for the light of the sun, and all give satisfactory results. I have myself made rather extensive use of the 'lime light,' and think it the best substitute for solar light with which I am familiar. But to use it continuously, day after day, is attended with considerable expense, and the frequent renewal of the supply of gas which it calls for is an inconvenience which one would gladly dispense with.

"These considerations have led some microscopists to use an oil lamp as the source of illumination, and very satisfactory photomicrographs with comparatively high power have been made with this cheap and convenient light. But in my experience the best illumination which I have been able to secure with an oil lamp has called for very long exposures when working with high powers, and, as most of my photomicrographs of bacteria are made with an amplification of one thousand diameters, I require a more powerful illumination than I have been able to secure in this way. And especially so because of the fact that a colored screen must be interposed, which shuts off a large portion of the actinic rays, on account of the staining agent usually employed in making my mounts. The most satisfactory staining agents for the bacteria are an aqueous solution of fuchsin, or of methylene blue, or of gentian violet; and all of these colors are so nearly transparent for the actinic rays at the violet end of the spectrum that a satisfactory photographic contrast cannot be obtained unless we shut off these rays by a colored screen.

"I am in the habit of using a yellow screen for my preparations stained with fuchsin or methylene blue, and have obtained very satisfactory results with the orthochromatic plates manufactured by Carbutt, of Philadelphia, and a glass screen coated with a solution of tropeolin dissolved in gelatin.

"But with such a screen, which shuts off a large portion of the actinic light and increases the time of exposure three- or fourfold, the use of an oil lamp becomes impracticable with high powers, on account of the feebleness of the illumination.

"These considerations have led me to experiment with gaslight, and the simple form of apparatus which I am about to describe is the result of these experiments. I have now had the apparatus in use for several months, during which time I have made a large number of very satisfactory photomicrographs of bacteria from fuchsin-stained preparations with an amplification of one thousand diameters. My photographs have been made with the three-millimetre oil-immersion apochromatic objective of Zeiss and his projection eyepiece No. 3. I use a large Powell and Lealand stand, upon the substage of which I have fitted an Abbe condenser. The arrangement of the apparatus will be readily understood by reference to the accompanying figure.

"A is the camera, which has a pyramidal bellows front supported by the

heavy block of wood B; this can be pushed back upon the baseboard which supports it, so as to allow the operator to place his eye at the eyepiece of the microscope. When it is brought forward an aperture of the proper size admits the outer extremity of the eyepiece and shuts off all light except that coming through the objective. C is the microscope, and D the Abbe condenser, supported upon the substage. E is a thick asbestos screen for protecting the microscope from the heat given off by the battery of gas burners F. This asbestos screen has an aperture of proper dimensions to admit the light to the condenser D. The gas burners are arranged in a series, with the flat portion of the flame facing the aperture in the asbestos screen E. The concave metallic mirror G is properly placed to reflect the light in the desired direction. I have not found any advantage in the use of a condensing lens other than the Abbe condenser upon the substage of the microscope. The focussing is accomplished by means of the rod I, which carries at one extremity a grooved wheel, H, which is connected with the fine adjustment screw of the microscope by means of a cord.

"The focussing wheel J may be slipped along the rod I to any desired position, and is retained in place by a set screw. The rod I is supported

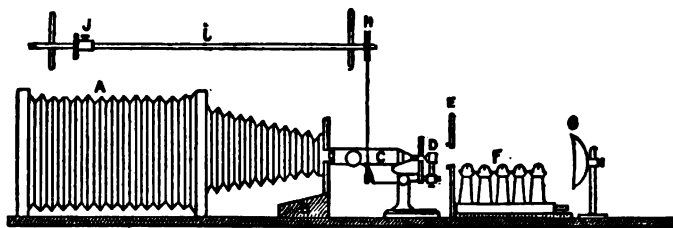


FIG. 74.

above the camera by arms depending from the ceiling, or by upright arms attached to the baseboard.

"I have lost many plates from a derangement of the focal adjustment resulting from vibrations caused by the passing of loaded wagons in the street adjoining the laboratory in which I work. This has been overcome to a great degree by placing soft rubber cushions under the whole apparatus."¹

Students who desire to perfect themselves in the art of making photomicrographs are advised to first make themselves familiar with the technique of photography with a landscape or portrait camera, and not to undertake the more difficult task of photographing bacteria until they know how to make a good negative and to judge whether an exposure has been too long or too short, etc.

¹ From Johns Hopkins University Circulars, vol. ix., No. 81, p. 72.

PLATE I.
STERNBERG'S BACTERIOLOGY.

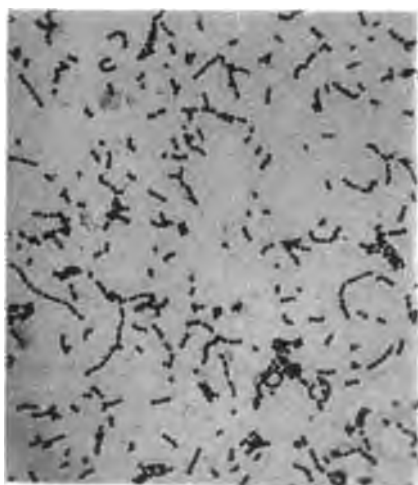


Fig. 1.



Fig. 2.

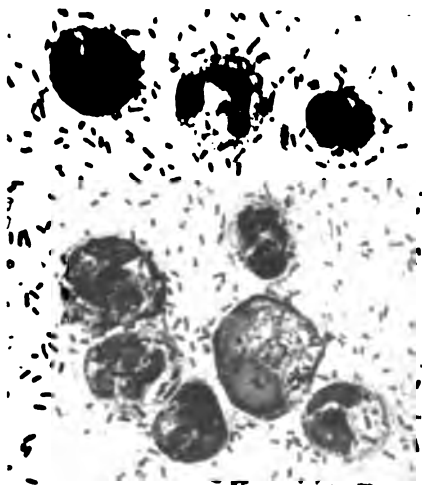


Fig. 3.

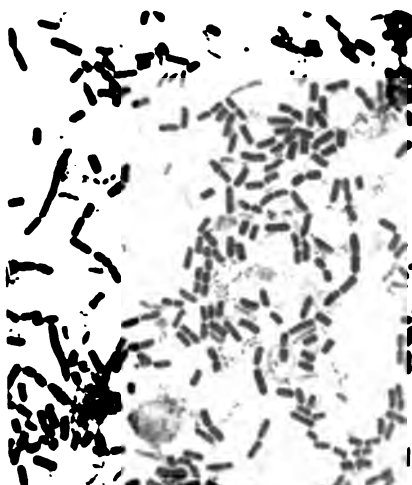


Fig. 4.

PHOTOMICROGRAPHS BY GAS LIGHT.

PLATE II.
STERNBERG'S BACTERIOLOGY



Fig. 1.



Fig. 2.



Fig. 3.



Fig. 4.



Fig. 5.



Fig. 6.



Fig. 7.



Fig. 8.

PLATE I.

PHOTOMICROGRAPHS OF BACTERIA MADE BY GASLIGHT.

FIG. 1.—*Streptococcus cadaveris*, from a culture in *agua coco*; stained with fuchsin. $\times 1,000$. (Sternberg.)

FIG. 2.—*Streptococcus Havaniensis*. $\times 1,000$. From a photomicrograph. (Sternberg.)

FIG. 3.—*Bacillus cuniculicida Havaniensis*, from peritoneal cavity of inoculated rabbit, showing leucocytes containing bacilli and free bacilli; stained with fuchsin. $\times 1,000$. (Sternberg.)

FIG. 4.—*Bacillus cadaveris*, smear preparation from yellow-fever liver kept for forty-eight hours in an antiseptic wrapping (Havana, 1889); stained with fuchsin. $\times 1,000$. (Sternberg.)

Note.—All of the above photomicrographs were made with the three-millimetre apochromatic hom. ol. im. objective and projection eye-piece of Zeiss.

PLATE II.

PHOTOGRAPHS OF COLONIES (IN ESMARCH ROLL TUBES) AND OF TEST-TUBE CULTURES.

FIG. 1.—Colonies of *Bacillus leporis lethalis*, in gelatin roll tube, end of forty-eight hours at room temperature. $\times 5$. (Sternberg.)

FIG. 2.—Colonies of *Bacillus coli similis* in gelatin roll tube, end of twenty-four hours at 22° C. $\times 10$. (Sternberg.)

FIG. 3.—Stick culture of *Bacillus coli similis* in nutrient gelatin, end of seven days at 20° C. (Sternberg.)

FIG. 4.—Stick culture of *Bacillus intestinus motilis* in nutrient gelatin, end of four days at 22° C. (Sternberg.)

FIG. 5.—Stick culture of *Bacillus leporis lethalis* in nutrient gelatin, end of eight days at 22° C. (Sternberg.)

FIG. 6.—Stick culture of *Micrococcus tetragenus versatilis* in nutrient gelatin, end of two weeks at 22° C. (Sternberg.)

FIG. 7.—Colonies of *Bacillus cuniculicida Havaniensis* in gelatin roll tube, end of forty-eight hours at 21° C. $\times 10$. (Sternberg.)

FIG. 8.—Colonies of *Bacillus coli communis* in gelatin roll tube, end of forty-eight hours at 22° C. $\times 10$. (Sternberg.)

PART SECOND.

GENERAL BIOLOGICAL CHARACTERS :

INCLUDING AN ACCOUNT OF THE ACTION OF ANTISEPTICS
AND GERMICIDES.

- I. STRUCTURE, MOTIONS, REPRODUCTION. II. CONDITIONS OF GROWTH.
III. MODIFICATIONS OF BIOLOGICAL CHARACTERS. IV. PRODUCTS OF
VITAL ACTIVITY. V. PTOMAINES AND TOXALBUMINS. VI. INFLUENCE
OF PHYSICAL AGENTS. VII. ANTISEPTICS AND DISINFECTANTS
—GENERAL ACCOUNT OF THE ACTION OF. VIII. ACTION OF
GASES AND OF THE HALOID ELEMENTS UPON BACTERIA.
IX. ACTION OF ACIDS AND ALKALIES. X. ACTION OF
VARIOUS SALTS. XI. ACTION OF COAL-TAR PRO-
DUCTS, ESSENTIAL OILS, ETC. XII. AC-
TION OF BLOOD SERUM AND OTHER OR-
GANIC LIQUIDS. XIII. PRACTICAL
DIRECTIONS FOR DISINFECTION.

PART SECOND.

I.

STRUCTURE, MOTIONS, REPRODUCTION.

THE bacteria are unicellular vegetable organisms, and consist of a *cell membrane* enclosing transparent and apparently structureless *protoplasm*. The very varied biological characters which distinguish different species make it evident, however, that there are essential differences in the living cell contents, although these differences are not revealed by our optical appliances. And among the bacteria, as in the cells of higher plants and animals, the peculiar biological characters of a species are transmitted to the cellular progeny of each individual cell. These characters are, however, subject to various modifications as a result of differing conditions of environment, as is the case with plants and animals higher in the scale of existence, and in this way more or less permanent varieties are produced. It is probable that among these lowly plants species are evolved more quickly, as a result of the laws of natural selection, in the struggle for existence, than among those of more complex organization. Still, this has not been proved, and, on the other hand, we have ample evidence that widely distributed species exist having very definite morphological and biological characters which enable us to recognize them wherever found.

It has generally been supposed that these simple vegetable cells are destitute of a nucleus, but a recent author (Fränkel) suggests the probability that a nucleus may exist, although it has not been demonstrated. This suggestion is based upon the fact that in staining bacteria very quickly it sometimes happens that a portion of the protoplasm is sharply differentiated by taking the stain more deeply than the remaining portion.

Sjöbring has recently (1892) made an investigation for the purpose of ascertaining the structure of bacterial cells. Various methods were employed, but the most satisfactory results were obtained by fixing with nitric acid, with or without alcohol, and without pre-

vious drying ; the preparations were then stained with carbol-methylene-blue or carbol-fuchsin solution ; they were decolorized with nitric acid and examined in glycerin or in water. By this procedure the author named was able to demonstrate two kinds of corpuscles. One of these may be seen just inside the cell wall ; it stains deeply with the carbol-fuchsin solution. The other lies in a position analogous to that occupied by the nucleus of vegetable cells higher in the scale, and resembles this both in its resting condition and in the process of indirect division.

In his address before the International Medical Congress of Berlin (1890) Koch says :

“ We had not succeeded, in spite of the constantly improving methods of staining and in spite of the use of objectives with constantly increasing angles of aperture, in learning more with reference to the interior structure of the bacteria than was shown by the original methods of staining. Only very recently new methods of staining appear to give us further information upon the structure of the bacteria, inasmuch as they serve to differentiate an interior portion of the protoplasm, which should probably be regarded as a nucleus, from an exterior protoplasmic envelope from which is given off the organ of locomotion, the flagellum.”

Although usually transparent, the protoplasm sometimes presents a granular appearance. The botanist Van Tieghem claims to have found chlorophyll grains in some water bacteria studied by him, and in the genus *Beggiatoa* grains of sulphur are found embedded in the protoplasm of certain species.

The cell membrane in certain species appears to be very flexible, as may be seen in those which have a sinuous motion. It is not easily recognized under the microscope, but by the use of reagents which cause the protoplasm to contract may be demonstrated—*e.g.*, by iodine solution. Outside of the true cell membrane a gelatinous envelope—so-called capsule—is sometimes seen. This may perhaps be, as claimed by some authors, nothing more than a jelly-like thickening of the outer layers of the cell wall. This jelly-like material causes the cells to adhere to each other, forming *zoöglæa* masses. In some cases the growth upon the surface of a culture medium is extremely viscid, and may be drawn out into long threads when touched with a platinum needle, owing to the gelatinous intercellular substance by which the cells are surrounded.

There is but little more to be said of the structure of these minute organisms, except to mention the fact that the motile species are provided with slender, whip-like appendages called *flagella*. The micrococci in general are not endowed with the power of executing spontaneous movements, and they are not provided with flagella.

But recently two motile species have been described, and in one of these—*Micrococcus agilis* of Ali-Cohen—the presence of flagella has been demonstrated.

Many of the bacilli and spirilla are actively motile, and the presence of flagella, which has long been suspected, has recently been demonstrated for a considerable number of species by Löffler and others.

It must be remembered that the molecular movement which is common to all minute particles suspended in a fluid is a vibratory motion *in situ*, which does not change the relative position of the moving particles. This so-called Brownian movement has frequently been mistaken for a vital motion, as has also the movement due to currents in the liquid in which non-motile organisms are suspended. The latter is to be distinguished by the fact that the microorganisms are all carried in one direction. This movement due to a current, in connection with the vibratory Brownian movement, is very deceptive, and it is often hard for a beginner in bacteriological study to convince himself that what he sees is not a vital movement. But in true vital movements we have progression in different directions, and the individual microorganisms approach and pass each other, often in a most vigorous and active manner, passing entirely across the field of view or changing direction in an abrupt way. Sometimes the motion is slow and deliberate, the bacillus progressing with a to-and-fro motion, as if propelled by a trailing flagellum; or it may be serpentine when the moving filament is flexible; or again it is a darting forward motion which is so rapid that the eye can scarcely follow the moving body. The spirilla have a rotary movement as well as a progressive one, and this is often extremely rapid. Sometimes bacilli spin around with a rotatory motion, as if they were anchored fast to a fixed point, as they may be by the flagellum being attached to the slide or cover glass. Frequently, in a pure culture, the individual bacilli may be seen to come to rest, and, after an interval of repose, to dart forward again in the most active way. Or we may find, on examining the same culture at different times, that upon one occasion there is no evidence of vital movements, and on another all of the bacilli are actively motile. These differences depend upon the age of the culture, temperature conditions, etc.

Reproduction by binary division is common to all of the bacteria, and in many species this is the only mode of reproduction known. When circumstances are favorable for rapid multiplication the individual cells grow in length, and a constriction occurs in the middle transverse to the long diameter. This becomes deeper, and after a time the cell is completely divided into two equal portions, which again divide in the same way. Separation may be complete, or the

cells may remain attached to each other, forming chains (streptococci) or articulated filaments (*scheinfäden* of the Germans).

The bacilli and spirilla divide only in a direction transverse to the long diameter of the cells, but among the micrococci division may occur either in one direction, forming chains; or in two directions, forming tetrads; or in three directions, forming "packets" of eight or more elements. The staphylococci, in which the cells do not remain associated, divide indifferently in any direction.

The rapidity of multiplication by binary division varies greatly in different species, and in the same species depends upon conditions relating to the culture medium, age of the culture, temperature, etc. Under favorable conditions bacilli have been observed to divide in twenty minutes, and it is a matter of common laboratory experience that colonies of considerable size and containing millions of bacilli may be developed from a single cell in twenty-four to forty-eight hours. A simple calculation will show what an immense number of cells may be produced in this time as a result of binary division occurring, for example, every hour. The progeny of a single cell would be at the end of twenty-four hours 16,777,220, and at the end of forty-eight hours the number would be 281,500,000,000.

Some of the earlier observers have noted the presence of oval or spherical refractive bodies in cultures containing bacilli; but that these were reproductive elements, although suspected, was not demonstrated until a comparatively recent date. Pasteur was one of the first to point out the fact that certain bacteria have two modes of reproduction—by fission and by the formation of *endogenous spores*; but the first careful study of the last-mentioned method was made by Koch in his classical study of the anthrax bacillus (1878), and by Cohn, who studied the formation of spores in *Bacillus subtilis*.

These reproductive bodies serve the same purpose in the preservation of species as the seeds of higher plants. They resist desiccation and may retain their vitality for months or years until circumstances are favorable to their development, when, under the influence of heat and moisture, they reproduce the vegetative form—bacillus or spirillum—with all of its biological and morphological characters. They are composed of condensed protoplasm which retains the vital characters of the soft protoplasm of the mother cell from which it has been separated; and it is evident that whether reproduction occurs by fission or by the formation of endogenous spores, the protoplasm of the cells in a pure culture of any microorganism is simply a separated portion of the protoplasm of the progenitors of these cells.

Some of the bacilli grow out into long filaments before the formation of spores occurs; and these filaments may be associated in bundles or intertwined in irregular masses. At first the protoplasm of the

filaments is homogeneous, but after a time it becomes segmented, and later the protoplasm of each segment becomes condensed into a spherical or oval refractive body, which is the spore. For a time these are retained in a linear position by the cell membrane of the filament (Fig. 75, *a*), but this is after a while dissolved or broken up and the spores are set free. In liquid cultures they sink to the bottom as a pulverulent precipitate, and upon the surface of a solid medium they form a layer which is usually of a white or yellowish-white color, and which, when examined under the microscope, in old cultures is found to consist almost entirely of shining spherical or oval bodies which do not stain, by the ordinary methods, with the aniline colors. While many of the bacilli during the stage of spore formation grow out into long filaments, others do not, and one or more spores make their appearance in rods of the ordinary length which characterizes the species. These may be located in the centre of the rod or at one extremity (Fig. 75, *b*). It sometimes occurs

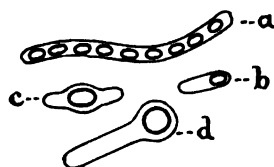


FIG. 75.

that when a single central spore is formed the rod becomes very much enlarged in its central portion, assuming a spindle shape (Fig. 75, *c*); or one extremity may be enlarged, producing forms such as are shown in Fig. 75, *d*. Some of the smaller spherical spores measure less than $0.5\ \mu$ in diameter, but they are, for the most part, oval bodies having a short diameter of 0.5 to $1\ \mu$ and a long diameter of one to two μ , or even more. They are enveloped in a cellular envelope which, according to some observers, consists of two layers—an exosporium and an endosporium.

The germination of spores has been studied by Prazmowski, Brefeld, and others. The process is as follows: By the absorption of water they become swollen and pale, losing their shining, refractive appearance. Later a little protuberance is seen upon one side or at one extremity of the spore, and this rapidly grows out to form a rod which consists of soft-growing protoplasm enveloped in a membrane which is formed of the endosporium or inner layer of the cellular envelope of the spore. The outer envelope, or exosporium, is cast off and may be seen in the vicinity of the newly formed rod (Fig. 76). Sometimes the vegetative cell emerges from one extrem-

ity of the oval spore, as shown at *a*, Fig. 76, and in other species the exosporium is ruptured and the bacillus emerges from the side, as seen at *b*.

The considerable resistance of these endogenous spores to desiccation, to heat, and to various chemical agents is an important fact both from a biological and from a hygienic point of view, and will be fully considered in a subsequent chapter. The fact that certain bacilli and spirilla do not withstand a temperature of 80° to 90° C., which does not destroy the vitality of known spores, leads to the inference that they do not form similar reproductive bodies. But reproductive elements of a different kind are described by some botanists as being produced during the development of these bacteria, and also of the micrococci. These are the so-called *arthrospores*. In the process of binary division certain cells in a chain may be observed to be somewhat larger than others and to refract light more strongly. The same may be true of certain cells in a culture in which the elements are not united in chains. These cells are believed



FIG. 76.

by De Bary and others to have greater resisting power to desiccation than the remaining cells in the culture, and to serve the purpose of reproductive elements.

It has generally been supposed that spore formation is most likely to occur when the pabulum for supporting the growth of the vegetative form is nearly exhausted. But, as pointed out by Fränkel, facts do not support this view, as many species form spores when conditions are most favorable for a continued development. An abundant supply of oxygen favors the formation of spores in aerobic species, and, in some instances at least, the temperature has an important influence upon spore formation. Thus the anthrax bacillus does not form spores at temperatures below 20° C.

The very interesting fact has been demonstrated by Lehman and by Behring that a species which usually forms spores may be so modified by certain influences that it is no longer capable of spore production, and that such an asporogenous variety may be cultivated for an indefinite time without showing any return to the stage of spore formation. This was effected in Behring's experiments by cultivating the anthrax bacillus in a medium containing some agent detrimental to the vitality of the vegetative cells, but not in sufficient quantity to restrain their development.

The *chemical composition* of the bacterial cells has been investigated by Nencki, Brieger, and others. Putrefactive bacteria cultivated in a two-per-cent solution of gelatin, and which produced an abundant intercellular substance connecting the cells in zoöglœa masses, were found by Nencki to have the following composition : Water, 84.26 per cent ; solids, 5.74 per cent, consisting of albumin 87.46 per cent, fat 6.41, ash 3.04, undetermined remnant 3.09. The albuminous substance, according to Nencki, is not precipitated by alcohol, and differs in its chemical composition from other known substances of this class. He calls it *mykoprotein* and gives the following as its chemical composition : C, 52.32 per cent ; H, 7.55 per cent ; N, 14.75 per cent. It contains no sulphur and no phosphorus. The spores of the anthrax bacillus, according to Nencki, do not contain mykoprotein, but a peculiar albuminous substance which he calls anthrax-protein. Brieger analyzed a gelatin culture of Friedländer's bacillus, with the following result : Water, 84.2 per cent ; solids, 5.8 per cent, containing 1.74 per cent of fats. After removal of the fat the solids gave an ash of 30.13 per cent ; this contains calcium phosphate, magnesium phosphate, sodium sulphate, and sodium chloride. The amount of nitrogen in the dried substance after removal of the fat was 9.75.

II.

CONDITIONS OF GROWTH.

BACTERIA only grow in presence of moisture, under certain conditions of temperature, and when supplied with suitable pabulum. As they do not contain chlorophyll, they cannot assimilate carbon dioxide, and light is not favorable to their development.

The aërobic species obtain *oxygen* from the air and cannot grow unless supplied with it. The anaërobic species, on the other hand, will not grow in the presence of oxygen, and must obtain this element, as they do carbon and nitrogen, from the organic material which serves them as food.

As a class the bacteria are supplied with nutriment by the higher plants and animals, the dead tissues of which they appropriate, and which it is their function to decompose, releasing the organic elements as simple compounds which may again be assimilated by the chlorophyll-producing plants.

Water is essential for the development of bacteria, and many species have their normal habitat in the waters of the ocean, of lakes, and of running streams; others thrive upon damp surfaces or in the interior of moist masses of organic material. Many species grow indifferently either in salt or fresh water, but it is probable that certain species will be found peculiar to the waters of the ocean. Some of the water bacteria multiply in the presence of an exceedingly minute amount of organic pabulum, or even in distilled water. This is shown by the experiments of Bolton and others. The author named tested two species of water bacteria (*Micrococcus aquatilis* and *Bacillus erythrosporus*) in the following manner: Ten cubic centimetres of distilled water in a test tube were infected with a small quantity of a culture of one of these microorganisms. A drop from this tube was transferred to the same quantity of distilled water in a second tube, and from this to a third. The number of bacteria in this tube No. 3 was now ascertained by counting, and it was put aside for two or three days, at the end of which time the number was again estimated by counting. In every case there was an enormous increase in the number of bacteria. In order to be sure that the dis-

tilled water was pure, it was distilled a second time in a clean glass retort, but the result was the same. Bolton remarks, with reference to these results: "If we seek to explain this remarkable fact we must remember, in the first place, what an extremely small absolute mass is represented by an enormous number of bacteria, and what a minute amount of material is required for the formation of this mass. In ten cubic centimetres of distilled water, in the experiment last referred to, there were about twenty million bacteria (two million per cubic centimetre). If we estimate the diameter of each at one μ , with a specific weight of 1, the absolute weight would be for the entire number one-one-hundredth of a milligramme—that is to say, a quantity which cannot be determined by any of our methods of weighing."

Bolton supposes that the small amount of organic pabulum required fell into the water in the shape of dust, or was attached to the walls of the test tube in spite of all the precautions taken.

Nitrogen is chiefly obtained from albuminoid substances, but Pasteur has shown that it may also be obtained from ammonia. This is shown by cultivating bacteria in a medium containing an ammonia salt, as in the following :

PASTEUR'S SOLUTION.

Distilled water,	100
Cane sugar,	10
Tartrate of ammonia,	1
Ashes of one gramme of yeast,	0.075

COHN'S SOLUTION.

Distilled water,	100
Tartrate of ammonia,	1
Ashes of yeast,	1

Many bacteria multiply abundantly in these solutions.

Carbon is obtained from the various organic substances containing it; among others, from starch, sugars, glycerin, organic acids and their salts, etc.

Temperature.—There are certain limits of temperature within which development may take place, but these differ greatly with different species. As a rule, growth is arrested when the temperature falls below 10° C. (50° F.), but some species multiply at a still lower temperature. Thus Bolton observed a very decided increase in certain water bacteria kept in an ice chest at 6° C., and other observers have witnessed development at the freezing temperature.

Most saprophytic bacteria grow within rather wide temperature limits, but the rapidity of development is greatest at a certain favorable temperature, which is usually between 25° and 30° C. The

parasitic species have a more restricted range, which approaches the normal temperature of the animals in which they habitually develop. At 40° C. (104° F.) growth, as a rule, ceases, but there are some notable exceptions to this rule.

Miquel some years ago found a bacillus in the water of the Seine which grew at a temperature of 69° to 70° C.; Van Tieghem reports having observed species in thermal waters capable of growth at a still higher temperature (74° C.); and Globig has more recently obtained from garden earth several species which multiplied at 65° C. Some of the species found by the last-named observer were even found to require a temperature of about 60° for their development; and yet this temperature is quickly fatal to a large number of the best known species.

Low temperatures, while arresting the growth of bacteria, do not destroy their vitality. This has been demonstrated by numerous experiments, in which they have been exposed for hours in a refrigerating mixture at -18° C. Frisch has even subjected them to a temperature of -87° C. by the evaporation of liquid carbon dioxide, and found that they still grew when placed in favorable conditions.

Parasitism.—The strict parasites grow only in the bodies of living animals, or in artificial media kept at a suitable temperature. As examples we may mention the bacillus of tuberculosis, the bacillus of leprosy, the micrococcus of gonorrhoea, the spirillum of relapsing fever. There is also a large class of *facultative parasites* which, when introduced into the body of a susceptible animal, multiply in it, and may continue to live as parasites so long as they are transferred from one animal to another, but which are also able to live as saprophytes independently of a living host. To this class belong the pus cocci, the bacillus of typhoid fever, the spirillum of cholera, and many others.

It seems extremely probable that the strict parasites were at one time capable of living a saprophytic existence, and that their restriction to a parasitic mode of life has been effected in course of time in accordance with the laws of natural selection. This view is supported by the fact that the tubercle bacillus, which has been regarded as a strict parasite, which can only be cultivated artificially under very special conditions, has been shown to be capable of modification in this regard to such an extent that when cultivated for a time in a favorable medium—bouillon with five per cent of glycerin—it will even grow in ordinary bouillon made from the flesh of a calf or a fowl (Roux).

Reaction of Medium.—Some bacteria grow readily in a medium having an acid reaction, while the slightest trace of acidity prevents the development of others. As a rule, the pathogenic species require a neutral or slightly alkaline culture medium.

While many species grow in various media and under various conditions of temperature, etc., others are greatly restricted in this regard; thus Bumm only succeeded in cultivating the gonococcus upon *human blood serum*, and even upon this was not able to carry it through a series of successive cultures. It is very probable that certain species can only grow in association with others which elaborate products necessary for their development.

Substances favorable for the growth of a particular species may restrain its development if present in too large an amount. Thus the phosphorescent bacilli multiply abundantly in a nutrient solution containing 2.5 per cent of sodium chloride; but this amount would restrain the development of some other species, and a considerable increase in the quantity of salt prevents the growth of all microorganisms. In the same way the addition of two per cent of glucose to culture solutions is favorable for the development of certain species, and especially for the anaërobic bacteria; but a concentrated solution of the same substance prevents the growth of all bacteria.

The influence of one species upon the growth of another has been studied by various bacteriologists, and especially by Sirotinin and by Freudenreich. When several species are associated in the same culture one may take the precedence and the others may develop later; or two or more species may develop at the same time; or the growth of one species may prevent the development of another, either (a) by exhausting the pabulum necessary for its growth or (b) by producing substances which inhibit the development of another species or destroy its vitality.

Freudenreich found, as a result of his numerous experiments, that the following species cause a change in bouillon which renders it unfit for the growth of other species: *Bacillus pyocyaneus*, *Bacillus cyanogenus*, *Bacterium phosphorescens*, *Bacillus prodigiosus*, *Spirillum cholerae Asiaticæ*. The following species do not cause such a change in bouillon as to render it unfit for the growth of other species: *Bacillus typhi abdominalis*, *Bacillus anthracis*, *Bacillus septicæmiæ hæmorrhagicæ*, *Spirillum tyrogenum*. The following have a decided antagonism: *Bacillus pyogenes fœtidus* prevents the growth of *Spirillum cholerae Asiaticæ*; *Micrococcus roseus* prevents the growth of *Micrococcus tetragenus*. The cholera spirillum will not grow in sterilized cultures of *Bacillus pyocyaneus*, or in bouillon which has served for a previous culture of the same microorganism (Kitasato). Other bacteria which fail to grow in bouillon which has already served for the cultivation of the same species are *Bacillus typhi abdominalis*, *Bacillus cyanogenus*, *Bacillus prodigiosus*, *Micrococcus roseus*, etc. (Freudenreich).

III.

MODIFICATIONS OF BIOLOGICAL CHARACTERS.

WE have already referred to the production of an asporogenous variety of the anthrax bacillus. This was effected by Behring by cultivation in media containing small amounts of hydrochloric acid, caustic soda, methyl violet, malachite green, and various other agents. This is only one of many instances of a change in biological characters due to changed conditions of environment. We have abundant experimental evidence that growth may occur under adverse conditions when the species is gradually habituated to these conditions. Thus the temperature limitations may be passed by successive cultivations at temperatures approaching these limits, and bacteria may grow in the presence of agents which in a given proportion have a complete restraining influence upon their development. For example, in the experiments of Kossiakoff, published in the *Annales* of the Pasteur Institute (vol. i.), it was found that the several species tested all became habituated to the presence of antiseptic agents in proportions which at first completely restrained their growth.

This modification of biological characters is well shown in the case of the chromogenic bacteria, some of which only form pigment under exceptionally favorable conditions of growth. It has been shown by several observers that non-chromogenic varieties of some of the best known chromogenic species may be produced by special methods of cultivation. Thus Wasserzug obtained a non-chromogenic variety of the bacillus of green pus (*Bacillus pyocyaneus*) by the action of time added to that of antiseptics. He says: "These two actions combined have permitted me to obtain cultures which remained without color in a durable way, and in which, consequently, the chromogenic function was abolished by heredity." In the case of a chromogenic bacillus obtained by the writer in Havana (my *Bacillus Havaniensis*), a non-chromogenic variety was obtained from a culture on nutrient agar which had been kept in a hermetically sealed glass tube for about a year. The variety preserved the morphological characters of the original stock, but, al-

though carried through successive cultures for a considerable period, did not regain its power to produce the brilliant carmine color which is the most striking character of the species. Katz, in cultivating the phosphorescent bacilli isolated by him from sea water at New South Wales, found that, after being propagated for some time in artificial media, their power to give off a phosphorescent light was diminished or temporarily lost. He also found that two species which when first cultivated did not liquefy gelatin, subsequently, after a year, caused liquefaction of the usual gelatin medium.

Modification shown in Cultures.—When bacteria have been subjected to the action of heat or chemical agents, without having their vitality completely destroyed, they often show diminished vigor of growth. Cultures which would ordinarily show an abundant development within twenty-four hours may not commence to grow for several days. For this reason, in disinfection experiments, it is necessary to test the question of destruction of vitality by leaving the cultures for a week or more under favorable conditions as to temperature. In plate cultures or Esmarch roll tubes a few colonies may develop in this tardy way, showing that there was a difference in the vital resisting power of the individual cells, some having survived while the majority were killed. This is well illustrated by Abbott's experiments upon the germicidal action of mercuric chloride as tested upon *Staphylococcus pyogenes aureus*. Irregularities in the results in experiments in which the conditions were identical having been noticed, Abbott inferred that this was due to a difference in the resisting power of individual cocci (arthrospores?). By making cultures from colonies which developed from these more resistant cocci, and again exposing the micrococci in these cultures to mercuric chloride in the proportion of 1:1,000 for a longer time and making new cultures from the surviving cocci, and so on, Abbott obtained cultures in which a majority of the cells survived exposure to a solution of the strength mentioned for ten to twenty minutes, whereas in his original culture most of the cocci were killed by this solution in five minutes.

These changes in vital resisting power enable us to comprehend other modifications which can only be detected by chemical or biological reactions. Thus the reducing power for various substances may be modified by changes in the conditions of environment. And among the pathogenic bacteria changes of a more or less permanent nature may be induced, which are shown by a modified degree of virulence when injected into susceptible animals.

Attenuation of Virulence may be effected by several methods, all of which depend upon subjecting the cultures to prejudicial influences of one kind or another.

Pasteur first announced, in 1880, that the microbe of fowl cholera

could be modified by special treatment in such a manner that it no longer produced a fatal form of the disease. He found that the virulence was greatest when cultures were made from fowls which had died from a chronic form of the disease, and that this virulence was not lost by successive cultivations in chicken bouillon, repeated at short intervals. But when an interval of more than two months was allowed to elapse without renewing the cultures, the virulence was diminished and fewer deaths occurred in fowls inoculated with such cultures. This diminution of virulence became more marked in proportion to the length of time during which a culture solution containing the microbe remained exposed to the action of the atmosphere, and at last all virulence was lost as a result of the death of the pathogenic microorganism. When the virus was preserved in hermetically sealed tubes it did not undergo this modification, but retained its full virulence for many months. According to Pasteur, the various degrees of modification of virulence resulting from prolonged exposure to the air may be preserved in successive cultures made at short intervals. Subsequent experiments with cultures of the anthrax bacillus gave similar results and enabled him to produce an "attenuated virus" for his protective inoculations.

In the case of the anthrax bacillus it was found that the spores retain their full virulence for years, and that the production of an attenuated virus required the exclusion of these reproductive elements. Cultivations were consequently made at a temperature of 42° to 43° C., at which point this bacillus is incapable of producing spores. Cultivation at this temperature for eight days gave an attenuated virus suitable for use in protective inoculations.

Attenuation by Heat.—Toussaint has shown that a similar modification of virulence may be produced by exposure for a short time to a temperature a little below that which destroys the vitality of the pathogenic organism. This is best accomplished, according to Chauveau, in the case of the bacillus of anthrax, by exposure for eighteen minutes to a temperature of 50° C. Exposure to this temperature for twenty minutes is said to completely destroy the vitality of the bacillus.

Attenuation by Antiseptic Agents.—The writer, in 1880, obtained evidence that attenuation of virulence may result from exposure to the action of antiseptic agents. In a series of experiments made to determine the comparative value of disinfectants, the blood of a rabbit recently dead from a form of septicæmia induced by the subcutaneous injection of my own saliva, and due to the presence of a micrococcus (*Micrococcus pneumoniae crouposæ*), was subjected to the action of various chemical agents, and subsequently injected into a rabbit to test the destruction of virulence. In the published report of these experiments the following statement is made :

"The most important source of error, however, and one which must be kept in view in future experiments, is the fact that a protective influence has been shown to result from the injection of virus the virulence of which has been modified, without being entirely destroyed, by the agent used as a disinfectant."

"Sodium hyposulphite and alcohol were the chemical reagents which produced the result noted in these experiments; but it seems probable that a variety of antiseptic substances will be found to be equally effective when used in proper proportion. Subsequent experiments have shown that neither of these agents is capable of destroying the vitality of the septic micrococcus in the proportion used (one per cent of sodium hyposulphite or one part of ninety-five-per-cent alcohol to three parts of virus), and that both have a restraining influence upon the development of this organism in culture fluids."¹

Cultivation in the Blood of an Immune Animal.—It has been recently shown by the experiments of Ogata and Jasuhara that when the anthrax bacillus is cultivated in the blood of an immune animal, such as the dog or the white rat, its pathogenic power is modified so that it no longer kills susceptible animals and may be used as a vaccine.

Pasteur had previously shown (1882) that the virus of rouget can be attenuated by passing it through rabbits.

Recovery of Virulence.—Pasteur has shown that when the virulence of a pathogenic organism has been modified it may be restored by successive inoculations into susceptible animals. Thus in the case of the anthrax bacillus a culture which would not kill an adult guinea-pig may be inoculated into a very young animal of the same species with a fatal result; and by inoculating the blood of this animal into another, and so on, the original virulence may be restored, so that a culture is obtained which will kill a sheep. In the same way the attenuated virus of fowl cholera may be restored to full vigor by inoculating a small bird—sparrow or canary—to which it is fatal. After several successive inoculations the virus resumes its original activity.

In general, pathogenic virulence is increased by successive inoculations into susceptible animals, and diminished by cultivation in artificial media under unfavorable conditions. Thus various pathogenic bacteria which have been cultivated in laboratories for a length of time are likely to disappoint the student if he makes inoculation experiments for the purpose of demonstrating their specific action as described in text books.

¹Quoted from "Bacteria," pages 207, 208, written in 1883.

IV.

PRODUCTS OF VITAL ACTIVITY.

ALL living cells, animal or vegetable, while in active growth, appropriate certain elements for their nutrition from the pabulum with which they are supplied, and at the same time excrete certain products which, in some cases at least, it is their special function to produce. In the higher plants and animals specialized cells excrete substances which are injurious to the economy of the individual, and secrete substances which are required to maintain its existence. As an example in animals we may mention the excretion of urea by the epithelium of the kidneys, the retention of which is fatal to the individual, and the gastric secretion which is essential for its continued existence. Among the higher plants we have an immense variety of substances formed in the cell laboratories, some of which are evidently useful for the preservation of the species, while others are perhaps to be considered simply as excretory products. The odorous volatile products given off by flowers are supposed to be useful to the plant in attracting insects by which cross-fertilization is effected. The various poisonous substances stored up in leaves and bark may serve to protect the plant from enemies, etc.

The minute plants with which we are especially concerned also produce a great variety of substances, some of which may be useful to the species in the struggle for existence. Thus the deadly ptomaines produced by some of the pathogenic bacteria serve to paralyze the vital resisting power of living animals and enable the parasitic invader to thrive at the expense of its host. In the present section we shall consider in a general way these various products of bacterial growth.

Pigment Production.—A considerable number of species are distinguished by the formation of pigment of various colors and shades. We have all of the shades of the spectrum from violet to red. The color, as a rule, is only produced in the presence of oxygen, and when the pigment-producing microorganisms are massed upon the surface of a solid culture medium the pigment production is often limited to the superficial portion of the mass. In some cases a soluble pigment is formed which is absorbed by the transpa-



Fig. 1.



Fig. 2.



Fig. 3.



Fig. 4.

Fig. 1. *Sarcina lutea*, agar culture.
 Fig. 2. *Bacillus prodigiosus*, agar culture.
 Fig. 3. *Bacillus pyocyaneus*, agar culture.
 Fig. 4. *Bacillus Havaniensis*, potato culture.

rent culture medium, coloring especially the upper portion, in stick cultures in nutrient gelatin or agar. This is the case with *Bacillus pyocyaneus*, which produces a blue pigment which has been isolated and carefully studied by Gessard and others. The pigment, which is called pyocyanin, is soluble in chloroform and crystallizes from a pure solution in long blue needles. Acids change the blue color to red, reducing substances to yellow. It resembles the ptomaines in its chemical reactions, being precipitated by platinum chloride and phosphomolybdic acid.

In some media the color produced by the *Bacillus pyocyaneus* (bacillus of green pus) is a fluorescent green. The recent studies of Gessard show that this is a different pigment. According to this author, cultures in a two-per-cent solution of peptone give a beautiful blue tint, the production of which is hastened by adding to the liquid five per cent of glycerin. In nutrient gelatin and agar cultures a fluorescent green color is developed, which, according to Gessard, is due to the presence of albumin. Peptone and gelatin are said to produce pyocyanin without the fluorescent-green pigment, and cultures in bouillon to give both this and pyocyanin. In milk the fluorescent-green color is first seen, but subsequently, when the casein has been peptonized by a diastase produced in the culture, pyocyanin is also formed. Several other microorganisms are known which produce a fluorescent-green color, due probably to the same pigment as is produced by the bacillus of green pus in albuminous media.

Babes claims to have obtained two pigments from cultures of the *Bacillus pyocyaneus* in addition to pyocyanin: one, soluble in alcohol, has by transmitted light a chlorophyll-green color, by reflected light it is blue; the other, insoluble in alcohol and chloroform, by transmitted light is of a dark orange-red, by reflected light a greenish-blue.

In Gessard's latest publication (1891) he shows that the production of pyocyanin or of the fluorescent-green pigment does not depend alone upon the culture medium, but that there are different varieties of the *Bacillus pyocyaneus*. He has succeeded in producing four distinct varieties—one which produces both pyocyanin and fluorescence, one which produces pyocyanin alone, one which produces the fluorescent-green pigment alone, and one which produces no pigment. The last-mentioned non-chromogenic variety was produced by subjecting the second variety to the action of heat. A temperature of 57° maintained for five minutes destroyed the power to produce pigment without destroying the vitality of the bacillus, which was propagated through successive cultures without regaining this power.

The well-known *Bacillus prodigiosus* (also described as a micrococcus) produces a red pigment which is insoluble in water but soluble in alcohol. By the addition of an acid the color becomes carmine and then violet, which is changed to yellow by an alkali. The color is said by Schottelius to be diffused in the young cells, and after the death of the cells to be present in their vicinity in the form of granules. The same author has shown that by subjecting the bacillus to special conditions a variety may be obtained which no longer produces pigment.

The conditions which govern the formation of pigment in the chromogenic bacteria are determined with comparative facility because the results of changed conditions are apparent to the eye; in the case of products which are not colored the difficulties attending the study of these conditions are much greater, but the results are in many instances more important. The following are among the best known pigment-producing (chromogenic) bacteria:

Staphylococcus pyogenes aureus (No. 1), *Staphylococcus pyogenes citreus* (No. 3), *Sarcina aurantiaca* (No. 226), *Sarcina lutea* (No. 227), *Bacillus cyanogenus* (No. 257), *Bacillus janthinus* (No. 267), *Bacillus fluorescens liquefaciens* (No. 277), *Bacillus indicus* (No. 283), *Bacillus pyocyaneus* (No. 95), *Bacillus prodigiosus* (No. 284), *Spirillum rubrum* (No. 429).

Liquefaction of Gelatin.—Many species of bacteria, when planted in a medium containing gelatin, cause a liquefaction of the gelatin in the immediate vicinity of the growing microorganisms, while many others multiply abundantly in the same medium without liquefying the gelatin. This character, as first shown by Koch, is an important one in the differential diagnosis of species which resemble each other in form and in other respects. It has no relation to pathogenic power, as some liquefying organisms are harmless saprophytes and some deadly disease germs, while, on the other hand, non-liquefying bacteria may be very pathogenic or quite innocent.

Liquefaction is produced by a soluble peptonizing ferment formed during the growth of the cells. This is shown by the fact that if a liquefying organism is cultivated in bouillon and the living cells removed by filtration or killed by heat, the power of liquefying gelatin remains in the culture fluid. This was first observed by Bitter (1886) and independently by the writer in 1887. In experiments made to determine the thermal death-point of various bacteria the writer found that when cultures of liquefying species were subjected to a temperature which killed the microorganisms, a few drops of the culture added to nutrient gelatin which had been liquefied by heat prevented it from subsequently forming a solid jelly when cold.

In a recent study of the ferments produced by bacteria which

cause liquefaction of gelatin—"tryptic enzymes"—made by Fermi, in the laboratory of the Hygienic Institute of Munich (1891), the following results were obtained :

The enzymes were not obtained pure, and their isolation from other proteids present in the cultures was found to be attended with great difficulties, but their ferment action was studied and was found to be influenced by various conditions.

All were destroyed by a temperature of 70° C., but the enzymes produced by various liquefying bacteria differed considerably as to the temperature which they were able to withstand. Some were destroyed by a temperature of 50° to 55° C.—*Bacillus megatherium*, *Bacillus ramosus*, *Staphylococcus pyogenes aureus* ; some by a temperature of 55° to 60° C.—*Bacillus subtilis*, *Bacillus pyocyaneus*, *Bacillus fluorescens liquefaciens*, *Sarcina aurantiaca* ; some by 65° to 70° C.—*Bacillus anthracis*, *Spirillum cholerae Asiaticæ*, *Spirillum* of Finkler and Prior, *Spirillum tyrogenum*.

These enzymes, like the previously known pepsin, trypsin, and invertin, do not dialyze.

Only a few of these bacteria enzymes acted upon fibrin, and no action was observed upon casein or upon egg albumen.

Their liquefying action upon gelatin was prevented by the action of sulphuric acid, and to a less degree by nitric acid, but was not interfered with by acetic acid.

The liquefying bacteria, as a rule, only produce enzymes when cultivated in a medium containing albumen.

These enzymes are not produced by a solution of the protoplasm of dead bacterial cells, but are a product of the vital activity of living cells.

Among the numerous liquefying bacteria known to bacteriologists we may mention the following species as deserving the student's special attention : *Staphylococcus pyogenes aureus* (No. 1), *Staphylococcus pyogenes albus* (No. 2), *Sarcina lutea* (No. 227), *Sarcina aurantiaca* (No. 226), *Bacillus anthracis* (No. 45), *Bacillus pyocyaneus* (No. 95), *Bacillus subtilis* (No. 379), *Bacillus indicus* (No. 283), *Bacillus prodigiosus* (No. 284), *Spirillum cholerae Asiaticæ* (No. 155), *Spirillum* of Finkler and Prior (No. 156), *Proteus vulgaris* (No. 97).

Production of Acids.—Numerous bacteria give an acid reaction to the media in which they are cultivated, and the acids produced are various—lactic, acetic, butyric, propionic, succinic, etc.

The power to produce an acid is well shown by adding to neutral or alkaline culture media a solution of litmus. The change in color due to the formation of an acid may be followed by the eye, and comparative tests may be made to aid in the differentiation of similar bacteria.

A considerable number of bacteria are able to produce *lactic acid* from milk sugar and other carbohydrates. One of these is considered the special lactic-acid ferment—*Bacillus acidi lactici*—and is the usual cause of the acid fermentation of milk. Pure cultures of this bacillus introduced into sterilized milk or solutions of milk sugar, cane sugar, dextrin, or mannite, give rise to the lactic-acid fermentation, in which carbonic acid is also set free. The process requires free access of oxygen, and progresses most favorably at a temperature of 35° to 40° C., ceasing at about 45°. In milk, coagulation of the casein occurs within fifteen to twenty-four hours after adding a small quantity of a pure culture of the lactic-acid bacillus. This is not due, however, to the acid fermentation, but to a ferment resembling that of rennet, which is produced by many different bacteria, some of which do not produce an acid reaction of the milk. Among the bacteria which produce lactic acid from milk sugar we may mention the staphylococci of pus, *Bacillus lactis aërogenes*, and *Bacillus coli communis*.

The formula showing the transformation of sugar into lactic acid is usually stated as follows : $C_6H_{12}O_6 = 2(HC_3H_5O_3)$.

Acetic acid is also produced from dilute solutions of alcohol by the action of a special bacterial ferment, which accumulates upon the surface of the fluid as a mycoderma, consisting almost entirely of the *Bacillus aceticus* (*Mycoderma aceti*). Free access of oxygen is required, and a temperature of about 33° C. is most favorable to the process. According to Duclaux, the "*Mycoderma aceti*" oxidizes the alcohol, in solutions containing it, so long as any is present, and when it is exhausted it oxidizes the acetic acid previously formed by oxidation of the alcohol, producing from it carbon dioxide and water.

The formation of acetic acid from alcohol is shown by the following formula : Ethyl alcohol $CH_3CH_2OH + O_2 = CH_3COOH + H_2O$.

Butyric acid is produced by a considerable number of bacteria, one of which, named *Bacillus butyricus*, has received the special attention of Prazmowski. This is strictly anaërobic. In solutions of starch, dextrin, sugar, or salts of lactic acid, when oxygen is excluded it produces butyric acid in considerable quantity, and at the same time carbon dioxide and hydrogen gas are set free. Duclaux gives the following formula of a solution containing lactate of lime in which the action of the butyric-acid ferment may be well studied :

Water,	8 to 10 litres.
Lactate of lime (pure),	225 grammes.
Phosphate of ammonia,	0.75 "
Phosphate of potash,	0.4 "
Sulphate of magnesia,	0.4 "
Sulphate of ammonia,	0.2 "

This is introduced into a flask with two necks, such as is shown in Fig. 77. Having filled the flask with the culture liquid, the bent neck is dipped into a porcelain dish containing the same. Heat is then applied both to flask and dish, and the liquid in each is kept in ebullition for half an hour. By this means the air is completely driven out of the flask. This is now allowed to cool, while the fluid in the shallow dish is kept hot, so that the liquid mounting from it into the flask shall be free from air. When the flask is full it is transferred to an incubating oven heated to 25° to 30° C., and the bent tube is immersed in a dish containing mercury. The little funnel attached to the upright tube is then filled with carbon dioxide and a culture of the butyric-acid bacillus is introduced into the funnel. By turning the stopcock in the upright tube a little of the culture is



FIG. 77.

admitted to the flask without admitting any air. Fermentation commences very soon, as is seen by the bubbles of gas given off. The liquid loses its transparency and the lactic acid is gradually consumed, butyrate of lime taking the place of the lactate.

Aërobic bacilli capable of producing butyric acid in culture solutions containing grape sugar or milk sugar have also been described by Liborius and by Hueppe.

Fitz has shown that in culture solutions containing glycerin the *Bacillus pyocyanus* produces butyric acid in addition to ethyl alcohol and succinic acid. *Bacillus Fitzianus* also produces some butyric acid in solutions containing glycerin, although the principal product of the fermentation caused by this microorganism is, according to Fitz, ethyl alcohol, twenty-nine grammes of which may be obtained from one hundred grammes of glycerin.

Botkin has recently (1892) described a "*Bacillus butyricus*" (No. 466) which he has not been able to identify positively with the butyric-acid ferment described by Prazmowski (No. 404). It is a widely distributed anaërobic bacillus, which he was able to obtain from milk or water containing it by placing it in the steam sterilizer for half an hour. The spores resisted this temperature and subsequently grew in anaërobic cultures, in a suitable medium, while all other bacteria and spores present were destroyed.

The writer has described a bacillus which causes active acid fermentation in culture solutions containing glycerin. The acid formed is volatile and is probably propionic acid—see *Bacillus acidiformans* (No. 93).

The Caucasian milk ferment—*Bacillus Kaukasicus*—produces a variety of products in the fermented milk which is a favorite drink among the Caucasians. The principal ones are ethyl alcohol, lactic acid, and carbon dioxide, but in addition to these small quantities of succinic, butyric, and acetic acids are formed. The inhabitants of the Caucasian mountains prepare this fermented drink in a very simple manner from the milk of cows or goats, to which they add the dried ferment collected from a receptacle in which the fermentation had previously taken place. Flügge gives the following directions for the preparation of this drink :

"Two methods may be employed. In the first the dry brown *kefir-körner* of commerce are allowed to lie in water for five to six hours until they swell; they are then carefully washed and placed in fresh milk, which should be changed once or twice a day until the *körner* become pure white in color and when placed in fresh milk quickly mount to the surface—in twenty to thirty minutes. One litre of milk is then poured into a flask and a full tablespoonful of the prepared *körner* added to it. It is allowed to stand open for five to eight hours; the flask is then closed and kept at 18° C. It should be shaken every two hours. At the end of twenty-four hours the milk is poured through a fine sieve into another flask, which must not be more than four-fifths full. This is corked and allowed to stand, being shaken from time to time. At the end of twenty-four hours a drink is obtained which contains but little CO₂ or alcohol. Usually it is not drunk until the second day, when, upon standing, two layers are formed, the lower milky, translucent, and the upper containing fine flakes of casein. When shaken it has a cream-like consistence. On the third day it again becomes thin and very acid.

"The second method is used when one has a good *kefir* of two or three days to start with. Three or four parts of fresh cow's milk are added to one part of this and poured into flasks which are allowed to stand for forty-eight hours with occasional shaking. When the drink is ready for use a portion (one-fifth to one-third) is left in the flask as ferment for a fresh quantity of milk. The temperature should be maintained at about 18° C.; but at the commencement a higher temperature is desirable. The *körner* should be carefully cleaned from time to time and broken up to the size of peas. The cleaned *körner* may be dried upon blotting paper in the sun or in the vicinity of a stove; when dried in the air they retain their power to germinate for a long time."

Fermentation of urea. The alkaline fermentation of urine is

effected by various microorganisms, but chiefly by the *Micrococcus ureæ*, the ferment action of which has been carefully studied by Pasteur, Duclaux, and others. The change which occurs under the action of the living ferment was determined by the chemist Dumas as long ago as 1830, but it remained for Pasteur to show that this change depends upon the presence and vital activity of a living microorganism.

The transformation of urea into carbonate of ammonia is shown by the following formula: $\text{COH}_2\text{N}_2 + 2\text{H}_2\text{O} = \text{CO}_2 + 2\text{NH}_3 + \text{H}_2\text{O} = (\text{NH}_4)_2\text{CO}_3$.

According to Van Tieghem, *Micrococcus ureæ* continues to grow in a liquid containing as much as thirteen per cent of carbonate of ammonia. It may be cultivated in an artificial solution of urea, with the addition of some phosphates, as well as in urine.

The *Bacillus ureæ* of Miquel has also the power of producing the alkaline fermentation of urine, but it does not thrive in so strong a solution of carbonate of ammonia.

A different micrococcus—*Micrococcus ureæ liquefaciens*—has also been studied in Flügge's laboratory which possesses the same power. According to Musculus, a soluble ferment may be isolated from urine which has undergone alkaline fermentation, which changes urea into carbonate of ammonia. He obtained it from urine containing considerable mucus, in a case of catarrh of the bladder. But Leube has shown that cultures of *Micrococcus ureæ* from which the micrococcus was removed by filtration through clay do not induce alkaline fermentation. The soluble ferment obtained by Musculus must therefore be from some other source.

Miquel has given special attention to the study of bacteria which produce alkaline fermentation in urine, and in addition to the species above mentioned has described the following: *Urobacillus Pasteuri* (No. 467), *Urobacillus Duclauxi* (No. 468), *Urobacillus Freudenreichi* (No. 469), *Urobacillus Maddoxi* (No. 470), *Urobacillus Schutzenbergi* (No. 471).

Viscous fermentation. A special fermentation which occurs sometimes in wines, and in the juices of bulbous roots containing glucose, and in milk, is produced by various bacteria. One of these is a micrococcus which has been described by Conn under the name of *Micrococcus lactis viscosus* (No. 195). The fermented juices become very viscous, owing to the formation of a gum-like product resembling dextrin; at the same time mannite and CO_2 are produced. The gum-like substance, called viscose by Béchamp, is soluble in cold water and is precipitated by alcohol. Recently (1892) Guillebeau has described a micrococcus and a bacillus which produce viscous fermentation in milk—*Micrococcus Freudenreichi* (No. 475) and

Bacillus Hessi (No. 476). A micrococcus producing viscous fermentation in milk has also been described by Schmidt-Mühlheim, and a bacillus by Löffler. *Bacillus mesentericus vulgatus* also produces a similar change in milk.

Marsh gas, CH_4 , is produced by the fermentation of cellulose, through the action of microorganisms the exact characters of which have not yet been determined. According to Tappeiner, there are two different fermentations of cellulose. The first occurs in a neutral one-per-cent flesh extract solution to which cotton or paper pulp has been added. The gases given off are CO_2 and CH_4 and small quantities of H_2S . The second fermentation occurs when an alkaline solution of flesh extract containing cellulose in suspension is used. The gases formed are CO_2 and H_2 . In both cases small quantities of aldehyde, isobutyric acid, and acetic acid are produced.

Hydrosulphuric acid, H_2S . This gas is produced during the growth of certain bacteria. The conditions governing its development have been studied by Holschewnikoff, who experimented with two species, one isolated by himself and one by Lindenborn, named respectively *Bacterium sulfureum* and *Proteus sulfureus*. The first-mentioned bacterium, when inoculated into eggs, produced within three or four days an abundant quantity of H_2S ; the other did not. Upon raw albumin both species produced but little, and on the yolk of egg a considerable amount of this gas. Upon cooked egg the action was the reverse. In peptone-bouillon the evolution of H_2S was abundant; in the absence of peptone, very slight.

Putrefactive fermentation. The putrefactive decomposition of albuminous material of animal and vegetable origin is effected by a great variety of microorganisms and gives rise to the formation of a great variety of products, some of which are volatile and are characterized by their offensive odors. According to Flügge, the first change which occurs consists in the transformation of the albumins into peptone, and this may be effected by a large number of different bacteria. Among the products of putrefactive fermentation known to chemists are the following substances: Carbon dioxide, hydrogen, nitrogen, hydrosulphuric acid (H_2S), phosphoretted hydrogen (PH_3), methane, formic acid, acetic acid, butyric acid, valerianic acid, palmitic acid, crotonic acid, glycolic acid, oxalic acid, succinic acid, propionic acid, lactic acid, amidostearic acid, leucin, ammonia, ammonium carbonate, ammonium sulphide, trimethylamine, propylamine, indol, skatol, tyrosin, neuridin, cadaverin, putrescin, cholin, neurin, peptotoxin, and various other volatile acids, ptomaines, etc.

The special products of putrefaction vary according to the nature of the material, the conditions in which it is placed, and the micro-

organisms present. One or the other of the bacteria concerned will take the precedence when circumstances favor its growth. Thus the aerobic bacteria cannot grow unless the putrefying material is freely exposed to atmospheric oxygen; the anaerobic species require its exclusion. Some saprophytic bacteria grow at a comparatively low temperature, others take the precedence when the temperature is high; some, no doubt, thrive only in presence of products evolved by other species, and are consequently associated with and dependent upon these species; some are restrained in their growth sooner than others by the products evolved as a result of their own vital activity or that of associated organisms; some grow in the presence of acids and give rise to an acid fermentation which wholly prevents the development of other species.

At the outset putrefaction is often attended with the presence of several species of micrococci and certain large bacilli, which are displaced later by short motile bacteria belonging to a group which includes several bacilli formerly described under the common name of *Bacterium termo*.

The malodorous volatile products of putrefaction are to a considerable extent produced by anaerobic species. For this reason these odors are more pronounced when masses of albuminous material undergo putrefaction in situations where the oxygen of the air has not free access or where it is displaced by carbon dioxide. The body of a dead animal, although freely exposed to the air, furnishes in its interior a suitable nidus for these anaerobic gas-forming species, and they may give rise to products of one kind, while aerobic species upon the surface of the mass induce different forms of putrefactive fermentation. In the bodies of living animals these anaerobic microorganisms are constantly present in the intestine, and after death they quickly invade the body and multiply at its expense under favorable conditions as to temperature. The surface decomposition due to aerobic bacteria occurs later and is not attended with the same putrefactive odors, the products evolved being of a simpler chemical composition— CO_2 , HN_3 . No doubt these aerobic bacteria, by consuming the oxygen and forming an atmosphere of carbon dioxide, help to make the conditions favorable for the continued development of the anaerobics in the interior of the organic mass; at the same time they find a suitable pabulum in some of the more complex products of decomposition occurring in the absence of oxygen. The gases produced in the interior of a putrefying mass are mainly CH_4 , H_2S , and H_2 .

Many of the bacteria of putrefaction are facultative anaerobics—that is to say, they are able to multiply either in the presence of oxygen or in its absence. The products evolved by these differ, no

doubt, according to whether they are or are not supplied with atmospheric oxygen.

The anaërobic bacteria concerned in putrefaction have as yet received comparatively little attention. Among the aërobics and facultative anaërobics the following are best known: *Micrococcus foetidus* (No. 189), *Bacillus saprogenes* I., II., and III., *Bacillus coprogenes foetidus* (No. 116), *Bacillus putrificus coli* (No. 324), *Proteus vulgaris* (No. 97), *Proteus Zenkeri* (No. 100), *Proteus mirabilis* (No. 99), *Bacillus pyogenes foetidus* (No. 72), *Bacillus fluorescens liquefaciens* (No. 277), *Bacillus pyocyanus* (No. 95), *Bacillus coli communis* (No. 89), *Bacillus janthinus* (No. 267).

Soluble Ferments.—Several species of bacteria produce soluble ferments capable of changing starch into maltose, dextrin, etc. Hueppe has shown that the lactic-acid bacillus produces a diastase, and Miller obtained from the human intestine a species which dissolves starch. Marcano, by filtering cultures of species capable of this ferment action through porcelain, was able to show that the effect is due to a soluble ferment, which must have been produced by the vital activity of the living microorganisms. Wortmann also obtained a diastase from culture liquids which was precipitated by alcohol and again dissolved in water; in slightly acid solutions it promptly converted starch into glucose. This is said to be produced in culture liquids only when these do not contain albumin. In the presence of albumin a peptonizing ferment was formed; in its absence, a diastase by which starch was dissolved to serve as pabulum for the bacteria present. These experiments were not made with pure cultures, and more exact researches in this direction are desirable.

A peptonizing ferment for gelatin is produced by a considerable number of bacteria, as stated under the heading "Liquefaction of Gelatin." The jellified albumin in cultures in blood serum is also liquefied by a peptonizing ferment produced by certain species of bacteria.

Some authors also speak of a soluble ferment capable of inverting cane sugar or milk sugar. According to Hueppe, such a ferment is produced by the *Bacillus acidi lactici*. A soluble ferment for cellulose is supposed by Flügge to be produced by several species—among others by *Bacillus butyricus* and by *Vibrio rugula*.

Several bacilli produce a soluble ferment capable of coagulating the casein of milk.

Reduction of Nitrates, and Nitrification.—The researches of Gayon, Dupetit, and others show that certain bacteria are able to reduce nitrates with liberation of ammonia and free nitrogen. This is effected in the absence of oxygen by anaërobic bacteria, and,

among others, by *Bacillus butyricus*. Certain aërobic bacteria also accomplish the same result. Thus Heræus obtained two species from water which reduced nitrates in a very decided manner. On the other hand, a number of species are known to oxidize ammonia, producing nitric acid. Schlösing and Münz, as a result of numerous experiments, arrived at the conclusion that in the soil nitrification is effected by a single species. But it is doubtful whether they worked with pure cultures, and more recent researches show that several, and probably many, different bacteria possess this power. According to Heræus, the following species, tested by him, oxidize ammonia: *Bacillus prodigiosus*, the cheese spirillum of Deneke, the Finkler-Prior spirillum, the typhoid bacillus, the anthrax bacillus, the staphylococci of pus. The oxidation does not always go to the point of forming nitrates, but nitrites may be formed in the soil (Duclaux). Warrington states that certain bacteria which formed nitrates in a suitable culture medium produced only nitrites when, after an interval of four or five months, some of the culture was transferred to a solution containing muriate of ammonia. The same author states that the process of nitrification occurs only in the dark.

The recent researches of Winogradsky, of the Franklands, and of Jordan show that the failure of earlier investigators to obtain the nitrifying bacteria from the soil in pure cultures was due to the fact that these bacteria do not grow in the usual culture media. By the use of certain saline solutions the authors named have succeeded in isolating nitrifying bacteria in pure cultures, or nearly so. It is still uncertain whether these investigators have obtained the same bacteria, but the microorganisms described by them, and obtained from widely distant sources, are similar in their morphological and biological characters, and at least belong to the same group (see Nos. 439, 440, 441). In his latest communication (September, 1891) Winogradsky arrives at the conclusion that the ferments which cause the oxidation of ammonia and production of nitrites are not capable of producing nitrates, but that other microorganisms are concerned in the oxidation of nitrites. In sterilized soil to which a pure culture of his nitromonas was added nitrites only were produced, and the presence of various microorganisms common in the soil did not result in the formation of nitrates so long as the specific ferment was absent to which this second oxidation is ascribed (nitrifying bacillus of Winogradsky, No. 451).

Phosphorescence.—Recently several different bacteria have been studied which, in pure cultures, give rise to phosphorescence in the medium in which they are cultivated. In gelatin cultures the light is sufficient in some instances to enable one to tell the time by a

watch in a perfectly dark room, and such cultures have even been photographed by their own light.

The phosphorescence is influenced by changes in the culture medium and by conditions of temperature, but we have no exact knowledge of the mode of its production. The *Bacillus phosphorescens* from sea water in the vicinity of the West Indies gives the most striking results, especially when planted upon the surface of cooked fish and placed in an incubating oven at 30° C. Two other species have been studied by Fischer—one obtained from the water of the harbor at Kiel, and the other a widely distributed species called by Fischer *Bacterium phosphorescens*. Katz (1891) has recently described several new species obtained by him from sea water and from phosphorescent fish in the markets at Sydney, New South Wales—*Bacillus smaragdino-phosphorescens*, *Bacillus argenteo-phosphorescens*, *Bacillus cyaneo-phosphorescens*, *Bacillus argenteo-phosphorescens liquefaciens* (Nos. 337, 338, 341, and 342).

V.

PTOMAINES AND TOXALBUMINS.

VARIOUS basic substances containing nitrogen, and in chemical constitution resembling the vegetable alkaloids, have been isolated by chemists from putrefying material and from cultures of the bacteria concerned in putrefaction, and also from certain pathogenic species. Some of these *ptomaines* are non-toxic, and others are very poisonous in minute doses (*toxines*). The toxic substances sometimes developed in milk, cheese, sausage, etc., are also of this nature, and are doubtless produced by the action of microorganisms. The pathogenic power of the bacteria which cause various infectious diseases in man and the lower animals has also been shown to result from the production of toxic ptomaines or of toxalbumins. Selmi first gave the name ptomaines to cadaveric alkaloids isolated by him, and Panum subsequently called attention to the fact that poisonous basic substances of this class are contained in putrefying material. Extended researches with reference to the ptomaines have since been made by numerous chemists, the most important being those of Bergmann, Schmiedeberg, Zuelzer and Sonnenschein, Hager, Otto, Selmi, Brieger, Gautier and Étard, and Vaughan.

For a full account of the history and chemical composition of the ptomaines the reader is referred to the valuable work of Vaughan and Novy ("Ptomaines and Leucomaines," Philadelphia, 1891). In the present volume we shall give a brief account only of some of the most important.

NON-TOXIC PTOMAINES.

Neuridin, $C_8H_{11}N_3$.—This is one of the most common of the alkaloids of putrefaction and was isolated by Brieger in 1884. It is obtained most abundantly from tissues containing gelatin. Very soluble in water, but insoluble in ether and absolute alcohol. Has a disagreeable odor.

Cadaverin, $C_8H_{11}N_3$.—Isomeric with neuridin; has a very disagreeable odor; forms a thick, transparent, syrupy liquid; is volatile, and can be distilled with steam without undergoing decomposition. When exposed to the air the base absorbs carbon dioxide and

forms a crystalline mass. Is produced in cultures of the cholera spirillum and of the spirillum of Finkler and Prior which have been kept for a month or more at 37° C.

Putrescin, $C_4H_{11}N_3$.—A base resembling cadaverin and commonly associated with it. Obtained by Brieger from various sources, most abundantly from substances containing gelatin and in the more advanced stages of putrefaction. It is obtained in the form of a hydrate, which is a transparent liquid having a boiling point of about 135° . With acids it forms crystalline salts.

Saprin, $C_4A_4N_2$.—Resembles cadaverin and is commonly associated with it in putrefying material. Isolated by Brieger.

Methylamine, CH_3NH_2 .—Obtained by Brieger from putrefying fish and from old cultures of the cholera spirillum.

Dimethylamine, $(CH_3)_2NH$.—Obtained by Brieger from putrefying gelatin and by Bocklisch from decomposing fish.

Trimethylamine, $(CH_3)_3N$.—Obtained from various sources, and by Brieger from cultures of the cholera spirillum and of the streptococcus of pus.

TOXIC PTOMAINES.

Neurin, $C_4H_{11}NO$.—First obtained by Liebreich in 1865 as a decomposition product of protagon from the brain. Obtained by Brieger from putrefying muscular tissue. When crystallized from an aqueous solution it forms five- or six-sided plates; from an alcoholic solution it crystallizes in the form of needles (Liebreich). This base is toxic in small doses. In frogs the injection of a few milligrammes produces paralysis of the extremities. Respiration is first arrested and the heart stops in diastole. Atropine appears to be a physiological antidote to the toxic effects of neurin. In rabbits it produces profuse salivation. The pupil is contracted by the direct application of a concentrated solution.

Cholin, $C_4H_{11}NO$.—First obtained from hog's bile by Strecker in 1862. Has been obtained by Brieger from various sources, including cultures of the cholera spirillum. It is also found widely distributed in the vegetable kingdom. May be prepared from the yolk of eggs by the method of Diakonow. Cholin is obtained in the form of a syrupy, alkaline liquid which combines with acids to form deliquescent salts. At first this base was not supposed to have toxic properties, but more recent researches have shown that in comparatively large doses it produces symptoms resembling those caused by minute doses of neurin.

Muscarin, $C_4H_{11}NO$.—This toxic principle of poisonous mushrooms has also been obtained by Brieger from putrefying fish. It may be produced artificially by the oxidation of cholin. In small doses it kills rabbits and frogs. In the rabbit it produces lacrymation and

salivation, the pupil is contracted, and the animal dies in convulsions. Frogs are completely paralyzed by the action of muscarin and die with arrest of the heart's action in diastole.

Peptotoxin.—The exact composition of this ptomaine has not been determined. Brieger obtained it during the early putrefaction of proteid substances and also from the artificial digestion of fibrin. It is very poisonous for frogs, which become paralyzed and die within fifteen or twenty minutes after the subcutaneous injection of a few drops of a dilute solution. Rabbits also are killed by doses of half a gramme to a gramme, the symptoms being paralysis of the posterior extremities and stupor. Peptotoxin is soluble in water, but insoluble in ether or chloroform. It is not destroyed by boiling.

Tyrotaxon.—First obtained by Vaughan in poisonous cheese, and subsequently by the same chemist and others in poisonous milk and ice cream. Chemically tyrotaxon is very unstable. It is decomposed when heated with water to 90° C. It is insoluble in ether. From sixteen kilogrammes of poisonous cheese Vaughan obtained 0.5 gramme of the poison. The symptoms produced in man by eating cheese or milk containing tyrotaxon are vertigo, nausea, vomiting, and severe rigors, with pain in the epigastrium, cramps in the legs, griping pain in the bowels attended with purging, numbness and a pricking sensation in the limbs, and great prostration.

Methylguanidin, $C_2H_5N_3$.—Obtained by Brieger from putrefying horseflesh which had been kept at a low temperature for several months. This base was previously known to chemists, having been obtained by the oxidation of creatin. By Bocklisch it has been obtained from impure cultures of the Finkler-Prior spirillum which had been kept for about a month. It is obtained as a colorless mass having an alkaline reaction, and which is quite deliquescent. Brieger gives the following account of the toxic action as tested on guinea-pigs in a dose of 0.2 gramme: The respiration increases in rapidity, the pupils dilate to the extreme limit, the animal has copious discharges of urine and fæces, the extremities become paralyzed, and at the end of about twenty minutes death occurs in convulsions.

Mytilotoxin.—Obtained by Brieger from poisonous mussels. The toxic action resembles that of curare.

Typhotoxin, $C_4H_7NO_2$.—Obtained by Brieger from bouillon cultures of the typhoid bacillus which had been kept for a week or more at a temperature of about 37.5° C. In mice and guinea-pigs this base produces salivation, rapid respiration, dilatation of the pupils, diarrhoea, and death in from twenty-four to forty-eight hours. It is believed by Brieger that the specific action of the typhoid bacillus is due to the production of this ptomaine.

A base which is isomeric with typhotoxin has been obtained by

Brieger from putrefying horseflesh which was kept at a low temperature for several months. Unlike it, however, the free base has an acid reaction, while typhotoxin is strongly alkaline. It differs also in its physiological action, being more toxic and producing convulsions; the heart is arrested in diastole. Typhotoxin, on the other hand, does not induce convulsions and the heart is arrested in systole.

Tetanin, $C_{11}H_{11}N_2O_4$.—Obtained by Brieger from impure cultures of the tetanus bacillus cultivated in bouillon in an atmosphere of hydrogen. (The tetanus bacillus is a strict anaërobic.) Obtained subsequently by the same chemist from the amputated arm of a patient with tetanus. This base has been obtained, by crystallization from hot alcohol, in clear yellow plates which are not very soluble in water. The hydrochloride is a deliquescent salt which dissolves readily in alcohol. When injected into guinea-pigs or mice in rather large doses, tetanin first causes the animal to fall into a lethargic condition, followed by increased rapidity of respiration and tetanic convulsions. In guinea-pigs opisthotonos is induced, together with the characteristic tetanic convulsions as seen in animals suffering from tetanus. Three other toxic bases have been obtained by Brieger from cultures of the tetanus bacillus, which cause similar symptoms. One—*tetanotoxin*—is given by Brieger the formula $C_8H_{11}N$. A second base, the composition of which has not been determined, is called *spasmotoxin*.

Cholera Ptomaines.—Brieger has obtained from pure cultures of the cholera spirillum several of the toxic ptomaines heretofore referred to—cadaverin, putrescin, cholin, methyl-guanidin. In addition to these he found two toxic substances which appear to be peculiar products of this microorganism. One induces cramps and muscular tremors in small animals, the other diarrhoea and symptoms of collapse.

Toxalbumins.—Recent researches by Brieger and Fränkel (1890) show that very toxic substances of a different nature are present in cultures of some of the pathogenic bacteria; these have been designated by the authors named "toxalbumins."

Roux and Yersin had previously shown that filtered cultures of the diphtheria bacillus contain a toxic substance which produces paralysis and death in guinea-pigs and rabbits. This substance has now been obtained in a pure state and its toxic action tested by the authors first named. It is destroyed by a temperature of $60^{\circ}C$., but remains in an active condition in cultures which have been sterilized by several hours' exposure to a temperature of 50° , or in those which have been passed through a clay filter. It is not volatile, and differs essentially from the ptomaines and also from the soluble ferments. It was obtained as a snow-white, amorphous mass which was ex-

tremely toxic in its action upon small animals. When injected into guinea-pigs in the proportion of two and one-half milligrammes to one kilogramme of body weight, it caused death after a considerable interval of time (from a few days to several weeks), during which the animal became emaciated and spreading abscesses and necrosis of the tissues occurred at the point of injection. This toxalbumin was obtained in a pure state by repeated precipitation from an aqueous solution by means of alcohol. It is produced most abundantly in cultures containing albumin, and old cultures are more toxic than recent ones. Chemical analysis gave the following result : C 45, 35, H 7, 13, N 16, 33, S 1, 39, O 29, 80. The authors remark, however, that the chemical characters have not yet been fully determined.

The same chemists have obtained toxic substances of a similar nature from cultures of the bacillus of typhoid fever, of the tetanus bacillus, of the *Staphylococcus aureus*, and of the cholera spirillum. Hankin had previously obtained a toxic "albumose" from cultures of the anthrax bacillus by precipitation with alcohol, drying, solution in water, and filtration through porcelain; and Christmas had obtained an albuminous substance from cultures of *Staphylococcus aureus* which produced pus formation when injected beneath the skin of rabbits or into the anterior chamber of the eye.

According to Brieger and Fränkel, these toxalbumins are divided into two principal groups, one of which is characterized by solubility in water, as in that produced by the diphtheria bacillus; and one in which the albumin is insoluble or but slightly soluble, as is the case with those obtained from cultures of the typhoid bacillus, the cholera spirillum, and the *Staphylococcus aureus*.

The toxalbumin from cholera cultures, obtained as pure as possible and suspended in water, when injected under the skin of a guinea-pig, caused its death in two or three days. It was not, however, toxic for rabbits, even when injected in considerable quantity.

On the contrary, the toxalbumin of the typhoid bacillus, which is dissolved with difficulty in water, was more poisonous for rabbits than for guinea-pigs. When injected subcutaneously into rabbits death usually occurred in eight to ten days. No notable pathological changes were observed at the autopsy.

The toxalbumin of *Staphylococcus aureus* killed rabbits and guinea-pigs within a few days, and in some cases at the end of twenty-four hours. The post-mortem appearances were necrosis or purulent breaking down of the tissues at the point of injection, with swelling and redness of the surrounding tissues and general inflammatory appearances. The toxalbumin of anthrax cultures resembles that of the diphtheria bacillus in being soluble in water. It was obtained by Brieger from the organs of animals recently dead from

anthrax. In a dry condition it has a grayish-white color and gives the reactions of albumins.

The toxalbumin of the tetanus bacillus is also soluble in water. It is best obtained in bouillon cultures containing glucose.

Quite recently (1891) G. and F. Klemperer have announced their success in obtaining a toxalbumin from cultures of *Micrococcus pneumoniae crouposæ* ("diplococcus pneumoniae"); this they propose to call pneumotoxin.

Koch's "Tuberculin."—This is a glycerin extract of the toxic substances present in cultures of the tubercle bacillus. Crude tuberculin is obtained from liquid cultures made in veal broth to which one per cent of peptone and four to five per cent of glycerin have been added. This culture liquid is placed in flasks and inoculated upon the surface with small masses from a pure culture of the tubercle bacillus. A tolerably thick and dry white layer is developed, which after a time covers the entire surface. At the end of six to eight weeks development ceases and the culture liquid is evaporated over a water bath to one-tenth its volume; this, after being filtered, constitutes the crude tuberculin. By precipitation with sixty-per cent alcohol Koch has obtained from this a white precipitate which has the active properties of the glycerin extract. This is soluble in water and in glycerin, and has the chemical reactions of an albuminous body.

Zuelzer has recently (1891) reported his success in isolating a toxic substance from tubercle cultures. The contents of tubes containing pure cultures of the bacillus are first treated with hot water acidulated with hydrochloric acid. This solution is filtered, evaporated, and then several times precipitated with platinum chloride. The double salt formed is decomposed by hydrosulphuric acid, after which the liquid is filtered and evaporated to dryness. A white, crystalline salt is thus obtained which is soluble in hot water. This salt was toxic for rabbits and guinea-pigs in doses of from one to three centigrammes. Death usually occurred in from two to four days. In guinea-pigs one centigramme injected subcutaneously caused, within a few minutes, a greatly increased frequency of respiration, an elevation of temperature, and protrusion of the eyeballs.

Mallein.—Kalwing, Preusse, and Pearson have obtained from cultures of the glanders bacillus a "lymph" which somewhat resembles the crude tuberculin of Koch. This was obtained by Preusse by treating old potato cultures of the glanders bacillus with glycerin and water. The extract was filtered several times and then sterilized in a steam sterilizer. This lymph injected into horses infected with glanders gives rise to a very decided elevation of temperature, while in horses free from this disease no such result follows.

VI.

INFLUENCE OF PHYSICAL AGENTS.

Heat.—We have already seen (Section II., Part Second) that the temperature favorable for the growth of most bacteria is between 20° and 40° C.; that some species are able to multiply at the freezing temperature, and others at as high a temperature as 60° to 70° C.; that, as a rule, the parasitic species require a temperature of 35° to 40°; and that low temperatures do not kill bacteria.

Frisch (1877) exposed various cultures to a temperature of -87° C., which he obtained by the evaporation of liquid CO₂, and found that micrococci and bacilli, after exposure to such a temperature, multiplied abundantly when again placed in favorable conditions. Prudden has also made extended experiments upon the influence of freezing. He found that while certain species resisted the freezing temperature for a long time, others failed to grow. Thus *Bacillus prodigiosus* did not grow after being frozen for fifty-one days; *Proteus vulgaris* was killed in the same time, and a slender, liquefying bacillus obtained from Croton aqueduct water was killed in seven days. *Staphylococcus pyogenes aureus* withstood freezing for sixty-six days, a fluorescent bacillus from Hudson River ice for seventy-seven days, and the bacillus of typhoid fever for one hundred and three days. Cultures made at intervals showed, however, a diminution in the number of bacteria. A similar diminution would perhaps have occurred in old cultures in which the pabulum for growth was exhausted, independently of freezing; for bacteria, like higher plants, die in time—which varies for different species—as a result of degenerative changes in the living protoplasm of the cells, and continued vitality in a culture depends upon continued reproduction.

Repeated freezing and thawing was found by Prudden to be more fatal to the typhoid bacillus than continuous freezing. Cultures were sterilized by being thawed out at intervals of three days and again refrozen, after repeating the operation five times.

Cadéac and Malet kept portions of a tuberculous lung in a frozen condition for four months, and found that at the end of this time tuberculosis was still produced in guinea-pigs by injecting a small quantity of this material.

In considering the influence of high temperatures we must take account of the very great difference in the resisting power of the vegetative cells and the reproductive elements known as spores, also of the fact as to whether dry or moist heat is used and the time of exposure.

Dry Heat.—When microorganisms in a desiccated condition are exposed to the action of heated dry air, the temperature required for their destruction is much above that required when they are in a moist condition or when they are exposed to the action of hot water or steam. This was thoroughly demonstrated by the experiments of Koch and Wolffhügel (1881). A large number of pathogenic and non-pathogenic species were tested, with the following general result : A temperature of 78° to 123° C. maintained for an hour and a half (over 100° for an hour) failed to kill various non-pathogenic bacteria, but was fatal to the bacillus of mouse septicæmia and that of rabbit septicæmia. To insure the destruction of all the species tested, in the absence of spores, a temperature of 120° to 128° C., maintained for an hour and a half, was required.

The spores of *Bacillus anthracis* and of *Bacillus subtilis* resisted this temperature and required to insure their destruction a temperature of 140° C. maintained for three hours. This temperature was found to injure most objects requiring disinfection, such as clothing and bedding. But the lower temperature which destroys microorganisms in the absence of spores (120° C. = 248° F.) can be used for disinfecting articles soiled with the discharges of patients with cholera, typhoid fever, or diphtheria, as the specific germs of these diseases do not form spores. It is probable also that it may be safely used to disinfect the clothing of small-pox patients, for we have experimental evidence that a lower temperature destroys the virulence of vaccine virus (90°–95° C.—Baxter).

In practical disinfection by means of dry heat it will be necessary to remember that it has but little penetrating power. In the experiments of Koch and Wolffhügel it was found that registering thermometers placed in the interior of folded blankets and packages of various kinds did not show a temperature capable of killing bacteria after three hours' exposure in a hot-air oven at 133° C. and above.

Moist Heat.—The thermal death-point of bacteria, in the absence of spores, is comparatively low when they are exposed to moist heat. The results of the writer's experiments are given below:

“In my temperature experiments I have taken great pains to insure the exposure of the test organisms to a uniform temperature, and have adopted ten minutes as the standard time of exposure. The method employed throughout has been as follows: From glass tubing having a diameter of about three-sixteenths of an inch I draw out in the flame of a Bunsen burner a number of capillary tubes, with an expanded extremity which serves as

an air chamber. A little material from a pure culture of the test organism is drawn into each of these capillary tubes by immersing the open extremity in the culture, after having gently heated the expanded end. The end of the tube is then hermetically sealed by heat. These tubes are immersed in a water bath maintained at the desired temperature for the standard time. The bath is kept at a uniform temperature by personal supervision. At the bottom of the vessel is a thick glass plate which prevents the thermometer bulb and capillary tubes, which rest upon it, from being exposed to heat transmitted directly from the bottom of the vessel. To further guard against this I am in the habit of applying the flame to the sides of the vessel, and a uniform temperature throughout the bath is maintained by frequent stirring with a glass rod. It is impossible to avoid slight variations, but by keeping my eye upon the thermometer throughout the experiment I have kept these within very narrow limits. . . . No attempt has been made to fix the thermal death-point within narrower limits than 2° C., and in the table the lowest temperature is given which has been found, in the experiments made, to destroy all of the microorganisms in the material subjected to the test. No doubt more extended experiments would result, in some instances, in a reduction of the temperature given as the thermal death-point for a degree or more. But the results as stated are sufficiently accurate for all practical purposes."¹

The results obtained in these experiments, for non-sporebearing bacteria, are given in the following table. The time of exposure was ten minutes, except for the cholera spirillum and the cheese spirillum of Deneke.

THERMAL DEATH-POINT OF BACTERIA.

	Centigrade.	Fahrenheit.
<i>Spirillum cholerae Asiaticæ</i>	52°	125.6° (4 m.)
<i>Spirillum tyrogenum</i> (cheese spirillum).....	52	125.6 (4 m.)
<i>Spirillum Finkler-Prior</i>	50	122.
<i>Bacillus typhi abdominalis</i>	56	138.8
<i>Bacillus of schweine-rothlauf</i> (rouget).....	58	136.4
<i>Bacillus murisepticus</i>	58	136.4
<i>Bacillus Neapolitanus</i> (Emmerich's bacillus).....	62	143.6
<i>Bacillus cavida</i>	62	143.6
<i>Bacillus pneumoniae</i> (Friedländer's).....	56	132.8
<i>Bacillus crassus sputigenus</i>	54	129.2
<i>Bacillus pyocyaneus</i>	56	132.8
<i>Bacillus indicus</i>	58	136.4
<i>Bacillus prodigiosus</i>	58	136.4
<i>Bacillus cyanogenus</i>	54	129.2
<i>Bacillus fluorescens</i>	54	129.2
<i>Bacillus acidi lactici</i>	56	132.8
<i>Staphylococcus pyogenes aureus</i>	58	136.4
<i>Staphylococcus pyogenes citreus</i>	62	143.6
<i>Staphylococcus pyogenes albus</i>	62	143.6
<i>Streptococcus pyogenes</i>	54	129.2
<i>Micrococcus tetragenus</i>	58	136.4
<i>Micrococcus Pasteuri</i>	53	125.6
<i>Sarcina lutea</i>	64	147.2
<i>Sarcina aurantiaca</i>	62	143.6

The following determinations of the thermal death-point of path-

¹ Quoted from the Report of the Committee on Disinfectants of the American Public Health Association, pages 136 and 152.

ogenic organisms have been made by the authors named : *Bacillus anthracis* (Chauveau), 54° C. ; *Bacillus mallei*—the bacillus of glanders—(Löffler), 55° C., *Bacillus gallinarum*—micrococcus of fowl cholera—(Salmon), 56° C. ; *Bacillus* of diphtheria (Löffler), 60° C.

In the writer's experiments the micrococcus of gonorrhœa was apparently killed by exposure for ten minutes to a temperature of 60° C.

"Some gonorrhœal pus from a recent case which had not undergone treatment was collected for me by my friend Dr. Rohé in the capillary glass tubes heretofore described. A microscopical examination of stained cover-glass preparations showed that this pus contained numerous 'gonococci' in the interior of the cells. Two of the capillary tubes were placed in a water bath maintained at 60° C. for ten minutes. The pus was then forced out upon two pledgets of cotton wet with distilled water. Two healthy men had consented to submit to the experiment, and one of these bits of cotton was introduced into the urethra of each and left *in situ* for half an hour. As anticipated, the result was entirely negative. For obvious reasons no control experiment was made to fix the thermal death-point within narrower limits.

"In connection with these experiments upon the thermal death-point of known pathogenic organisms, it is of interest to inquire whether the virulence of infectious material, in which it has not been demonstrated that this virulence is due to a microorganism, is destroyed by a correspondingly low temperature. Evidently, if this proves to be the case, it will be a strong argument in favor of the view that we have to deal with a microorganism in these diseases also. We have experimental proof that a large number of pathogenic organisms are killed by exposure for ten minutes to a temperature of 55° to 60° C. But, so far as I am aware, this low temperature would not be likely to destroy any of the poisonous chemical products which might be supposed to be the cause of infective virulence, leaving aside the fact that such chemical products have no power of self-multiplication, and, therefore, could not be the independent cause of an infectious disease."

"*Vaccine Virus*.—Carstens and Coert have experimented upon the temperature required to destroy the potency of vaccine virus. In a paper read at the International Medical Congress in 1879 they report, as a result of their experiments, that the maximum degree of heat to which fresh vaccine virus can be exposed without losing its virulence probably varies between 52° and 54° C. Fresh animal vaccine heated to 52° C. for thirty minutes does not lose its virulence. Fresh animal vaccine heated to 54.5° for thirty minutes loses its virulence.

"*Rinderpest*.—According to Semmer and Raupach, exposure for ten minutes to a temperature of 55° C. destroys the virulence of the infectious material in this disease.

"*Sheep-pox*.—The authors last mentioned have also found that the same temperature—55° C. for ten minutes—destroys the virulence of the blood of an animal dead from sheep-pox.

"*Hydrophobia*.—Desiring to fix the thermal death-point of the virus of hydrophobia, I obtained, through the kindness of Dr. H. C. Ernst, a rabbit which had been inoculated, by the method of trephining, with material which came originally from Pasteur's laboratory. The rabbit sent me showed the first symptom of paralytic rabies on the eighth day after inoculation. It died on the eleventh day (March 2d, 1887), and I at once proceeded to make the following experiment :

"A portion of the medulla was removed and thoroughly mixed with

¹ Since this was written Brieger has isolated a toxalbumin from cultures of the diphtheria bacillus which is destroyed by a temperature of 60° C., but resists 50°.

sterilized water. The milky emulsion was introduced into four capillary tubes, such as had been used in my experiments heretofore recorded. Two of these tubes were then placed for ten minutes in a water bath, the temperature of which was maintained at 60° C. Four rabbits were now inoculated by trephining, two with the material exposed to 60° C. for ten minutes, and two with the same material from the capillary tube not so exposed. The result was as definite and satisfactory as possible. The two control rabbits were taken sick, one on March 10th and one on the 11th; both died with the characteristic symptoms of paralytic rabies on the third day. The two rabbits inoculated with material exposed to 60° C. remained in perfect health. On the 26th of March one of these rabbits was again inoculated, by trephining, with material from the medulla of a rabbit just dead from hydrophobia. This rabbit died from paralytic rabies on the 8th of April. Its companion remains in perfect health.

"A second experiment was made in the same way on the 14th of March. Two rabbits were inoculated with material exposed for ten minutes to a temperature of 50° C.; two with material exposed to 55° C.; and two control rabbits with material not so exposed. One of the rabbits inoculated with material exposed to 50° C., and one of the control rabbits, died on the 25th; the other rabbit inoculated with the material exposed to 50°, the other control, and one inoculated with material exposed to 55°, on the 26th. The second rabbit inoculated with material exposed to 55° died five days later with the characteristic symptoms of the disease. These experiments show, then, that the virus of hydrophobia is destroyed by a temperature of 60° C., and that 55° C. fails to destroy it, the time of exposure being ten minutes."

The experimental data given show that the pathogenic bacteria tested and different kinds of virus are all killed by a temperature of 60° C. or below; some, like the cholera spirillum and *Micrococcus pneumoniae crouposae*, failing to grow after exposure to as low a temperature as 52° C. for four minutes. By extending the time a still lower temperature will effect the same result. Thus, according to Chauveau, the anthrax bacillus is killed by twenty minutes' exposure to a temperature of 50° C.; and Brieger sterilizes cultures of the diphtheria bacillus, to obtain the soluble toxalbumin produced in them, by exposure for several hours to 50° C. A temperature of 60° has been found to decompose the toxalbumin. The non-pathogenic bacteria tested have, as a rule, a higher thermal death-point—58° C. for *Bacillus prodigiosus*, 64° C. for *Sarcina lutea*, etc.

It is a remarkable fact that certain bacteria not only are not destroyed at higher temperatures than this, but are able to multiply at a temperature of 65° to 70° C. Thus Miquel, in 1881, found in the waters of the Seine a motionless bacillus which grew luxuriantly in bouillon at a temperature of 69° to 70° C. Van Tieghem has also cultivated several different species at about the same temperature, and more recently Globig has obtained from the soil several species which grow at temperatures ranging from 50° to 70° C.

The resisting power of spores to heat also varies in different species; but the spores of known pathogenic bacteria are quickly destroyed by a temperature of 100° C. (212° F.). In the writer's experi-

¹ Report of the Committee on Disinfectants (op. cit.). p. 147.

ments the spores of *Bacillus anthracis* and of *Bacillus alvei* failed to grow after exposure to a temperature of 100° C. for four minutes, and only a few colonies developed after two minutes' exposure to this temperature. The thermal death-point of spores of the "wurtzel bacillus" and of *Bacillus butyricus* (of Hueppe) was the same—100° C. for four minutes.

Schill and Fischer, in 1884, made a number of experiments to determine the thermal death-point of *Bacillus tuberculosis*. They found that five minutes' exposure to a temperature of 100° C. in steam destroyed the vitality of the bacillus in sputum in five minutes. When the time was reduced to two minutes a negative result from inoculation was obtained in two guinea-pigs, but one inoculated at the same time became tuberculous. My own experiments and those of Yersin, made since, lead me to think that there may have been some cause of error in this experiment of Schill and Fischer, and that the thermal death-point of the spores of *Bacillus tuberculosis* is considerably below the boiling point of water. I inoculated guinea-pigs with tuberculous sputum subjected for ten minutes to the following temperatures: 50°, 60°, 70°, 80°, 90° C. The animal inoculated with material exposed to 50° died from tuberculosis at the end of seven weeks. None of the others developed tuberculosis.

Yersin exposed an old culture in glycerin bouillon, in which many of the bacilli contained spores—"très nettes"—to the following temperatures: 55°, 60°, 65°, 70°, 75°, 80°, 85°, 90°, 100° C. "At the end of ten days the bacilli heated to 55° gave a culture in glycerin bouillon; those exposed to 60° grew after twenty-two days; none of the bacilli heated above 70° gave any development. This experiment, repeated a great number of times, has always given us the same result." Voelsh, who has studied the same question, reports as the result of his experiments that the tubercle bacillus in sputum was not destroyed by heating to 100° C. Further experiments will be required to reconcile these contradictory results.

While the spores of the pathogenic bacteria mentioned are destroyed by the boiling point of water within a few minutes, certain non-pathogenic species resist this temperature for hours. Thus Globig obtained a bacillus from the soil the spores of which required five and one-half to six hours' exposure to streaming steam for their destruction. These spores survived exposure for three-quarters of an hour in steam under pressure at from 109° to 113° C. They were destroyed, however, by exposure for twenty-five minutes in steam at 113° to 116°, and in two minutes at 127°.

In the practical application of steam for disinfecting purposes it must be remembered that, while steam under pressure is more effective than streaming steam, it is scarcely necessary to give it the pre-

ference, in view of the fact that all known pathogenic bacteria and their spores are quickly destroyed by the temperature of boiling water ; and also that superheated steam is less effective than moist steam. When confined steam in pipes is "superheated" it has about the same germicidal power as hot dry air at the same temperature. This is shown by the experiments of Esmarch, who found that anthrax spores were killed in streaming steam in four minutes, but were not killed in the same time by superheated steam at a temperature of 141°C .

Desiccation.—Cultures of bacteria kept in a moist condition retain their vitality for a considerable time, which varies greatly with different species. The writer has found that a culture of the typhoid bacillus preserved in a hermetically sealed glass tube retained its vitality for eighteen months, as did also *Bacillus prodigiosus*, *Bacillus cavica*, and some others. According to Kitasato, the cholera spirillum may be preserved in a moist state for seven months ; other bacteria die out in a month or two, but, as a rule, vitality is preserved for several months at least.

Spores in a desiccated condition preserve their vitality for a great length of time. But desiccation is quickly fatal to some of the pathogenic bacteria, and especially so to the cholera spirillum. Koch, in his earlier experiments, found that his "comma bacillus" did not grow after being dried upon a cover glass for three hours. Kitasato, in experiments made since, found that a bouillon culture dried upon a thin glass cover was incapable of development after three hours' time, but that cultures in nutrient agar or gelatin survived for two days, probably on account of the thicker layer formed and the longer time required for complete desiccation. Pfuhl has found that the typhoid bacillus dried upon a cover glass retains its vitality for eight to ten weeks, and Löffler states that the diphtheria bacillus resists desiccation for four or five months. Cadéac and Malet produced tuberculosis in guinea-pigs by injecting material from the lung of a tuberculous cow which had been kept in the form of a dried powder for nearly five months ; at a later date the virulence was lost.

Light.—Downes and Blunt, in a communication made to the Royal Society of London in 1877, first called attention to the fact that light has an injurious effect upon bacteria, and that cultures may be sterilized by exposure to direct sunlight.

Tyndall, in experiments made in the clear sunlight of the Alps, verified the fact that the development of bacteria was restrained in cultures during their exposure, but failed to obtain evidence that vitality was destroyed.

In 1885 Duclaux took up the subject with pure cultures of various

bacteria, and showed that by prolonged exposure to direct sunlight the spores of various bacilli lose their capacity to germinate. About the same time Arloing published his researches upon the influence of light upon the development of anthrax spores. He found that the anthrax bacillus was not restrained in its growth by diffused lamp-light, but its growth was retarded by an intense gaslight. Spore formation was more abundant in darkness than in red light, and more abundant in red than in white light. When a screen was interposed between the culture and the source of light, consisting of an aqueous solution of hæmatoglobin, the growth of the bacilli and of spores was much more luxuriant than in white light. In yellow light it was less abundant than in red. The blue and violet rays were still less favorable for the growth of the bacillus and the development of spores. The pathogenic power of cultures was not especially influenced by exposure to white gaslight. In subsequent experiments with sunlight Arloing found that two hours of exposure to the July sun sufficed to destroy the vitality of anthrax spores, but that a considerably longer exposure (twenty-six to thirty hours) was necessary when the spores had been allowed to germinate in a suitable culture medium. Cultures which were not exposed long enough to destroy the vitality of the bacilli were retarded in their growth, and subsequent exposure for a shorter time (nine to ten hours) completely sterilized them. Cultures which were weakened in their reproductive energy by exposure to sunlight were also "attenuated" as to their pathogenic power and could be used as a vaccine in protective inoculations. According to Arloing, the effect produced results from the action of the full sunlight and cannot be obtained by the use of monochromatic light.

The experiments of Strauss seemed to give support to the view advanced by Nocard that in Arloing's experiments spores did not really exhibit a less degree of resisting power than the vegetating bacilli, but that in fact they commenced to vegetate before they were killed. Strauss placed anthrax spores in sterilized distilled water and in bouillon, and found that, under the same conditions of exposure, the bouillon cultures were sterilized in direct sunlight in nine hours, while the spores suspended in distilled water grew when transferred to a suitable medium. This was accounted for on the supposition that the bouillon furnishes the necessary pabulum for the development of the spores and that distilled water does not.

Arloing combats this view and has published additional experiments which seem to disprove it. He placed small flasks containing anthrax spores in bouillon in the direct rays of the sun in February. Some of the flasks were placed upon a block of ice which reduced the temperature to 4° C.; the others were not so placed, and the tempe-

rature, in the open air where all were exposed, was 11° C. All of the spores failed to grow after an exposure of four hours. When exposed in water the time of exposure was longer.

Roux has shown that the light also has an effect upon the culture medium, and that sterilized bouillon which has been exposed to direct sunlight for some hours restrains the development of anthrax spores subsequently introduced into it, but not of the growing bacilli. His experiments show that access of oxygen is a necessary factor in the sterilization of cultures by sunlight.

In the experiments of Momont (1892) dry anthrax spores were found to resist the action of light for a long time, but moist spores, freely exposed to the air, failed to grow after forty-four hours' exposure to sunlight. In the absence of spores, anthrax bacilli in a moist condition, when freely exposed to the air, failed to grow after exposure to sunlight for half an hour to two hours; but in the absence of air the same bacilli were not destroyed at the end of fifty hours' exposure.

Duclaux, in 1885, experimented upon several different species of micrococci and bacilli, with the following results: Recent cultures of micrococci in bouillon, which preserved their vitality for more than a year in a dark place, were sterilized in July by exposure in the sun for fifteen days, and two species even within two or three days (micrococcus of biskra and micrococcus of pemphigus). Bacilli showed greater resisting power.

Still more recently Gaillard has conducted a series of experiments, under the direction of Arloing, which confirm the results previously obtained by observers heretofore named as to the germicidal power of sunlight in the presence of oxygen.

Geisler (1892), in experiments made upon the typhoid bacillus, found that all portions of the solar spectrum except the red rays exercised a restraining influence upon the development of this bacillus. The electric light gave a similar result. The most decided effect was produced by rays from the violet end of the spectrum. The restraining influence appears, from the researches of Geisler, not to be due solely to the direct action of light upon the development of the bacilli, but also to changes induced in the gelatin culture medium employed in his experiments.

In his address before the International Medical Congress of Berlin, 1890, Koch states that the tubercle bacillus is killed by the action of direct sunlight in a time varying from a few minutes to several hours, depending upon the thickness of the layer exposed. Diffused daylight also has the same effect, although a considerably longer time of exposure is required—when placed close to a window, from five to seven days.

In view of these facts we may conclude, with Duclaux, that sunlight is one of the most potent and one of the cheapest agents for the destruction of pathogenic bacteria, and that its use for this purpose is to be remembered in making practical hygienic recommendations. The popular idea that the exposure of infected articles of clothing and bedding in the sun is a useful sanitary precaution is fully sustained by the experimental data relating to the action of heat, desiccation, and sunlight.

Electricity.—Cohn and Mendelssohn, in 1879, attempted to determine the effect of the galvanic current upon bacteria. Cultures were placed in U-tubes through which a constant current was passed. A feeble current was found to be without effect. A strong current from two elements, maintained for twenty-four hours, restrained development in the vicinity of the positive pole, but this was probably due to the highly acid reaction which the culture liquid acquired. When a current from five elements was used for twenty-four hours the liquid was sterilized, but this may have been due to the decided changes produced in the chemical composition of the culture liquid rather than to the direct action of the galvanic current.

The same may be said of the similar results obtained in later experiments by Apostoli and Laquerrière, and by Prochownick and Spaeth. The last-mentioned investigators found that the positive pole had a more decided effect than the negative, and that the effect depended upon the intensity and duration of the current. A current of fifty milliamperes passed for a quarter of an hour did not kill *Staphylococcus pyogenes aureus*, but a current of sixty milliamperes maintained for the same time did. The spores of *Bacillus anthracis* required a current of two hundred to two hundred and thirty milliamperes during an hour or two. In these experiments the cultures in gelatin were attached to the strips of platinum serving as the two poles, and these were immersed in a solution of sodium chloride. As chlorine was disengaged at the positive pole, the germicidal action is attributed to this gas rather than to the direct action of the current upon the living microorganisms.

The more recent researches of Spilker and Gottstein, made with an induction current from a dynamo machine, are more valuable in estimating the power of this agent to destroy the vitality of bacteria. The current was passed through a spiral wire which was wrapped around a test tube of glass, containing the microorganism to be tested, suspended in distilled water. In a first experiment *Bacillus prodigiosus*, suspended in sterilized distilled water and contained in test tubes having a capacity of two hundred and fifty cubic centimetres, was subjected to a current having an energy of $2.5 \text{ ampères} \times 1.25 \text{ volts}$ for twenty-four hours. The temperature did not go above

30° C. No development occurred when the microörganism tested was subsequently planted in nutrient gelatin. Further experiments gave a similar result. It was found that stronger currents were effective in shorter time; but in no case was sterilization effected in less than an hour.

VII.

ANTISEPTICS AND DISINFECTANTS.

GENERAL ACCOUNT OF THE ACTION OF.

THE term *antiseptic* is used by some authors to designate an agent which destroys the vitality of the microorganisms which produce septic decomposition, and others of the same class. We prefer to restrict the use of the term to those agents which restrain the development of such microorganisms without destroying their vitality. The complete destruction of vitality is effected by *germicides* or disinfectants. Material containing the germs of infectious diseases is infectious material, and we disinfect it by the use of agents which destroy the living disease germs or pathogenic bacteria which give it its infecting power. Such an agent is a disinfectant. But we extend the use of this term to germicides in general—that is, to those agents which kill non-pathogenic bacteria as well as to those which destroy disease germs. All disinfectants are also antiseptics, for agents which destroy the vitality of the bacteria of putrefaction arrest the putrefactive process ; and these agents, in less amount than is required to completely destroy vitality, arrest growth and thus act as antiseptics. But all antiseptics are not germicides. Thus a concentrated solution of salt or of sugar will prevent the putrefactive decomposition of organic material, animal or vegetable ; but these agents do not destroy the vitality of the germs of putrefaction. In a certain degree of concentration they are antiseptics and are largely used for the preservation of meats and vegetables. In the same way many mineral salts in solutions of various strengths act as antiseptics, and some of these in still stronger solutions are disinfectants. Thus mercuric chloride, when introduced into a culture solution in the proportion of 1 : 300,000, will restrain the development of anthrax spores, but to insure the destruction of these spores a solution of 1 : 1,000 must be used. As a rule, the difference between restraining action—antiseptic—and germicidal power—disinfectant—is not so great as this. We give below some recent determinations by Boer which illustrate this point, the test organism being the bacillus of typhoid fever in a culture in bouillon twenty-four hours old :

	Restrains.	Kills.
Hydrochloric acid... ..	1 : 2100	1 : 800
Sulphuric acid... ..	1 : 1550	1 : 500
Silver nitrate... ..	1 : 50000	1 : 4000
Sodium arseniate... ..	1 : 6000	1 : 250
Carbolic acid... ..	1 : 400	1 : 200

Method of Determining Antiseptic Value.—To determine the restraining or antiseptic power of an agent for a particular micro-organism, the agent is dissolved in a definite proportion in a suitable culture medium, which is then inoculated with a pure culture of the test organism and placed in favorable circumstances—as to temperature—for its growth. At the same time a control experiment is made by placing another portion of the same culture medium, inoculated with the same microorganism, in the same conditions, but without the addition of the antiseptic agent. If development occurs in the control experiment and not in the culture medium containing the antiseptic, the failure to grow must be attributed to the presence of this agent. Having made a preliminary experiment, we are guided by the result in further experiments to determine the exact amount required to restrain development under the same conditions. Or we may make a series of experiments in the first instance. The problem being, for example, to determine the antiseptic value of carbolic acid for the typhoid bacillus, we may add this agent to a definite amount of bouillon in test tubes in the proportion of 1 : 100, 1 : 200, 1 : 300, 1 : 400, 1 : 500. In experiments with volatile agents the bouillon, in test tubes or small flasks, must be sterilized in advance, and the antiseptic agent introduced by means of a sterilized pipette with great care to prevent the accidental contamination of the nutrient medium. In experiments with non-volatile agents it will be best to sterilize the culture medium after the antiseptic has been added. Next we inoculate the liquid in each flask with a pure culture of the test organism. The flasks are then placed in an incubating oven at 35° to 37° C. At the same time a control, not containing any carbolic acid, is placed in the oven. At the end of twenty-four hours the control will be found to be clouded, showing an abundant multiplication of the bacillus. Taking the result of Boer above given, we would expect to find all of the solutions clear except that containing 1 : 500. This too might remain clear for some days and finally “break down,” for experience shows that when we pass the point at which a permanent restraining influence is exerted there may be a temporary restraint or retardation of development. For this reason we must continue the experiment for a considerable time—not less

than two weeks. Having found that 1:400 and below prevents development, and 1:500 does not, we may make further experiments to determine the antiseptic power within narrower limits; but this is hardly necessary from a practical point of view.

In these experiments the result will be influenced by several circumstances, as follows:

(a) *By the composition of the nutrient medium.* This is a very important factor, especially in determining the antiseptic value of certain metallic salts. The presence of a considerable quantity of albumin, for example, reduces greatly the antiseptic power of mercuric chloride, silver nitrate, creolin, etc. The presence of a substance chemically incompatible, as, for example, sodium chloride in testing nitrate of silver, will of course neutralize antiseptic action.

(b) *The nature of the test organism.* Within certain limits an antiseptic for one microorganism of this class restrains the development of all, but there are wide differences in the ability of different species to grow in the presence of different chemical agents. Some grow readily in the presence of a considerable amount of free acid, others are restrained by a slightly acid reaction of the medium in which they are placed. The *Bacillus acidi lactici*, for example, can thrive in the presence of a considerable amount of the acid which is a product of its growth, but there is a limit to its power of developing in the presence of this and other acids. So, too, *Micrococcus ureæ*, which causes the alkaline fermentation of urine, grows in the presence of a considerable amount of carbonate of ammonia, but is finally restrained in its growth by this alkaline salt. The following determinations by Boer show the difference in the antiseptic power of hydrochloric acid for certain pathogenic bacteria: *Bacillus* of anthrax (without spores), 1:3,400; diphtheria bacillus, 1:3,400; glanders bacillus, 1:700; typhoid bacillus, 1:2,100; cholera spirillum, 1:5,500. It will be noted that the cholera spirillum is restrained in its growth by about one-eighth the amount of hydrochloric acid which is required to prevent the development of the bacillus of glanders. The typhoid bacillus has a special tolerance for carbolic acid, etc.

(c) *The temperature at which the experiment is made.* At the temperature most favorable for growth a greater proportion of the antiseptic agent is required than at unfavorable temperatures—lower or higher.

(d) *The restraining influence for spores* is much greater than for the vegetative form of bacteria.

Methods of Determining Germicide Value.—The disinfecting power of a chemical agent is determined by allowing it to act for a given time, in a definite proportion, on a pure culture of a given

microorganism, and then testing the question of loss of vitality by culture experiments or by inoculations of infectious disease germs into susceptible animals.

The *test by cultivation* is the most reliable, but in making it several points must be kept in view. Naturally the conditions must be such as are favorable for the growth of the particular microorganism which serves as the test; and we must allow a considerable time for the development of the test organism, for it often happens that its vital activity has been weakened without being completely destroyed, and that growth will occur after an interval of several days, while in the control experiment it has perhaps been seen at the end of twenty-four hours. Another most important point is the fact that some of the disinfecting agent is necessarily carried over with the test organisms when these are transferred to a nutrient medium to ascertain whether they will grow, and this may be in sufficient amount to restrain their development and lead to the mistaken inference that they have been killed. This is especially true of mercuric chloride, which restrains the development of spores in very minute amounts. Spores which have been subjected to its action in comparatively strong solutions, when transferred to a culture medium may fail to grow because of the restraining influence of the mercuric chloride carried over at the same time. For this reason liquid cultures are to be preferred in experiments of this kind. When the test organisms are planted in a solid culture medium the chemical agent is left associated with them; in a liquid culture, on the other hand, it is diluted, and the microorganisms, being distributed through the nutrient medium, have the disinfecting agent washed from their surface. In the case of mercuric chloride, however, the experiments of Geppert show that the agent is so attached to spores which have been subjected to its action that ordinary washing does not suffice. Moreover, spores which have been exposed to the action of mercuric chloride without being killed are restrained in their growth by a much smaller proportion of the corrosive sublimate than is required for spores not so exposed—according to Geppert, by 1 part in 2,000,000. Geppert therefore proposes, in experiments with this agent, to neutralize the mercuric chloride which remains attached to the test organisms by washing these in a solution of ammonium sulphide, by which the sublimate is precipitated as an inert sulphide.

With most agents simple dilution will serve the purpose of preventing an erroneous inference from the restraining influence of the chemical agent being tested. If we carry, by means of a platinum loop, one or two *öse* into five to ten cubic centimetres of bouillon, the dilution will usually be beyond the restraining influence of the

germicidal agent ; but we may carry the dilution still further, to be on the side of safety, by inoculating a second tube containing the same amount of sterile bouillon from the first, carrying over in the same way one or two öse. We will still be very sure to have a considerable number of the microorganisms to test the question of the destruction of vitality. Instead of bouillon we may use liquefied flesh-peptone-gelatin, which gives us the same advantage as to dilution of the disinfecting agent ; and after inoculating two tubes as above indicated, we may make Esmarch roll tubes by turning them upon a block of ice. The development of colonies will show that there was a failure to disinfect ; their absence, after a proper interval, will be evidence of the germicidal action of the agent employed.

Koch's Method.—In 1881 Koch published his extended experiments made to determine the germicidal power of various chemical agents as tested upon anthrax spores. His method consisted in exposing silk threads, to which the dried spores were attached, in a solution of the disinfecting agent, and at intervals transferring one of these threads to a solid culture medium. The precaution was taken to wash the thread in distilled water when the agent tested was supposed to be likely to restrain development. In these experiments a standard solution of the disinfecting agent was used, and the time of exposure was varied from a few hours to many days.

The Writer's Method.—In the writer's experiments, made in 1880 and subsequently, a different method has been adopted. The time has been constant—usually two hours—and the object has been to find the minimum amount of various chemical agents which would destroy the test organisms in this time ; and instead of subjecting a few of the test organisms attached to a silk thread to the action of the disinfecting agent, a certain quantity of a recent culture—usually five cubic centimetres—has been mixed with an equal quantity of a standard solution of the germicidal agent. Thus five cubic centimetres of a 1 : 200 solution of carbolic acid would be added to five cubic centimetres of a recent culture of the typhoid bacillus, for example, and after two hours' contact one or two öse would be introduced into a suitable nutrient medium to test the question of disinfection. In the case given the result obtained would be set down as the action of a solution of carbolic acid in the proportion of 1 : 400, for the 1 : 200 solution was diluted by the addition of an equal quantity of the culture.

Other experimenters have adopted still a different method. Instead of using a considerable and definite quantity of a culture containing the test organism, they introduce one or two öse from such a culture into a solution containing a given proportion of the disinfectant ; then after exposure for a given time the nutrient medium is inoculated.

These different methods give results which cannot be directly compared one with another, for to obtain corresponding results we must have identical conditions.

Test by Inoculation into Susceptible Animals.—In testing the action of disinfectants upon anthrax spores and other infectious disease germs, we may inoculate the microorganisms, after exposure to the disinfectant, into a susceptible animal. This method was adopted by the writer in a series of experiments in 1881, but he has not since employed it, for reasons set forth in his paper giving an account of these experiments.

“*First.* The test organism may be modified as regards reproductive activity without being killed; and in this case a modified form of disease may result from the inoculation, of so mild a character as to escape observation. *Second.* An animal which has suffered this modified form of the disease enjoys protection, more or less perfect, from future attacks, and if used for a subsequent experiment may, by its immunity from the effects of the pathogenic test organism, give rise to the mistaken assumption that this had been destroyed by the action of the germicidal agent to which it had been subjected.”¹

In experiments to determine the value of an agent as a disinfectant, no matter by what method, the following conditions, which influence the result, should be kept in view :

(a) *The difference in vital resisting power of different species of bacteria.* As a rule, the pathogenic species have rather less resisting power than the common saprophytes, and the micrococci have greater resisting power than many of the bacilli. The difference in the vital resisting power of some of the best known pathogenic species is shown in the following table, which we have made up from determinations made by Boer—cultures in bouillon twenty-four hours old ; time of exposure, two hours.

	Hydrochloric Acid.	Caustic Soda.	Chloride of Gold and Sodium.	Nitrate of Silver.	Carbolic Acid.
Anthrax bacillus.....	1 : 1100	1 : 450	1 : 8000	1 : 20000	1 : 300
Diphtheria bacillus.....	1 : 700	1 : 300	1 : 1000	1 : 2500	1 : 300
Glanders bacillus	1 : 200	1 : 150	1 : 400	1 : 4000	1 : 300
Typhoid bacillus	1 : 800	1 : 190	1 : 500	1 : 4000	1 : 200
Cholera spirillum.....	1 : 1350	1 : 150	1 : 1000	1 : 4000	1 : 400

(b) *The presence or absence of spores.* The reproductive elements known as spores have a far greater resisting power to chemical agents, as well as to heat, than have the vegetative cells. In

¹ Quoted from article on “Germicides and Disinfectants,” in “Bacteria,” p. 212.

practical disinfection, therefore, it is important to know what disease germs form spores and what do not. The following are known to form spores: The bacillus of anthrax, the bacillus of tetanus, the bacillus of malignant œdema, the bacillus of symptomatic anthrax, the bacillus of foul brood (infectious disease of bees). The following, so far as is known, do not form spores: The pus cocci (*Staphylococcus pyogenes albus*, *aureus*, and *citreus*, and *Streptococcus pyogenes*), the micrococcus of pneumonia, the bacillus of typhoid fever, the bacillus of glanders, the bacillus of diphtheria, the spirillum of cholera, the spirillum of relapsing fever.

Many agents which kill the growing bacteria are incapable of destroying the vitality of spores, and others only do so in much stronger solutions or after a long exposure to their action.

(c) *The number of bacteria to be destroyed.* This is an essential factor which has often been overlooked by those making experiments. To destroy the bacteria carried over to five cubic centimetres of distilled water by means of a platinum loop, is a very different matter from destroying the immensely greater number in five cubic centimetres of a recent bouillon culture.

(d) *The nature and quantity of associated material.* The oxidizing disinfectants, like permanganate of potash and chloride of lime, not only act upon the bacteria, destroying them by oxidation, but upon all organic matter with which they come in contact, and at the same time the disinfecting agent is destroyed in the chemical reaction, which is a quantitative one. The presence, therefore, of organic material in association with the bacteria is an important factor, and if this is in excess the disinfectant may be neutralized before the living bacteria are destroyed. Other substances which precipitate the disinfecting agent in an insoluble form, or decompose it, must of course have the same effect. Thus the presence of sodium chloride in a culture medium would be an important circumstance if nitrate of silver was the agent being tested, as the insoluble chloride would be precipitated. And in the case of mercuric chloride and certain other metallic salts the presence of albumin very materially influences the result. Van Ermengem states that the cholera spirillum in bouillon is destroyed in half an hour by mercuric chloride in the proportion of 1:60,000, while in blood serum 1:800 was required to destroy it in the same time.

(e) *The time of exposure* is also an important factor. Some agents act very promptly, while others require a considerable time to effect the destruction of bacteria exposed to their action. Thus a solution of chloride of lime containing 0.12 per cent destroys the typhoid bacillus and the cholera spirillum in five minutes, and the anthrax bacillus in one minute (Nissen). On the other hand,

quicklime (milk of lime) requires a contact of several hours to insure the destruction of pathogenic bacteria.

(f) *The temperature at which the exposure is made* has a material influence upon the result. This is shown by the experiments of Henle and of Nocht.

(g) *The degree of dilution of the disinfecting agent* is also a matter of importance. This is especially true of solutions of acids and alkalies. When a silk thread to which bacteria are attached is suspended in an acid solution the essential point is the degree of acidity, and not the quantity of acid in the entire solution. But if a solution of permanganate of potash, or any other active oxidizing agent, is used, the principal question is not the degree of dilution, but the amount of the disinfecting agent present in the solution used. A grain of potassium permanganate dissolved in two fluidounces of distilled water would probably kill just as many bacteria as if it were dissolved in half a fluidounce, although the time required for disinfection might be longer.

From what has been said it is evident that the simple statement that a certain agent is a germicide in a certain proportion has but little scientific value, unless we are made acquainted with the conditions under which its germicidal action has been tested.

VIII.

ACTION OF GASES AND OF THE HALOID ELEMENTS UPON BACTERIA.

Oxygen.—Free oxygen is essential for the development of a large number of species of bacteria—aërobics ; and it completely prevents the growth of others—anaërobics. Many bacteria, even when freely exposed in a desiccated condition to the action of atmospheric oxygen, retain their vitality for a long time. The gradual loss of pathogenic power which Pasteur has shown occurs in cultures of the anthrax bacillus and the micrococcus of fowl cholera, is ascribed by him to exposure to oxygen, and as proof of this he states that cultures kept in hermetically sealed tubes do not lose their virulence in the same degree. But other circumstances may influence the result. Thus some of the products of growth which accumulate in culture fluids have an injurious effect upon the vitality of the bacteria which produced them, and in time may cause a complete destruction of vitality. In cultures exposed to the air these products would be in a more concentrated solution from the gradual evaporation of the culture liquid. It must also be remembered that light in the presence of oxygen is a germicidal agent.

The experiments of Fränkel show that the aërobic bacteria grow abundantly in the presence of pure oxygen, and some species even more so than in ordinary air. *Micrococcus prodigiosus*, however, appeared to be unfavorably affected by pure oxygen, inasmuch as it did not produce pigment so readily as when cultivated in ordinary air.

Nascent oxygen is a very potent germicidal agent, as will be seen in our account of such oxidizing disinfectants as potassium permanganate and the hypochlorite of lime.

Ozone.—It was formerly supposed that ozone would prove to be a most valuable agent for disinfecting purposes ; but recent experiments show that it is not so active a germicide as was anticipated, and that from a practical point of view it has comparatively little value.

Lukaschewitsch found that one gramme in the space of a cubic metre failed to kill anthrax spores in twenty-four hours. The cholera spirillum in a moist state was killed in this time by the same amount, but fifteen hours' exposure failed to destroy it. Ozone for these experiments was developed by means of electricity.

Wyssokowicz found that the presence of ozone in a culture medium restrained the development of the anthrax bacillus, the bacillus of typhoid fever, and others tested, but concludes that this is rather due to the oxidation of bases contained in the nutrient medium than to a direct action upon the pathogenic bacteria.

Sonntag, in his carefully conducted experiments, in which a current of ozonized air was made to pass over silk threads to which were attached anthrax spores, had an entirely negative result. The anthrax bacillus from the spleen of a mouse, and free from spores, was then tested, also with a negative result, even after exposure to the ozonized air for twenty minutes at a time on four successive days. In another experiment several test organisms (*Bacillus anthracis*, *Bacillus pneumoniae* of Friedländer, *Staphylococcus pyogenes aureus*, *Staphylococcus pyogenes albus*, *Bacillus murisepticus*, *Bacillus crassus sputigenus*) were exposed on silk threads for twenty-four hours in an atmosphere containing 4.1 milligrammes of ozone to the litre of air (0.19 volumes per cent). The result was entirely negative. When the amount was increased to 13.53 milligrammes per litre the anthrax bacillus and *Staphylococcus pyogenes albus* failed to grow after twenty-four hours' exposure. The conclusion reached by Nissen, from his own experiments and a careful consideration of those previously made by others, is that ozone is of no practical value as a germicide in therapeutics or disinfection.

Hydrogen.—This gas has no injurious effect upon bacteria, as is shown by the fact that the anaërobic and facultative anaërobic species grow readily in an atmosphere of pure hydrogen.

Hydrogen peroxide in solution in water is a valuable antiseptic and deodorant, but its value as a germicide has been very much overestimated. Miquel, in his experiments to determine the antiseptic value of various agents, places H_2O_2 third in the list of "substances eminently antiseptic," and states that it prevents the development of the bacteria of putrefaction in the proportion of 1:20,000.

In the writer's experiments (1885) a solution was used which contained at first 4.8 per cent of H_2O_2 , and five per cent of sulphuric acid which was added by the chemist who prepared the solution, to prevent loss of the hydrogen peroxide. At the end of a month the amount of H_2O_2 was again estimated, and found to be 3.98 per cent. Five weeks later the proportion was 2.4 per cent. Tested upon "broken-down" beef tea, this solution was found to destroy the vitality of the bacteria of putrefaction contained in it, in two hours' time, in the proportion of thirty per cent (about 1.2 per cent of H_2O_2). Anthrax spores were killed in the same time by a twenty-per-cent solution (0.8 per cent H_2O_2). Tested upon a pure culture of pus cocci, it was active in the proportion of ten per cent (0.4 per cent of

H_2O_2); a solution containing 0.24 per cent of H_2O_2 failed to kill pus cocci. But the solution used in these experiments contained also five per cent of sulphuric acid, which by itself kills micrococci in the proportion of 1:200. My conclusion was that, unless the chemists can furnish more concentrated solutions which will keep better than that with which I experimented, we are not likely to derive any practical benefit from the use of hydrogen peroxide as a disinfectant.

Altehofer more recently has experimented with a solution containing 9.7 per cent of H_2O_2 , and reports the following results: He added to ninety-eight cubic centimetres of hydrant water two cubic centimetres of a bouillon culture of the typhoid bacillus, and to this was added sufficient of his aqueous solution of H_2O_2 to make the proportion present 1:1,000. At the end of twenty-four hours the bacillus was proved by culture experiments to be killed. Water containing the cholera spirillum, treated in the same way, was not entirely sterilized, as a few colonies developed in Esmarch roll tubes; but the general result of his experiments was that the ordinary water bacteria, and the pathogenic bacteria named (cholera, typhoid) when suspended in water, required for their destruction exposure for twenty-four hours in a solution containing one part of H_2O_2 in one thousand of water.

Carbon Dioxide.—The experiments of Fränkel show that certain bacteria grow in an atmosphere of CO_2 , as well as in the air; among these are the bacillus of typhoid fever and the pneumonia bacillus of Friedländer. Other species are slightly restricted in their growth, e.g. *Bacillus prodigiosus*, *Proteus vulgaris*. Still others grow only when the temperature is elevated, including the pus cocci and the bacillus of swine pest. Most of the saprophytic bacteria failed to grow in an atmosphere of CO_2 , although their vitality was not destroyed by it. Certain pathogenic species were, however, killed by the action of this gas, among others the cholera spirillum, *Bacillus anthracis*, and *Staphylococcus pyogenes aureus*.

Leone and Hochstetter had previously reported that certain bacteria are injuriously affected by CO_2 . Fränkel also found that the growth of strictly anaërobic species was restricted in an atmosphere of carbon dioxide. The aërobic species which failed to grow in pure CO_2 grew abundantly when a little atmospheric oxygen was admitted. In the experiments of Frankland the cholera spirillum and the Finkler-Prior spirillum failed to develop in an atmosphere of CO_2 , and at the end of eight days were no longer capable of growth when the carbon dioxide was replaced with atmospheric air.

Carbonic Oxide.—Frankland's experiments show that an atmosphere of this gas is not favorable to the growth of the cholera spirillum or of the Finkler-Prior spirillum, although it did not entirely

prevent development, and after seven days' exposure the spirilla were not all killed, although a comparatively small number of colonies developed. *Bacillus pyocyaneus* failed to grow in an atmosphere of CO, but when air was admitted, at the end of seven or eight days, abundant development occurred.

Methane, CH₄.—We have no exact experiments to determine the action of marsh gas in a pure state on bacteria, but the experiments of Kladakis upon illuminating gas may be taken as representing approximately what might be expected from exposure in pure CH₄. An analysis of the gas used in his experiments showed it to contain 37.97 per cent of hydrogen, 39.37 per cent of methane (CH₄), 9.99 per cent of nitrogen, 4.29 per cent of ethene (C₂H₄), 3.97 per cent of carbonic oxide (CO), 0.61 per cent of oxygen, and 0.41 per cent of carbon dioxide. As hydrogen and nitrogen are neutral, and carbonic oxide is shown by the experiments of Frankland not to act as a germicide after several days' exposure to its action, the positive results obtained in the experiments of Kladakis may be ascribed to the presence of CH₄ (39.37 per cent) or of C₂H₄ (4.29 per cent), or of both together.

A large number of microorganisms were tested, and among these *Proteus vulgaris* alone grew in an atmosphere of illuminating gas. The others not only failed to grow in such an atmosphere, but were destroyed by it. Cultures of *Bacillus anthracis*, *Staphylococcus pyogenes aureus*, and *Spirillum cholerae Asiaticæ* were sterilized in half an hour by the action of this gas. The gas was also found to be unsuitable for anaerobic cultures.

Nitrous Oxide, N₂O.—The experiments of Frankland, made upon the cholera spirillum, the spirillum of Finkler-Prior, and the bacillus of green pus, gave results similar to those obtained with CO, viz., seven days' exposure in an atmosphere of this gas failed to destroy the test organisms, but completely restrained the growth of *Bacillus pyocyaneus* and interfered materially with the development of the two species of spirillum without entirely preventing it.

Nitrogen Dioxide, NO.—Frankland found that his test organisms were quickly killed by this gas (*Bacillus pyocyaneus*, *Spirillum cholerae Asiaticæ*, *Spirillum Finkler-Prior*).

Hydrosulphuric Acid, H₂S.—In the experiments of Frankland this gas proved to be quickly fatal to the bacteria tested (*Bacillus pyocyaneus*, *Spirillum cholerae Asiaticæ*, *Spirillum Finkler-Prior*). On the other hand, Grauer found that this gas did not exercise any injurious influence upon the tubercle bacillus, the bacillus of anthrax, the typhoid bacillus, or the cholera spirillum, after the exposure of these microorganisms in a current of the gas for an hour.

It has been shown by the experiments of Holschewnikoff and

others that certain species of bacteria cause an abundant evolution of H_2S as a result of their development in an albuminous medium (*Bacillus sulfureus* and *Proteus sulfureus*).

Sulphur Dioxide, SO_2 .—Very numerous experiments have been made with this gas, owing to the fact that it has been extensively used in various parts of the world for the disinfection of hospitals, ships, apartments, clothing, etc.

In the writer's experiments, made in 1880, dry vaccine virus on ivory points was disinfected by exposure for twelve hours in an atmosphere containing one volume per cent of this gas, and liquid virus, exposed in a watch glass, by one-third of this amount. Subsequent experiments (1885) showed that pus micrococci were killed by exposure for eighteen hours in a dry atmosphere containing twenty volumes per cent of SO_2 , but that four volumes per cent failed. In the presence of moisture this gas has considerably greater germicidal power than this, owing, no doubt, to the formation of the more active agent, sulphurous acid (H_2SO_3). But in a pure state anhydrous sulphur dioxide does not destroy spores. The writer has shown that the spores of *Bacillus anthracis* and *Bacillus subtilis* are not killed by contact for some time with liquid SO_2 (liquefied by pressure). Koch exposed various species of spore-bearing bacilli in a disinfection chamber for ninety-six hours, the amount of SO_2 at the outset of the experiment being 6.13 volumes per cent, and at the end 3.3 per cent. The result was entirely negative.

But in the absence of spores the anthrax bacillus, in a moist condition, attached to silk threads, was destroyed in thirty minutes in an atmosphere containing one volume per cent.

In another of Koch's experiments the amount of SO_2 in the disinfection chamber was at the outset 0.84 per cent, and at the end of twenty-four hours 0.55 per cent. An exposure of one hour in this atmosphere killed anthrax bacilli attached to silk threads, in a moist condition; but four hours' exposure failed to kill *Bacillus prodigiosus* growing on potato, while twenty-four hours' exposure was successful. A similar result was obtained with *Bacillus pyocyaneus*.

Thinot, as a result of experiments made in 1890, arrives at the conclusion that the specific germs of tuberculosis, glanders, farcy of cattle, typhoid fever, cholera, and diphtheria are destroyed by twenty-four hours' exposure in an atmosphere containing SO_2 developed by the combustion of sixty grains of sulphur per cubic metre. This amount corresponds closely with that fixed by the Committee on Disinfectants of the American Public Health Association on the experimental evidence obtained by the writer in 1885. But the committee insisted upon the presence of moisture and made the time of exposure twelve hours—"exposure for twelve hours to an atmosphere con-

taining at least four volumes per cent of this gas in the presence of moisture."

Chlorine.—The haloid elements are active germicidal agents, especially chlorine on account of its affinity for hydrogen, and the consequent release of nascent oxygen when it comes in contact with microorganisms in a moist condition. And for the same reason this agent is a much more active germicide in the presence of moisture than in a dry condition. The experiments of Fischer and Proskauer showed that when dried anthrax spores were exposed for an hour in an atmosphere containing 44.7 per cent of dry chlorine they were not destroyed; but if the spores were previously moistened and were exposed in a moist atmosphere for the same time, four per cent was effective, and when the time was extended to three hours one per cent destroyed their vitality. The anthrax bacillus, in the absence of spores, was killed by exposure in a moist atmosphere containing 1 part to 2,500, the time of exposure being twenty-four hours, and the same amount was effective for *Micrococcus tetragenus*; the streptococcus of erysipelas and the micrococcus of fowl cholera were killed in three hours by 1:2,500, and in twenty-four hours by 1:25,000. The bacillus of mouse septicæmia and the tubercle bacillus were killed in one hour by 1:200.

In the writer's experiments (1880) four children were vaccinated with virus from ivory points which had been exposed for six hours in an atmosphere containing one-half per cent of chlorine; also with four points, from the same lot, not disinfected. Vaccination was unsuccessful in every case with the disinfected points, and successful with those not disinfected. Koch found that anthrax spores failed to grow after twenty-four hours' exposure in chlorine water. In the experiments of De la Croix to determine the antiseptic power of this agent, it was found that when present in unboiled beef infusion in the proportion of 1:15,600 no development of bacteria occurred. Miquel gives the antiseptic value of chlorine as 1:4,000.

Chloroform.—Immersion for one hundred days in chloroform does not destroy the vitality of anthrax spores (Koch). This agent is without effect on the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). Salkowski found that the anthrax bacillus in the absence of spores, and the cholera spirillum, were killed by being immersed in chloroform water for half an hour. Kirchner reports still more favorable results. In his experiments a one-per-cent solution killed the cholera spirillum in less than a minute, and a one-quarter-per-cent solution in an hour. But the typhoid bacillus required at least one-half per cent acting for an hour.

Iodine.—In the writer's experiments (1880) iodine in aqueous solution with potassium iodide was found to be fatal to *Micrococcus*

pneumonise crouposa in the proportion of 1 : 1,000, and to the staphylococci of pus in 1 : 500—time of exposure two hours. Iodine water was found by Koch to destroy the vitality of anthrax spores in twenty-four hours, but a two-per-cent solution in alcohol failed to destroy anthrax spores in forty-eight hours. In the experiments of Schill and Fischer twenty hours' contact with a solution of the strength of 1 : 500 failed to destroy the virulence of tuberculous sputum, as tested by inoculation experiments. The antiseptic value of iodine is given by Miquel as 1 : 4,000.

Bromine.—Fischer and Proskauer have studied the action of bromine vapor upon various microorganisms. They found that exposure for three hours in a dry atmosphere to three per cent does not destroy the tubercle bacillus in sputum or the spores of anthrax. But when the atmosphere is saturated with moisture 1 : 500 is effective; and when the time of exposure was extended to twenty-four hours, 1 : 3,500. A two-per-cent solution destroys the vitality of anthrax spores in twenty-four hours (Koch). Bromine vapor is an active agent for the destruction of the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). Miquel gives the antiseptic value of bromine as 1 : 1,666, which is considerably below that of chlorine and iodine.

Iodine Trichloride.—According to Behring, we possess in this agent a disinfectant which possesses the potency of free chlorine and iodine without having their disadvantages. As prepared by O. Riedel it is a yellowish-red powder of penetrating odor. It remains unchanged for weeks in concentrated aqueous solution (five per cent). A one-per-cent solution destroys anthrax spores suspended in water almost instantly, and a 0.2-per-cent solution within a few minutes. Anthrax spores in blood serum are killed by a one-per-cent solution in forty minutes (Behring). Langenbuch found that a solution of 1 : 1,000 kills spores in a short time, and that when added to nutrient gelatin in the proportion of 1 : 1,200 it restrains the development of bacteria.

Iodoform.—Numerous experiments have been made with this agent, which show that it has little, if any, germicidal power; but it acts to some extent as an antiseptic. Tilanus reports that the tubercle bacillus will not grow in glycerin-agar cultures to which a small quantity of iodoform has been added, and that a pure culture of the tubercle bacillus was not killed in six days by exposure to iodoform vapor, but that after six weeks' exposure it failed to grow. The experiments of Neisser and of Buchner show that while most bacteria are not injuriously affected by exposure to iodoform vapor, the cholera spirillum and the Finkler-Prior spirillum are restrained in their growth by such exposure. When plate cultures of the cholera

spirillum were placed under a bell jar beside iodoform powder no development occurred, but when they were removed colonies developed, showing that the spirilla were not killed.

Iodoform Ether, according to Yersin, is fatal to the tubercle bacillus in one-per-cent solution in five minutes. Cadéac and Meunier found that a saturated solution required thirty-six hours to kill the bacillus of typhoid fever.

Iodol.—In experiments made by the writer (1885) this agent was found to be without germicidal power. Riedlin found it without any action, even upon the cholera spirillum.

Hydrofluoric Acid, HF_l.—From a series of experiments made with this gas, Grancher and Chautard arrive at the conclusion that "the direct and prolonged action of hydrofluoric acid upon the tubercle bacillus diminishes its virulence but does not kill it."

IX.

ACTION OF ACIDS AND ALKALIES.

Sulphuric Acid, H_2SO_4 .—The experiments of Koch (1881) showed that anthrax spores were still capable of growing after exposure in a one-per-cent solution of sulphuric acid for twenty days. In the writer's experiments (1885) a four-per-cent solution failed to destroy the spores of *Bacillus subtilis* in four hours, and an eight-per-cent solution was found to be required for the sterilization of culture fluids containing spores; but the multiplication of the bacteria of putrefaction was prevented by the presence of this acid in a culture solution in the proportion of 1 : 800. Pus micrococci were destroyed by exposure for two hours in a solution containing 1 : 200.

The experiments of Boer show that there is a considerable difference in the resisting power of different pathogenic bacteria. The time of exposure being two hours, cultures in bouillon twenty-four hours old gave the following results :

	Restrains development.	Destroys vitality.
Anthrax bacillus.....	1 : 2550	1 : 1300
Diphtheria bacillus... ..	1 : 2050	1 : 500
Glanders bacillus.....	1 : 750	1 : 200
Typhoid bacillus.....	1 : 1550	1 : 500
Cholera spirillum	1 : 7000	1 : 1300

Leitz, in his studies relating to the bacillus of typhoid fever, reports the following results : The dejections of typhoid patients, mixed with an equal proportion of the disinfecting solution, were sterilized by a five-per-cent solution of sulphuric acid in three days. A pure culture was sterilized in fifteen minutes by two per cent, and in five minutes by five per cent.

Sulphurous Acid, H_2SO_3 .—In the writer's experiments (1885) micrococci were destroyed in two hours by 1 : 2,000 by weight of SO_2 added to water. Kitasato found that solutions of sulphurous acid in the proportion of 0.28 per cent killed the typhoid bacillus, and 0.148 per cent the cholera spirillum. De la Croix found that one

gramme of SO_2 added to two thousand of bouillon prevents the development of putrefactive bacteria and after a time destroys the vitality of these bacteria. The writer found that pus cocci failed to grow in a culture solution containing one part of SO_2 in five thousand of water.

Nitric Acid, HNO_3 .—In the writer's experiments an eight-per-cent solution which contained 0.819 gramme of HNO_3 in each cubic centimetre sterilized broken-down beef tea containing spores, and five per cent failed to do so. Kitasato, in experiments upon the cholera spirillum and typhoid bacillus, obtained results corresponding with those obtained with hydrochloric acid—0.2 per cent destroyed vitality at the end of four or five hours. In these experiments the acid used contained 0.35 gramme HNO_3 in one cubic centimetre.

Nitrous Acid.—In the writer's experiments on vaccine virus (1880) exposure for six hours in an atmosphere containing one per cent of nitrous acid destroyed the virulence of dried virus upon ivory points.

Hydrochloric Acid, HCl .—Anthrax spores are destroyed in ten days by a two-per-cent solution, but not in five days (Koch). Tested upon broken-down beef tea containing spores of *Bacillus subtilis*, it was effective in two hours in the proportion of fifteen per cent, but failed in ten per cent (Sternberg). In the experiments of Kitasato this acid destroyed the typhoid bacillus in five hours in the proportion of 0.2 per cent, and the cholera spirillum in 0.132 per cent—the acid used contained 0.26 gramme HCl in one cubic centimetre. We give the more recent determinations of Boer in tabular form. Its germicidal power was tested upon bouillon cultures which had been kept for twenty-four hours in an incubating oven; time of exposure to the action of the acid solution, two hours.

	Restrains development.	Destroys vitality.
Anthrax bacillus	1:3400	1:1100
Diphtheria bacillus.....	1:3400	1:700
Glanders bacillus.....	1:700	1:200
Typhoid bacillus	1:2100	1:300
Cholera spirillum	1:5500	1:1350

Chromic Acid.—In Koch's experiments a one-per-cent solution destroyed anthrax spores in from one to two days. In the proportion of 1:5,000 it prevents the development of putrefactive bacteria (Miquel).

Osmic Acid.—A solution of one per cent kills anthrax spores in twenty-four hours (Koch). It is an antiseptic in the proportion of 1:6,666 (Miquel).

Phosphoric Acid.—Exposure for four or five hours to a solution

containing 0.3 per cent destroys the typhoid bacillus, and 0.183 per cent the cholera spirillum (Kitasato). The acid used contained 0.152 gramme H_3PO_4 in one cubic centimetre.

Acetic Acid.—A five-per-cent solution failed to kill anthrax spores after five days' exposure (Koch). In Abbott's experiments glacial acetic acid in fifty-per-cent solution failed in two hours to kill anthrax spores, but micrococci were killed by two hours' exposure to a one-per-cent solution. A solution of 1 : 300 of glacial acetic acid destroys the cholera spirillum in half an hour (Van Ermengem). In the proportion of 0.25 per cent it restrains the growth of the typhoid bacillus, and 0.3 per cent destroys its vitality after five hours' exposure ; the cholera spirillum fails to grow in presence of 0.132 per cent and is destroyed by 0.2 per cent (Kitasato).

Lactic Acid.—The bacillus of typhoid fever is killed in five hours by a solution containing 0.4 per cent, the cholera spirillum by 0.3 per cent (Kitasato).

Citric Acid.—The bacillus of typhoid fever is killed in five hours by 0.43 per cent, the cholera spirillum by 0.3 per cent (Kitasato). The cholera spirillum is killed in half an hour by 1 : 200 (Van Ermengem).

Oxalic Acid.—The typhoid bacillus requires a solution of 0.36 per cent, the cholera spirillum one of 0.28 per cent, to destroy vitality in five hours (Kitasato).

Boracic Acid.—In the writer's experiments (1883) a saturated solution failed to kill pus cocci in two hours. A five-per-cent solution failed to destroy anthrax spores in five days (Koch). The typhoid bacillus is killed in five hours by 2.7 per cent, the cholera spirillum by 1.5 per cent (Kitasato). According to Arloing, Cornevin, and Thomas, the fresh virus of symptomatic anthrax requires exposure to a twenty-per-cent solution for forty-eight hours for the destruction of vitality. Boracic acid acts as an antiseptic in the proportion of 1 : 143 (Miquel).

Salicylic Acid.—In the writer's experiments this agent was dissolved by the addition of sodium bichromate, which by itself has no germicidal power. A two-per-cent solution was found to destroy pus cocci in two hours. Dissolved in oil or in alcohol a five-per-cent solution does not destroy anthrax spores (Koch). Micrococci are destroyed by solutions containing 1 : 400 (Abbott). The typhoid bacillus is killed in five hours by 1.6 per cent, the cholera spirillum by 1.3 per cent (Kitasato). A one-per-cent solution destroys *Micrococcus Pasteuri* in half an hour (Sternberg). It is an antiseptic in the proportion of 1 : 1,000 (Miquel). A solution of 2.5 per cent kills the tubercle bacillus in six hours (Yersin). In the proportion of 1 : 300 it destroys the cholera spirillum in half an hour (Van Ermengem).

Benzoic Acid.—According to Miquel, this acid restrains the development of putrefactive bacteria when present in bouillon in the proportion of 1:909. In the proportion of 1:2,000 it retards the development of anthrax spores (Koch).

Formic Acid.—The typhoid bacillus is restrained in its growth by 0.25 per cent, and is killed in five hours by 0.35 per cent, the cholera spirillum by 0.22 per cent (Kitasato).

Tannic Acid.—A solution of one per cent kills *Micrococcus Pasteuri* in the blood of a rabbit in half an hour (Sternberg). A five-per-cent solution failed in ten days to destroy anthrax spores (Koch). A twenty-per-cent solution failed in two hours to destroy the vitality of spores of the anthrax bacillus or of *Bacillus subtilis* (Abbott). Micrococci are destroyed by 1:400, and 1:800 failed (Abbott). A twenty-per-cent solution has no effect upon the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). A solution of 1.66 per cent kills the typhoid bacillus in five hours, and 1.5 per cent the cholera bacillus in the same time (Kitasato). It restrains the development of putrefactive bacteria in the proportion of 1:207 (Miquel).

Tartaric Acid.—A twenty-per-cent solution of this acid fails, after two hours' exposure, to destroy the spores of *Bacillus anthracis* or *Bacillus subtilis*. Micrococci are killed by two hours' exposure in a solution containing 1:400 (Abbott).

Malic Acid.—This was found by Kitasato to correspond with citric acid in its germicidal power.

Valerianic Acid.—A five-per-cent solution in ether failed in five days to destroy anthrax spores (Koch).

Oleic Acid.—A solution of five per cent in ether does not destroy anthrax spores in five days (Koch).

Thymic Acid.—In the proportion of 1:500 this acid prevents the putrefactive decomposition of beef tea (Miquel).

Butyric Acid.—Five days' immersion in this acid failed to destroy anthrax spores (Koch).

Arsenious Acid.—A one-per-cent solution destroys the vitality of anthrax spores in ten days, but failed to do so in six days (Koch). In the proportion of 1:166 it prevents putrefactive changes in bouillon (Miquel).

Gallic Acid.—Abbott found this acid to destroy the bacteria in broken-down beef tea in the proportion of 2.37 per cent, but it failed to destroy anthrax spores in two hours in the same proportion. Micrococci were killed in two hours by 1:142, while 1:250 failed.

ALKALIES.

Potassium Hydroxide, KHO.—In the writer's experiments a ten-per-cent solution of caustic potash was fatal to pus cocci, and an

eight-per-cent solution failed—two hours' exposure. Exposure for twenty-four hours to a ten-per-cent solution failed to kill the tubercle bacillus (Schill and Fischer). A solution of one per cent kills the anthrax bacillus, the bacillus of rothlauf, and several others (Jäger). The addition of 0.14 per cent restrains the development of the typhoid bacillus, and 0.18 per cent kills this bacillus in four or five hours; the cholera spirillum failed to grow in cultures containing 0.18 per cent and was killed by 0.237 per cent in the same time (Kitasato).

Sodium Hydroxide, NaHO .—The experiments of Jäger and of Kitasato show that soda has about the same germicidal power as caustic potash. Boer obtained the following results with bouillon cultures after two hours' exposure: Anthrax bacillus, 1:450; diphtheria bacillus, 1:300; glanders bacillus, 1:150; typhoid bacillus, 1:190; cholera spirillum, 1:150. In about one-half the amount required to destroy vitality the development of the above-named bacteria was prevented. In the proportion of 1:56 it acts as an anti-septic (Miquel).

Ammonia, NH_3 .—In Kitasato's experiments the typhoid bacillus was destroyed in five hours by 0.3 per cent of NH_3 , and the cholera spirillum by about the same amount. Boer obtained the following results, the time of exposure being two hours: Anthrax bacillus, 1:300; diphtheria bacillus, 1:250; glanders bacillus, 1:250; typhoid bacillus, 1:200; cholera spirillum, 1:350. The growth of the anthrax bacillus and of the diphtheria bacillus in culture solutions was prevented by 1:650.

Calcium Hydroxide, Ca_2HO .—According to Kitasato, the typhoid bacillus and the cholera spirillum, in bouillon cultures, are killed in four or five hours by the addition of 0.1 per cent of calcium oxide. Liborius had previously reported still more favorable results, but his bouillon cultures were largely diluted with distilled water. From a practical point of view the experiments of Pfuhr are more valuable. Calcium hydrate was added to the dejections of typhoid patients. When added in the proportion of three per cent sterilization was effected in six hours, and by six per cent in two hours. When milk of lime containing twenty per cent of calcium hydrate was used the results were still more favorable, the typhoid bacillus and cholera spirillum being killed in one hour by the addition of two per cent of the disinfectant. The practical value of lime-wash applied to walls has been determined by Jäger. Silk threads soaked in cultures of various pathogenic bacteria were attached to boards and the lime-wash applied with a camel's-hair brush. Anthrax bacilli (without spores), the glanders bacillus, *Staphylococcus pyogenes aureus*, and several other pathogenic bacteria were killed by a single application after twenty-four hours, but the tubercle bacillus was not

killed by three successive applications. In the writer's experiments (1885) the typhoid bacillus and *Staphylococcus pyogenes aureus* were killed in two hours by a solution containing 1:40 of calcium oxide, and 1:80 failed. Spores of the anthrax bacillus and of several other spore-forming species were not killed by two hours' exposure to a milk of lime containing twenty per cent of calcium oxide.

X.

ACTION OF SALTS.

WHILE some of the metallic salts, and especially those of mercury, silver, and gold, have remarkable germicidal power, others, even in concentrated solutions, do not destroy the vitality of bacteria exposed to their action. For convenience of reference we shall consider the agents in this group in alphabetical order, but first we give Miquel's tables of antiseptic value. This author recognizes the importance of experiments to determine the restraining power of chemical agents for various species of pathogenic bacteria, but says: "As to me, faithful to a plan I adopted at the outset, I will treat the subject in a more general manner by making known simply the minimum weight of the substances capable of preventing the evolution of any bacteria or germs. The method adopted is very simple. To a liquid always comparable to itself it is sufficient at first to add a known weight of the antiseptic and some atmospheric germs or adult bacteria, and to vary the quantity of the antiseptic until the amount is ascertained which will preserve indefinitely the liquid from putrefaction. In order to obtain germs of all kinds in a dry state it suffices to take them, where they are most abundant, in the dust collected in the interior of houses or of hospitals; and to procure a variety of adult bacteria we may take the water of sewers."

SUBSTANCES EMINENTLY ANTISEPTIC.

	Efficient in the proportion of—
Mercuric iodide,	1: 40000
Silver iodide,	1: 33000
Hydrogen peroxide,	1: 20000
Mercuric chloride,	1: 14300
Silver nitrate,	1: 12500

SUBSTANCES VERY STRONGLY ANTISEPTIC.

Osmic acid,	1: 6666
Chromic acid,	1: 5000
Chlorine,	1: 4000
Iodine,	1: 4000
Chloride of gold,	1: 4000
Bichloride of platinum,	1: 3333
Hydrocyanic acid,	1: 2500

Bromine,	1:1666
Cupric chloride,	1:1428
Thymol,	1:1340
Cupric sulphate,	1:1111
Salicylic acid,	1:1000

SUBSTANCES STRONGLY ANTISEPTIC.

Benzoic acid,	1:909
Potassium bichromate,	1:909
Potassium cyanide,	1:909
Aluminum chloride,	1:714
Ammonia,	1:714
Zinc chloride,	1:526
Mineral acids,	1:500 to 1:333
Thymic acid,	1:500
Lead chloride,	1:500
Nitrate of cobalt,	1:476
Sulphate of nickel,	1:400
Nitrate of uranium,	1:356
Carbolic acid,	1:333
Potassium permanganate,	1:285
Lead nitrate,	1:277
Alum,	1:222
Tannin,	1:207

SUBSTANCES MODERATELY ANTISEPTIC.

Bromhydrate of quinine,	1:182
Arsenious acid,	1:166
Boracic acid,	1:143
Sulphate of strychnia,	1:143
Arsenite of soda,	1:111
Hydrate of chloral,	1:107
Salicylate of soda,	1:100
Ferrous sulphate,	1:90
Caustic soda,	1:66

SUBSTANCES FREELY ANTISEPTIC.

Perchloride of manganese,	1:40
Calcium chloride,	1:25
Sodium borate,	1:14
Muriate of morphia,	1:13
Strontium chloride,	1:12
Lithium chloride,	1:11
Barium chloride,	1:10
Alcohol,	1:10

SUBSTANCES VERY FEEBLY ANTISEPTIC.

Ammonium chloride,	1:9
Potassium arsenite,	1:8
Potassium iodide,	1:7
Sodium chloride,	1:6
Glycerin (sp. gr. 1.25),	1:4
Ammonium sulphate,	1:4
Sodium hyposulphite,	1:3

ANTISEPTIC AND GERMICIDAL VALUE OF VARIOUS SALTS,
ARRANGED ALPHABETICALLY.

Alum.—Antiseptic in the proportion of 1 : 222 (Miquel).

Aluminum Acetate.—According to De la Croix, this salt is an antiseptic in the proportion of 1 : 6,310. Kuhn found it to be antiseptic in 1 : 5,250.

Aluminum Chloride.—Antiseptic in the proportion of 1 : 714 (Miquel).

Ammonium Carbonate.—When present in the proportion of 1 : 125 it restrains the development of typhoid bacilli, and in five hours' time it kills these bacilli in the proportion of 1 : 100; the cholera spirillum is killed in the same time by 1 : 77 (Kitasato).

Ammonium Chloride.—Antiseptic in the proportion of 1 : 9 (Miquel). A five-per-cent solution does not kill anthrax spores in twenty-five days (Koch).

Ammonium Fluosilicate.—The bacillus of anthrax and of typhoid fever fail to grow in nutrient gelatin containing 1 : 1,000, and a two-per-cent solution kills anthrax spores in one-quarter to three-quarters of an hour (Faktor).

Ammonium Sulphate.—Antiseptic in the proportion of 1 : 4 (Miquel). A five-per-cent solution failed in two days to kill anthrax spores, but was effective in five days (Koch).

Barium Chloride is an antiseptic in the proportion of 1 : 10 (Miquel).

Calcium Chloride is an antiseptic in the proportion of 1 : 25 (Miquel). A saturated solution does not destroy anthrax spores (Koch).

Calcium Hypochlorite.—This is a powerful germicidal agent and has great value as a practical disinfectant. Good chloride of lime contains from twenty-five to thirty per cent of available chlorine as hypochlorite. The experiments made by the Committee on Disinfectants of the American Public Health Association in 1885 showed that a solution containing 0.25 per cent of chlorine as hypochlorite is an effective germicide, even when allowed to act only for one or two minutes. In Bolton's experiments a solution of chloride of lime of 1 : 2,000 (available chlorine 0.015) destroyed the typhoid bacillus and the cholera spirillum in two hours. For the destruction of anthrax spores a one-per-cent solution was required (available chlorine 0.3 per cent). Nissen found that the typhoid bacillus and the cholera spirillum are destroyed with certainty in five minutes by a solution containing 0.12 per cent, anthrax bacilli in one minute by 0.1 per cent, *Staphylococcus pyogenes aureus* in one minute by 0.2 per cent, anthrax spores in thirty minutes by a

five-per-cent solution and in seventy minutes by a one-per-cent solution. Experiments made by the same author upon the sterilization of fæces showed that 0.5 per cent to one per cent could be relied upon to destroy the typhoid bacillus or the cholera spirillum in fæces in ten minutes.

Chloral Hydrate.—Antiseptic in the proportion of 1 : 107 (Miquel). A twenty-per-cent solution destroys pus cocci in two hours (Sternberg).

Cupric Chloride.—Antiseptic in the proportion of 1 : 1,428 (Miquel).

Cupric Sulphate.—Antiseptic in the proportion of 1 : 111 (Miquel). Kills the cholera spirillum in the proportion of 1 : 3,000 in ten minutes (Nicati and Rietsch). Destroys the cholera spirillum in bouillon cultures in less than half an hour in 1 : 600, and in four hours in 1 : 1,000; cultures in blood serum require 1 : 200 (Van Ermengem). A solution of 1 : 20 kills the typhoid bacillus in ten minutes (Leitz). This salt failed, in the writer's experiments, to kill the spores of *Bacillus anthracis* and *Bacillus subtilis* in two hours' time in a twenty-per-cent solution. In Koch's experiments a five-per-cent solution failed to kill anthrax spores in ten days. Kills pus micrococci in two hours in the proportion of 1 : 200 (Sternberg). In Bolton's experiments made for the Committee on Disinfectants of the American Public Health Association the following results were obtained: Recent cultures in bouillon, time of exposure two hours : *Bacillus* of typhoid fever, 1 : 200; cholera spirillum, 1 : 500; *Bacillus pyocyaneus*, 1 : 200; Brieger's bacillus, 1 : 200; Emmerich's bacillus, 1 : 200; *Staphylococcus pyogenes aureus*, 1 : 100; *Staphylococcus pyogenes citreus*, 1 : 100; *Staphylococcus pyogenes albus*, 1 : 200; *Streptococcus pyogenes*, 1 : 500. When ten per cent of dried egg albumin was added to a recent culture in bouillon of the typhoid bacillus the amount required to insure sterilization was 1 : 10.

In the report of the Committee on Disinfectants of the American Public Health Association this agent is recommended in "a solution of two to five per cent for the destruction of infectious material *not containing spores*." The experimental data above given show that this is a liberal allowance for material which does not contain an excessive amount of albumin. In the experiments of Leitz the typhoid bacillus in cultures was destroyed in ten minutes by a five-per-cent solution.

Ferric Chloride.—A five-per-cent solution failed in two days to destroy anthrax spores, but was effective in five days (Koch).

Ferrous Sulphate.—In the writer's experiments (1883) a solution of twenty per cent failed to destroy micrococci and putrefactive bacteria. In a more recent experiment ten per cent failed to kill pus

cocci, but was fatal to *Micrococcus tetragenus*—two hours' exposure. Koch found that a five-per-cent solution failed to destroy anthrax spores in six days. Exposure to a twenty-per-cent solution for forty-eight hours does not destroy the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). In the experiments of Jäger immersion in a solution of 1 : 3 destroyed the infective virulence of certain pathogenic bacteria (fowl cholera, rothlauf, glanders), as tested by injection into mice, but failed to kill anthrax spores and tubercle bacilli. The antiseptic power of ferrous sulphate is placed by Miquel at 1 : 90. In the writer's experiments 1 : 200 prevented the development of micrococci and of putrefactive bacteria in bouillon placed in the incubating oven for forty-eight hours. Leitz found that a five-per-cent solution required three days' exposure for the destruction of the typhoid bacillus.

Gold Chloride.—Antiseptic in the proportion of 1 : 4,000 (Miquel). Boer has made extended experiments with the chloride of gold and sodium. We give his results below. In his disinfection experiments a bouillon culture which had been in the incubating oven for twenty-four hours was used, and the time of exposure was two hours.

	Restrains development.	Destroys vitality.
Anthrax bacillus	1 : 40000	1 : 8000
Diphtheria bacillus	1 : 40000	1 : 1000
Glanders bacillus	1 : 15000	1 : 400
Typhoid bacillus	1 : 20000	1 : 500
Cholera spirillum	1 : 25000	1 : 1000

Lead Chloride.—Antiseptic in the proportion of 1 : 500 (Miquel).

Lead Nitrate.—Antiseptic in the proportion of 1 : 277 (Miquel).

Lithium Chloride.—Antiseptic in the proportion of 1 : 11 (Miquel).

Manganese Protochloride.—Antiseptic in the proportion of 1 : 40 (Miquel).

Mercuric Chloride.—Koch's experiments (1881) gave the following results : A solution of 1 : 1,000 destroys anthrax spores in a few minutes, and 1 : 10,000 is effective after a more prolonged exposure. The writer (1884) obtained similar results—1 : 10,000 destroyed the spores of *Bacillus anthracis* and of *Bacillus subtilis* in two hours. More recent experiments indicate that failure to grow in culture solutions cannot be accepted as evidence of the destruction of vitality in the case of spores exposed to the action of this agent, unless due precautions are taken to exclude the restraining influence of the small amount of mercuric chloride which remains attached to the spores. Koch had ascertained that the development of spores is restrained by

the presence of 1 : 300,000 in a culture medium, and Geppert has recently shown that even so small an amount as 1 : 2,000,000 will prevent the development of spores the vitality of which has been reduced by the action of a strong solution (1 : 1,000). When this restraining action is entirely neutralized by washing the spores in a solution containing ammonium sulphide it requires, according to Geppert, a solution of 1 : 1,000 acting for one hour to completely destroy the vitality of anthrax spores. Fränkel found that a solution of 1 : 1,000 was effective in half an hour. The typhoid bacillus, the bacillus of mouse septicaemia, and the cholera spirillum, in bouillon cultures and in cultures in flesh-peptone-gelatin, are destroyed in two hours by 1 : 10,000 ; but in a bouillon culture to which ten per cent of dried egg albumin was added a one-per-cent solution was required to destroy the typhoid bacillus in the same time (Bolton). According to Van Ermengem, cultures of the cholera spirillum in bouillon are sterilized in half an hour by 1 : 60,000, but cultures in blood serum require 1 : 800 to 1 : 1,000. In experiments upon tuberculous sputum Schill and Fischer found that exposure of fresh sputum to an equal amount of a 1 : 2,000 solution for twenty-four hours failed to disinfect it, as shown by inoculation experiments in guinea-pigs. The antiseptic power of mercuric chloride is given by Miquel as 1 : 14,300. In the writer's experiments 1 : 33,000 was found to prevent the development of putrefactive bacteria in bouillon, but a minute bacillus contained in broken-down beef infusion multiplied, after several days, in 1 : 20,000. The pus cocci were restrained in their development by 1 : 30,000.

In Behring's experiments the anthrax bacillus and cholera spirillum were killed in one hour by 1 : 100,000 when the temperature was 36° C., but at a temperature of 3° C. the proportion required was 1 : 25,000. The same author states that at 22° C. *Staphylococcus aureus* in bouillon is not always killed in twenty-five minutes by 1 : 1,000.

In a recent series (1891) of experiments Abbott has shown that a 1 : 1,000 solution does not always destroy *Staphylococcus pyogenes aureus* in five minutes. He says: "Frequently all the organisms would be destroyed after five minutes' exposure, but almost as often a certain few would resist for that length of time, and even longer, going in some cases to ten, twenty, and even thirty minutes."

According to Yersin, a solution of 1 : 1,000 kills the tubercle bacillus in one minute.

We might add considerably to the experimental data given, but the results already recorded are sufficient to show the value of this agent as an antiseptic and germicide, and justify its use for general purposes of disinfection in the proportion of 1 : 500 or 1 : 1,000 for material containing spores, and in the proportion of 1 : 2,000 to

1 : 5,000 for pathogenic bacteria in the absence of spores; due regard being had to the fact that the presence of albumin very materially reduces its germicidal potency, and that it may be decomposed and neutralized by alkalies and their carbonates, by hydrosulphuric acid, and by many other substances.

The albuminate of mercury, as has been shown by Lister, is soluble in an excess of albumin, and, according to Behring, is just as effective as an aqueous solution containing the same amount of sublimate when dissolved in an albuminous liquid like blood serum (?).

In practice the addition of a mineral acid to sublimate solutions, or of sodium, potassium, or ammonium chloride, is to be recommended, to prevent the precipitation of the mercuric chloride by albumin in fluids containing it. Behring recommends the addition of five parts of sodium or potassium chloride to one of the sublimate. Such a solution is more stable than a simple solution of sublimate, and no precipitate is formed by the addition of alkalies or by albumin.

The same result is obtained, according to La Place, by the addition of five parts of hydrochloric or tartaric acid to one part of sublimate in aqueous solution.

Mercuric Cyanide, $\text{Hg}(\text{CN})_2$, and the *Oxycyanide* of mercury have been tested, with the following results : *Staphylococcus aureus* is destroyed in five minutes by 1 : 100, in one hour by 1 : 1,000, in two hours by 1 : 1,500 (Chibret). The development of *Bacillus anthracis* in culture solutions is prevented by the presence of cyanide of mercury in the proportion of 1 : 25,000, and by the oxycyanide by 1 : 16,000 (Behring).

Boer obtained the following results with the oxycyanide—cultures in bouillon, twenty-four hours in incubating oven, time of exposure two hours :

	Restrained development.	Destroyed vitality.
Anthrax bacillus.....	1 : 80000	1 : 40000
Diphtheria bacillus.....	1 : 80000	1 : 40000
Glanders bacillus.....	1 : 60000	1 : 30000
Typhoid bacillus.....	1 : 60000	1 : 30000
Cholera spirillum.....	1 : 90000	1 : 60000

Mercuric Iodide.—The antiseptic value of this salt is placed by Miquel at 1 : 40,000, which is more than double that given by the same author to the bichloride. In the writer's experiments upon the antiseptic value of salts and oxides of mercury the following results were obtained :

	Active.	Failed.
Biniiodide of mercury.....	1 : 20000	1 : 40000
Bichloride	1 : 15000	1 : 20000
Protiodide.....	1 : 10000	1 : 20000
Yellow oxide.....	1 : 1000	1 : 2000
Black oxide.....	1 : 500	1 : 1000

Morphia Hydrochlorate.—Antiseptic in the proportion of 1 : 13 (Miquel).

Nickel Sulphate.—Antiseptic in the proportion of 1 : 400 (Miquel).

Platinum Bichloride.—Antiseptic in the proportion of 1 : 3,333 (Miquel).

Potassium Acetate.—A saturated solution of this salt failed to kill anthrax spores in ten days (Koch).

Potassium Arsenite.—In the writer's experiments Fowler's solution failed to kill micrococci in two hours in the proportion of four per cent. Miquel places the antiseptic value of potassium arsenite at 1 : 8.

Potassium Bichromate.—A five-per-cent solution failed in two days to destroy anthrax spores (Koch). Efficient as an antiseptic in the proportion of 1 : 909 (Miquel).

Potassium Bromide.—The bacillus of typhoid fever and the cholera spirillum fail to grow in culture solutions containing 9 to 10.6 per cent, and are killed in four or five hours by ten to twelve per cent (Kitasato).

Potassium Carbonate.—The development of the typhoid bacillus and of the cholera spirillum is prevented by 0.74 to 0.81 per cent, and these bacteria are killed in five hours by 1 per cent (Kitasato).

Potassium Chlorate.—In the writer's experiments a four-per-cent solution failed in two hours to kill *Micrococcus Pasteuri*. A five-per-cent solution failed in six days to destroy anthrax spores (Koch).

Potassium Chromate.—A five-per-cent solution failed to kill anthrax spores in five days (Koch).

Potassium Cyanide.—Antiseptic in the proportion of 1 : 909 (Miquel).

Potassium Iodide.—A solution of five per cent does not destroy anthrax spores in eighty days (Koch). Putrefactive bacteria in broken-down beef infusion are not destroyed by two hours' exposure in a twenty-per-cent solution (Sternberg). The typhoid bacillus and the cholera spirillum do not grow in culture solutions containing

eight per cent, and are destroyed by five hours' exposure to 9.23 per cent (Kitasato). Antiseptic in the proportion of 1 : 7 (Miquel).

Potassium Permanganate.—In the writer's experiments (1881) a two-per-cent solution was required to destroy *Micrococcus Pasteuri* in the blood of a rabbit. In later experiments pus cocci in bouillon were killed by 1 : 833—time of exposure two hours. One per cent was found by Koch not to destroy anthrax spores in two days, but five per cent was effective in one day. The glanders bacillus is destroyed in two minutes by a one-per-cent solution (Löffler). The experiments of Jäger show that a one-per-cent solution is not reliable for the destruction of anthrax bacilli and other pathogenic bacteria tested, but a five-per-cent solution was effective. The tubercle bacillus was not, however, killed by exposure in a five-per-cent solution. According to Miquel, permanganate of potash is an antiseptic in the proportion of 1 : 285.

Quinine Hydrobromate.—Antiseptic in the proportion of 1 : 182 (Miquel).

Quinine Hydrochlorate.—Antiseptic in the proportion of 1 : 900 (Ceri). Quinine dissolved with hydrochloric acid destroys anthrax spores in ten days in one-per-cent solution (Koch).

Quinine Sulphate.—The writer found that in the proportion of 1 : 800 quinine prevents the development of various micrococci and bacilli. A ten-per-cent solution does not destroy the bacilli of symptomatic anthrax (Arloing, Cornevin, and Thomas).

Silver Nitrate.—Miquel places nitrate of silver next to mercuric chloride as an antiseptic, effective in the proportion of 1 : 12,500. Behring also places it next to bichloride as an antiseptic and germicide, and says that it is even superior to this salt in albuminous fluids. He reports that it prevents the development of anthrax spores when present in a culture liquid in the proportion of 1 : 80,000, and in the proportion of 1 : 10,000 destroys these spores in forty-eight hours. We give below the result of recent experiments by Boer, in which the time of exposure was two hours :

	Restrains development.	Destroys vitality.
Anthrax bacillus.....	1 : 60000	1 : 20000
Diphtheria bacillus.....	1 : 60000	1 : 2500
Glanders bacillus.....	1 : 75000	1 : 4000
Typhoid bacillus.....	1 : 50000	1 : 4000
Cholera spirillum.....	1 : 50000	1 : 4000

Silver Chloride.—A solution of chloride of silver in hyposulphite of soda is much less effective as an antiseptic than nitrate of silver.

Behring found that to prevent the development of anthrax spores a solution of 1 : 8,000 was required.

Sodium Borate.—In the writer's experiments a saturated solution of borax was found to be without germicidal power. A twenty-per-cent solution does not destroy the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). A five-per-cent solution failed to destroy anthrax spores in fifteen days (Koch). Antiseptic in the proportion of 1 : 14 (Miquel).

Sodium Carbonate.—A solution of 2.2 per cent restrains the growth of the typhoid bacillus, and of 2.47 per cent of the cholera spirillum. The first-named bacillus is killed by four or five hours' exposure in a 2.47-per-cent solution, and the cholera spirillum by 3.45 per cent (Kitasato).

Sodium Chloride.—A saturated solution failed in forty-eight hours to destroy the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). A saturated solution failed in forty days to destroy anthrax spores (Koch). A saturated solution failed in twenty hours to destroy the tubercle bacillus in fresh sputum (Schill and Fischer). In the writer's experiments a five-per-cent solution failed to kill *Micrococcus Pasteuri* in blood. Antiseptic in the proportion of 1 : 6 (Miquel). According to Forster, the bacillus of typhoid fever, the bacillus of rouget, and the streptococcus of pus are not killed by several weeks' exposure in strong solutions of sodium chloride, but the cholera spirillum is destroyed in a few hours. Cultures of the tubercle bacillus are not sterilized in two months by a saturated solution; and tuberculous organs from an ox, preserved in a solution of salt, did not lose their power of infecting susceptible animals inoculated with material from the diseased tissue. The flesh of swine which died of rothlauf was found by Petri to still contain the bacillus in a living condition after having been preserved in brine for a month.

Sodium Hyposulphite.—In the writer's experiments a saturated solution failed in two hours to kill micrococci and bacilli. Exposure for forty-eight hours to a fifty-per-cent solution does not destroy the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas). Antiseptic in the proportion of 1 : 3 (Miquel).

Sodium Sulphite.—The results with a saturated solution of this salt were, in the writer's experiments, entirely negative.

Tin Chloride.—A one-per-cent solution acting for two hours destroyed the bacteria in putrefying bouillon, while 0.8 per cent failed (Abbott).

Zinc Chloride.—In the writer's experiments 1 : 200 destroyed *Micrococcus Pasteuri* in two hours, but a two-per-cent solution was required to kill pus cocci in the same time; spores of *Bacillus anthracis*

were not destroyed by two hours' exposure in a ten-per-cent solution, but a solution of five per cent killed the spores of *Bacillus subtilis* in the same time. Koch found that anthrax spores germinated after being immersed in a five-per-cent solution for thirty days. The development of *Bacillus prodigiosus* is only slightly retarded by exposure for sixteen hours in a one-per-cent solution. Antiseptic in the proportion of 1 : 526 (Miquel).

Zinc Sulphate.—In the writer's first experiments a twenty-per-cent solution failed to destroy in two hours micrococci obtained from the pus of an acute abscess. In later experiments a micrococcus from the same source resisted two hours' exposure to a ten-per-cent solution, but *Micrococcus tetragenus* was destroyed by this amount. Broken-down beef infusion mixed with an equal quantity of a forty-per-cent solution was not sterilized after two hours' contact. In Koch's experiments anthrax spores were found to germinate after having been immersed for ten days in a five-per-cent solution.

XI.

ACTION OF COAL-TAR PRODUCTS, ESSENTIAL OILS, ETC.

IN the present section we shall consider the action upon bacteria of a variety of organic products, and for convenience will arrange them alphabetically.

Acetone.—Anthrax spores grow freely after two days' exposure to the action of this agent; at the end of five days their development is feeble (Koch).

Alcohol.—In the writer's experiments ninety-five-per-cent alcohol did not destroy the bacteria (spores) in broken-down beef tea in forty-eight hours. *Micrococcus Pasteuri* was destroyed by two hours' exposure in a twenty-four-per-cent solution; pus cocci required a forty-per-cent solution. Koch found that absolute alcohol had no effect upon anthrax spores exposed to its action for one hundred and ten days. Schill and Fischer found that when tuberculous sputum was mixed with an equal amount of absolute alcohol its infecting power was not destroyed in twenty-four hours, but that in the proportion of five parts to one of sputum it was effective in destroying the tubercle bacillus, as proved by inoculation experiments. Yersin found that in pure cultures the tubercle bacillus is killed by five minutes' exposure to the action of absolute alcohol.

Aniline Dyes.—Recent researches have shown that some of the aniline colors possess very decided germicidal power. Stilling found that solutions of methyl violet containing 1:30,000 exercise a restraining influence upon the development of putrefactive bacteria and pus cocci, and that these microorganisms are destroyed by solutions containing 1:2,000 to 1:1,000. Methyl violet has been placed in the market by Merck under the name of pyoktanin. Jänicke reports the following results with pyoktanin: *Staphylococcus pyogenes aureus* was restrained in its development by solutions containing 1:2,000,000, *Bacillus anthracis* by 1:1,000,000, *Staphylococcus pyogenes* by 1:333,300, *Spirillum cholerae Asiaticæ* by 1:62,500, *Bacillus typhi abdominalis* by 1:5,000. In blood serum stronger solutions were required (1:500,000 for *Staphylococcus pyogenes aureus*). *Staphylococcus pyogenes aureus*, *Streptococcus pyogenes*, and *Bacillus*

anthracis were killed in thirty seconds by 1 : 1,000, the typhoid bacillus by the same amount in thirty minutes. Boer found malachite green to be still more effective than methyl violet. In his experiments upon bouillon cultures twenty-four hours old, with two hours' exposure to the action of the disinfectant, he obtained the following results :

MALACHITE GREEN.

	Restrains development.	Destroys vitality.
Anthrax bacillus	1 : 120000	1 : 40000
Diphtheria bacillus	1 : 40000	1 : 8000
Glanders bacillus	1 : 5000	1 : 300
Typhoid bacillus	1 : 5000	1 : 300
Cholera spirillum	1 : 100000	1 : 5000

METHYL VIOLET (PYOKTANIN).

	Restrains development.	Destroys vitality.
Anthrax bacillus	1 : 70000	1 : 5000
Diphtheria bacillus	1 : 10000	1 : 2000
Glanders bacillus	1 : 2500	1 : 150
Typhoid bacillus	1 : 2500	1 : 150
Cholera spirillum	1 : 30000	1 : 1000

Aniline Oil.—According to Riedlin, the addition of 1 : 5 of aniline water prevents the development of all bacteria in nutrient gelatin.

Aromatic Products of Decomposition.—Klein has tested the germicidal power of phenylpropionic and phenylacetic acids. He finds that anthrax spores resist both of these acids, in the proportion of 1 : 400, for two days, but in the absence of spores anthrax bacilli are quickly killed by a solution of this strength. Certain non-pathogenic micrococci were not killed by exposure for twenty-five minutes to 1 : 200. The caseous matter of pulmonary tuberculosis infected guinea-pigs after exposure for ninety-six hours to 1 : 200.

Aseptol.—A ten-per-cent aqueous solution kills anthrax spores in ten minutes, and a three- to five-per-cent solution is a reliable disinfectant in the absence of spores (Hueppe).

Benzene, C₆H₆.—Exposure in benzol for twenty days failed to destroy the vitality of anthrax spores (Koch).

Camphor.—Alcohol saturated with camphor has no effect upon the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas).

The experiments of Cadéac and Meunier show that camphor (oil of, or tincture ?) has but little germicidal power. The typhoid ba-

cillus and cholera spirillum were only destroyed after eight to ten days' exposure to the action of camphor ("essence").

Carbolic Acid.—Tested upon anthrax spores, Koch found a one-per-cent solution to be without effect after fifteen days' exposure; a two-per-cent solution retarded development but did not completely destroy vitality in seven days; a three-per-cent solution was effective in two days. In the absence of spores Koch found that a one-per-cent solution quickly destroys the vitality of anthrax bacilli. He recommends a five-per-cent solution for the destruction of the "comma bacillus" in the discharges of cholera patients, and a two-per-cent solution for the disinfection of surfaces soiled with such discharges. In the writer's experiments 1:200 destroyed *Micrococcus Pasteuri* in two hours; and pus cocci were destroyed by 1:125, while 1:200 failed. Davaine showed by inoculation experiments that anthrax bacilli in fresh blood are destroyed by being exposed to the action of a one-per-cent solution for one hour. A two-per-cent solution destroys the dried virus of symptomatic anthrax in forty-eight hours (Arloing, Cornevin, and Thomas). Solutions in oil or in alcohol have been shown by Koch to be less effective than aqueous solutions. Thus a five-per-cent solution in oil failed to destroy anthrax spores in one hundred and ten days, and the same solution failed to kill the bacilli, in the absence of spores, in less than six days. A five-per-cent solution in alcohol did not destroy anthrax spores in seventy days. Schill and Fischer found that a three-per-cent solution destroyed the infecting power of tuberculous sputum, as shown by inoculation into guinea-pigs, in twenty-four hours, while solutions of one and two per cent failed. Bolton's experiments gave the following results, the test organisms being in fresh bouillon cultures and the time of exposure two hours: The cholera spirillum, the bacillus of typhoid fever, the bacillus of schweinerotlauf, Brieger's bacillus, the bacillus of green pus, and the pus cocci (*Staphylococcus pyogenes aureus*, *albus*, and *citreus*, and *Streptococcus pyogenes*) were all killed by a solution of one per cent, while in a majority of the experiments a one-half-per-cent (1:200) solution failed. Cultures of the typhoid bacillus in flesh-peptone-gelatin gave the same result (1:100 with two hours' exposure), and the addition of ten per cent of dried egg albumin to bouillon cultures did not influence the result.

The experiments of La Place show that the addition of hydrochloric acid to a disinfecting solution containing carbolic acid greatly increases its germicidal power for spores. Thus it is stated that "two per cent of crude carbolic acid with one per cent of pure hydrochloric acid destroyed anthrax spores in seven days, while two per cent of carbolic acid or one per cent of hydrochloric acid alone did

not destroy these spores in thirty days. A four-per-cent solution of crude carbolic acid with two per cent of hydrochloric acid destroyed spores in less than an hour; four per cent of carbolic acid alone did not destroy them in twelve days. Van Ermengem reports that in his experiments the cholera spirillum in chicken bouillon was killed in less than half an hour by 1 : 600, and that in blood serum 1 : 400 was effective. Nicati and Rietsch fix the germicidal power for the cholera spirillum as 1 : 200, the time of exposure being ten minutes; Ramon and Cajal, 1 : 50. Boer gives the following results, the time of exposure being two hours, cultures in bouillon twenty-four hours old :

	Restrains development.	Destroys vitality.
Anthrax bacillus.....	1 : 750	1 : 300
Diphtheria bacillus.....	1 : 500	1 : 300
Glanders bacillus.....	1 : 500	1 : 300
Typhoid bacillus.....	1 : 400	1 : 200
Cholera spirillum.....	1 : 600	1 : 400

Leitz reports the following results : The dejections of patients suffering from typhoid fever, mixed in equal quantity with the disinfecting solution, were sterilized by a five-per-cent solution of carbolic acid in three days. Pure cultures of the typhoid bacillus were sterilized in fifteen minutes by a five-per-cent solution.

In the experiments of Nocht upon anthrax spores it was found that while at the room temperature these spores were not destroyed by several days' exposure in a five-per-cent solution, they were destroyed in three hours by the same solution at a temperature of 37.5°.

Carbolic acid prevents putrefactive changes in bouillon when present in the proportion of 1 : 333 (Miquel). The tubercle bacillus is killed in thirty seconds by a five-per-cent solution, and in one minute by a one-per-cent solution (Yersin).

Coffee Infusion.—Experiments have been made by Heim and by Lüderitz on the antiseptic power of an infusion of coffee. The first-named author found that anthrax bacilli no longer developed after three hours' exposure in a ten-per-cent solution, but spores were not killed at the end of a week. Streptococci in a bouillon culture required twenty-four hours' exposure, and the staphylococci of pus were not destroyed in this time. Lüderitz found that a three-per-cent infusion restrained the growth in nutrient gelatin of the typhoid bacillus, and a five-per-cent infusion killed the bacillus in two days; the cholera spirillum failed to grow in presence of one per cent, and a solution of this strength killed it in seven hours; Staphylococcus

pyogenes aureus was prevented from developing by two per cent, and was killed in six days by a five-per-cent solution; *Streptococcus pyogenes* was prevented from growing by one per cent, and killed by a ten-per-cent solution in one day; *Proteus vulgaris* did not grow in presence of 2.5 per cent, and was killed in two days by ten per cent. The question as to what constituent of the infusion of roasted coffee was the active germicidal agent was not determined, but the authors referred to agree that it was not caffeine.

Creolin.—This is a coal-tar product which resembles crude carbolic acid in appearance, but smells rather like tar than like phenol. It makes a milky emulsion with water, which has been proved by numerous experiments to possess very decided germicidal power, being superior to carbolic acid. The first careful test of the germicidal power of this agent was made by Esmarch, who found that a solution of 1:200 killed the cholera spirillum in a minute, the typhoid bacillus at the end of several days. Anthrax spores were not destroyed in twenty days by a five-per-cent solution, but this solution killed the tubercle bacillus attached to silk threads which were immersed in it for a short time, and also disinfected tuberculous sputum. Behring has shown that in albuminous liquids creolin is less effective than carbolic acid. In blood serum 1:175 was required to restrain the development of staphylococci, and 1:100 to destroy the same in ten minutes. Van Ermengem, as a result of numerous experiments, arrived at the conclusion that creolin is a cheap and useful disinfecting agent, in a five-per-cent solution, for various pathogenic organisms. Kaupe reports that in his experiments a ten-per-cent solution killed anthrax spores in twenty-four hours. According to Boer, a solution of 1:5,000 destroys anthrax bacilli in bouillon cultures in two hours, 1:2,000 diphtheria bacilli, 1:300 the glanders bacillus, 1:250 the typhoid bacillus, and 1:3,000 the cholera spirillum.

Creosote.—This agent was found by the writer to be fatal to micrococci in the proportion of 1:200. In the proportion of one per cent it failed, after twenty hours' exposure, to destroy tubercle bacilli in sputum (Schill and Fischer). A saturated aqueous solution does not destroy the tubercle bacillus in cultures in twelve hours (Yersin). Guttman, in extended experiments upon various pathogenic organisms, found that development was prevented by 1:3,000 to 1:4,000. A solution containing 1:300 killed *Bacillus pyocyaneus* and *Bacillus anthracis* in one minute, *Bacillus prodigiosus* in two minutes, and the Finkler-Prior spirillum in one minute in the proportion of 1:600.

Cresol.—This is a dark, reddish-brown, transparent fluid, somewhat thinner than creolin, and, like it, having an odor of tar. It forms an emulsion with water, which is not so stable as that formed

by creolin. Of the three cresols, ortho-, meta-, and paracresol, the second was found by Fränkel to be most active. This author states that the addition of sulphuric acid adds greatly to its germicidal power. A four-per-cent solution, containing equal parts of cresol and H_2SO_4 , killed anthrax spores in less than twenty-four hours. In Behring's experiments a solution containing ten per cent of each killed anthrax spores in eighty minutes, and five per cent of each in one hundred minutes, while an eighteen-per-cent solution of sulphuric acid alone did not kill them in twenty-four hours. In the experiments of Jäger a two-per-cent solution destroyed the tubercle bacillus in cultures and in sputum. As a result of his experiments Behring concludes that cresol has no advantage over carbolic acid as a germicide for the destruction of spores. Tested upon *Staphylococcus aureus*, *Streptococcus erysipelatos*, and *Bacillus pyocyaneus*, Fränkel found that a solution of 0.3 per cent destroyed these microorganisms in five minutes, while a two-per-cent solution of carbolic acid required fifteen minutes' contact to accomplish the same result.

Disinfektol.—This is a coal-tar product similar to creolin which has been recommended in Germany for disinfecting purposes. It is an oily, dark-brown fluid having a specific gravity of 1.086. It forms an emulsion with water, which has a slightly alkaline reaction. It has been tested upon typhoid stools by Uffelmann and by Beselin. The last-named author gives the following summary of the results obtained: An emulsion of five per cent of disinfektol equals in value, for the disinfection of the liquid discharges of typhoid patients, 12.5 per cent of creolin, thirty-three per cent of hydrochloric acid, five per cent of carbolic acid, 1 : 500 of mercuric chloride.

Ether.—Anthrax spores may germinate after being immersed in sulphuric ether for eight days (Koch). The tubercle bacillus is destroyed by ten minutes' exposure to the action of ether (Yersin).

Essential Oils.—Chamberlain has made an extended series of experiments to determine the antiseptic power of the vapor of volatile oils. A large number of essential oils tested were found to prevent the development of the anthrax bacillus, while a few did not. At the end of six days the tubes were opened and the oil absorbed by the culture liquid allowed to evaporate. Cultures were now obtained from all except the following, which, it was inferred, had destroyed the vitality of the spores: Angelica, cinnamon of China, cinnamon of Ceylon, geranium of France, geranium of Algeria, origanum.

Cadéac and Meunier have also made extended experiments upon the typhoid bacillus and the bacillus of glanders, for the purpose of determining the germicidal power of agents of this class. Their method consisted in the introduction of a sterilized platinum needle into a pure culture of the test organism, in immersing it in the

essential oil for a certain time, and then making with it a puncture in a suitable solid culture medium. Their results are given below for the typhoid bacillus.

Essences which kill the bacillus after a contact of less than twenty-four hours:

	At the end of—
Cinnamon of Ceylon,	12 minutes.
Cloves,	25 "
Eugenol,	30 "
Thyme,	35 "
Wild thyme,	35 "
Verbena of India,	45 "
Geranium of France,	50 "
Origanum,	75 "
Patchouly,	80 "
Zedoary,	2 hours.
Absinthe,	4 "
Sandalwood,	12 "

The following were effective in from twenty-four to forty-eight hours: Cumin, caraway, juniper, matico, galbanum, valerian, citron, angelica, celery, savin, copaiba, pepper, turpentine, opoponax, rose, chamomile; the following required from two to four days: *Illicium*, sassafras, tuberose, coriander; the following from four to eight days: Calamus, sage, fennel, mace, cascarilla, orange of Portugal; the following in eight to ten days: Mint, nutmeg, rosemary, carrot, mustard, anise, onion, marjoram, bitter almonds, cherry laurel, myrtle, lavender, eucalyptus, cedar, cajuput, wintergreen, camphor.

Riedlin reports as the result of his experiments that the essential oils which have the greatest antiseptic value are oil of lavender, eucalyptus, rosemary, and cloves.

Eucalyptol.—Chabaunes and Perret found that a five-per-cent solution of eucalyptol is without effect upon tubercle bacilli in sputum. According to Behring, eucalyptol is about four times less active as a disinfectant than carbolic acid.

Glycerin has no action upon the virus of symptomatic anthrax (Arloing, Cornevin, and Thomas), and is inert as regards the spores of anthrax (Koch). Glycerin prevents putrefactive decomposition in bouillon when present in the proportion of 1:4 (Miquel). Roux has shown that the addition of five per cent of glycerin to a culture medium is favorable to the growth of the tubercle bacillus; it is also appropriated as pabulum by various other species.

Hydroxylamin.—Heinisch found that the development of the anthrax bacillus is prevented by 1:77 of hydroxylamin hydrochlorate, and of the diphtheria bacillus by 1:75. In these experiments a solution of soda was added to release the hydroxylamin. Marpmann found that 1:100 preserved milk without change for four

to six weeks, and that alkaline fermentation of urine was prevented by 1:1,000.

Indol.—When added in excess to water this agent failed to destroy anthrax spores in eighty days (Koch).

Lanolin.—According to Gottstein, various microorganisms tested by him failed to grow in cultures after having been in contact with pure lanolin for five to seven days.

Naphthol.—In the proportion of 1:10,000 naphthol prevents the development of the glanders bacillus, the anthrax bacillus, the typhoid bacillus, the micrococcus of fowl cholera, of *Staphylococcus aureus* and *albus*, and of several other microorganisms tested by Maximovitch. The same author states that although insoluble in cold water, water at 70° C. dissolves 0.44 in one thousand parts. When urine is shaken up with naphthol in powder it does not undergo fermentation.

In the experiments of Foote hydronaphthol was found to show some germicidal power in the proportion of 1:2,300, but the conclusion is reached that a saturated aqueous solution (1:1,150) does not equal a one-per-cent solution of carbolic acid or of creolin.

Olive Oil.—Anthrax spores germinate after having been immersed for ninety days in pure olive oil (Koch).

Oil of Mustard.—Koch found that the development of anthrax spores is prevented by 1:33,000.

Oil of Peppermint.—A five-per-cent solution in alcohol failed in twelve days to destroy anthrax spores, but the development of these spores is restrained by 1:33,000 (Koch).

Oil of Turpentine destroys anthrax spores in five days, but failed to do so in one day (Koch). The development of anthrax spores is prevented by 1:75,000 (Koch). The addition of 1:200 to nutrient gelatin prevents the development of bacteria (Riedlin). An excess of oil of turpentine added to a liquefied gelatin culture of *Staphylococcus aureus* does not destroy this micrococcus in five hours (v. Christmas-Dirckinck-Holmfeld).

Skatol in excess in water has no germicidal power, as tested upon anthrax spores (Koch).

Smoke.—The researches of Beu show that meats which have been preserved by smoking commonly contain living bacteria capable of growing in culture media; and Petri has shown that pork which has been salted for a month and then smoked for fourteen days may still contain the bacillus of rothlauf in a living condition, as shown by inoculation experiments. It was not until about six months after smoking that the bacillus failed to give evidence of vitality.

Thymol.—A five-per-cent solution in alcohol does not destroy

anthrax spores in fifteen days, but the development of these spores is retarded by a solution of 1 : 80,000 (Koch). The anthrax bacillus and staphylococci fail to grow in culture media containing 1 : 3,000 (Samter). The tubercle bacillus is destroyed by contact with thymol for three hours (Yersin). Thymol has about four times less germicidal power than carbolic acid (Behring). Antiseptic in the proportion of 1 : 1,340 (Miquel).

Tobacco Smoke.—Tassinari found that tobacco smoke restrains the development of bacteria, and that certain species failed to develop after exposure for half an hour in an atmosphere of tobacco smoke—spirillum of cholera and Friedländer's bacillus.

XII.

ACTION OF BLOOD SERUM AND OTHER ORGANIC LIQUIDS.

Blood Serum.—Bacteriologists have long been aware of the fact that many species of bacteria, when injected into the circulation of a living animal, soon disappear from the blood, and that the blood of such an animal a few hours after an injection of putrefactive bacteria, for example, does not contain living bacteria capable of developing in a suitable nutrient medium. Wyssokowitsch, in an extensive series of experiments, has shown that non-pathogenic bacteria injected into the circulation may be obtained in cultures from the liver, spleen, kidneys, and bone marrow after they have disappeared from the blood, but that, as a rule, those present in these organs have lost their vitality, as shown by culture experiments, in a period varying from a few hours to two or three days. According to the theory of Metschnikoff, this destruction of bacteria in the blood and tissues of a living animal is effected by the cellular elements, and especially by the leucocytes, which pick up and digest these vegetable cells very much as an amoeba disposes of similar microorganisms which serve it as food. Some such theory seemed necessary to account for the disappearance of bacteria from the blood before the demonstration was made that the serum of the circulating fluid, quite independently of its cellular elements, possesses very decided germicidal power.

Von Fodor first (1887) called attention to the fact that anthrax bacilli may be destroyed by freshly drawn blood; and Nuttall (1888) in an extended series of experiments, showed that various bacteria are destroyed within a short time by the fresh blood of warm-blooded animals. Thus the anthrax bacillus in rabbit's blood was usually killed in from two to four hours when the temperature was maintained at 37°-38° C., and the same result was obtained with pigeon's blood at 41° C. But when the blood was allowed to stand for a considerable time, or was heated for forty-five minutes at 45° C., it served as a culture fluid, and an abundant development of anthrax bacilli occurred in it. *Bacillus subtilis* and *Bacillus me*

therium were also destroyed in two hours by fresh rabbit's blood, but it was without action on *Staphylococcus pyogenes aureus*, which at a temperature of 37.5°C . was found to have increased in numbers at the end of two hours. Further researches by Nissen and Behring show that there is a wide difference in the blood of different animals as to germicidal power, and that certain bacteria are promptly destroyed, while other species are simply restrained for a time in their development or are not affected. Thus Nissen found that the cholera spirillum, the bacillus of anthrax, the bacillus of typhoid fever, and Friedländer's pneumococcus were killed, while *Staphylococcus pyogenes aureus* and *albus*, the streptococcus of erysipelas, the bacillus of fowl cholera, the bacillus of rothlauf, and *Proteus hominis* were able to multiply in rabbit's blood after having been restrained for a short time in their development. In the case of the cholera spirillum a period of ten to forty minutes sufficed for the complete destruction of a limited number, but when the number exceeded 1,200,000 per cubic centimetre they were no longer destroyed with certainty, and after five hours an increase occurred. The anthrax bacillus was commonly destroyed within twenty minutes and the typhoid bacillus at the end of two hours. In the experiments of Behring and Nissen it was found that the most pronounced germicidal effect upon the anthrax bacillus was obtained from the blood of the rat, an animal which has a natural immunity against anthrax; while the blood of the guinea-pig, a very susceptible animal, had no restraining effect and served as a favorable culture medium for the anthrax bacillus. And the remarkable fact was demonstrated that when the blood of a rat was added to the blood of the guinea-pig in the proportion of 1 : 8, it exercised a decided restraining influence upon the growth of the anthrax bacillus. Later researches have shown that cultivation in the blood of an immune animal causes an attenuation of the virulence of an anthrax culture (Ogata and Jasuhara); and also that the injection of the blood of a frog or rat—naturally immune—into a susceptible animal which has been inoculated with a virulent culture of the anthrax bacillus, will prevent the death of the inoculated animal.

Buchner has shown that the germicidal power of the blood of dogs and rabbits does not depend upon the presence of the cellular elements, but is present in clear serum which has been allowed to separate from the clot in a cool place. Exposure for an hour to a temperature of 55°C . destroys the germicidal action of serum as well as of blood; the same effect is produced by heating to 52°C . for six hours or to 45.6°C . for twenty hours. The germicidal power of blood serum is not destroyed by freezing and thawing, but is lost after it has been kept for some time. Buchner's experiments led

him to the conclusion that the germicidal power of fresh blood serum depends upon the presence of some albuminous body present in it. This view is sustained by the researches of Ogata, who has obtained from the blood of dogs and other animals a glycerin extract of a "ferment" which is insoluble in alcohol or in ether and which has germicidal properties.

It has been demonstrated by several experimenters that *other albuminous fluids* possess a similar germicidal power. Thus Nuttall found that a pleuritic exudation from man destroyed the anthrax bacillus in an hour, the aqueous humor of a rabbit in two hours. Wurz has experimented with fresh egg albumin, and found that the anthrax bacillus failed to grow after having been exposed for an hour to the action of albumin from a hen's egg; other bacteria tested were not killed so promptly, but a decided germicidal action was manifested. Prudden has shown that the albuminous fluid obtained from a hydrocele, or from the abdominal cavity in ascites, possesses similar germicidal power; and Fokker has demonstrated that *fresh milk* destroys the vitality of certain bacteria which induce an acid fermentation of this fluid.

The results heretofore referred to induced Hankin to experiment with *cell globulin* obtained from the spleen or lymphatic glands of a dog or cat. This is extracted by means of a solution of chloride of sodium, the solution is filtered, and the globulin precipitated by the addition of alcohol. The precipitate is washed and again dissolved in salt solution. The result showed that this cell globulin possesses germicidal power similar to that of blood serum.

Urine.—The experiments of Lehmann show that fresh urine has a decided germicidal power for the cholera spirillum and the anthrax bacillus, and no doubt for other bacteria as well. To what constituent of the urine this is due has not been determined.

XIII.

PRACTICAL DIRECTIONS FOR DISINFECTION.

THE Committee on Disinfectants of the American Public Health Association (appointed in 1884), after an extended investigation with reference to the germicidal value of various agents, in a final report submitted in 1887 submits the following "Conclusions":

The experimental evidence recorded in this report seems to justify the following conclusions:

The most useful agents for the destruction of spore-containing infectious material are—

1. *Fire.* Complete destruction by burning.
2. *Steam under pressure.* 105° C. (221° F.) for ten minutes.
3. *Boiling in water* for half an hour.
4. *Chloride of lime.*¹ A four-per-cent solution.
5. *Mercuric chloride.* A solution of 1:500.

For the destruction of infectious material which owes its infecting power to the presence of microörganisms not containing spores, the committee recommends—

1. *Fire.* Complete destruction by burning.
2. *Boiling in water* for ten minutes.
3. *Dry heat.* 110° C. (230° F.) for two hours.
4. *Chloride of lime.* A two-per-cent solution.
5. *Solution of chlorinated soda.*² A ten-per-cent solution.
6. *Mercuric chloride.* A solution of 1:2,000.
7. *Carbolic acid.* A five per-cent solution.
8. *Sulphate of copper.* A five-per-cent solution.
9. *Chloride of zinc.* A ten-per-cent solution.
10. *Sulphur dioxide.*³ Exposure for twelve hours to an atmosphere containing at least four volumes per cent of this gas in presence of moisture.

The committee would make the following recommendations with reference to the practical application of these agents for disinfecting purposes:

FOR EXCRETA.

(a) In the sick-room:

1. Chloride of lime in solution, four per cent.

In the absence of spores:

2. Carbolic acid in solution, five per cent.
3. Sulphate of copper in solution, five per cent.

¹ Should contain at least twenty-five per cent of available chlorine.

² Should contain at least three per cent of available chlorine.

³ This will require the combustion of between three and four pounds of sulphur for every thousand cubic feet of air space.

- (b) In privy vaults:
 1. Mercuric chloride in solution, 1:500.¹
 2. Carbolic acid in solution, five per cent.
- (c) For the disinfection and deodorization of the surface of masses of organic material in privy vaults, etc.:
 - Chloride of lime in powder.

FOR CLOTHING, BEDDING, ETC.

- (a) Soiled underclothing, bed linen, etc.:
 1. Destruction by fire, if of little value.
 2. Boiling for at least half an hour.
 3. Immersion in a solution of mercuric chloride of the strength of 1:2,000 for four hours.
 4. Immersion in a two-per-cent solution of carbolic acid for four hours.
- (b) Outer garments of wool or silk, and similar articles, which would be injured by immersion in boiling water or in a disinfecting solution:
 1. Exposure in a suitable apparatus to a current of steam for ten minutes.
 2. Exposure to dry heat at a temperature of 110° C. (230° F.) for two hours.
- (c) Mattresses and blankets soiled by the discharges of the sick:
 1. Destruction by fire.
 2. Exposure to superheated steam, 105° C. (221° F.), for ten minutes.
(Mattresses to have the cover removed or freely opened.)
 3. Immersion in boiling water for half an hour.

FURNITURE AND ARTICLES OF WOOD, LEATHER, AND PORCELAIN.

Washing, several times repeated, with—

1. Solution of carbolic acid, two per cent.

FOR THE PERSON.

The hands and general surface of the body of attendants of the sick, and of convalescents, should be washed with—

1. Solution of chlorinated soda diluted with nine parts of water, 1:10.
2. Carbolic acid, two-per-cent solution.
3. Mercuric chloride, 1:1,000.

FOR THE DEAD.

Envelop the body in a sheet thoroughly saturated with—

1. Chloride of lime in solution, four per cent.
2. Mercuric chloride in solution, 1:500.
3. Carbolic acid in solution, five per cent.

FOR THE SICK-ROOM AND HOSPITAL WARDS.

- (a) While occupied, wash all surfaces with—
 1. Mercuric chloride in solution, 1:1,000.
 2. Carbolic acid in solution, two per cent.
- (b) When vacated, fumigate with sulphur dioxide for twelve hours, burning at least three pounds of sulphur for every thousand cubic feet of air space in the room; then wash all surfaces with one of the above-mentioned disinfecting solutions, and afterward with soap and hot water; finally throw open doors and windows, and ventilate freely.

¹ The addition of an equal quantity of potassium permanganate as a deodorant, and to give color to the solution, is to be recommended. [The writer no longer indorses this recommendation. See his paper on "The Disinfection of Excreta," appended.]

FOR MERCHANDISE AND THE MAILS.

The disinfection of merchandise and of the mails will only be required under exceptional circumstances; free aëration will usually be sufficient. If disinfection seems necessary, fumigation with sulphur dioxide will be the only practicable method of accomplishing it without injury.

RAGS.

(a) Rags which have been used for wiping away infectious discharges should at once be burned.

(b) Rags collected for the paper-makers during the prevalence of an epidemic should be disinfected, before they are compressed in bales, by—

1. Exposure to superheated steam of 105° C. (221° F.) for ten minutes.
2. Immersion in boiling water for half an hour.

SHIPS.

(a) Infected ships at sea should be washed in every accessible place, and especially the localities occupied by the sick, with—

1. Solution of mercuric chloride, 1:1,000.
2. Solution of carbolic acid, two per cent.

The bilge should be disinfected by the liberal use of a strong solution of mercuric chloride.

(b) Upon arrival at a quarantine station, an infected ship should at once be fumigated with sulphurous acid gas, using three pounds of sulphur for every thousand cubic feet of air space; the cargo should then be discharged on lighters; a liberal supply of the concentrated solution of mercuric chloride (four ounces to the gallon) should be thrown into the bilge, and at the end of twenty-four hours the bilge water should be pumped out and replaced with pure sea water; this should be repeated. A second fumigation, after the removal of the cargo, is recommended; all accessible surfaces should be washed with one of the disinfecting solutions heretofore recommended, and subsequently with soap and hot water.

FOR RAILWAY CARS.

The directions given for the disinfection of dwellings, hospital wards, and ships apply as well to infected railway cars. The treatment of excreta with a disinfectant, before they are scattered along the tracks, seems desirable at all times in view of the fact that they may contain infectious germs. During the prevalence of an epidemic of cholera this is imperative. For this purpose the standard solution of chloride of lime is recommended.

DISINFECTION BY STEAM.

The Committee on Disinfectants, in the above-quoted "Conclusions," recommends the use of "steam under pressure, 105° C. (221° F.), for ten minutes" for the destruction of spore-containing infectious material. The spores of all known pathogenic bacteria are destroyed by a temperature of 100° C. maintained for five minutes, and in view of this fact the temperature fixed by the committee is ample, and to exact a higher temperature or longer exposure would be unreasonable. But in practical disinfection the temperature required to destroy infectious material is not the only question to be considered. Economy in the construction and operation of the steam disinfecting apparatus must have due attention, and an important point relates

to the penetration of porous, non-conducting articles, such as rolls of blankets, clothing, etc. These points have been the subject of numerous experimental investigations, and the principles involved have been elucidated, especially by the investigations of Esmarch (1887), of Budde (1889), and of Teuschner (1890).

It has been shown that streaming steam is more effective than confined steam at the same temperature, because it penetrates porous objects more quickly. Also that superheated, "dry" steam is not as effective as flowing steam at 100° C.; on the other hand, it corresponds in effectiveness with dry air, and the temperature must be raised to 140° to 150° C. in order to quickly destroy the spores of bacilli.

Esmarch's investigations show that streaming steam penetrates porous objects, like rolled blankets, more readily than confined steam; but the later researches of Budde and of Teuschner show that a temperature of 100° C. is more rapidly reached in the interior of such rolls when the flowing steam is under pressure. With the same pressure (fifteen pounds) a temperature of 100° C. was reached in two and one-half minutes when the steam was flowing, and in eleven minutes by steam at rest (Budde). Intermittent pressure was not found by Budde to present any advantages over continuously flowing steam; on the contrary, the time of penetration was longer.

Teuschner, whose investigations are the most recent, arrives at the following conclusions:

1. Strongly superheated steam is not to be recommended for practical disinfection. On the contrary, a slight superheating of the steam, such as occurs in the apparatus of Schimmel, is not objectionable.

2. Those forms of apparatus in which the steam enters from above are much safer and quicker in their disinfecting action than those in which this is not the case. In the construction of such apparatus care must be taken, in order to secure penetration of the objects, that the air and steam have a free escape below.

3. Disinfection is hastened by previously warming the apparatus.

4. The most rapid disinfecting action is secured by the use of streaming steam in a state of tension (under pressure).

5. Objects which have been in contact with fatty or oily substances require a longer time for disinfection than those which have not.

6. To accomplish disinfection it is necessary to expel, as completely as possible, all air from the objects to be disinfected, and also to secure a sufficient condensation of the steam.

7. The condensation of the steam advances in a sharply defined line from the periphery to the centre of porous objects.

8. The temperature necessary for disinfection is only found in the zone where condensation has already taken place.

9. Only a few centimetres from the zone in which the temperature is 100° C.—when disinfection is incomplete—there may be places in which the temperature is 40° C. or more below the boiling point.

DISINFECTION OF THE HANDS.

The importance of a reliable method of disinfecting the hands of surgeons, obstetricians, and nurses after they have been in contact with infectious material from wounds, puerperal discharges, etc., is now fully recognized, and some surgeons consider it necessary to completely sterilize the hands before undertaking any surgical operation which will bring them in contact with the freshly-cut tissues. The numerous experiments which have been made with a view to ascertaining the best method of accomplishing such sterilization of the hands show that it is by no means a simple matter to effect it, and especially to insure the destruction of microorganisms concealed beneath the finger nails. Fürbringer, in an extended series of experiments (1888), found that a preliminary cleansing with soap and a brush was even more important than the degree of potency of the disinfecting wash subsequently applied. He recommends the following procedure :

1. Remove all visible dirt from beneath and around the nails.
2. Brush the spaces beneath the nails with soap and hot water for a minute.
3. Wash for a minute in alcohol (not below eighty per cent), and, before this evaporates, in the following solution :
4. Wash thoroughly for a minute in a solution containing 1 : 500 of mercuric chloride or three per cent of carbolic acid.

Roux and Reynés tested the above method of Fürbringer, and found that it gave better results than others previously proposed, although not always entirely successful in securing complete sterilization.

Boll has recently (1890) reported favorable results from the following method :

1. Cleanse the finger nails from visible dirt with knife or nail scissors.
2. Brush the hands for three minutes with hot water and potash soap.
3. Wash for half a minute in a three-per-cent solution of carbolic acid, and subsequently in a 1 : 2,000 solution of mercuric chloride.
4. Rub the spaces beneath the nails and around their margins with iodo-form gauze wet in a five-per-cent solution of carbolic acid.

Welch, as a result of extended experiments made at the Johns Hopkins Hospital, recommends the following procedure :

1. The nails are kept short and clean.
2. The hands are washed thoroughly for several minutes with soap and water, the water being as warm as can be comfortably borne, and being frequently changed. A brush sterilized by steam is used. The excess of soap is washed off with water.
3. The hands are immersed for one or two minutes in a warm saturated solution of permanganate of potash and are rubbed over thoroughly with a sterilized swab.

4. They are then placed in a warm saturated solution of oxalic where they remain until complete decolorization of the permanganate occurs.

5. They are then washed off with sterilized salt solution or water.

6. They are immersed for two minutes in sublimate solution, 1 : 5000.

The bacteriological examination of the skin thus treated yields all uniformly negative results, the material for the cultures being taken underneath and around the nails. This is the procedure now employed in the gynecological and surgical wards of the hospital.

THE DISINFECTION OF EXCRETA.

The following paper by the present writer was read before the Section on State Medicine at the last (1891) meeting of the American Medical Association :

The Committee on Disinfectants appointed by the American Public Health Association in 1884, in its final report submitted in 1887, gives the following *general directions* :

"*Disinfection of Excreta, etc.*—The infectious character of the dejecta of patients suffering from cholera and from typhoid fever is well established and this is true of mild cases and of the earliest stages of these diseases as well as of severe and fatal cases. It is probable that epidemic dysentery, tuberculosis, and perhaps diphtheria, yellow fever, scarlet fever, and typhoid fever, may also be transmitted by means of the alvine discharges of the sick. It is, therefore, of the first importance that these should be disinfectcd. In cholera, diphtheria, yellow fever, and scarlet fever all vomited material should also be looked upon as infectious. And in tuberculous diphtheria, scarlet fever, and infectious pneumonia the sputa of the sick should be disinfected or destroyed by fire. It seems advisable also to disinfect the urine of patients sick with an infectious disease with one of the disinfecting solutions below recommended.

"Chloride of lime, or bleaching powder, is perhaps entitled to the place for disinfecting excreta, on account of the rapidity of its action.

"The following standard solution is recommended :

"*Dissolve chloride of lime of the best quality,¹ in pure water, in the proportion of six ounces to one gallon.* Use one quart of this solution for the disinfection of each discharge in cholera, typhoid fever, etc."² Mix well and leave in the vessel for at least one hour before throwing into the privy or water closet.

"The same directions apply to the disinfection of vomited matters. Infectious sputum should be discharged directly into a cup half-full of the disinfectant. A five-per-cent solution of carbolic acid may be used instead of the chloride of lime solution, the time of exposure to the action of the disinfectant being four hours" (op. cit., pp. 237, 238).

The object of this paper is to inquire whether these recommendations which were based upon the experimental data available at the time they were made, are sustained by subsequent investigations; and whether other agents have been shown to possess superior advantages for the purpose in view.

But first we desire to call attention to another portion of the report of

¹ Good chloride of lime should contain at least twenty-five per cent of available chlorine (page 92). It may be purchased by the quantity at three and one-half cents per pound. The cost of the standard solution recommended is therefore but more than one cent a gallon. A clear solution may be obtained by filtration and decantation, but the insoluble sediment does no harm and this is an unnecessary refinement.

² For a very copious discharge use a larger quantity.

Committee on Disinfectants. On page 236 the following definition of disinfection and disinfectants is given:

"The object of disinfection is to prevent the extension of infectious diseases by destroying the specific infectious material which gives rise to them. This is accomplished by the use of disinfectants. There can be no partial disinfection of such material; either its infecting power is destroyed or it is not. In the latter case there is a failure to disinfect. *Nor can there be any disinfection in the absence of infectious material.*"

I have italicized the last sentence because I wish to call especial attention to it. I am frequently asked, "What is the best disinfectant to put into a water closet?" Now, if a closet or privy vault is resorted to only by healthy persons and no infectious material has been thrown into it, there is nothing in it to disinfect, and the recommendation of the Committee on Disinfectants does not apply to it at all. It may smell badly, and in this case the bad odor may be neutralized by the use of deodorants; or we may prevent the putrefactive decomposition of its contents, and thus prevent the formation of the offensive gases given off as a result of such decomposition, by the use of antiseptics. But to accomplish this it is not necessary to sterilize the entire contents by the use of active germicidal agents.

A solution of sulphate of iron or of chloride of zinc is a useful antiseptic and deodorizing agent, and the Committee on Disinfectants, in making its recommendations, did not intend to discourage the use of such agents. But exact experimental data showed that these agents could not be depended upon for the destruction of infectious disease germs, and the recommendations made related to disinfection in the strict and proper use of the term as above defined. This definition is now accepted by sanitarians in all parts of the world, but many practising physicians still use the term disinfectant as synonymous with deodorant. For example, I find in a recent sanitary periodical, under the heading "Medical Excerpt," an item copied from the *American Journal of Obstetrics*, to which the name of a distinguished gynecologist is attached, in which the following statement is made with reference to a much-advertised so-called "disinfectant": "As a disinfectant I have used it in my house for over a year with great satisfaction." Now, the agent referred to has been proved by exact experiments to have comparatively little disinfecting power, although it is a very good deodorant. According to our definition, "the object of disinfection is to prevent the extension of infectious diseases by destroying the specific infectious material which gives rise to them." Are we to suppose that the distinguished gynecologist above quoted had such infectious material in his house "for over a year" at the time he was employing "with great satisfaction" the agent he recommends? If not, the term was improperly employed, for "there can be no disinfection in the absence of infectious material." I wish to emphasize this point, because I have reason to believe that, in the army at least, the recommendation of the Committee on Disinfectants has led to the substitution of chloride of lime for cheaper deodorants and antiseptic agents—and especially for sulphate of iron—in latrines which are frequented only by healthy persons and consequently need no disinfection. The amount of chloride of lime issued from the Medical Purveying Depot at San Francisco during the past six months for use at military posts on the Pacific coast is more than double the amount of sulphate of iron; but there has been no epidemic of an infectious disease, and probably comparatively little call for the use of a disinfecting agent in the sick-room. We quote again from the report of the Committee on Disinfectants:

"In the sick-room we have disease germs at an advantage, for we know where to find them as well as how to kill them. Having this knowledge, not to apply it would be criminal negligence, for our efforts to restrict the extension of infectious diseases must depend largely upon the proper use of disinfectants in the sick-room" (op. cit., p. 237).

"The injurious consequences which are likely to result from such misapprehension and misuse of the word disinfectant will be appreciated when

it is known that recent researches have demonstrated that many of the agents which have been found useful as deodorizers or as antiseptics are entirely without value for the destruction of disease germs.

"This is true, for example, as regards the sulphate of iron, or copper salt which has been extensively used with the idea that it is a valuable antiseptic. As a matter of fact, sulphate of iron in saturated solution does not destroy the vitality of disease germs, or the infecting power of materials containing them. This salt is, nevertheless, a very valuable antiseptic, its low price makes it one of the most available agents for the arrest of putrefactive decomposition" (op. cit., p. 237).

Chloride of lime is also a valuable antiseptic and deodorant, and I know of no objection to substituting it for sulphate of iron other than the question of cost. The first cost of chloride of lime, by the quantity, is about double that of sulphate of iron, but practically the difference is much greater, because it is necessary to preserve the chloride of lime in air-tight packages. When exposed to the air it deteriorates in value very rapidly. It is, therefore, necessary to pack it in air-tight receptacles which will not be injured by the corrosive action of free chlorine, and in comparatively small quantities so that the contents of a package may be used soon after it is opened.

We now proceed to consider the experimental data relating to the germicidal value of *chloride of lime*.

The Committee on Disinfectants gave it "the first place for disinfecting excreta, on account of the rapidity of its action." This recommendation is based upon experimental data obtained in the pathological laboratory of the Johns Hopkins University, under the writer's direction, and is sustained by recent experiments made in Germany.

The experiments of Bolton, made for the Committee on Disinfectants in 1886, gave the following results: The time of exposure being two hours, typhoid bacillus and cholera spirillum in bouillon cultures were killed by a solution containing one part to one thousand parts of water (containing 0.1 per cent of available chlorine). Anthrax spores were killed in the same time by a solution containing 0.3 per cent of available chlorine. Typhoid bacilli were sterilized by a two-per-cent solution, and in several instances by a half-per-cent solution; but some resistant spores of non-pathogenic bacilli survived in two experiments in which a solution of 1:100 was used. In bouillon cultures to which ten per cent of dried egg albumin had been added, the typhoid bacillus was destroyed by one-half per cent (1:200).

Nissen, whose experiments were made in Koch's laboratory in 1890, found that anthrax spores were destroyed in thirty minutes by a five-per-cent solution, and in seventy minutes by a one-per-cent solution. In his experiments the typhoid bacillus and the cholera spirillum were destroyed with certainty in five minutes by a solution containing 0.12 per cent (1:833); anthrax bacillus in one minute by 1:1,000; *Staphylococcus pyogenes aureus* in one minute by 1:500. Experiments made by the same author on the sterilization of faeces showed that one per cent could be relied upon to destroy the bacillus of typhoid fever and the spirillum of cholera in faeces in ten minutes.

Carbolic Acid.—The Committee on Disinfectants says: "A five-per-cent solution of carbolic acid may be used instead of the chloride of lime solution, the time of exposure to the action of the disinfectant being four hours. This recommendation is made in view of the fact that in those diseases in which it is most important to disinfect the excreta the specific germ does not form spores. This is now believed to be true of the typhoid bacillus, the spirillum of cholera, the bacillus of diphtheria, the bacillus of glanders, the streptococcus of erysipelas; and it has been shown by exact experiments that all of these pathogenic bacteria are destroyed in two hours by a one-per-cent solution, or less, of this agent.

Spores require for their destruction a stronger solution and a longer time. Koch found a one-per-cent solution to be without effect on anthrax spores after fifteen days' exposure; a two-per-cent solution retarded their develop-

ment, but did not destroy their vitality in seven days; a three-per cent solution was effective in two days. According to Nocht, at a temperature of 37.50° C. anthrax spores are killed by a five-per cent solution in three hours.

Carbolic acid possesses the advantage of not being neutralized by the substances found in excreta, or by the presence of albumin. Thus Bolton found that the addition of ten per cent of dried albumin to a bouillon culture of the typhoid bacillus did not materially influence the result, the bacillus being destroyed in two hours by a one-per cent solution.

This agent, then, is firmly established as a valuable disinfectant for excreta, but we still give the preference to the standard solution of chloride of lime of the Committee on Disinfectants for use in the sick-room, "on account of the rapidity of its action," and also on account of its comparative cheapness.

At the International Sanitary Conference at Rome (1885) the writer, who was associated with Dr. Koch on the Committee on Disinfectants, presented the claims of chloride of lime, and in the recommendations of the committee it was placed beside carbolic acid with the following directions:

"Carbolic acid and chloride of lime are to be used in aqueous solution.

"Weak solutions, carbolic acid, two per cent; chloride of lime, one per cent.

"Strong solutions, carbolic acid, five per cent; chloride of lime, four per cent."

The strong solutions were to be used for the disinfection of excreta.

Creolin, a coal-tar product, which is a syrupy, dark-brown fluid with the odor of tar, has during the past three years received much attention from the German bacteriologists. It is probably the same product which was tested under the writer's direction for the Committee on Disinfectants, in 1885, under the name of "Little's soluble phenyle." It stood at the head of the "Commercial Disinfectants" tested. The experiments made in Germany show that it is not so active for spores as carbolic acid, but that it very promptly kills known pathogenic bacteria, in the absence of spores, in solutions of two per cent or less. Eisenberg found that a solution of two per cent killed all test organisms within fifteen minutes. Esmarch found it especially fatal to the cholera spirillum, which was killed by solutions of 1:1,000 in ten minutes. The typhoid bacillus showed much greater resisting power—a one-half-per cent solution failed after ten minutes' exposure. The pus cocci was still more resistant. Behring has shown that the presence of albumin greatly diminishes its germicidal power. As a deodorant it is superior to carbolic acid, and on this account is to be preferred in the sick-room. A recently prepared emulsion may be used to disinfect the liquid excreta of cholera or typhoid patients, in the proportion of four per cent, two hours' time being allowed for the action of the disinfectant. The experiments of Jäger upon pure cultures of the tubercle bacillus attached to silk threads were successful in destroying the infecting power of these cultures, as tested by inoculation into the anterior chamber of the eye of a rabbit, when solutions of two per cent were used.

The value of this agent as a disinfectant is then fully established; as to its cost in comparison with the agents heretofore mentioned I am not informed.

Quicklime.—Experiments made in Koch's laboratory in 1887 by Liborius led him to place a high value upon recently burned quicklime as a disinfectant. More recent experiments by Jäger, Kitasato, Pfuhl, and others have shown that this agent has considerable germicidal power in the absence of spores, and that the value which has long been placed upon it for the treatment of excrementitious material in latrines, etc., and as a wash for exposed surfaces, is justified by the results of exact experiments made upon known pathogenic bacteria. The germicidal power of lime is not interfered with by the presence of albuminous material, but is neutralized by phosphates, carbonates, and other bases, and by carbonic acid.

In the writer's experiments a saturated aqueous solution of calcium oxide

failed to kill typhoid bacilli; but when suspended in water in the proportion of 1 : 40 by weight this bacillus was killed at the end of two hours. Antiseptics were not killed in the same time by a lime wash containing two per cent by weight of pure calcium oxide. According to Kitasato typhoid bacillus and the cholera spirillum in bouillon cultures are destroyed by the addition of one-tenth per cent of calcium oxide. Pfuhl experimented upon sterilized fæces to which pure cultures of the typhoid bacillus and cholera spirillum were added. The liquid discharges of patients with typhoid fever or diarrhoea were used for the purpose. He found that sterilization was effected at the end of two hours by adding fragments of calcium hydrate in the proportion of six per cent, and that three per cent was effective in six hours. When a milk of lime was used which could be thoroughly mixed with the dejecta the result was still more favorable. A standard preparation of milk of lime containing twenty per cent of calcium hydrate killed typhoid bacillus and the cholera spirillum in one hour when added to liquid fæces in the proportion of two per cent.

The experiments with this agent show that time is an important factor and that much longer exposures, as well as stronger solutions, are required to destroy pathogenic bacteria than is the case with chloride of lime. For this reason we still give the last-named agent the preference for the disinfection of excreta in the sick-room. But in latrines the time required to accomplish disinfection is of less importance, and *we are disposed to give recently burned quicklime the first place for the disinfection of excreta in privies, vaults or on the surface of the ground.* It may be applied in the form of a milk of lime, prepared by adding gradually eight parts, by weight, of quicklime to one part of calcium hydrate. This must be freshly prepared, or protected from the air to prevent the formation of the inactive carbonate of lime.

According to Behring, lime has about the same germicidal value as other caustic alkalies, and destroys the cholera spirillum and the bacilli of typhoid fever, of diphtheria, and of glanders after several hours' exposure in the proportion of fifty cubic centimetres *normal-lauge* per litre. Vaseline, ashes or lye of the same alkaline strength may therefore be substituted for quicklime.

Finally, it must not be forgotten that we have a ready means of disinfecting excreta in the sick-room or its vicinity by the application of boiling water. Exact experiments, made by the writer and others, show that the death-point of the following pathogenic bacteria, and of the kinds of fungi mentioned is below 60° C. (140° F.): Spirillum of cholera, bacillus of anthrax, bacillus of typhoid fever, bacillus of diphtheria, bacillus of glanders, diplococcus of pneumonia (*Micrococcus Pasteuri*), streptococcus of erysipelas, staphylococci of pus, micrococcus of gonorrhoea, vaccine virus, sheep virus, hydrophobia virus. Ten minutes' exposure to the temperature mentioned may be relied upon for the disinfection of material containing any of these pathogenic organisms, except the anthrax bacillus when in the stage of spore formation. The use, therefore, of *boiling water in the proportion of three or four parts to one part of the material to be disinfected* may be safely recommended for such material. Or, better still, a ten-per-cent solution of sulphate of iron or of chloride of zinc at the boiling point may be used in the same way (three parts to one). This will have a higher boiling point than water, and will serve at the same time as a deodorant. During an epidemic of cholera or typhoid fever such a solution might be kept in a proper receptacle in the vicinity of hospital wards containing patients, and would serve to conveniently, promptly, and cheaply disinfect all excreta.

DISINFECTION IN DIPHTHERIA.

At the meeting of the Tenth International Medical Congress at Berlin (1890) Löffler made an important communication upon

measures to be taken to prevent the spread of diphtheria. His conclusions are summarized as follows :

1. The cause of diphtheria is the diphtheria bacillus, which is found in the secretions of the affected mucous membrane.

2. With this secretion it is distributed outside of the body and may be deposited upon anything in the vicinity of the sick.

3. Those sick with diphtheria carry about bacilli capable of infecting others so long as there is the slightest trace of diphtheritic deposit, and even for several days after such deposit has disappeared.

4. Those sick with diphtheria are to be rigidly isolated so long as the diphtheria bacilli are present in their secretions. Children who have been sick with diphtheria should be kept from school for at least four weeks.

5. The diphtheria bacilli may preserve their vitality in dried fragments of diphtheritic membrane for four or five months. Therefore all objects which may have been exposed to contact with the excretions of those sick with diphtheria, such as linen, bedclothing, utensils, clothing of nurses, etc., should be disinfected by boiling in water or treated with steam at 100° C. In the same way the rooms occupied by diphtheria patients are to be carefully disinfected. The floors should be repeatedly scrubbed with hot sublimate solution (1:1,000) and the walls rubbed down with bread.

The recommendation made by Löffler with reference to rubbing down the walls of an infected apartment with bread is based upon the experiments of Esmarch (1887), as a result of which he arrived at the conclusion that this is the most reliable method of removing bacteria attached to the walls of an apartment. Fresh bread is used, and, after having been used, is destroyed by burning. We judge that this method would be especially applicable to painted surfaces or to walls covered with paper. For plastered walls the liberal application of lime wash is probably the safest method of disinfection.

PART THIRD.

PATHOGENIC BACTERIA.

- I. MODES OF ACTION. II. CHANNELS OF INFECTION. III. SUSCEPTIBILITY AND IMMUNITY. IV. PYOGENIC BACTERIA. V. BACTERIA IN CROUPOUS PNEUMONIA. VI. PATHOGENIC MICROCOCCI NOT DESCRIBED IN SECTIONS IV. AND V. VII. THE BACILLUS OF ANTHRAX. VIII. THE BACILLUS OF TYPHOID FEVER. IX. BACTERIA IN DIPHTHERIA. X. BACTERIA IN INFLUENZA. XI. BACILLI IN CHRONIC INFECTIOUS DISEASES. XII. BACILLI WHICH PRODUCE SEPTICÆMIA IN SUSCEPTIBLE ANIMALS. XIII. PATHOGENIC AÆROBIC BACILLI NOT DESCRIBED IN PREVIOUS SECTIONS. XIV. PATHOGENIC ANAÆROBIC BACILLI. XV. PATHOGENIC SPIRILLA. XVI. BACTERIA IN INFECTIOUS DISEASES NOT PROVED TO BE OF PARASITIC ORIGIN. XVII. CLASSIFICATION OF PATHOGENIC BACTERIA.

PART THIRD.

PATHOGENIC BACTERIA.

I.

MODES OF ACTION.

MANY of the saprophytic bacteria are pathogenic for man, or for one or more species of the lower animals, when by accident or experimental inoculation they obtain access to the body ; these may be designated *facultative parasites*. Other species which, for a time at least, are able to lead a saprophytic mode of life have their normal habitat in the bodies of infected animals, in which they produce specific infectious diseases. To this class belong the cholera spirillum, the anthrax bacillus, the bacillus of typhoid fever, and various other microorganisms which are the cause of specific infectious diseases in some of the lower animals. These we may speak of as *parasites* and *facultative saprophytes*. Still others are *strict parasites* and do not find the conditions for their development outside of the bodies of the animals which they infest, except under the special conditions in which bacteriologists have succeeded in cultivating some of them. The best known strict parasites are the tubercle bacillus, the bacillus of leprosy, the spirillum of relapsing fever, and the micrococcus of gonorrhœa.

There can be but little doubt that even the strict parasites, at some time in the past, were also saprophytes, and that the adaptation to a parasitic mode of life was gradually effected under the laws of natural selection. In a previous chapter (Section III., Part Second) we have referred to the modifications in biological characters which may occur as a result of special conditions of environment. Thus we may obtain non-chromogenic varieties of species which usually produce pigment, or non-pathogenic varieties of bacteria which are usually pathogenic. There is also evidence that the tubercle bacillus, a strict

parasite, may be so modified, by cultivation for successive generations in a culture medium containing glycerin, that it will finally grow in ordinary beef infusion, thus showing a tendency to adapt itself to a saprophytic mode of life.

Some of the saprophytic bacteria are indirectly pathogenic because of their power to multiply in articles of food, such as milk, cheese, fish, sausage, etc., and there produce poisonous ptomaines which, when these articles are ingested, give rise to various morbid symptoms, such as vomiting, gastric and intestinal irritation, fever, etc. Or similar symptoms may result from the multiplication of *bacteria producing toxic ptomaines in the alimentary canal*. No doubt gastric and intestinal disorders are largely due to this cause and may be induced by a variety of saprophytic bacteria when they establish themselves in undue numbers in any portion of the alimentary tract. In Asiatic cholera the same thing occurs, but with more fatal results from the introduction of the East Indian cholera germ discovered by Koch. This is pathogenic for man, because it is able to multiply rapidly in the human intestine, and there produces a toxic substance which, being absorbed, gives rise to the morbid phenomena of the disease. The spirillum itself does not enter the blood or invade the tissues, except to a limited extent in the mucous coat of the intestine, and the true explanation of its pathogenic power is no doubt that which has been given.

Other microorganisms invade the tissues and multiply in certain favorable localities, but have not the power of developing in the blood, in which they are only found occasionally and in very small numbers or not at all. Thus the typhoid bacillus locates itself in the intestinal glands, in the spleen, and in the liver, forming colonies to a limited extent, and evidently not finding the conditions extremely favorable for its growth, inasmuch as it does not take complete possession of these organs. The symptoms which result from its presence are doubtless partly due to local irritation, disturbance of function, and, in the case of the intestinal glands, necrotic changes induced by it. But in addition to this its pathogenic action depends upon the production of a poisonous ptomaine which has been isolated and studied by the German chemist Brieger (typhotoxine).

Certain saprophytic bacteria, when injected beneath the skin of a susceptible animal, multiply at the point of inoculation and invade the surrounding tissues, giving rise in some instances to the formation of a local abscess, in others to an infiltration of the tissues with bloody serum, and in others to extensive necrotic changes. These local changes are due not simply to the mechanical presence of the microorganisms which induce them, but to chemical products evolved during the growth of these pathogenic bacteria. Indeed, their patho-

genic power evidently depends, in some instances at least, upon these toxic products of their growth, by which the vital resisting power of the tissues is overcome.

Among the bacteria which in this way produce extensive local inflammatory and necrotic changes are certain anaërobic species found in the soil and in putrefying material, such as the bacillus of malignant oedema and the writer's *Bacillus cadaveris*. The bacillus of symptomatic anthrax, an infectious disease of cattle, acts in the same way. All of these produce toxic substances which have a very pronounced local action upon the tissues invaded by them. Other bacteria, while they develop chiefly in the vicinity of the point of entrance—by accident or by inoculation—produce a potent toxic substance which gives rise to general symptoms of a serious character, such as tetanic convulsions (bacillus of tetanus) or intense fever and nervous phenomena (micrococcus of erysipelas). Again, the local irritation resulting from the presence of parasitic bacteria may primarily give rise to the formation of new growths having a low grade of vitality, which later may undergo necrotic changes, as in tuberculosis, glanders, and leprosy. In this case constitutional symptoms are not present, or are of a mild character during the development of these new formations, which apparently result from the local action of substances eliminated during the growth of the parasite, rather than from its simple presence. This is an inference based upon the fact that non-living particles, or even living parasites, as in trichinosis, do not produce similar new growths composed of cells; but become encysted in a fibrous capsule.

In pneumonia we have a local process in which one or more lobes of the lung are invaded by a pathogenic micrococcus (*Micrococcus pneumoniae croupose*) which induces a fibrinous exudation that completely fills the air cells. How far the symptoms of the disease are due to the local inflammation and disturbance of function, and to what extent they may be due to the absorption of a soluble toxic substance evolved as a result of the growth of the micrococcus, has not been determined. But the mild character of the general symptoms when a limited area of lung tissue is involved leads to the inference that the pathogenic power of this particular pathogenic microorganism is chiefly exercised locally.

The pus cocci and various other saprophytic bacteria, when introduced beneath the skin, give rise to the *formation of abscesses*, unattended by any very considerable general disturbance; and also to secondary purulent accumulations—metastatic abscesses.

That this is not due simply to their mechanical presence is shown by the fact that powdered glass and other inert substances, when thoroughly sterilized, do not give rise to pus formation when intro-

duced beneath the skin or injected into the cavity of the abdomen. On the other hand, it has been demonstrated by the experiments of Grawitz, De Bary, and others that certain chemical substances which act as local irritants when brought in contact with the tissues may induce pus formation quite independently of microorganisms: nitrate of silver, oil of turpentine, and strong liquor ammoniæ have been shown to possess this power. And it has been demonstrated by the recent experiments of Buchner that sterilized cultures of a long list of different bacteria—seventeen species tested—give rise to supuration when introduced into the subcutaneous tissues.

Buchner has further shown that this property of inducing pus formation resides in the dead bacterial cells and not in soluble products present in the cultures. For the clear fluid obtained by passing these sterilized cultures through a porcelain filter gave a negative result, while the bacteria retained by the filter, although no longer capable of development, having been killed by heat, invariably caused suppuration.

Individuals suffering from malnutrition are more susceptible to invasion by specific disease germs or by the common pus cocci than are those in vigorous health. Thus the sufferers from starvation, from crowd poisoning, sewer-gas poisoning, etc., are not only liable to be early victims during the prevalence of an epidemic disease, but are very subject to abscesses, boils, ulcers, etc. A slight abrasion in such an individual, inoculated by the ever-present pus cocci, may give rise to an obstinate ulcer or a phlegmonous inflammation.

In the same way some of the ordinary saprophytes, which usually have no pathogenic power, may be pathogenic for an animal whose strength is reduced by disease or injury. Thus necrotic changes may occur in injured tissues, or in those which have a deficient blood supply—from occlusion of an artery, for example—due to the presence of putrefactive bacteria which are incapable of development in the circulation of a healthy animal or in healthy tissues. We may also have a *progressive gangrene*, due to infection of wounds by bacteria which are able to invade healthy tissues. This is seen in the so-called hospital gangrene, which is undoubtedly due to microorganisms, although the species concerned in its production has not been determined, owing to the fact that modern bacteriologists have had few, if any, opportunities for studying it. The history of the disease, its rapid extension in infected surgical wards, the extensive sloughing which occurs within a few hours in previously healthy wounds, and the effect of deep cauterization by the hot iron, nitric acid, or bromine in arresting the progress of the disease, all support this view of its etiology. Whether it is due to a specific pathogenic micro-

organism, or to exceptional pathogenic power acquired by some one of the common bacteria which infest suppurating wounds, cannot be determined in the absence of exact experiments by modern methods. But the latter view has seemed to the writer the most probably correct. There are many facts which go to show that pathogenic virulence may be increased by cultivation in animal fluids, and where wounded men are brought together under unfavorable sanitary conditions, as has been the case where hospital gangrene has made its appearance, it may be that some common saprophyte acquires the power of invading the exposed tissues instead of simply feeding upon the secretions which bathe its surface.

Koch has described a progressive tissue necrosis in mice, due to a streptococcus, which he first obtained by inoculating a mouse in the ear with putrid material. The morbid process is entirely local and rapidly progressive, causing a fatal termination in about three days, without invasion of the blood.

In *diphtheritic inflammations* of mucous membranes we have a local invasion of the tissues and a characteristic plastic exudation. In true diphtheria the local inflammation and necrotic changes in the invaded tissues are not sufficient to account for the serious general symptoms, and we now have experimental evidence that the diphtheria bacillus produces a very potent toxic substance to which these symptoms are no doubt largely due. The diphtheria bacillus of Löffler appears to be the cause of the fatal malady which goes by this name, but undoubtedly other microorganisms may be concerned in the formation of diphtheritic false membranes. In certain forms of diphtheria, and especially when it occurs as a complication of scarlet fever, measles, and other diseases, the Klebs-Löffler bacillus is absent, and a streptococcus, which appears to be identical with *Streptococcus pyogenes*, is found in considerable numbers and is probably the cause of the diphtheritic inflammation. An epidemic of diphtheria occurring among calves was studied by Löffler, and is ascribed by him to his *Bacillus diphtheriæ vitulorum*. The same bacteriologist has shown that the diphtheria of chickens and of pigeons is due to a specific bacillus which differs from that found in human diphtheria, and which he calls *Bacillus diphtheriæ columbarum*.

Recently Prof. Welch has studied the histological lesions produced by filtered cultures of the diphtheria bacillus. Cultures in glycerin-bouillon, several weeks old, were filtered through porcelain, and the sterile filtrate was injected beneath the skin of guinea-pigs. One cubic centimetre of this filtrate was injected into a guinea-pig on the 10th of December, and two cubic centimetres more on the 14th of the same month. The animal succumbed at the end of

three weeks and five days after the first inoculation. At the autopsy "the lymphatic glands of the inguinal and axillary regions were found to be enlarged and reddened; the cervical glands were swollen and the thyroid gland was greatly congested. There was a considerable excess of clear fluid in the peritoneal cavity. Both layers of the peritoneum were reddened, the vessels of the visceral layer being especially injected. The spleen was enlarged to double the average size; it was mottled, and the white follicles were distinctly outlined against the red ground. The liver was dark in color and contained much blood. . . . The kidneys were congested and the cut surface was cloudy. . . . The pericardial sac was distended with clear serum. Under the epicardium were many ecchymotic spots. The lungs exhibited areas of intense congestion or actual hæmorrhage into the tissues. . . . The histological lesions in this case are identical with those observed by us in connection with the inoculation of the living organisms."

To what extent non-specific *catarrhal inflammations* of mucous membranes are caused by the local action of microorganisms has not been determined, but in gonorrhœa the proof is now considered satisfactory that the "gonococcus" of Neisser is the cause of the intense local inflammation and purulent discharge. In this disease the action of the pathogenic microorganism seems to be limited to the tissues invaded by it, as there is no general systemic disturbance indicating the absorption of a toxic ptomaine.

Chronic catarrhal inflammations appear, in some cases at least, to be kept up by the presence of microorganisms, which are always found in the discharges from inflamed mucous surfaces.

The influence of microorganisms, and especially of the pus cocci, in preventing the prompt healing of wounds, is now well established. An extensive suppurating wound or collection of pus, especially if putrefactive bacteria are present, causes fever and nervous symptoms, due to the absorption of toxic products. More intense general symptoms result from the presence of the streptococcus of pus than from the less pathogenic staphylococci; this is seen in erysipelatous inflammations and in puerperal metritis due to the presence of this micrococcus. Like the other pus cocci, the *Streptococcus pyogenes* does not invade the blood, but when introduced into the subcutaneous tissues it induces a local inflammatory process, with a tendency to pus formation, and it invades the neighboring lymph channels, in which the conditions appear to be especially favorable for its multiplication.

Finally, certain pathogenic bacteria, when introduced into the bodies of susceptible animals, quickly invade the blood and multiply in it. In so doing they necessarily interfere with its physiological

functions by appropriating for their own use material required for the nutrition of the tissues ; and at the same time toxic substances are formed which play an important part in the production of the morbid phenomena, which in this class of diseases very commonly lead to a fatal result. The pathogenic bacteria which invade the blood may also, in certain cases, give rise to local necrosis and disturbance of function in various organs in a mechanical way by blocking up the capillaries.

The invasion of the blood which occurs in anthrax and in various forms of septicæmia in the lower animals, induced by subcutaneous inoculation with pure cultures of certain pathogenic bacteria, does not generally immediately follow the inoculation. Usually a considerable local development first occurs, which gives rise to more or less inflammation of the invaded tissues, and very commonly to an effusion of bloody serum in which the pathogenic microörganism is found in great numbers. Even in susceptible animals the blood seems to offer a certain resistance to invasion, which is overcome after a time by the vast number of the parasitic host located in the vicinity of the point of inoculation, aided probably by the toxic substances developed as a result of their vital activity.

The experiments of Cheyne (1886) seem to show that in the case of very pathogenic species, like the anthrax bacillus or Koch's bacillus of mouse septicæmia, a single bacillus introduced subcutaneously may produce a fatal result in the most susceptible animals, while greater numbers are required in those which are less susceptible. Thus a guinea-pig succumbed to general infection after being inoculated subcutaneously with anthrax blood diluted to such an extent that, by estimation, only one bacillus was present in the fluid injected ; and a similar result in mice was obtained with *Bacillus murisepticus*. In the case of the microbe of fowl cholera (*Bacillus septicæmiæ hæmorrhagicæ*) Cheyne found that for rabbits the fatal dose is 300,000 or more, that from 10,000 to 300,000 cause a local abscess, and that less than 10,000 produce no appreciable effect. The common saprophyte *Proteus vulgaris* was found to be pathogenic for rabbits when injected into the dorsal muscles in sufficient numbers. But, according to the estimates made, 225,000,000 were required to cause death, while with doses of from 9,000,000 to 112,000,000 a local abscess was produced, and less than 9,000,000 gave an entirely negative result.

Secondary infections occurring in the course of specific infectious diseases are of common occurrence. Thus a pneumonia may be developed in the course of an attack of measles or of typhoid fever ; or infection by the common pus cocci in the course of scarlet fever, typhoid fever, mumps, etc., may give rise to local abscesses,

to endocarditis, etc. Again, *mixed infection* may be induced by injecting simultaneously into susceptible animals two species of pathogenic bacteria.

Bumm, Bockhart, and others have reported cases of mixed gonorrhœal infection in which the pyogenic micrococci gave rise to abscesses in the glands of Bartholin, to cystitis, parametritis, or to "gonorrhœal inflammation" of the knee joint. Babes gives numerous examples of mixed infection in scarlet fever and in other diseases of childhood. Anton and Fütterer have studied the question of secondary infection in typhoid fever. Karlinski has reported a case of secondary infection with anthrax in a case of typhoid fever, infection occurring by way of the intestine. Many other examples of secondary or mixed infection are recorded in the recent literature of bacteriology and clinical medicine, but enough has been said to call attention to the importance of the subject.

II.

CHANNELS OF INFECTION.

WE have abundant evidence that susceptible animals may be infected by the injection of various pathogenic bacteria beneath the skin, and accidental infection *through an open wound or abrasion of the skin* is the common mode of infection in tetanus, erysipelas, hospital gangrene, and the "traumatic infectious diseases" generally. Other infectious diseases, like anthrax and glanders, are frequently transmitted in the same way. We have also satisfactory evidence that tuberculosis may be transmitted to man by the accidental inoculation of an open wound; and in view of the fact that susceptible animals are readily infected in this way, it would be strange if it were otherwise.

The question whether infection may occur through *the unbroken skin* has been studied by several bacteriologists and an affirmative result obtained. Thus Schimmelbusch produced pustules upon the thigh in two young persons suffering from pyæmia by rubbing upon the surface a pure culture of *Staphylococcus pyogenes aureus* which he had obtained from the pus of a furuncle. The same author also succeeded in infecting rabbits and guinea-pigs with anthrax, and rabbits with rabbit septicæmia, by rubbing pure cultures upon the uninjured skin. Similar results had previously been reported by Roth, who also showed that infection might occur through the uninjured mucous membrane of the nose. Machnoff also succeeded in infecting guinea-pigs with anthrax through the uninjured skin of the back, and, as a result of subsequent microscopical examination of stained sections, arrived at the conclusion that the principal channel through which infection was accomplished was the hair follicles. Braunschweig, in a series of experiments in which he introduced various pathogenic bacteria into the conjunctival sac of mice, rabbits, and guinea-pigs, obtained a negative result with the anthrax bacillus, the bacillus of mouse septicæmia, the bacillus of chicken cholera, and *Micrococcus tetragenus*; but the bacillus obtained by Ribbert from the intestinal diphtheria of rabbits gave a positive result in five mice, two guinea-pigs, and a rabbit.

Infection through the *mucous membrane of the intestine* no doubt occurs in certain diseases. This is believed to be a common mode of the infection of sheep and cattle with anthrax, and probably also in the infectious disease of swine known as hog cholera. The anthrax bacillus would be destroyed by the acid secretions of the stomach, but if spores are present in food ingested they will reach the intestine. The experiments of Korkunoff do not, however, support the view that infection is likely to occur in this way. In a series of experiments upon white mice fed with bread containing a quantity of anthrax spores the result was uniformly negative, but exceptionally infection occurred in rabbits. The same author obtained positive results in rabbits fed with food to which a pure culture of the bacillus of chicken cholera had been added.

Buchner, in experiments upon mice and guinea-pigs fed with material containing anthrax spores, obtained a positive result in four out of thirty-three animals. This is no doubt the usual mode of infection in typhoid fever in man.

Infection may also occur through the *mucous membrane of the respiratory organs*. This has been demonstrated by several bacteriologists, and especially by the experiments of Buchner, who mixed dried anthrax spores with lycopodium powder or pulverized charcoal, and caused mice and guinea-pigs to respire an atmosphere containing this powder in suspension. In a series of sixty-six experiments fifty animals died of anthrax, nine of pneumonia, and seven survived. That infection did not occur through the mucous membrane of the alimentary canal was proved by comparative experiments in which animals were fed with double the quantity of spores used in the inhalation experiments. Out of thirty-three animals fed in this way but four contracted anthrax. That infection occurred through the lungs was also demonstrated by the microscopical examination of sections and by culture experiments, which showed that the lungs were extensively invaded, while in many cases the spleen contained no bacilli. Positive results were also obtained with cultures of the anthrax bacillus not containing spores, which the animals were made to inhale in the form of spray. But in this case a considerable quantity was required, and a sero-fibrinous pneumonia was usually produced as well as general infection; the inhalation of small quantities gave no result. Positive results in rabbits were also obtained by causing them to inhale considerable quantities of a spray containing the bacillus of chicken cholera.

The fact that large quantities of a liquid culture of these virulent bacilli were required to infect very susceptible animals by way of the pulmonary mucous membrane, and that Buchner failed to cause the infection of these animals with small quantities of a pure culture

inhaled in the form of spray, indicates that this is not a common mode of infection in the absence of spores. This view receives further support from the experiments of Hildebrandt, who made tracheal fistulæ in three rabbits, and, after the wound had entirely healed, injected into the trachea of each a pure culture of the anthrax bacillus, which was proved to be virulent by inoculation in mice or guinea-pigs. All of the animals remained in good health. On the other hand, three rabbits which received in the same way a pure culture of the bacillus of rabbit septicæmia died as a result of general infection.

That man may be infected with anthrax by way of the respiratory organs seems to be well established. In England the disease known as "wool-sorter's disease" results from infection in this way among workmen engaged in sorting wool, which is liable to contain the spores of the anthrax bacillus when obtained from the skin of an animal which has fallen a victim to this disease. That infection occurs through the lungs is shown by the fact that these organs are first involved, the disease being, in fact, a pulmonic anthrax.

While these experiments prove the possibility of infection through the respiratory mucous membrane, other experiments made by Hildebrandt show that under ordinary circumstances bacteria suspended in the air do not reach the trachea in rabbits, but are deposited upon the mucous membrane of the mouth, nares, and fauces. In healthy rabbits the tracheal mucus was, as a rule, found to be free from bacteria, while they were very numerous in mucus obtained from the mouth or nares. But when a rabbit was made to inhale for half an hour an atmosphere charged with the spores of *Aspergillus fumigatus* their presence in the lungs was demonstrated by cultivation, the animal being killed for the purpose half an hour after the inhalation experiment.

III.

SUSCEPTIBILITY AND IMMUNITY.

No questions in general biology are more interesting, or more important from a practical point of view, than those which relate to the susceptibility of certain animals to the pathogenic action of certain species of bacteria, and the immunity, natural or acquired, from such pathogenic action which is possessed by other animals. It has long been known that certain infectious diseases, now demonstrated to be of bacterial origin, prevail only or principally among animals of a single species. Thus typhoid fever, cholera, and relapsing fever are diseases of man, and the lower animals do not suffer from them when they are prevailing as an epidemic. On the other hand, man has a natural immunity from many of the infectious diseases of the lower animals, and diseases of this class which prevail among animals are frequently limited to a single species. Again, several species, including man, may be susceptible to a disease, while other animals have a natural immunity from it. Thus tuberculosis is common to man, to cattle, to apes, and to the small herbivorous animals, while the carnivora are, as a rule, immune; anthrax may be communicated by inoculation to man, to cattle, to sheep, to guinea-pigs, rabbits, and mice, but the rat, the dog, carnivorous animals, and birds are generally immune; glanders, which is essentially a disease of the equine genus, may be communicated to man, to the guinea-pig, and to field mice, while house mice, rabbits, cattle, and swine are to a great extent immune.

In addition to this general race immunity or susceptibility we have individual differences in susceptibility or resistance to the action of pathogenic bacteria, which may be either natural or acquired. As a rule, *young animals are more susceptible than older ones*. Thus in man the young are especially susceptible to scarlet fever, whooping cough, and other "children's diseases," and after forty years of age the susceptibility to tubercular infection is very much diminished. Among the lower animals it is a matter of common laboratory experience that the very young of a susceptible species may be infected when inoculated with an "attenuated culture" which older animals of the same species are able to resist.

Considerable differences as to susceptibility may also exist among adults of the same species. In man these differences in individual susceptibility to infectious diseases are frequently manifested. Of a number of persons exposed to infection in the same way, some may escape entirely while others have attacks differing in severity and duration. In our experiments upon the lower animals we constantly meet with similar results, some individuals proving to be exceptionally resistant. Exceptional susceptibility or immunity may be to some extent a family characteristic or one of race. Thus the negro race is decidedly less subject to yellow fever than the white race, and this disease is more fatal among the fair-skinned races of the north of Europe than among the Latin races living in tropical or sub-tropical regions. On the other hand, small-pox appears to be exceptionally fatal among negroes and dark-skinned races generally.

A very remarkable instance of race immunity is that of Algerian sheep against anthrax, a disease which is very fatal to other sheep.

In the instances mentioned *race immunity* is probably an acquired tolerance due to natural selection and inheritance. If, for example, a susceptible population is exposed to the ravages of small-pox, the least susceptible individuals will survive and may be the parents of children who will be likely to inherit the special bodily characters upon which this comparative immunity depends. The tendency of continuous or repeated exposure to the same pathogenic agent will evidently be to establish a race tolerance; and there is reason to believe that such has been the effect in the case of some of the more common infectious diseases of man, which have been noticed to prevail with especial severity when first introduced among a virgin population, as in the islands of the Pacific, etc.

In the same way we may explain the immunity which carnivorous animals have for anthrax and various forms of septicæmia to which the herbivora are very susceptible when the pathogenic germ is introduced into their bodies by inoculation. From time immemorial the carnivora have been in the habit of fighting over the dead bodies of herbivorous animals, some of which may have fallen a prey to these infectious germ diseases, and in their fighting they receive wounds, inoculated with the infectious material from these bodies, which would be fatal to a susceptible animal. If at any time in the past a similar susceptibility existed among the carnivora, with individual differences as to resisting power, it is evident that there would be a constant tendency for the most susceptible individuals to perish and for the least susceptible to survive.

But if we admit this to be a probable explanation of the immunity of carnivorous animals from septic infection, we have not yet explained the precise reason for the immunity enjoyed by the

selected individuals and their progeny. The essential difference between a susceptible and immune animal depends upon the fact that in one the pathogenic germ, when introduced by accident or experimental inoculation, multiplies and invades the tissues or blood, where, by reason of its nutritive requirements and toxic products, it produces changes in the tissues and fluids of the body inconsistent with the vital requirements of the infected animal; while in the immune animal multiplication does not occur or is restricted to a local invasion of limited extent, and in which after a time the resources of nature suffice to destroy the parasitic invader.

Now the question is, upon what does this essential difference depend? Evidently upon conditions favorable or unfavorable to the development of the pathogenic germ; or upon its destruction by some active agent present in the tissues or fluids of the body of the immune animal; or upon a neutralization of its toxic products by some substance present in the body of the animal which survives infection.

What, then, are the unfavorable conditions which may be supposed to prevent development in immune animals? In the first place, the temperature of the body may not be favorable. Certain pathogenic bacteria are only able to develop within very narrow temperature limits, and, if all other conditions were favorable, could not be expected to multiply in the bodies of cold-blooded animals. Or the temperature of warm-blooded animals, and especially of fowls, may be above the point favorable for their development. This is the explanation offered by Pasteur of the immunity of fowls, which are usually refractory against anthrax; and in support of this view he showed an experiment that when chickens are refrigerated after inoculation, being partly immersed in cold water, they are liable to become infected and to perish. But, as pointed out by Koch, the sparrow, which has a temperature as high as that of the chicken, may contract anthrax without being refrigerated. We must not, therefore, too hastily conclude that the success in Pasteur's experiment depended alone upon a reduction of the body heat. Gibier has shown that the anthrax bacillus may multiply in the bodies of frogs and fish, if these are kept in water having a temperature of 35°. But the anthrax bacillus grows within comparatively wide temperature limits, while other pathogenic bacteria are known to have more restricted temperature range and would be more decidedly influenced by this factor—*e.g.*, the tubercle bacillus.

The composition of the body fluids, and especially their reaction, is probably a determining factor in some instances. Thus Behr has ascribed the failure of the anthrax bacillus to develop in the white rat, which possesses a remarkable immunity against anthrax.

to the highly alkaline reaction of the blood and tissue juices of this animal. Behring claims to have obtained experimental proof of the truth of this explanation by feeding white rats on an exclusively vegetable diet or by adding acid phosphate of lime to their food, by which means this excessive alkalinity of the blood is diminished. Rats so treated are said to lose their natural immunity, and to die as a result of inoculation with virulent cultures of the anthrax bacillus.

The recent experiments of Nuttall, Behring, Buchner, and others have established the fact that recently drawn blood of various animals possesses decided germicidal power, and Buchner has shown that this property belongs to the fluid part of the blood and not to its cellular elements. It has also been shown that aqueous humor, the fluid of ascites, and lymph from the dorsal lymph sac of a frog possess the same power. This power to kill bacteria is destroyed by heat, and is lost when the blood has been kept for a considerable time, but it is not neutralized by freezing. Further, this power to destroy bacteria differs greatly for different species, being very decided in the case of certain pathogenic bacteria, less so for others, and absent in the case of certain common saprophytes. Behring has also shown that the blood of different animals differs considerably in this regard, and that the blood of the rat and of the frog, which animals have a natural immunity against anthrax, is especially fatal to the anthrax bacillus. The experiments made show that this germicidal power is very prompt in its action, but that it is limited as to the number of bacteria which can be destroyed by a given quantity of blood serum. When the number is excessive, development occurs after an interval during which a limited destruction has taken place. It would appear that the element in the blood to which this germicidal action is due is neutralized in exercising this power; and as, independently of this, blood serum is an excellent culture medium for bacteria, an abundant development takes place when the destruction has been incomplete.

Buchner has ascribed this remarkable property of blood serum to the presence of some albuminoid substance, the exact nature of which he was not able to determine; and quite recently Hankin (1891) has published the results of his interesting researches confirming this view. From the spleen and blood serum of rats he has isolated a globulin possessing germicidal properties, to which he ascribes the power of rat's blood to destroy anthrax bacilli, without, however, rejecting the view that the excessive alkalinity of the blood of this animal may be a factor in producing this result. The globulin obtained by him is insoluble in water or in alcohol and does not dialyze.

In a recent communication (1892) Brieger, Kitasato, and Wasser-

mann have published the results of their interesting experiments with a bouillon made from the thymus gland of the calf. It was found that the tetanus bacillus cultivated in this bouillon did not form spores and had comparatively little virulence. Mice or rabbits inoculated with it in small doses—0.001 to 0.2 cubic centimetre for a mouse—proved to be subsequently immune. And the blood serum of an immune rabbit injected into the peritoneal cavity of a mouse—0.1 to 0.5 cubic centimetre—was found to give it immunity from the pathogenic action of a virulent culture of the tetanus bacillus. Similar results were obtained with several other pathogenic bacteria cultivated in the thymus bouillon—spirillum of cholera, bacillus of diphtheria, typhoid bacillus. We give here the directions for preparing the thymus bouillon as used by the authors named :

Two or three thymus glands are chopped into small pieces immediately after they are taken from the animal. An equal part of distilled water is added to the mass and stirred for some time ; it is then placed in an ice chest for twelve hours. The juices are now expressed through gauze by means of a flesh press. A clouded, slimy fluid is obtained, which constitutes a stock solution. This is diluted with water, and a certain quantity of carbonate of soda is added to the solution before sterilization. By this means coagulation and precipitation of the active substance from the thymus gland are avoided. The exact amount of water and of sodium carbonate required to prevent precipitation must be determined by experiment, as it differs for different glands. Usually an equal portion of water and sufficient soda solution to turn litmus paper feebly blue will give the desired result. The liquid is now heated in a large flask, which is left for fifteen minutes in the steam sterilizer. The liquid is allowed to cool and then filtered through fine linen to remove any suspended coagula ; the filtrate has a milky opalescence. It is now placed in test tubes and again sterilized. The active principle is precipitated by the addition of a few drops of acetic acid.

Additional facts bearing upon this important question have been developed by the experiments of Ogata and Jasuhara, which show that the anthrax bacillus, when cultivated in the blood of an immune animal (rat, dog, or frog), becomes attenuated as to its pathogenic power, and that such cultures injected into a susceptible animal give rise to a mild attack followed by immunity. Moreover, the injection of a small amount—one drop—of blood from a frog or a dog into a mouse, made before or after inoculating it with a virulent culture of the anthrax bacillus, was found to protect the animal from a fatal attack, and, after its recovery from the mild attack resulting from the injection, it proved to be immune. The protective influence was exercised when the blood was injected as long as seventy-two hours before the inoculation, or five hours after ; and it was not lost when the blood was kept for weeks in a cool place. But subjected to a temperature of 45° C. for an hour completely destroyed its power to protect inoculated mice from a fatal attack of anthrax.

Similar results have been reported by Behring and Kitasato.

experiments made by them relating to acquired immunity from the pathogenic action of the bacillus of tetanus. The blood of animals which had been made immune was injected into a susceptible animal, and at the end of twenty-four hours it was inoculated with a virulent culture. The result was negative, while control experiments made with the same culture gave a uniformly fatal result. So small a quantity as 0.2 cubic centimetre of blood from an immune rabbit, injected into the cavity of the abdomen of a mouse, was sufficient to protect it from the fatal effects of a virulent culture of the tetanus bacillus injected twenty-four hours later. Further, these bacteriologists have shown that the toxic substances present in a filtered culture of the tetanus bacillus are neutralized by admixture with the blood of an immune rabbit. A culture ten days old was sterilized by filtration; 0.0001 cubic centimetre of the filtrate was found to kill a mouse with certainty in less than two days. Of this filtered culture one cubic centimetre was added to five cubic centimetres of blood serum from an immune rabbit. At the end of twenty-four hours four mice received each 0.2 cubic centimetre of the mixture, containing more than three hundred times the fatal dose of the filtered culture. All of these mice survived the injection and proved subsequently to be immune for virulent tetanus bacilli, while four control mice, each of which was inoculated with 0.0001 cubic centimetre of the same filtrate unmixed with blood, perished within thirty-six hours. The blood of rabbits not immune was without effect in neutralizing the toxic substances in a filtered culture of the tetanus bacillus, as was also the blood of children, calves, sheep, and horses.

The same bacteriologists have obtained similar results by mixing the blood of an animal which had an acquired immunity against the poison of the diphtheria bacillus with filtered cultures of this bacillus. The toxic substances present are neutralized by such admixture, but, according to Behring, the bacilli themselves are not destroyed by the blood of an immune animal.

It has also been shown by experiment that naturally immune animals may be infected by the addition of certain substances to cultures of pathogenic bacteria. Thus Arloing was able to induce symptomatic anthrax in animals naturally immune by mixing with his cultures various chemical substances, such as carbolic acid, pyrogallie acid, and especially lactic acid (twenty per cent). Leo has shown that white mice, which are not subject to the pathogenic action of the glanders bacillus, may be rendered susceptible by feeding them for some time upon phloridzin, which gives rise to an artificial diabetes and causes the tissues to be impregnated with sugar.

Before discussing the rationale of *acquired immunity* a statement of certain established facts will be desirable.

In the infectious diseases of man involving the system generally, a single attack commonly confers immunity from subsequent attacks. This is true of the eruptive fevers, of typhoid fever, of yellow fever, of mumps, of whooping cough, and, to some extent at least, of syphilis. But it seems not to be the case in epidemic influenza (*la grippe*), in croupous pneumonia, or in Asiatic cholera, in which diseases second attacks not infrequently occur. In localized infectious diseases such as diphtheria, erysipelas, and gonorrhœa one attack is not protective. Croupous pneumonia and Asiatic cholera should perhaps be grouped with diphtheria and erysipelas as local infections with constitutional symptoms resulting from the absorption of toxic products. But typhoid fever, mumps, and whooping cough, in which one attack gives immunity, are also localized infectious diseases.

We are therefore not able to group infectious diseases into two classes, in one of which there is a general infection followed by immunity, and in the other a local infection without subsequent immunity. Indeed, in the eruptive fevers and specific febrile infectious diseases generally *the immunity following an attack is not absolute*. Second attacks of small-pox, of scarlet fever, and of yellow fever occur occasionally, although a large majority of those who suffer an attack of one of these diseases have an immunity for life. On the other hand, in the diseases mentioned in which one attack is not generally recognized as protecting from future attacks, it is probable that a certain degree of immunity, of limited duration perhaps, is acquired. In localized infection, as in gonorrhœa or erysipelas, the invaded tissues appear after a time to acquire a certain tolerance to the pathogenic action of the invading parasite, and no doubt recovery from these diseases would in many cases occur, after a time, without medical interference. In diphtheria, cholera, and epidemic influenza second attacks do not often occur during the same epidemic, and there is reason to believe that a recent attack affords a certain degree of immunity.

That immunity may result from a comparatively mild attack as well as from a severe one is a matter of common observation in the case of small-pox, scarlet fever, yellow fever, etc.; and since the discovery of Jenner we have in vaccination a simple method of producing immunity in the first-mentioned disease. The acquired immunity resulting from vaccination is not, however, as complete or as permanent as that which results from an attack of the disease.

These general facts relating to acquired immunity from infectious diseases constituted the principal portion of our knowledge with reference to this important matter up to the time that Pasteur (1880) demonstrated that in the disease of fowls known as chicken cholera,

which he had proved to be due to a specific microorganism, a mild attack followed by immunity may be induced by inoculation with an "attenuated virus"—i.e., by inoculation with a culture of the pathogenic microorganism the virulence of which had been so modified that it gave rise to a comparatively mild attack of the disease in question. Pasteur's original method of obtaining an attenuated virus consisted in exposing his cultures for a considerable time to the action of atmospheric oxygen. It has since been ascertained that the same result is obtained with greater certainty by exposing cultures for a given time to a temperature slightly below that which would destroy the vitality of the pathogenic microorganism, and also by exposure to the action of certain chemical agents (see Part Second, p. 124).

Pasteur at once comprehended the importance of his discovery, and inferred that what was true of one infectious germ disease was likely to be true of others. Subsequent researches, by this savant and by other bacteriologists, have justified this anticipation, and the demonstration has already been made for a considerable number of similar diseases—anthrax, symptomatic anthrax, rouget.

A virus which has been attenuated artificially—by heat, for example—may be cultivated through successive generations without retaining its original virulence. As this virulence depends, to a considerable extent at least, upon the formation of toxic products during the development of the pathogenic microorganism, we naturally infer that diminished virulence is due to a diminished production of these toxic substances.

There is reason to believe that a natural attenuation of virulence may occur in pathogenic bacteria which are able to lead a saprophytic existence during their multiplication external to the bodies of living animals, and the comparatively mild character of some epidemics is probably due to this fact.

Again, cultivation within the body of a living animal may, in certain cases, cause a diminution in the virulence of a pathogenic microorganism. Thus Pasteur and Thuiller have shown that the *microbe* of rouget when inoculated into a rabbit kills the animal, but that its pathogenic virulence is nevertheless so modified that a culture made from the blood of a rabbit killed by it is a suitable "vacine" for the pig.

On the other hand, we have experimental evidence that the virulence of attenuated cultures may be reestablished by passing them through the bodies of susceptible animals. Thus a culture of the bacillus of rouget, attenuated by having been passed through the body of a rabbit, is restored to its original virulence by passing it through the bodies of pigeons. And a culture of the anthrax bacillus

which will not kill an adult guinea-pig may be fatal to a very young animal of the same species or to a mouse, and the bacillus cultivated from the blood of such an animal will be found to have greatly increased virulence.

In Pasteur's inoculations against anthrax "attenuated" cultures are employed which contain the living pathogenic germ as well as the toxic products developed during its growth. Usually two inoculations are made with cultures of different degrees of attenuation—that is to say, with cultures in which the toxic products are formed in less amount than in virus of full power. The most attenuated virus is first injected, and after some time the second vaccine, which if injected first might have caused a considerable mortality. The animal is thus protected from the pathogenic action of the most virulent cultures.

Now, it has been shown by recent experiments that a similar immunity may result from the injection into a susceptible animal of the toxic products contained in a virulent culture, independently of the living bacteria to which they owe their origin. Chauveau, in 1880, ascertained that if pregnant ewes are protected against anthrax by inoculation with an attenuated virus, their lambs, when born, also give evidence of having acquired an immunity from the disease. As the investigations of Davaine seemed to show that the anthrax bacillus cannot pass through the placenta from the mother to the fœtus, the inference seemed justified that the acquired immunity of the latter was due to some soluble substance which could pass the placental barrier. More recent researches by Strauss and Chamberlain, Malvoz and Jacquet, and others, show that the placenta is not such an impassable barrier for bacteria as was generally believed at the time of Chauveau's experiments, so that these cannot be accepted as establishing the inference referred to. But we have more recent experimental evidence which shows that immunity may result from the introduction into the bodies of susceptible animals of the toxic substances produced by certain pathogenic bacteria. The first satisfactory experimental evidence of this important fact was obtained by Salmon and Smith in 1886, who succeeded in making pigeons immune from the pathogenic effects of cultures of the bacillus of hog cholera by inoculating them with sterilized cultures of this bacillus. In 1888 Roux reported similar results obtained by injecting into susceptible animals sterilized cultures of the anthrax bacillus. As already stated, Behring and Kitasato have quite recently reported their success in establishing immunity against virulent cultures of the bacillus of tetanus and the diphtheria bacillus by inoculating susceptible animals with filtered, germ-free cultures of these pathogenic bacteria.

In Pasteur's inoculations against hydrophobia, made subsequently to infection by the bite of a rabid animal, an attenuated virus is introduced subcutaneously in considerable quantity by daily injections, and immunity is established during the interval—so-called period of incubation—which usually occurs between the date of infection and the development of the disease. That the immunity in this case also depends upon the introduction of a chemical substance present in the desiccated spinal cord of rabbits which have succumbed to rabies, which is used in these inoculations, is extremely probable. But, as the germ of rabies has not been isolated or cultivated artificially, this has not yet been demonstrated. Wooldridge claims to have made susceptible animals immune against anthrax by inoculating them with an aqueous extract of the testicle or of the thymus gland of healthy animals.

We may mention also the interesting results obtained by Emmerich, Freudenreich, and others, who have shown that an anthrax infection in a susceptible animal inoculated with a virulent culture may be made to take a modified and non-fatal course by the simultaneous or subsequent inoculation of certain other non-pathogenic bacteria—streptococcus of erysipelas, *Bacillus pyocyaneus*.

In a series of experiments made by the writer about two years ago evidence was obtained that, under certain circumstances, immunity from the effects of one pathogenic bacillus may be obtained by the previous injection of a pure culture of a different species. In the experiments referred to injections into the cavity of the abdomen of a culture of *Bacillus pyocyaneus* protected rabbits from the lethal effects of *Bacillus cuniculicida* Havaniensis, when subsequently injected into the cavity of the abdomen in such amount (one cubic centimetre of a bouillon culture) as invariably proved fatal in rabbits not protected by such injections.

Before considering the theories which have been offered in explanation of acquired immunity it is desirable to call attention to certain observations which have been made during the past few years relating to "chemiotaxis."

The term *chemiotaxis* was first used by Pfeffer to designate the property, observed by himself and others, which certain living cells exhibit with reference to non-living organic material, and by virtue of which they approach or recede from certain substances. The chemiotaxis is said to be positive when the living cell approaches, and negative when it recedes from, a chemical substance. As examples of this we may mention the approach of motile bacteria to nutrient material or to the surface of a liquid medium where they find the oxygen required for their vital activities; and of leucocytes to certain substances when these are introduced beneath the skin of warm-

or cold-blooded animals. This subject has recently received much attention and has been studied especially by Ali-Cohen, Massart and Bordet, Gabritchevski, and others.

According to Gabritchevski, the following substances have a negative chemiotaxis for the leucocytes : Sodium chloride in ten-per-cent solution, alcohol in ten-per-cent solution, quinine, lactic acid, glycerin, chloroform, bile. On the other hand, a positive chemiotaxis is excited by sterilized or non-sterilized cultures of various bacteria. This is shown by the fact that when a small capillary tube, closed at one end, which contains the substance to be tested, is introduced beneath the skin of an animal, the leucocytes are repelled from the tube by certain substances, while those which incite positive chemiotaxis cause them to enter the tube in great numbers. The experiments of Buchner seem to show that the positive chemiotaxis induced by sterilized cultures of bacteria introduced beneath the skin of an animal, is due to the proteid contents of the cells rather than to the chemical products elaborated as a result of their vital activity. But that such chemical products may, in some instances at least, produce a positive chemiotaxis independently of the bacteria is shown by the experiments of Gabritchevski with filtered cultures of *Bacillus pyocyaneus*—confirmed by Massart and Bordet.

An important observation made by Bouchard, and confirmed by Massart and Bordet, is the following: When a tube containing a culture of *Bacillus pyocyaneus* is introduced beneath the skin of a rabbit it is found, at the end of a few hours, to contain a great number of leucocytes. But if immediately after its introduction ten cubic centimetres of a sterilized culture of the same bacillus are injected into the circulation through a vein, very few leucocytes enter the tube introduced beneath the skin—that is, the chemiotaxis of the leucocytes for the bacilli contained in the tube has been neutralized by injecting a considerable quantity of the soluble products of the same bacillus into the circulation.

Buchner, having shown that the bacterial cells contain a proteid substance which attracts the leucocytes, experimented with various other proteids and found that gluten, casein from wheat, and legumin from peas had a similar effect. Starch has no effect, but a mass of flour, made from wheat or from peas, introduced beneath the skin of a rabbit or of a guinea-pig, with antiseptic precautions, in the course of a day or two is enveloped and penetrated by immense numbers of leucocytes. If, instead of introducing these substances which induce positive chemiotaxis beneath the skin, they are injected into the circulation, Buchner has shown that a great increase in the number of leucocytes occurs.

THEORIES OF IMMUNITY.

Exhaustion Theory.—For a time Pasteur supported the view that during an attack of an infectious disease the pathogenic micro-organism, in its multiplication in the body of a susceptible animal, exhausts the supply of some substance necessary for its development, that this substance is not subsequently reproduced, and that consequently the same pathogenic germ cannot again multiply in the body of the protected animal. This view is sustained in a memoir published in the *Comptes Rendus* of the French Academy in 1880, in which Pasteur says :

"It is the life of a parasite in the interior of the body which produces the malady commonly called '*choléra des poules*,' and which causes death. From the moment when this culture (*i.e.*, the multiplication of the parasite) is no longer possible in the fowl the sickness cannot appear. The fowls are then in the constitutional state of fowls not subject to be attacked by the disease. These last are as if vaccinated from birth for this malady, because the foetal evolution has not introduced into their bodies the material necessary to support the life of the microbe, or these nutritive materials have disappeared at an early age.

"Certainly one should not be surprised that there may be constitutions sometimes susceptible and sometimes rebellious to inoculation—that is to say, to the cultivation of a certain virus—when, as I have announced in my first note, one sees a preparation of beer yeast made, exactly like one from the muscles of fowls (*bouillon*), to show itself absolutely unsuited for the cultivation of the parasite of fowl cholera, while it is admirably adapted to the cultivation of a multitude of microscopic species, notably to the *bactérie charbonneuse* (*Bacillus anthracis*).

"The explanation to which these facts conduct us, as well of the constitutional resistance of some individuals as of the immunity produced by protective inoculations, is only natural when we consider that every culture, in general, modifies the medium in which it is effected—a modification of the soil when it relates to ordinary plants; a modification of plants and animals when it relates to their parasites; a modification of our culture liquids when it relates to *mucédinées*, *vibrioniens*, or ferments.

"These modifications are manifested and characterized by the circumstance that new cultivations of the same species in these media become promptly difficult or impossible. If we sow *chicken bouillon* with the microbe of fowl cholera, and, after three or four days, filter the liquid in order to remove all trace of the microbe, and subsequently sow anew in the filtered liquid this parasite, it will be found quite powerless to resume the most feeble development. The liquid, which is perfectly limpid after being filtered, retains its limpidity indefinitely.

"How can we fail to believe that by cultivation in the fowl of the attenuated virus we place its body in the state of this filtered liquid which can no longer cultivate the microbe? The comparison can be pushed still further; for if we filter the *bouillon* containing the microbe in full development, not on the fourth day of culture, but on the second, the filtered liquid will still be able to support the development of the microbe, although with less energy than at the outset. We comprehend, then, that after a cultivation of the modified (*attenué*) microbe in the body of the fowl we may not have removed from all parts of its body the aliment of the microbe. That which remains will permit, then, a new culture, but in a more restricted measure.

"This is the effect of a first inoculation; subsequent inoculations will

remove progressively all the material necessary for the development of the parasite."

In discussing this theory, in a paper published in the *American Journal of the Medical Sciences* (April, 1881), the writer says:

"Let us see where this hypothesis leads us. In the first place, we must have a material of small-pox, and a material of measles, and a material of scarlet fever, etc., etc. Then we must admit that each of these different materials has been formed in the system and stored up for these emergencies—attacks of the diseases in question—for we can scarcely conceive that they were all packed away in the germ cell of the mother and the sperm cell of the father of each susceptible individual. If, then, these peculiar materials have been formed and stored up during the development of the individual, how are we to account for the fact that no new production takes place after an attack of any one of the diseases in question?"

"Again, how shall we account for the fact that the amount of material which would nourish the small-pox germ, to the extent of producing a case of confluent small-pox, may be exhausted by the action of the attenuated virus (germ) introduced by vaccination? Pasteur's comparison of a fowl protected by inoculation with the microbe of fowl cholera, with a culture fluid in which the growth of a particular organism has exhausted the pabulum necessary for the development of additional organisms of the same kind, does not seem to me to be a just one, as in the latter case we have a limited supply of nutriment, while in the former we have new supplies constantly provided of the material—food—from which the whole body, including the hypothetical substance essential to the development of the disease germ, was built up prior to the attack. Besides this we have a constant provision for the elimination of effete and useless products.

"This hypothesis, then, requires the formation in the human body, and the retention up to a certain time, of a variety of materials which, so far as we can see, serve no purpose except to nourish the germs of various specific diseases, and which, having served this purpose, are not again formed in the same system, subjected to similar external conditions, and supplied with the same kind of nutriment."

It is unnecessary to discuss this hypothesis any further, inasmuch as it is no longer sustained by Pasteur or his pupils, and is evidently untenable.

The Retention Theory, proposed by Chauveau (1880), is subject to similar objections. According to this view, certain products formed during the development of a pathogenic microorganism in the body of a susceptible animal accumulate during the attack and are subsequently retained, and, being prejudicial to the growth of the particular microorganism which produced them, a second infection cannot occur. Support for this theory has been found by its advocates in the fact that various processes of fermentation are arrested after a time by the formation of substances which restrain the development of the microorganisms to which they are due. But in the case of a living animal the conditions are very different, and it is hard to conceive that adventitious products of this kind could be retained for years, when in the normal processes of nutrition and excretion the tissues and fluids of the body are constantly undergoing change. Certainly the substances which arrest ordinary processes of ferment-

tation by their accumulation in the fermenting liquid, such as alcohol, lactic acid, phenol, etc., would not be so retained. But we cannot speak so positively with reference to the toxic albuminous substances which recent researches have demonstrated to be present in cultures of some of the best known pathogenic bacteria. It is difficult, however, to believe that an individual who has passed through attacks of half a dozen different infectious diseases carries about with him a store of as many different chemical substances produced during these attacks, and sufficient in quantity to prevent the development of the several germs of these diseases. Nor does the experimental evidence relating to the action of germicide and germ-restraining agents justify the view that a substance capable of preventing the development of one microorganism should be without effect upon others of the same class; but if we accept the retention hypothesis we must admit that the inhibiting substance produced by each particular pathogenic germ is effective only in restraining the development of the microbe which produced it in the first instance.

Pasteur discusses this hypothesis in his paper from which we have already quoted, as follows :

"We may admit the possibility that the development of the microbe, in place of removing or destroying certain matters in the bodies of the fowls, adds, on the contrary, something which is an obstacle to the future development of this microbe. The history of the life of inferior beings authorizes such a supposition. The excretions resulting from vital processes may arrest vital processes of the same nature. In certain fermentations we see anti-septic products make their appearance during, and as a result of, the fermentation, which put an end to the active life of the ferments and arrest the fermentations long before they are completed. In the cultivation of our microbe, products may have been formed the presence of which, possibly, may explain the protection following inoculation.

"Our artificial cultures permit us to test the truth of this hypothesis. Let us prepare an artificial culture of the microbe, and after having evaporated it, *in vacuo*, without heat, let us bring it back to its original volume by means of fresh chicken bouillon. If the extract contains a poison for the life of the microbe, and if this is the cause of its failure to multiply in the filtered liquid, the new liquid should remain sterile. Now, this is not the case. We cannot, then, believe that during the life of the parasite certain substances are produced which are capable of arresting its ulterior development."

This experiment of Pasteur appears to be conclusive so far as the particular pathogenic microorganism referred to is concerned; and we may say, in brief, that more recent investigations do not sustain the view that acquired immunity is due to the retention of products such as are formed by pathogenic bacteria in artificial culture media, and which act by destroying these bacteria or restraining their development when they are introduced into the bodies of immune animals.

Moreover, if we suppose that the toxic substances which give pathogenic power to a particular microorganism are retained in the

body of an immune animal, we must admit that the animal has acquired a tolerance to the pathogenic action of these toxic substances for their presence no longer gives rise to any morbid phenomena. And this being the case, we are not restricted to the explanation that immunity depends upon a restraining influence exercised upon the microbe when subsequently introduced.

Another explanation offers itself, viz., that *immunity depends upon an acquired tolerance to the toxic products of pathogenic bacteria*. This is a view which the writer has advocated in various published papers since 1881. In a paper contributed to the *American Journal of the Medical Sciences* in April, 1881, it is presented in the following language :

"The view that I am endeavoring to elucidate is that, during a mortal fatal attack of one of the specific diseases, the cellular elements implicated which do not succumb to the destructive influence of the poison acquire a tolerance to this poison which is transmissible to their progeny, and which is the reason of the exemption which the individual enjoys from future attacks of the same disease."¹

In my chapter on "Bacteria in Infectious Diseases," in "Bacteria," published in the spring of 1884, but placed in the hands of publishers in 1883, I say:

"It may be that the true explanation of the immunity afforded by a mortal attack of an infectious germ disease is to be found in an acquired tolerance to the action of a chemical poison produced by the microorganism, and consequent ability to bring the resources of nature to bear to restrict invasion by the parasite."

The "resources of nature" are referred to in the same chapter as follows :

"The hypothesis of Pasteur would account for the fact that one individual suffers a severe attack and another a mild attack of an infectious disease after being subjected to the influence of the poison under identical circumstances, by the supposition that the pabulum required for the development of this particular poison is more abundant in the body of one individual than in the other. The explanation which seems to us more satisfactory is that the vital resistance offered by the cellular elements in the bodies of these two individuals was not the same for this poison. It is well known that in conditions of lowered vitality resulting from starvation, profuse discharges, or any other cause, the power to resist disease poisons is greatly diminished, and, consequently, that the susceptibility of the same individual differs at different times.

"From our point of view, the blood, as it is found within the vessels of a living animal, is not simply a culture fluid maintained at a fixed temperature, but under these circumstances is a tissue, the histological elements of which present a certain vital resistance to pathogenic organisms which may be introduced into the circulation.

"If we add a small quantity of a culture fluid containing the bacteria to putrefaction to the blood of an animal, withdrawn from the circulation in a proper receptacle and maintained in a culture oven at blood heat, we will find that these bacteria multiply abundantly, and evidence of putrefaction

¹ "What is the Explanation of the Protection from Subsequent Attacks, resulting from an Attack of Certain Diseases, etc.?" *American Journal of the Medical Sciences*, April, 1881, p. 376.

decomposition will soon be perceived. But if we inject a like quantity of the culture fluid with its contained bacteria into the circulation of a living animal, not only does no increase and no putrefactive change occur, but the bacteria introduced quickly disappear, and at the end of an hour or two the most careful microscopical examination will not reveal the presence of a single bacterium. This difference we ascribe to the vital properties of the fluid as contained in the vessels of a living animal; and it seems probable that the little masses of protoplasm known as white blood corpuscles are the essential histological elements of the blood, so far as any manifestation of vitality is concerned. *The writer has elsewhere (1881) suggested that the disappearance of the bacteria from the circulation, in the experiment referred to, may be effected by the white corpuscles, which, it is well known, pick up, after the manner of amœbæ, any particles, organic or inorganic, which come in their way. And it requires no great stretch of credulity to believe that they may, like an amœba, digest and assimilate the protoplasm of the captured bacterium, thus putting an end to the possibility of its doing any harm.*

"In the case of a pathogenic organism we may imagine that, when captured in this way, it may share a like fate if the captor is not paralyzed by some potent poison evolved by it, or overwhelmed by its superior vigor and rapid multiplication. In the latter event the active career of our conservative white corpuscle would be quickly terminated and its protoplasm would serve as food for the enemy. It is evident that in a contest of this kind the balance of power would depend upon circumstances relating to the *inherited* vital characteristics of the invading parasite and of the invaded leucocyte."

In the same chapter the writer quotes from his paper on acquired immunity, published in 1881, as follows :

"The difficulties into which this hypothesis [the exhaustion theory of Pasteur] leads us certainly justify us in looking further for an explanation of the phenomena in question. This explanation is, I believe, to be found in the peculiar properties of the protoplasm, which is the essential framework of every living organism. The properties referred to are the tolerance which living protoplasm may acquire to certain agents which, in the first instance, have an injurious or even fatal influence upon its vital activity ; and the property which it possesses of transmitting its peculiar qualities, inherent or acquired, through numerous generations, to its offshoots or progeny.

"Protoplasm is the essential living portion of the cellular elements of animal and vegetable tissues; but as our microscopical analysis of the tissues has not gone beyond the cells of which they are composed, and is not likely to reveal to us the complicated molecular structure of the protoplasm, upon which, possibly, the properties under consideration depend, it will be best, for the present, to limit ourselves to a consideration of the living cells of the body. These cells are the direct descendants of the pre-existent cells, and may all be traced back to the sperm cell and the germ cell of the parents. Now, the view which I am endeavoring to elucidate is that, during a non-fatal attack of one of the specific diseases, the cellular elements implicated, which do not succumb to the destructive influence of the poison, acquire a tolerance to this poison which is transmissible to their progeny, and which is the reason of the exemption which the individual enjoys from future attacks of the same disease.

"The known facts in regard to the hereditary transmission by cells of acquired properties make it easy to believe in the transmission of such a tolerance as we imagine to be acquired during the attack ; and if it is shown by analogy that there is nothing improbable in the hypothesis that such a tolerance is acquired, we shall have a rational explanation, not of heredity and of the mysterious properties of protoplasm, but of the particular result under consideration. The transmission of acquired properties is shown in the budding and grafting of choice fruits and flowers, produced by cultiva-

tion, upon the wild stock from which they originated. The acquired properties are transmitted indefinitely; and the same sap which on one twig nourishes a sour crab apple, on another one of the same branch is elaborated into a delicious pippin.

"The tolerance to narcotics—opium and tobacco—and to corrosive poisons—arsenic—which results from a gradual increase of dose, may be cited as an example of acquired tolerance by living protoplasm to poisons which at the outset would have been fatal in much smaller doses.

"The immunity which an individual enjoys from any particular disease must be looked upon as a power of resistance possessed by the cellular elements of those tissues of his body which would yield to the poison in the case of an unprotected person."

This theory of immunity, advanced by the author in 1881, has received considerable support from investigations made since that date, and especially from the experimental demonstration by Salmon, Roux, and others that, as suggested in the paper from which I have quoted, immunity may result from the introduction into the body of a susceptible animal of the soluble products of bacterial growth—filtered cultures.

The theory of vital resistance to the toxic products evolved by pathogenic bacteria is also supported by numerous experiments which show that natural or acquired immunity may be overcome when these toxic products are introduced in excess, or when the vital resisting power of the animal has been reduced by various agencies.

Thus Bouchard has shown that very small doses of a pure culture of the *Bacillus pyocyaneus* are fatal to rabbits, when at the same time a considerable quantity of a filtered culture of the same bacillus is injected into a vein. The animal could have withstood the filtered culture alone or the bacilli injected beneath its skin; but when its vital resisting power (paralysis of phagocytes?) has been partially overcome by the filtered culture injected into a vein the bacilli multiply abundantly and a fatal result follows.

The same result may be obtained by injecting sterilized cultures of a different microorganism. Thus Roger has shown that the rabbit, which has a natural immunity against symptomatic anthrax, succumbs to infection when inoculated with a culture of the bacillus of this disease, if at the same time it receives an injection of a sterilized or non-sterilized culture of *Bacillus prodigiosus*.

Monti has succeeded in killing animals with old and attenuated cultures of the *Streptococcus pyogenes* or of *Staphylococcus pyogenes aureus*, by injecting at the same time a culture of *Proteus vulgaris*. A similar result may be obtained by subjecting animals to physical agencies which reduce the vital resisting power of the tissues. Thus Nocard and Roux found by experiment that an attenuated culture of the anthrax bacillus, which was not fatal to guinea-pigs, killed these animals when injected into the muscles of the thigh after they had been bruised by mechanical violence. Charrin

and Roger found that white rats, which are unsusceptible to anthrax, became infected and frequently died if they were exhausted, previous to inoculation, by being compelled to turn a revolving wheel for a considerable time. Pasteur found by experiment that fowls, which have a natural immunity against anthrax, become infected and perish if they are subjected to artificial refrigeration after inoculation. This has been confirmed by the more recent experiments of Wagner (1890). According to Canalis and Morpurgo, pigeons which are enfeebled by inanition easily contract anthrax as a result of inoculation. Arloing states that sheep which have been freely bled contract anthrax more easily than others; and Serafini found that when dogs were freely bled the bacillus of Friedländer, injected into the trachea or the pleural cavity, entered and apparently multiplied to some extent in the blood, whereas without such previous bleeding they were not to be found in the circulating fluid.

Again, as already stated in a previous section, the simultaneous injection of certain chemical substances overcomes the vital resisting power of the tissues or fluids of the body in such a way that infection and death may occur as a result of inoculations into animals which have a natural or acquired immunity against the pathogenic microorganism introduced. Thus Arloing, Cornevin, and Thomas have shown that rabbits succumb to symptomatic anthrax when lactic acid is injected at the same time with the bacillus into the muscles. Nocard and Roux have obtained the same result by injecting various other substances, and their experiments show that the result is due to the injurious effects of the substance injected upon the tissues, and not to an increased virulence on the part of the pathogenic bacillus. The experiments of Léo are of a similar nature. By injecting phloridzin into rats he caused them to lose their natural immunity against anthrax. Certain anæsthetic agents have also been shown to produce a similar result. Platania communicated anthrax to immune animals—dogs, frogs, pigeons—by bringing them under the influence of curare, chloral, or alcohol; and Wagner obtained a similar result in his experiments on pigeons to which he had administered chloral.

More direct experimental evidence in favor of the view under consideration is that obtained by Beumer in his experiments with sterilized cultures of the typhoid bacillus. He found that after the repeated injection of non-lethal doses mice were able to resist an amount of this toxine which was fatal to animals of the same species not so treated. But, on the other hand, Gamaléia found, in his experiments upon guinea-pigs which had been made immune against the pathogenic action of a spirillum, called by him *Vibrio Metschnikovi*, that these animals have no increased tolerance for the toxic

products of this microorganism. Although immune against infection by the living microbe, they were killed by the same quantity of a sterilized culture as was fatal to guinea-pigs which had not been rendered immune.

Charrin has obtained similar results in experiments with filtered cultures of *Bacillus pyocyaneus*. Rabbits which had an artificial immunity against the pathogenic action of the bacillus were killed by doses of a sterilized culture such as were fatal to other rabbits of the same size not immune. In subsequent experiments by Charrin and Gameléia "vaccinated" rabbits were found to be even more susceptible to the toxic action of filtered cultures than were those not vaccinated. Recently (1891) Metschnikoff has followed up this line of experiment, and has shown that when considerable amounts of filtered cultures of *Bacillus pyocyaneus* are injected subcutaneously in rabbits a certain tolerance to the toxic action of the same cultures is established in some instances. But his results do not give any substantial support to the view that immunity depends upon an acquired tolerance to the toxic action of the chemical products contained in cultures of the pathogenic bacteria with which he experimented—*Bacillus pyocyaneus* and *Vibrio Metschnikovi*.

In view of the results of experimental researches above recorded and of other recent experiments which show that, in certain cases at least, acquired immunity depends upon the formation of an antitoxine in the body of the immune animal, we are convinced that the theory of immunity under discussion, first proposed by the writer in 1881, cannot be accepted as a sufficient explanation of the facts in general. At the same time we are inclined to attribute considerable importance to acquired tolerance to the toxic products of pathogenic bacteria as one of the factors by which recovery from an infectious disease is made possible and subsequent immunity established.

The "vital-resistance theory" of the present writer, as set forth in the above-quoted extracts from his published papers, is essentially the same as that advocated by Buchner at a later date (1883). Buchner supposes that during the primary infection, when an animal recovers, a "reactive change" has been produced in the cells of the body which enables it to protect itself from the pathogenic action of the same microorganism when subsequently introduced.

Of course when we ascribe immunity to the "vital resistance" of the cellular elements of the body, we have not explained the *modus operandi* of this vital resistance or "reactive change," but we have simply affirmed that the phenomenon in question depends upon some acquired property residing in the living cellular elements of the body. We have suggested that that which has been acquired is a tolerance to the action of the toxic products produced by patho-

genic bacteria. But, as already stated, in the light of recent experiments this theory now appears to us to be untenable as a general explanation of acquired immunity.

The Theory of Phagocytosis.—The fact that in certain infectious diseases due to bacteria the parasitic invaders, at the point of inoculation or in the general blood current, are picked up by the leucocytes and in properly stained preparations may be seen in their interior, has been known for some years. In mouse septicæmia—an infectious disease described by Koch in his work on “Traumatic Infectious Diseases,” published in 1878—the slender bacilli which are the cause of the disease are found in large numbers in the interior of the leucocytes. Koch says, in the work referred to: “Their relation to the white blood corpuscles is peculiar; they penetrate these *and multiply in their interior*. One often finds that there is hardly a single white corpuscle in the interior of which bacilli cannot be seen. Many corpuscles contain isolated bacilli only; others



FIG. 78.—Bacillus of mouse septicæmia in leucocytes from blood of mouse (Koch).

have thick masses in their interior, the nucleus being still recognizable; while in others the nucleus can be no longer distinguished; and, finally, the corpuscle may become a cluster of bacilli, breaking up at the margin—the origin of which one could not have explained had there been no opportunity of seeing all the intermediate steps between the intact white corpuscle and these masses” (Fig. 78). It will be noted that in the above quotation Koch affirms that the bacilli penetrate the leucocytes and multiply in their interior. Now, the theory of phagocytosis assumes that the bacilli are picked up by the leucocytes and destroyed in their interior, and that immunity depends largely upon the power of these “phagocytes” to capture and destroy living pathogenic bacilli.

The writer suggested this as an hypothesis as long ago as 1881, in a paper read before the American Association for the Advancement of Science, in the following language:

“It has occurred to me that possibly the white corpuscles may have the office of picking up and digesting bacterial organisms which

by any means find their way into the blood. The propensity exhibited by the leucocytes for picking up inorganic granules is well known, and that they may be able not only to pick up but to assimilate, and so dispose of, the bacteria which come in their way, does not seem to me very improbable, in view of the fact that amoebae which resemble them so closely, feed upon bacteria and similar organisms."¹

At a later date (1884) Metschnikoff offered experimental evidence in favor of this view, and the explanation suggested in the above quotation is commonly spoken of as the *Metschnikoff theory*.

The observations which first led Metschnikoff to adopt this view were made upon a species of daphnia which is subject to fatal infection by a torula resembling the yeast fungus. Entering with the food, this fungus penetrates the walls of the intestine and invades the tissues. In certain cases the infection does not prove fatal, owing, Metschnikoff asserts, to the fact that the fungus cells are seized upon by the leucocytes, which appear to accumulate around the invading parasite (chemiotaxis) for this special purpose. If they are successful in overpowering and destroying the parasite the animal recovers; if not, it succumbs to the general infection which results. In a similar manner, Metschnikoff supposes, pathogenic bacteria are destroyed when introduced into the body of an immune animal. The colorless blood corpuscles, which he designates *phagocytes*, accumulate at the point of invasion and pick up the living bacteria, as they are known to pick up inorganic particles injected into the circulation. So far there can be no doubt that Metschnikoff is right. The presence of bacteria in the leucocytes in considerable numbers, both at the point of inoculation and in the general circulation, has been repeatedly demonstrated in animals inoculated with various pathogenic bacteria. The writer observed this in his experiments, made in 1881, in which rabbits were inoculated with cultures of his *Micrococcus Pasteurii*, and it was this observation which led him to suggest the theory which has since been so vigorously supported by Metschnikoff. But the presence of a certain number of bacteria within the leucocytes does not prove the destructive power of these cells for living pathogenic organisms. As urged by Weigert, Baumgarten, and others, it may be that the bacteria were already dead when they were picked up, having been destroyed by some agency outside of the blood cells. As heretofore stated, we have now experimental evidence that blood

¹ "A Contribution to the Study of Bacterial Organisms commonly found upon Exposed Mucous Surfaces and in the Alimentary Canal of Healthy Individuals." Illustrated by photomicrographs. Proceedings of the American Association for the Advancement of Science, 1881, Salem, 1882, xxx., 89-94. Also in Studies from the Biological Laboratory, Johns Hopkins University, Baltimore, vol. ii., No. 2, 1883.

serum, quite independently of the cellular elements contained in it in the circulation, has decided germicidal power for certain pathogenic bacteria, and that the blood serum of the rat and other animals which have a natural immunity against anthrax is especially fatal to the anthrax bacillus.

Numerous experiments have been made during the past two or three years with a view to determining whether pathogenic bacteria are, in fact, destroyed within the leucocytes after being picked up, and different experimenters have arrived at different conclusions. In the case of mouse septicæmia, already alluded to, and in gonorrhœa, one would be disposed to decide, from the appearance and arrangement of the pathogenic bacteria in the leucocytes, that they are not destroyed, but that, on the other hand, they multiply in the interior of these cells, which in the end succumb to this parasitic invasion. In both of the diseases mentioned we find leucocytes so completely filled with the pathogenic microorganisms that it is difficult to believe that they have all been picked up by a voracious phagocyte, which has stuffed itself to repletion, while numerous other leucocytes from the same source and in the same microscopic field of view have failed to capture a single bacillus or micrococcus. Moreover, the staining of the parasitic invaders, and the characteristic arrangement of the "gonococcus" in stained preparations of gonorrhœal pus, indicate that their vitality has not been destroyed in the interior of the leucocytes or pus cells, and we can scarcely doubt that the large number found in certain cells is due to multiplication *in situ* rather than to an unusual activity of these particular cells. But in certain infectious diseases, and especially in anthrax, the bacilli included within the leucocytes often give evidence of degenerative changes, which would support the view that they are destroyed by the leucocytes, unless these changes occurred before they were picked up, as is maintained by Nuttall and others. We cannot consider this question as definitely settled, but, in view of the importance attached to the theory of phagocytosis by many pathologists and bacteriologists, we reproduce here a recent paper by Metschnikoff in which his views are fully set forth :

LECTURE ON PHAGOCYTOSIS AND IMMUNITY.¹

It is not possible to study the bacteriology of disease without noticing that, while in many cases the invading microorganisms are to be found solely in the fluids of the body, in not a few affections they present themselves in the interior of certain cells, and this either partially—some being within the cells, others free in the blood plasma and the lymph that bathes the various tissues—or exclusively, all the bacteria that are visible being

¹ Delivered at the Institut Pasteur, December 29th, 1890, by Dr. Elias Metschnikoff, Chef de Service de l'Institut Pasteur, Paris; late Professor of Zoölogy in the University of Odessa.

intracellular. Many of the facts bearing upon the terms of this relationship between tissue cell and microorganism are now well known, yet it is worth while to recapitulate the more important, in order to show that from them it is possible to gain a general law; and what is more, that from a study of such facts some insight may be gained into the phenomena of immunity.

It may, in the first place, be postulated that whenever a microorganism is discoverable within a cell its passage thither has been by means of protoplasmic or amoeboid movements, either on the part of the microbe or of the cell itself. The first alternative is the rarer, although it certainly exists, and of this the malarial parasite affords an excellent example; for here in the amoeboid stage of its existence the hæmatozoon makes its way into the interior of a cell that possesses no active movements of its own, namely, the red blood corpuscle, and from the substance of this corpuscle the parasite gains its nourishment. Other sporozoa furnish instances almost equally good. More commonly, however, as in the case of all bacteria, where we have to deal with microorganisms which, even when mobile, are destitute of protoplasmic appendages, it is the cells which play the active part; certain cells include the parasites. Of such the amoebiform leucocyte of the blood and lymph is the most typical example, capable, as it is, of sending out pseudopodia in all directions, while a closely allied form is the cell of the splenic pulp. But there are also cells—as, for instance, those forming the endothelial lining of the vessels—which are very definitely fixed, which nevertheless can give off protoplasmic processes from their free surface and so capture and include bacteria.

All these may be spoken of as *phagocytes*, and may be divided into the two broad groups of *fixed* phagocytes (endothelial cells, etc.) and *free* (leucocytes). Not that the terms "phagocyte" and "leucocyte" are synonymous for of the latter three main forms may be distinguished, of which one is practically immobile and never takes up bacteria. This is the *lymphocyte*, characterized by its relatively small size, its large single nucleus, and the small amount of surrounding protoplasm. The two remaining (phagocytic) forms are, *first*, the large uninuclear leucocyte, whose prominent nucleus is at times lobed or reniform, which stains well with aniline dyes and possesses much protoplasm and active amoeboid movements—the *macrophage*—and *second*, the *microphage*, a small form, also staining well, but either multinuclear or with one nucleus in the process of breaking up. If now we compare the endothelial cells with these, it is evident that their properties connect them closely with the macrophage; and, in fact, there is now little or no doubt that a very large proportion of the macrophages are of endothelial origin.

Leaving aside the subject of amoeboid microbes and their life within animal cells, it is to the phagocytes and their relation to the bacteria that I wish specially to draw your attention.

Taking as wide a view as possible of this relationship, we can first determine that *the more malignant the microorganism the rarer is its presence within the phagocyte*. Thus in those which of all diseases are the most rapidly fatal—in chicken cholera affecting birds and rabbits, in hog cholera ("cholera des porcs") given to pigeons and rabbits, in the anthrax of mice and other specially sensitive animals, in the "septicémie vibrienne" of guinea-pigs and birds, and in yet other diseases of peculiarly swift course—the corresponding bacteria are only very exceptionally to be found within the cells, but remain free in the neighborhood of their introduction and thence invade the blood. For all the above-mentioned diseases are not localized, but, on the contrary, present the characters of general acute septicæmia, causing death within twenty to thirty-six hours, or, in certain cases, even within six hours.

And when we pass to those diseases in which the bacteria are to be found either in part or almost wholly within the phagocytes, the same law still applies; for in such cases the disease has lost its suddenness, tending to have a slower course, or, indeed, to be of a chronic nature. Even in those

affections in which an acute course is accompanied by considerable phagocytosis, the fatal termination is far from occurring at the same early period as in the diseases recorded above. Thus mouse septicæmia, characterized as it is by frequent intracellular bacteria, has a duration in the mouse two and a half times as long as that of anthrax in the same animal. But in general a well-marked phagocytosis is associated with diseases presenting an essentially chronic development; it is in affections such as tuberculosis, leprosy, rhinoscleroma, glanders, that the specific bacteria are most readily taken up by the phagocytes; it is here that, at the seat of the disease, we meet with innumerable macrophages—epithelioid cells in which lie the individual microorganisms.

Further, if we consider the phenomena associated with the resolution of an infectious disease, this inverse relationship between the malignancy of the malady and the occurrence of phagocytosis is, if possible, yet more clearly demonstrated. Notice, for instance, what obtains during the progress of relapsing fever, a malady still fairly common in Russia and other Slavonic countries, and one which, while presenting many difficulties to the bacteriologist, in that the specific spirochæte has so far resisted cultivation, and in that it cannot be communicated to the ordinary animals of the laboratory, is nevertheless in many respects not ill-adapted for our present purpose. Here, during the sudden access of the fever, the spirilla are present in the blood in enormous numbers; they all are free in the plasma, and not a single intracellular spirillum is to be met with. During the apyretic stage (and in the monkey this is, at the same time, the stage of resolution) not a single free spirillum is discoverable in the blood, while the phagocytes of the spleen contain the microbes. The like phenomena repeat themselves in all those cases where it is possible to follow the fate of the microorganisms of acute disease during the stage of recovery. Thus rats and pigeons very frequently survive an attack of anthrax, and, where this occurs, the bacteria, which at the commencement of the disease were for the most part free, now, during resolution, are for the most part included within leucocytes and splenic phagocytes.

Nor is this all. Analogous phenomena as a rule attend immunity, which most often is but *recovery in operation from the very onset of a disease*. The more closely one studies this condition of immunity the more is one led to the conviction that immunity and recovery are very intimately connected; that one can pass by slight gradations from the resolution of disease to the production of immunity. So it is that, in inoculating refractory animals with the microbe to whose action they have been rendered immune, it is found that the parasite *begins to develop, but that from the onset a reaction on the part of the organism shows itself*, accompanied by a considerable emigration of leucocytes, which soon include the bacteria in great numbers.

This relationship of phagocytosis to acquired immunity is in the highest degree instructive. Where a given species of animal is specially sensitive to the onslaught of one or other microorganism, there, during the course of the disease, the phagocytes are inoperative, including none, or almost none, of the bacteria. On the other hand, when by previous vaccination these animals have been rendered refractory, their phagocytes have acquired the property of including the same bacteria. As an example of this I may cite the action of the bacillus of anthrax and of the *Vibrio Metschnikovi*. In ordinary rabbits the development of anthrax is only followed by a very feeble phagocytosis, while in vaccinated rabbits this phagocytosis is very extensive. Corresponding but yet more strongly marked differences are to be made out between the unvaccinated guinea-pig—an animal most readily affected by the vibronic septicæmia—and the guinea-pig vaccinated against the same; after inoculation with the *Vibrio Metschnikovi* none of the vibrios are to be found in the cells of the former; in the latter the phagocytes are simply replete with the microbes.

The facts enumerated thus far would seem to prove that there exists a certain antagonism between the microbes and the phagocytes, and this view is confirmed by the fact that in general the microbes find the interior of the

phagocytes an unfavorable medium for their development and continued existence. Very often it is possible to determine absolutely that the parasites are killed within the phagocytes; after inoculating refractory animals with bacteria, an afflux of white corpuscles toward the region of inoculation, followed by the inclusion of the bacteria and by their death, is seen to occur. These stages can be well followed where the anthrax bacilli are taken into the phagocytes of animals that are, or have been rendered, immune. This occurs also with a long series of other microorganisms studied in this connection, and, among others, in the case of the tubercle bacillus invading animals that are more or less resistant. The giant cells of tuberculosis are, in fact, huge multinuclear phagocytes, and here the intracellular destruction of the bacilli is the more clearly demonstrable, inasmuch as the microorganisms exhibit such very evident signs of degeneration; the bacilli swell, their enveloping membrane becomes much thickened and highly refractive, and at the same time the contents lose their power of fixing the staining material, so that eventually, nothing is left but slightly yellowish forms, recalling, in proportions and position, the enlarged bacilli; and these shadowy bodies unite into small masses of an amber-like appearance. Analogous transformations never being observable outside the phagocytes—that is to say, either in cultures or in caseating masses—these changes may well be regarded as due to a specific action upon the part of the giant cells.

The broad fact that the invasion of the organism by microbes most often induces, on the one hand, an inflammatory reaction with its associated emigration of leucocytes, and that, on the other hand, the phagocytes are capable of including and destroying the invaders, leads us to admit that *the afflux of phagocytes to the invaded region, and their bactericidal properties, are mechanisms which serve to ward off bacterial attack and to maintain the integrity of the organism.* Where the phagocytes do not, either immediately or eventually, intervene, but leave the field free to the microbes, these multiply without hindrance and succeed in killing the animal within, it may be, an excessively short period. Thus the microorganism of hog cholera, which is left quite untouched, kills the pigeon in the course of a few hours—often within five hours after inoculation; chicken cholera kills not only pigeons but also rabbits in an equally short period. In other diseases, in which the phagocytes appear upon the scene in relatively large numbers, and even include the microorganisms, the latter gain the day whenever and wherever the phagocytes are incapable of destroying them or of preventing their growth.

This manifest bactericidal action is to be compared with the phenomenon of intracellular digestion characteristic of amœboid cells in general, and of leucocytes and other microbic phagocytes in particular. These cells have the power of digesting with ease red corpuscles and other organized elements, just as have the amœbæ proper and other protozoa. Among the last are many which have been found to include and transform bacteria exactly the same way as do the phagocytes of the higher animals.

Now, in determining the intervention or non-intervention of the leucocytes in this war between the organism and the bacteria, a very great part is played by the sensitiveness of these cells to external influences, and especially to the chemical composition of their environment. The leucocytes are powerfully attracted by many microorganisms and the resultants of their growth, and as powerfully repelled by others and their resultants, as it is expressed, they have a *positive chemiotaxis* for certain microbes and a *negative chemiotaxis* for others. The existence of these chemiotactic properties has been so clearly proved of late by the researches of Leber, Metschnikoff, and Bordet, and Gabritschewski that I need not enter into a fuller explanation of the subject here. Where *negative chemiotaxis* manifests itself, the parasites, being shunned by the white corpuscles, the parasites freely propagate themselves and induce the death of their host. Nevertheless this chemiotaxis is not immutable, and the cells can become accustomed to substances from which they shrank at first—a negative may thus be transformed into

positive chemiotactic state. Such obtains in acquired immunity; the cells which in the unvaccinated animal never included the bacteria, now in the vaccinated take them up readily. . . .

There is not a single portion of the theory which I have just expounded but has encountered a lively opposition. Even the fundamental fact that the phagocytes are capable of including the microbes has had doubts thrown upon it; it has been held that the latter insinuate themselves into the former. Only after successive series of observations upon the phagocytes and the living microbes has it been proved that assuredly it is the phagocytes which, by the aid of their pseudopodia, themselves include the microorganisms. The observer can see the whole process in the case of immobile bacilli—can see the leucocyte approach, send out pseudopodia, and gradually include the individual bacillus. Or, conversely, in cases of negative chemiotaxis, one can, in blood taken from the monkey during the access of relapsing fever, observe the actively moving spirilla come into contact with a leucocyte, and even become attached by one end to its surface; yet, however active the movement, one never finds that the spirillum succeeds in piercing the surface and gaining an entrance. If it be suggested that this entry may take place in consequence of the force of active growth and elongation of bacilli, then, apart from the fact that here but one set of cases is embraced, it can be determined that this force is too feeble—it can be seen that, during the active growth of the anthrax organism in the blood, the elongating chains of bacilli curve in and out between the corpuscles, but never penetrate the cells.

From another side the objection has been formulated that in many cases the organism gets rid of its invaders without the aid of the phagocytes. According to those who support this objection, this happens in the anthrax of pigeons (Czaplewski) and of refractory rats (Behring, Franck), in symptomatic anthrax of various refractory animals (Rogowicz), and in the septicæmia of vaccinated guinea-pigs, due to the *Vibrio Metschnikovi* (R. Pfeiffer). A reëxamination of the cases here adduced has, however, shown that in each a very considerable phagocytosis can be proved, and that the negative results of the above observers have been due to insufficient methods of observation.

While accepting that the phagocytes do truly absorb the microorganisms, other opponents of the theory have urged that these cells are only capable of including microorganisms already killed by other means, and that living microbes are solely to be found within the cells in those cases where there has been a fatal ending—in tuberculosis, mouse septicæmia, and so on. Against this may be brought the fact determined by Lubarsch, that the phagocytes of several animals, refractory to anthrax, take up living bacilli that have been injected, with greater eagerness than they include those which have been killed before injection. But, further, this objection may be disposed of by direct observation of bacteria undergoing development from within the interior of phagocytes after the latter have been destroyed by a substance which is at the same time a favorable medium for bacterial growth—as, for instance, beef broth. Such observations have been made upon pigeons rendered immune to anthrax.

During the last year or two great stress has been laid upon the fact that the body humors themselves possess most marked bactericidal properties, and, in fact, against the theory of phagocytosis has been brought another, based upon this power of the humors to destroy the microorganisms. Observer after observer has remarked that in blood plasma, defibrinated blood, blood serum, and in the blood as a whole, in the removed aqueous humor and other fluids and exudations of the body, many species of bacteria perish after a longer or shorter interval; and forthwith an endeavor has been made to find in these facts some elucidation of the phenomena of immunity. Yet the more deeply one examines into the question the more one is convinced that no relationship exists between the two. Thus it happens often that the bactericidal property is more developed in susceptible species than

in refractory ; so, with regard to the anthrax bacilli, in the very sensitive rabbit the bactericidal properties of the humors are more pronounced than they are in the refractory dog ; and Behring and Nissen, the two who most simultaneously first drew our attention to these phenomena, in the combined research, recently published, admit that, as against the bacteria of anthrax, pneumonia, and diphtheria, this bactericidal property exists to the same degree in the juices of animals of the same species, whether they be susceptible or have been rendered immune. Often, again, it has been determined that the blood removed from the organism has a greater power of destroying bacteria than it has within the organism. A small quantity of blood withdrawn from the body will, in certain instances, kill a mass of bacilli greater than that which, injected into the circulation, would inevitably cause death. Evidently, therefore, in this bactericidal influence external vascular phenomena enact an important rôle—phenomena, that is, which have no connection with what occurs in the living refractory organism.

From another point of view strong arguments have been directed against this theory of the tissue fluids. It has been shown, especially by the researches of M. Haffkine, that the death of the bacteria transported into organic fluids is largely due to the sudden change of medium, and that, passing from one medium to another by successive slight modifications of the fluid of growth, it is easy to make bacteria live in fluids which, when the change of environment has been abrupt, swiftly lead to their destruction.

In order to gain an idea as to the part played in the refractory animal the fluids and the phagocytes respectively, the endeavor has been made to separate the two by placing under the skin of frogs (which are naturally immune to anthrax) minute packets formed of filter paper or of animal membrane, and containing the bacilli. The paper, while permitting the passage of fluid, wards off the wandering amoeboid cells for a certain time. Shielded in this way from the phagocytes, though exposed to the action of the juices, the bacilli grow well and produce the characteristic felted mass of anthrax filaments. Baumgarten has not been able to confirm this experiment, but Hueppe and Lubarsch have repeatedly verified it.

But it is not even necessary to take these precautions in order to assure one's self that anthrax spores germinate in the juices of refractory animals. Recently, for instance, M. Trapeznikoff has found that, when these spores are injected into the dorsal lymph sac of the frog, they constantly tend to develop into bacilli, whose further growth is stopped by the phagocytes which include them, along with such spores as have not had time to germinate. Eventually the bacilli so absorbed are digested by their hosts, while the included spores remain intact, although incapable of giving birth to new bacilli for so long a time as the phagocytes remain alive. And I might adduce other similar cases. Such a comparative examination proves that in the living body the bactericidal property resides in the phagocytes and not in the fluids.

Still, it may be urged that possibly these cells, which can thus devour and destroy the living microbes, are only in a position to attack bacteria whose virulence has already been lessened by other means. Were this so, the microbes present in a refractory organism should behave, not like parasites, but as simple, inoffensive saprophytes. Hence these microbes—powerless to produce upon a refractory soil the toxic substances which render them pathogenic and dangerous—should easily be included and destroyed ; that, according to this hypothesis, which has frequently been brought forward, the phagocytes play a purely secondary and dependent part, waiting until the microbes are weakened before they seize upon them. In favor of this view the fact has been cited that certain microorganisms cultivated in the blood, or serum, of vaccinated animals become attenuated, so that they no longer induce a fatal disease. The *Bacillus anthracis* grown in the blood of vaccinated sheep no longer kills rabbits, and, according to Roger, the *Streptococcus erysipelas* grown in the blood of vaccinated rabbits on

occasions a slight and passing disturbance in susceptible members of the same species. But here again we are dealing with fluids withdrawn from the body, and so modified in various ways. Let us make an observation more strictly to the point. Take, for instance, a rabbit vaccinated against anthrax and inoculate it with anthrax bacilli, thus allowing these to exist directly within the refractory organism. Such bacilli as are not destroyed preserve their virulence for a sufficiently long period, and it is possible to kill a guinea-pig with a drop of exudation, taken from the region of injection thirty hours after subcutaneous inoculation, eight days after inoculation into the anterior chamber of the eye. A sojourn of so long duration within the vaccinated organism, then, has not deprived the microbes of their virulence, although twenty-four hours suffice to completely attenuate the bacilli cultivated in the removed blood of vaccinated sheep.

Years ago it was established in M. Pasteur's laboratory that the refractory organism, instead of being an unfavorable soil for the preservation of virulence, tends the rather to reinforce this property. To exalt the virulence of an attenuated microorganism, one always employs, not animals very susceptible to the specific disease, but those which are slightly susceptible, or it may be, under many circumstances, refractory. In this manner the most active anthrax virus has usually been obtained by passage through birds, notably fowls; the greatest virulence of chicken cholera was gained by passage through the vaccinated cock; and quite recently M. Malm has shown that passage of the anthrax bacillus through the organisms of dogs, which of all mammals are the most refractory in this respect, increases its virulence in a most remarkable manner, so that the general law may be laid down that an organism which is but slightly susceptible or is refractory is able not only to preserve, but even to exalt, the virulence of bacteria. The principal argument in favor of the hypothesis that pathogenic microorganisms become simple inoffensive saprophytes when they find themselves in a refractory region, loses therefore its *raison d'être*.

M. Bouchard, in his objection to the theory of phagocytosis, may be regarded as introducing but a modification of this hypothesis. He holds that pathogenic bacteria placed under favorable conditions give rise to substances which hinder the inflammatory process, and that only when these inhibitory substances are inadequately represented do the cells intervene. When, therefore, the organism rendered refractory by vaccination becomes an unfavorable soil for the production of these inhibitory bodies, the bacteria can no longer prevent the inflammatory reaction; free emigration of the leucocytes ensues, these cells seize upon the impotent microbes and put a stop to their further growth. In this theory the part played by the phagocytes is again secondary, depending upon a dearth of anti-inflammatory substance.

If the theory could be accepted in certain cases, it is nevertheless inapplicable as a general rule. In all those affections which are characterized by the absence of leucocytes upon the field of battle there is certainly no lack of inflammation. The very reverse obtains. In anthrax affecting small mammals, just as in the vibronic septicæmia of pigeons and guinea-pigs, and other analogous diseases, we find that there is a very distinct dilatation of the vessels, accompanied by great exudation; the inflammatory reaction is well marked; nothing is wanting save the determination of the white corpuscles. Or, employing yet further that affection which is, as it were, the touchstone of the bacteriologist, a still clearer proof of our contention is to be gained if we inoculate a rabbit on the one ear with a small quantity of virulent, on the other with a like quantity of attenuated, anthrax virus. In the course of a few hours the external signs of inflammation are far more conspicuous in the former; the vessels are greatly enlarged and there is literally a huge exudation of clear serous fluid into the part; in the latter the external signs are less prominent, but examination of the seat of inoculation shows it to be packed with leucocytes. Consequently, the phenomenon we are discussing is to be explained, not by an absence of the inflammatory process, but much more satisfactorily by a negative chemiotaxis of the

leucocytes, which, instead of being attracted by the bacterial products, repelled; where the animal is vaccinated or refractory a much slighter inflammation is sufficient to produce an abundant emigration of the cocytes.

Recently Behring has brought forward another view which would explain immunity in a wholly different way. According to him, the bacteria can live, and even preserve their virulence, in the refractory organism, but the *toxines* excreted by them now undergo a modification so as to be rendered completely inoffensive for the animal. And to this "toxic property" of the organism is to be attributed the essential quality of the immune state. It is impossible to pronounce upon the arguments that have led up to this theory, for as yet they have not been circumstantially set forth; but already one can declare that such a theory is in no wise applicable to the phenomena of immunity in general. In three diseases remarkable for their pronounced toxic character—vibronic septicæmia, pyocyanic sepsis, and hog cholera affecting the rabbit—as shown by the experiments of Charrin, Gamaléia, and Selander, the *toxines* are so little attacked by the refractory organism that the same quantity of these substances (freed from bacteria) suffices to kill an animal very susceptible to one or other disease, and an animal vaccinated against it and thus completely immune. So, non-fatal doses of these *toxines* produce in animals of the two categories the same febrile and inflammatory reactions. The proof is clear that there is no special destruction of *toxines* in the refractory animal, and that the "toxic property," if it exists, is not one whit more developed after vaccination than before. Passing in review all these counter theories, we see that each of them can only be applied to a certain number of facts; in some an attenuating or even bactericidal influence of the juices is relied upon, in others an anti-inflammatory action, in yet others a toxicide property. Still the phagocytic reaction is the only constant in all those cases of immunity, and recovery that have as yet been sufficiently studied, and while certain of the factors mentioned (the attenuating and toxicide properties) do not in the least touch upon the continued existence or otherwise of the microorganisms, the bactericidal power of the phagocyte puts an end to the parasite itself, thus at a given moment prevents further manifestation of its virulence, and preserves the animal attacked at a time when the toxicide properties would have been found wanting, and the microbe remaining alive would consequently gain the upper hand.

But while thus placing before you the important part played by the phagocytes, I do not wish it to be thought that these cells are unaided in this contest by other defensive means possessed by the organism. This is far from being my view. Thus, in the febrile reaction, we see a puissant auxiliary very definitely favoring the work of the phagocytes. This febrile action has only to be inhibited—as was done by M. Pasteur in the anthrax of fowls—and animals naturally refractory to the affection succumb to the ravages of the bacilli. It is not possible at the present time to state fully and accurately all these influences which are associated in aiding phagocytic action, but already we have the right to maintain that, *in the property of its amœboid cells to include and to destroy microorganisms, the animal body possesses a formidable means of resistance and defence against these infectious agents.*¹

We are disposed to agree with Metschnikoff in his final conclusion, as above stated in italics. But in view of experimental evidence, to be referred to later, we cannot accept the so-called Metschnikoff theory as a sufficient explanation for the facts relating to natural and acquired immunity in general, and must regard phagocytosis simply as a factor which, in certain infectious diseases,

¹ From the British Medical Journal.

pears to play an important part in enabling immune animals to resist invasion by pathogenic bacteria.

Going back to the demonstrated fact that susceptible animals may be made immune by inoculating them with the toxic products produced during the growth of certain pathogenic bacteria, we may suppose either that immunity results from the continued presence of these toxic products in the body of the inoculated animal, or from a tolerance acquired at the time of the inoculation and subsequently retained—by transmission from cell to cell, as heretofore suggested. Under the first hypothesis—retention theory—immunity may be explained as due to a continued tolerance on the part of the cellular elements of the body to the toxic substances introduced and retained; or to the effect of these retained toxic products in destroying the pathogenic bacteria, or in neutralizing their products when these are subsequently introduced into the body of the immune animal. We cannot understand how toxic substances introduced in the first instance can neutralize substances of the same kind introduced at a later date. There is something in the blood of the rat which, according to Behring, neutralizes the toxic substances present in a filtered culture of the tetanus bacillus; but whatever this substance may be, it is evidently different from the toxic substance which it destroys, and there is nothing in chemistry to justify the supposition last made. Is it, then, by destroying the pathogenic microorganism that these inoculated and retained toxic products preserve the animal from future infection? Opposed to this supposition is the fact that the blood of an animal made immune in this way, when removed from the body, does not prove to have increased germicidal power as compared with that of a susceptible animal of the same species. Again, these same toxic substances in cultures of the anthrax bacillus, the tetanus bacillus, the diphtheria bacillus, etc., do not destroy the pathogenic germ after weeks or months of exposure. And when we inoculate a susceptible animal with a virulent culture of one of these microorganisms, the toxic substances present do not prevent the rapid development of the bacillus; indeed, instead of proving a germicide, they favor its development, which is more abundant and rapid than when attenuated cultures containing less of the toxic material are used for the inoculation. In view of these facts we are unable to adopt the view that acquired immunity results from the direct action of the products of bacterial growth, introduced and retained in the body of the immune animal, upon the pathogenic microorganism when subsequently introduced or upon its toxic products.

But there is another explanation which, although it may appear *a priori* to be quite improbable, has the support of recent experimental evidence. This is the supposition that *some substance is formed*

in the body of the immune animal which neutralizes the toxic products of the pathogenic microörganism. How the presence of these toxic products in the first instance brings about the formation of an "antitoxine" by which they are neutralized is still a mystery, but that such a substance is formed appears to be proved by the recent experiments of Ogata, Behring and Kitasato, Tizzoni and Cattani, G. and F. Klemperer, and others.

Ogata and Jasuhara, in a series of experiments made in the Hygienic Institute at Tokio (1890), discovered the important fact that the blood of an animal immune against anthrax contains some substance which neutralizes the toxic products of the anthrax bacillus. When cultures were made in the blood of dogs, frogs, or of white rats, which animals have a natural immunity against anthrax, they were found not to kill mice inoculated with them. Further experiments showed that mice inoculated with virulent anthrax culture did not succumb to anthrax septicæmia if they received at the same time a subcutaneous injection of a small quantity of the blood of an immune animal. So small a dose as one drop of frog's blood or one half drop of dog's blood proved to be sufficient to protect a mouse from the fatal effect of an anthrax inoculation. And the protective inoculation was effective when made as long as seventy-two hours before or five hours after infection with an anthrax culture. Further, it was found that mice which had survived anthrax infection as a result of this treatment were immune at a later date (after several weeks) when inoculated with a virulent culture of the anthrax bacillus.

Behring and Kitasato have obtained similar results in their experiments upon tetanus and diphtheria, and have shown that the blood of an immune animal, added to virulent cultures before inoculation into susceptible animals, neutralizes the pathogenic power of these cultures.

They have shown by experiment that the blood of a rabbit which has an acquired immunity against tetanus, mixed with the virulent filtrate from a culture of the tetanus bacillus, neutralizes its toxic power. One cubic centimetre of this filtrate was mixed with five cubic centimetres of serum from the blood of an immune rabbit and allowed to stand for twenty-four hours; 0.2 cubic centimetre of this injected into a mouse was without effect, while 0.0001 cubic centimetre of the filtrate without such admixture was infallibly fatal to mice. The mice inoculated with this mixture remained immune for forty to fifty days, after which they gradually lost their immunity. The blood or serum from an immune rabbit, when preserved in a dark, cool place, retained its power of neutralizing the tetanus toxin in albumin for about a week, after which time it gradually lost the

power. The blood of chickens, which have a natural immunity against tetanus, was found not to have a similar power. Behring and Kitasato have also shown that the serum of a diphtheria-immune rabbit destroys the potent toxalbumin in diphtheria cultures. It does not, however, possess any germicidal power against the diphtheria bacillus.

Ogata, in a recent publication (1891), reports that he has succeeded in isolating from the blood of dogs and of chickens a substance to which he ascribes the immunity of these animals from certain infectious diseases, and the power of their blood to protect susceptible animals from the same diseases. This substance is soluble in water and in glycerin, but insoluble in alcohol or ether, by which it is precipitated without being destroyed. Its activity is neutralized by acids, but not by weak alkaline solutions. Ogata supposes the substance isolated by him to be the active agent in blood serum by which certain pathogenic bacteria are destroyed, as shown by the experiments of Nuttall, Buchner, and others. Hankin had previously isolated an albuminoid substance from the spleen and blood of the rat, to which he ascribes the immunity of this animal from anthrax. This substance, according to the author named, is a globulin; it is insoluble in alcohol and in distilled water, and does not dialyze.

Tizzoni and Cattani ascribe the protection of animals which have acquired an immunity against tetanus to the presence of an albuminous substance which they call the tetanus-antitoxine. This they have isolated from the blood of immune animals. They arrive at the conclusion that it is a globulin, or a substance which is carried down with the globulin precipitate, and that it is different from the globulin, above referred to, obtained by Hankin from animals immune against anthrax.

G. and F. Klemperer have recently (1891) published an important memoir in which they give an account of their researches relating to the question of immunity, etc., in animals subject to the form of septicæmia produced by the *Micrococcus pneumoniae crouposæ*. They were able to produce immunity in susceptible animals by introducing into their bodies filtered cultures of this micrococcus, and proved by experiment that this immunity had a duration of at least six months. They arrive at the conclusion that the immunity induced by injecting filtered cultures is not directly due to the toxic substances present in these cultures, but that they cause the production in the tissues of an antitoxine which has the power of neutralizing their pathogenic action. The toxic substance present in cultures of the "diplococcus of pneumonia" they call "pneumotoxine"; the substance produced in the body of an artificially immune animal, by

which this pneumotoxine is destroyed if subsequently introduced, to call "anti-pneumotoxine."

Emmerich, in a communication made at the recent (1891) International Congress for Hygiene and Demography, in London, reports results which correspond with those of G. and F. Klemperer as far as the production of immunity is concerned, and also gives an account of experiments made by Dönissen in which the injection of twenty to twenty-five cubic centimetres of blood or expressed tissue juices, filtered through porcelain, from an immune rabbit into an unprotected rabbit, subsequently to infection with a bouillon culture of "*diplococcus pneumoniae*," prevented the development of fatal septicaemia. Even when the injection was made twelve to fourteen hours after infection, by inhalation, the animal recovered.

Emmerich and Mastrbaum had previously reported similar results in experiments made upon mice with the *Bacillus erysipelatos* (rothlauf bacillus). White mice are very susceptible to the pathogenic action of this bacillus. But mice which, subsequently to infection, were injected with the expressed and filtered tissue juices of an immune rabbit, recovered, while the control animals succumbed. According to Emmerich, the result in these experiments was due to a destruction of the pathogenic bacilli in the bodies of the infected animals; and the statement is made that at the end of eight hours after the injection of the expressed tissue juices all bacilli in the body of the infected animal were dead. The same liquid did not, however, kill the bacilli when added to cultures external to the body of the animal. The inference, therefore, seems justified that the result depends, not upon a substance present in the expressed juices of the immune animal, but upon a substance formed in the body of the animal into which these juices are injected.

We have, however, an example of induced immunity in which the result appears to depend directly upon the destruction of the pathogenic microorganism in the body of the immune animal. In guinea-pigs which have an acquired immunity against *Vibrio Metchnikovi* the blood serum has been proved to possess decided germicidal power for this "*vibrio*," whereas it multiplies readily in the blood serum of non-immune guinea-pigs (Behring and Nissen).

There is experimental evidence that animals may acquire an artificial immunity against the toxic action of certain toxalbumins from other sources than bacterial cultures. Thus Sewell (1887) has shown that a certain degree of tolerance to the action of rattlesnake venom may be established by inoculating susceptible animals with small doses of the "*hemialbumose*" to which it owes its toxic potency. In this connection we may remark that there is some evidence to show that persons who are repeatedly stung by certain poisonous

insects—mosquitoes, bees—acquire a greater or less degree of immunity from the distressing local effects of their stings.

Recently (1891) Ehrlich, of Berlin, has reported his success in establishing immunity in guinea-pigs against two toxalbumins of vegetable origin: one—ricin—from the castor-oil bean (*Ricinus communis*), the other—abrin—from the jequirity bean. The toxic potency of ricin is somewhat greater than that of abrin, and it is estimated by Ehrlich that one gramme of this substance would suffice to kill one and a half millions of guinea-pigs. When injected beneath the skin, in dilute solution, it produces intense local inflammation, resulting in necrosis of the tissues. Mice are less susceptible than guinea-pigs and are more easily made immune. This is most readily effected by giving them small and gradually increasing doses with their food. As a result of this treatment the animal resists subcutaneous injections of two hundred to four hundred times the fatal dose for animals not having this artificial immunity. The fatal dose of abrin is about double that of ricin. When injected into mice in the proportion of one cubic centimetre to twenty grammes of body weight a solution of one part in one hundred thousand of water proved to be a fatal dose. The local effects are also less pronounced when solutions of abrin are used; they consist principally in an extensive induration of the tissues around the point of injection and a subsequent falling off of the hair over this indurated area. When introduced into the conjunctival sac, however, abrin produces a local inflammation in smaller amounts than ricin, a solution of 1:800 being sufficient to cause a decided but temporary conjunctivitis. Solutions of 1:50 or 1:100 of either of these toxalbumins, introduced into the eye of a mouse, give rise to a panophthalmitis which commonly results in destruction of the eye. But in mice which have been rendered immune by feeding them for several weeks with food containing one of these toxalbumins, no reaction follows the introduction into the eye of the strongest possible solution, or of a paste made by adding abrin to a little ten-per-cent salt solution. Ehrlich gives the following explanation of the remarkable degree of immunity established in his experiments by the method mentioned:

“All of these phenomena depend, as may be easily shown, upon the fact that the blood contains a body—antiabrin—which completely neutralizes the action of the abrin, probably by destroying this body.”

In a more recent paper Ehrlich has given an account of subsequent experiments which show that the young of mice which have an acquired immunity for these vegetable toxalbumins may acquire immunity from the ingestion of the mother's milk; and also that immunity against tetanus may be acquired in a very brief time by young mice through their mother's milk. In his tetanus experi-

ments Ehrlich used blood serum from an immune horse to give immunity to the mother mouse when her young were already seven to ten days old. Of this blood serum two cubic centimetres were injected at a time on two successive days. The day after the first injection one of the sucklings received a tetanus inoculation by means of a splinter of wood to which spores were attached. The animal remained in good health, while a much larger control mouse inoculated in the same way died of tetanus at the end of twenty-four hours. Other sucklings, inoculated at the end of forty-eight and seventy-two hours after the mother had received the injection of blood serum, likewise remained in good health, while other control mice died.

A most interesting question growing out of these extraordinary experimental results at once presents itself: Does the animal which is immune for one of these toxalbumins also exhibit immunity as regards the toxic action of the other? This question Ehrlich has answered. His experiments show that animals which are immune against one of these substances are quite as susceptible to the toxic action of the other as if they did not possess this immunity—*i.e.*, the antitoxine of ricin does not destroy abrin, and *vice versa*. As an illustration of this fact he states that in one experiment a rabbit was made immune against ricin to such an extent that the introduction into its eye of this substance in powder produced no inflammatory reaction; but the subsequent introduction of a solution of abrin in 1:10,000 caused a violent inflammation.

Evidently these facts are of the same order as those relating to immunity from infectious diseases, and, taken in connection with the experimental data previously referred to, give strong support to the view that the morbid phenomena in all diseases of this class are due to the specific toxic action of substances resembling the toxalbumins already discovered; and that acquired immunity from any one of these diseases results from the formation of an antitoxine in the body of the immune animal.

Hankin calls these substances produced in the bodies of immune animals "defensive proteids," and proposes to classify them as follows: First, those occurring naturally in normal animals, which he calls *sozins*; second, those occurring in animals that have acquired an artificial immunity—these he calls *phylaxins*. Each of these classes of defensive proteids is further subdivided into those which act upon the pathogenic microorganism itself and those which act upon its toxic products. These subclasses are distinguished by the prefixes *myco* and *toxo* attached to the class name.

In accordance with this classification a *mycosozin* is a defensive

proteid, found in the body of a normal animal, which has the power of destroying bacteria.

A *toxosozin* is a defensive proteid, found in the body of a normal animal, which has the power of destroying the toxic products of bacterial growth.

A *mycophylaxin* is a defensive proteid produced in the body of an animal which has an acquired immunity for a given infectious disease, which has the power of destroying the pathogenic bacteria to which the disease is due.

A *toxophylaxin* is a defensive proteid produced in the body of an animal which has an acquired immunity for a given infectious disease, which has the power of destroying the toxic products of the pathogenic bacteria to which the disease is due.

Buchner had previously proposed the name "alexines" for these defensive proteids.

The importance of the experimental evidence above referred to in explaining the phenomena of natural and acquired immunity is apparent. The facts stated also suggest a rational explanation of recovery from an attack of an acute infectious disease. But the idea that during such an attack an antidote to the disease poison is developed in the tissues is yet so novel, and the experimental evidence in support of this view is of such recent date, that it would be premature to accept this explanation as applying to immunity in general. It seems difficult to believe that an individual who has passed through attacks of measles, mumps, whooping cough, scarlet fever, small-pox, etc., has in his blood or tissues a store of the antitoxine of each of these diseases, formed during the attack and retained during the remainder of his life, or continuously produced so long as the immunity lasts. Moreover, in those diseases to which the experimental evidence above recorded relates—diphtheria, tetanus, pneumonia—as they occur in man, no lasting immunity has been shown to result from a single attack, and in this regard they do not come into the same class with the eruptive fevers and other diseases in which a single attack usually protects during the lifetime of the individual.

In those instances in which acquired immunity has been shown to be due to the production in the body of the immune animal of an antitoxine, it is still uncertain whether there is a continuous production of the protective proteid, or whether that formed during the attack remains in the body during the subsequent immunity. The latter supposition appears at first thought improbable; but when we remember that the protective proteids which have been isolated by Hankin from the blood and spleen of rats, and by Tizzoni and Cattani from the blood of animals made immune against tetanus, do

not dialyze, it does not seem impossible that these substances might be retained indefinitely within the blood vessels. On the other hand, the passage of the tetanus antitoxine into the mother's milk, as shown by Ehrlich's experiments upon mice, indicates a continuous supply, otherwise the immunity of the mother would soon be lost.

The writer has recently (May, 1892) obtained experimental evidence that the blood of vaccinated, and consequently immune, calves contains something which neutralizes the specific virulence of vaccine virus, both bovine and humanized. Four drops of blood serum from a calf which had been vaccinated two weeks previously, mixed with one drop of liquid lymph recently collected in a capillary tube, after contact for one hour was used to vaccinate a calf; the same animal was also vaccinated with lymph, preserved on three quills, which was mixed with four drops of serum from the immune calf and left for one hour. The result of these vaccinations was entirely negative, while vaccinations upon the same calf made with virus from the same source, and mixed with the same amount of blood serum from a non-immune calf, gave a completely successful and typical result.

The experimental evidence detailed gives strong support to the view that *acquired immunity depends upon the formation of antitoxines in the bodies of immune animals*. As secondary factors it is probable that tolerance to the toxic products of pathogenic bacteria and phagocytosis have considerable importance, but it is evident that the principal rôle cannot be assigned to these agencies.

PLATE IV.

FIGS. 1, 2, and 3.—Leucocytes from the spleen of an inoculated monkey, containing *Spirillum Obermeieri*. (Soudakewitch.)

FIGS. 4 and 5.—Leucocytes ("macrophages") from a preparation of muscle from a pigeon which succumbed to an anthrax inoculation. In Fig. 4 the bacilli are deeply stained; in Fig. 5 they are pale. (Metschnikoff.)

FIG. 6.—Leucocyte from a frog seventy-two hours after the injection of anthrax spores. (Trapeznikoff.)

FIGS. 7 and 8.—Leucocytes from a chicken four hours after the injection of anthrax spores. (Trapeznikoff.)



Fig. 1.



Fig. 2.



Fig. 3.



Fig. 4.



Fig. 5.



Fig. 6.

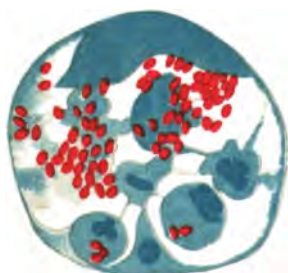


Fig. 7.

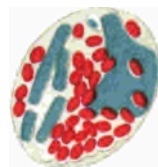


Fig. 8.

PHAGOCYTES.

IV.

PYOGENIC BACTERIA.

THE demonstration made by Ogston, Rosenbach, Passet, and others that micrococci are constantly present in the pus of acute abscesses, led to the inference that there can be no pus formation in the absence of microorganisms of this class. But it is now well established, by the experiments of Grawitz, De Bary, Steinhaus, Scheurlen, Kaufmann, and others, that this inference was a mistaken one, and that certain chemical substances introduced beneath the skin give rise to pus formation quite independently of bacteria. Among the substances tested which have given a positive result are nitrate of silver, oil of turpentine, strong liquor ammoniæ, cadaverin, etc. The demonstration has also been made by numerous investigators that cultures of pus cocci, when sterilized by heat, still give rise to pus formation when injected subcutaneously. This was first established by Pasteur in 1878, who found that sterilized cultures of his "microbe générateur du pus" induced suppuration as well as cultures containing the living microbe. This fact has since been confirmed, as regards the pus staphylococci and various bacilli, by a number of bacteriologists. Wyssokowitsch produced abscesses containing sterile pus by injecting subcutaneously agar cultures of the anthrax bacillus sterilized by heat. Buchner obtained similar results in a series of forty experiments from the injection of sterilized cultures of Friedländer's bacillus ("pneumococcus"), and has shown that the pus-forming property belongs to the bacterial cells and not to a soluble chemical substance produced by them. When cultures were filtered by means of a Chamberlain filter the clear fluid which passed through the porous porcelain was without effect, while the dead bacteria retained by the filter produced aseptic pus infiltration in the subcutaneous tissues within forty-eight hours after having been injected. Subsequent experiments gave similar results with seventeen different species tested, including *Staphylococcus pyogenes aureus*, *Staphylococcus cereus flavus*, *Sarcina aurantiaca*, *Bacillus prodigiosus*, *Bacillus Fitzianus*, *Bacillus subtilis*, *Bacillus coli communis*, *Bacillus acidi lactici*, etc. From the experi-

ments made to determine the exact cause of pus formation following the injection of sterilized cultures Buchner arrives at the conclusion that it is due to the albuminous contents of the bacterial cells.

While it is demonstrated that a large number of microorganisms, either living or in sterilized cultures, may give rise to the formation of pus, the extended researches of Rosenbach, Passet, and other bacteriologists show that few species are usually concerned in the formation of acute abscesses, furuncles, etc., in man. Of these two most important, by reason of their frequent occurrence and pyogenic power, are *Staphylococcus pyogenes aureus* and *Streptococcus pyogenes*; next to these comes *Staphylococcus pyogenes albus*, and the following species are occasionally found: *Staphylococcus pyogenes citreus*, *Staphylococcus cereus flavus*, *Staphylococcus cereus albus*, *Micrococcus tenuis*, *Bacillus pyogenes foetidus*, *Micrococcus tetragenus*, *Micrococcus pneumoniae crouposa*. Two or more species are often found in the same abscess; thus Passet, in thirty-three cases of acute abscess, found *Staphylococcus aureus* and *Streptococcus pyogenes* alone in eleven, *albus* alone in four, *albus* and *citreus* in two, *Streptococcus pyogenes* alone in eight, *albus* and *Streptococcus pyogenes* in one, and *albus*, *citreus*, and *Streptococcus pyogenes* in one. Hoffa found, in twenty-two cases of inguinal bubo, *aureus* in ten, *albus* in nine, and *citreus* in three. Bumm, in ten cases of puerperal mastitis, found *aureus* in seven and *Streptococcus pyogenes* in three. Rosenbach found staphylococci alone sixteen times, *Streptococcus pyogenes* alone fifteen times, staphylococci and streptococci associated five times, and *Micrococcus tenuis* three times in thirty-nine acute abscesses and phlegmons examined by him.

Robb and Ghriksy have shown that under the most rigid antiseptic treatment microorganisms are constantly found attached to sutures when these are removed from wounds made by the surgeon, and that a skin abscess frequently results from the presence of the most common of these microorganisms—*Staphylococcus epidermidis albus*.

The authors named state their conclusions as follows:

"A wound, at some time of its existence, always contains organisms. They occur either on the stitches or in the secretions.

"The number of bacteria is influenced by the constricting action of ligatures or drainage tube, or anything interfering with the circulation of the tissues.

"The virulence of the organisms present will influence the progress of the wound.

"The body temperature is invariably elevated if the bacteria are virulent; and, indeed, in cases where many of the less virulent organisms are found, almost without exception there is some rise of temperature."

The organism most frequently found—*Staphylococcus epidermidis*

dis albus—has but slight virulence. Out of forty-five cases in which a bacteriological examination was made this micrococcus was obtained in pure cultures in thirty-three; in five cases it was associated with *Staphylococcus pyogenes aureus*, in one case with *Streptococcus pyogenes*, in three cases *Streptococcus pyogenes* was obtained alone.

In abscesses resulting from inflammation of the middle ear the micrococcus commonly known under the name of "*diplococcus pneumoniae*"—*Micrococcus pneumoniae crouposæ*—has been obtained in pure cultures in a considerable number of cases when the pus has been examined immediately after paracentesis of the tympanic membrane. We shall not, however, describe this among the pyogenic bacteria, but will give an account of it in the following section (*Bacteria in Croupous Pneumonia, etc.*). *Bacillus pyocyaneus*, which is described by some authors among the pyogenic bacteria, is found only in the pus of open wounds, where its presence is evidently accidental. We shall describe it among the chromogenic saprophytes.

1. STAPHYLOCOCCUS PYOGENES AUREUS.

Synonym.—*Micrococcus* of infectious osteomyelitis (Becker).

Observed by Ogston (1881) in the pus of acute abscesses, but not differentiated from the associated staphylococci and the streptococcus of pus. Obtained by Becker from the pus of osteomyelitis (1883). Isolated from the pus of acute abscesses and accurately described by Rosenbach (1884) and by Passet (1885).

The *Staphylococcus pyogenes aureus* is a facultative parasite, and is the most common pyogenic micrococcus found in suppurative processes generally. But it is also a common and widely distributed saprophyte, which finds the conditions necessary for its existence on the external surface of the human body and of moist mucous membranes. This is shown by the researches of numerous bacteriologists. Thus Ullmann found it upon the skin and in the secretions of the mouth of healthy persons, and also in the dust of occupied apartments, in water, etc.; Bockhart obtained it in cultures from the surface of the body and from the dirt beneath the finger nails of healthy persons; Biondi, Vignal, and others in the salivary secretions; B. Fränkel in mucus from the pharynx; Von Besser and Wright in nasal mucus; Escherich in the alvine discharges of healthy infants; C. Fränkel in the air; and Lübbert in the soil. Its presence in the air, in water, or in the soil is, however, quite exceptional, and is probably to be considered the result of accident, its normal habitat as a saprophyte appearing to be rather upon the surface of the body and of mucous membranes.

Morphology.—Spherical cells having a diameter of $0.7\ \mu$ (Hadelich) to $0.9\ \mu$ ($0.87\ \mu$ Passet), solitary, in pairs, or in irregular groups, occasionally in chains of three or four elements or in groups



FIG. 79.—*Staphylococcus pyogenes aureus*, from a drawing by Rosenbach.

of four. The dimensions vary somewhat in different culture media, being larger in a favorable than in an unfavorable medium. The individual cells, as pointed out by Hadelich, consist of two hemispherical portions separated from each other by a very narrow cleft, which is not visible when the cells are deeply stained, but may be demonstrated, with a high power, by staining for a short time (two minutes or less) in a solution of fuchsin in aniline water.

This micrococcus *stains* quickly in aqueous solutions of the basic aniline colors, and may also be stained with acid carmine and hæmatoxylin. It is not decolorized by iodine solution when stained with methyl violet—Gram's method.

Biological Characters.—*Staphylococcus pyogenes aureus* grows either in the presence or absence of oxygen, and is consequently *facultative anaërobic*. It multiplies rapidly at a temperature of 20°C . in milk, flesh infusions, and various other liquid media, and in nutrient gelatin or agar. It *liquefies gelatin*, and in staphylococcal cultures liquefaction occurs all along the line of puncture, forming a pouch which is largest above and at the end of three or four days has extended to the full capacity of the test tube at the surface. The liquefied gelatin in this pouch is at first opaque from the presence of little agglomerations of micrococci in suspension, but after a time these are deposited and the gelatin becomes transparent. During the period of active growth the cocci accumulate near the surface of the gelatin, and, in contact with the air, the characteristic *golden-yellow pigment* is produced. By the subsidence of the colored mass of cocci from this superficial stratum a yellow deposit is gradually formed at the bottom of the pouch of liquefied gelatin (Fig. 80). This pigment, which is the principal character distinguishing the micrococcus under consideration from certain other liquefying staphylococci, is only formed in the presence of oxygen. Upon the surface of nutrient agar development occurs in the form of a moist, shining layer, with more or less wavy outlines, having at first a pale-yellow color, which soon deepens to an orange- or golden-yellow. The colonies which develop upon agar plates are spherical and opaque, and usually acquire the golden-yellow color within a few days. Colonies on gelatin plates or in Esmarch roll tubes first appear as small white dots, which later are more or less granular in appearance and present the yellow color, especially towards the centre; but, owing to the extensive liquefaction of the gelatin caused by them, their develop-

ment can only be followed for two or three days. Upon potato, at a temperature of 35° to 37° C., a rather thick, moist layer of considerable extent forms at the end of twenty-four to forty-eight hours; this is also at first of a pale-yellow, and later of an orange-yellow color. The temperature mentioned is most favorable for the rapid development of this micrococcus, although multiplication may occur at a comparatively low temperature and is tolerably abundant at the ordinary room temperature.

Cultures of the "golden staphylococcus," and especially those upon potato, give off a peculiar odor which resembles that of sour paste. When cultivated in milk it gives rise to the formation of lactic and butyric acids and to coagulation of the casein. No poisonous ptomaines or toxalbumins have been isolated from cultures of this micrococcus, but, like other liquefying bacteria, it forms a soluble peptonizing ferment, by which gelatin may be liquefied independently of the living microorganism. While the *Staphylococcus aureus* gives rise to the production of acids—principally lactic acid—in media containing glucose or lactose, it has also been shown by Brieger that ammonia is one of the products of its vital activity. Unlike some other pathogenic bacteria, it is able to grow in a medium having a distinctly acid reaction. A non-poisonous basic substance has been isolated by Brieger from old cultures in meat infusion which differs from any of the ptomaines obtained by him from other sources.

The thermal death-point of this micrococcus, in recent cultures in flesh-peptone-gelatin, as determined by the writer, is between 56° and 58° C., the time of exposure being ten minutes. When in a desiccated condition a much higher temperature is required— 90° to 100° C.—for its destruction; and it retains its vitality for more than ten days when dried upon a cover glass (Passet). It retains its vitality for a long time in cultures in nutrient gelatin or agar, and may grow when transplanted from such cultures even at the end of a year.

Very numerous experiments have been made to determine the proportion of various chemical agents required to destroy the vitality or to restrain the growth of this important pyogenic micrococcus. The extended researches of Lübbert (1886) with reference to the antiseptic power of agents added to a suitable culture medium—nutrient gelatin—gave the following results: Development was pre-



FIG. 80.—Gelatin culture of *Staphylococcus pyogenes aureus* (Baumgarten).

vented by the agents named in the proportion given : Nitric acid, 1 : 797 ; phosphoric acid, 1 : 750 ; boracic acid, 1 : 327 ; oxalic acid, 1 : 433 ; acetic acid, 1 : 720 ; citric acid, 1 : 433 ; lactic acid, 1 : 350 ; benzoic acid, 1 : 400 ; salicylic acid, 1 : 655 ; iodine dissolved with potassium iodide, 1 : 1,100 ; arsenite of potash, 1 : 733 ; mercuric chloride, 1 : 81,400 ; chloral hydrate, 1 : 133 ; carbolic acid, 1 : 814 ; thymol, 1 : 11,000 ; resorcin, 1 : 122 ; hydrochinon, 1 : 353 ; kairin, 1 : 407 ; antipyrin, 1 : 26 ; muriate of quinine, 1 : 550 ; muriate of morphia, 1 : 60. For the destruction of vitality very much larger amounts are required. In Bolton's experiments (1887) a one-per-cent solution of carbolic acid was successful after two hours' exposure, but two per cent failed to completely destroy vitality in the same time ; one per cent of sulphate of copper was also successful, and but a single colony developed after exposure to a solution of 1 : 200. In the experiments of Gärtner and Plagge the *Staphylococcus aureus* in bouillon cultures is said to have been killed in a few seconds (eight) by a solution of mercuric chloride of the proportion of 1 : 1,000 ; Behring found it was killed by the acid sublimate solution of La Place, in the proportion of 1 : 1,000, in ten minutes ; Tarnier and Vignal found that a solution of 1 : 1,000 was successful in two minutes. Abbott (1891) has shown that in the same culture there may be a considerable difference in the resisting power of the cocci, and that while frequently all are destroyed in five minutes by a 1 : 1,000 solution, it occurs quite as frequently that some may survive after an exposure of ten, twenty, and even thirty minutes.

Pathogenesis.—Subcutaneous inoculation with a small quantity of a culture of *Staphylococcus pyogenes aureus* is without result in rabbits, guinea-pigs, or mice, but when a considerable quantity is injected beneath the skin of a rabbit or a guinea-pig an abscess is produced, which usually results in recovery, but may give rise to general infection and the death of the animal. Injection into a vein or into the cavity of the abdomen in the animals mentioned usually induces a fatal result within a few days. The most characteristic pathological changes are found in the kidneys, which contain numerous small collections of pus and under the microscope present the appearances resulting from embolic nephritis. Many of the capillaries and some of the smaller arteries of the cortex are plugged up with thrombi consisting of micrococci. Metastatic abscesses may also be found in the joints and muscles. The micrococci may be recovered in pure cultures from the blood and the various organs ; but they are not numerous in the blood, and a simple microscopical examination will often fail to demonstrate their presence.

Animals frequently survive the injection of a small quantity of

a pure culture made directly into the circulation, and there is evidence that the pathogenic potency of this micrococcus may vary considerably as a result of conditions relating to its origin and cultivation in the animal body or in artificial media. When injected in considerable quantities it may be obtained in cultures from the urine, but not sooner than six or eight hours after the injection, and not until the formation of purulent foci in the kidneys has already occurred (Wyssokowitsch).

The pyogenic properties of this micrococcus have been demonstrated upon man by the experiments of Garré, of Bockhart, and of Bumm. The first-named observer inoculated a small wound at the edge of one of his finger nails with a minute quantity of a pure culture, and a subepidermal, purulent inflammation extending around

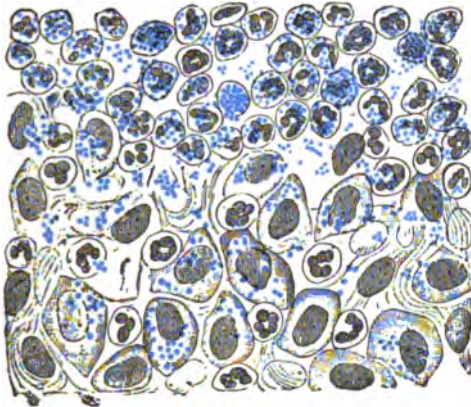


FIG. 81.—Vertical section through a subcutaneous abscess caused by inoculation with staphylococci, in the rabbit, forty-eight hours after infection; margin towards the normal tissue. $\times 950$. (Baumgarten.)

the margin of the nail resulted from the inoculation. *Staphylococcus aureus* was recovered in cultures from the pus thus formed. A more extensive and extremely satisfactory experiment was subsequently made by Garré, who applied a considerable quantity of a pure culture obtained from the above-mentioned source—third generation—to the uninjured skin of his left forearm. At the end of four days a large carbuncle, surrounded by isolated furuncles, developed at the point where the culture had been applied. This ran the usual course, and it was several weeks before it had completely healed. No less than seventeen scars remained to give evidence of the success of the experiment.

In Bockhart's experiments a similar but milder result was obtained, the conditions having been somewhat different. A small

quantity of an agar culture was suspended in 0.5-per-cent salt solution, and this was rubbed upon the uninjured skin of the left arm. By gentle scratching with a disinfected finger nail the epiderm was removed in places over the area to which the micrococcus had been applied. As a result of this procedure numerous impetiginous pustules and occasionally a genuine furuncle developed. Portions of the skin containing the smaller pustules were excised and examined microscopically. As a result of this examination Bumm concluded that the cocci penetrate by way of the hair follicles, the sebaceous and sudoriparous glands, or, where the epidermis had been removed by scratching, directly to the deeper layers of the skin.

In Bumm's experiments, made upon himself and several other persons, *Staphylococcus aureus* suspended in sterilized salt solution was injected beneath the skin. An abscess resulted in every case.

The very extended researches made by bacteriologists during the past five or six years show that the golden staphylococcus is the most common pyogenic microorganism. Its presence has been demonstrated not only in furuncles and carbuncles, but also in various pustular affections of the skin and mucous membranes—impetigo, sycosis, phlyctenular conjunctivitis; in purulent conjunctivitis, inflammation of the lacrymal sac; in acute abscesses formed in the lymphatic glands, the parotid gland, the tonsils, the mammae, and in metastatic abscesses and purulent collections in the joints; in pyæmia; in infectious osteomyelitis; and in ulcerative endocarditis. The evidence relating to its presence and etiological import in the last-mentioned affections demands special consideration.

Infectious osteomyelitis appears from the researches of Becker, Rosenbach, Krause, Passet, and others, to be usually due to the presence of *Staphylococcus aureus*, although Kraske has shown that in certain cases this is associated with other microorganisms. Becker, who obtained this micrococcus from the pus of osteomyelitis in a human, was the first to show by experiment that the same affection might be induced in rabbits by injecting cultures of the micrococcus into the circulation, after having crushed or fractured a bone in one of the legs. The animal usually died in from twelve to fourteen days and presented the usual appearances of osteomyelitis at the fracture point. The abundant yellowish-white pus contained the golden staphylococcus which was described by Becker, and subsequently known in the bacteriological laboratories of Germany as the "micrococcus of infectious osteomyelitis." Becker's experimental results have been confirmed by Krause and Rosenbach; and Rodet, by injecting smaller quantities of a culture into the circulation, has succeeded in producing an osteomyelitis without previous injury to the bone.

Ulcerative endocarditis has been shown by the researches of numerous bacteriologists to be occasionally accompanied by a mycotic invasion of the affected tissues by the golden staphylococcus; in other cases *Streptococcus pyogenes* is present. The researches of Weichselbaum, and of E. Fränkel and Sängér, also show that it is present in a certain proportion of the cases, at least, of endocarditis verrucosa, although in smaller numbers. That the diseased condition of the cardiac valves in ulcerative endocarditis is due to mycotic invasion is now generally admitted and is supported by experimental evidence. Rosenbach first (1873) produced an endocarditis in lower animals by mechanical injury to the cardiac valves, effected by introducing a sound through the aorta. Following his method, Wyszokowitsch (1885), after injuring the cardiac valves in rabbits, injected into the circulation pure cultures of various bacteria. He obtained positive results with *Staphylococcus aureus* and *Streptococcus pyogenes* only. When these micrococci were injected into the trachea or subcutaneously the result was negative, as was the case when very few cocci were injected into a vein, or when two days or more were allowed to elapse after injury to the cardiac valves. Subsequently Weichselbaum, Prudden, and Fränkel and Sängér obtained confirmatory results, thus establishing the fact that when the valves are first injured mechanically (or chemically—Prudden) the injection into a vein of a pure culture of *Staphylococcus aureus* gives rise to a genuine ulcerative endocarditis. It has been further shown by Ribbert that the same result may be obtained without previous injury to the valves by injecting into a vein the staphylococcus from a potato culture suspended in water. In his experiments not only the micrococci from the surface but the superficial layer of the potato was scraped off with a sterilized knife and mixed with distilled water; and the successful result is ascribed to the fact that the little agglomerations of micrococci and infected fragments of potato attach themselves to the margins of the valves more readily than isolated cocci would do. In these experiments the mitral and tricuspid valves were affected, while the semilunar valves remained intact. In ulcerative endocarditis it is evident that cocci detached from the diseased valves must find their way into the circulation. As a matter of fact, masses of micrococci are carried away by the blood stream and form emboli in various parts of the body, which become secondary foci of infection and give rise to local necrotic changes and accumulations of pus. While this undoubtedly occurs, it is generally admitted that the mycotic infection of the cardiac valves is usually a secondary affection, resulting from the transportation of micrococci in the blood current from some other infected focus. But there is no general development of micrococci in the cir-

culating fluid, and in man, as in animals infected experimentally, microscopic examination of the blood for microorganisms usually gives a negative result. Culture experiments may, however, demonstrate their presence. Thus recent investigations by Netter, Eberberg, and others show that the pus cocci are usually present in blood in small numbers, as demonstrated by culture experiments in septic infection from wounds.

2. STAPHYLOCOCCUS PYOGENES ALBUS.

Isolated by Rosenbach (1884) from the pus of acute abscesses, which it is sometimes the only microorganism present, and sometimes associated with other pus cocci. In thirty-three acute abscesses examined by Passet (1885) it was associated with *Staphylococcus aureus* in eleven, with *Staphylococcus citreus* in two, with *Streptococcus pyogenes* in one, with both *Staphylococcus citreus* and *Streptococcus pyogenes* in one, and was obtained alone from four.

In its *morphology* this micrococcus is identical with the previous one, but it is distinguished from it by the absence of pigment, and by being somewhat less pathogenic. Surface cultures upon nutrient agar or potato have a milk-white color. It liquefies gelatin in the same way as does the golden staphylococcus, but the deposit at the bottom of the liquefied gelatin is without color. In the temperature conditions favorable to its growth, and in its biological characteristics generally, with the exceptions noted, it is not to be distinguished from the species previously described. According to Flügge, it is more common than *aureus* among many of the lower animals.

Pathogenesis.—Fortunati has tested the comparative pathogenic power of *Staphylococcus aureus* and *Staphylococcus albus* by inoculations into the cornea of rabbits. A purulent infiltration of the cornea and panophthalmitis resulted when *Staphylococcus aureus* was inoculated upon the surface of the cornea by scratching with an infected needle, but inoculations made in the same way with *Staphylococcus albus* healed spontaneously or gave rise to a perforating ulcer. After paracentesis of the cornea with an instrument infected with *Staphylococcus aureus* panophthalmitis developed in thirty hours; the same result occurred at the end of sixty to seventy-two hours when the instrument was infected with *Staphylococcus albus*. When a sterilized instrument was used the result was negative. In bacteriological researches made by Gallenga, in cases of panophthalmitis in man, *Staphylococcus albus* was found in ten cultures and *Staphylococcus aureus* in nine.

Staphylococcus Epidermidis Albus (Welch).

The recently published researches of Welch show that a white staphylococcus, probably identical with *Staphylococcus pyogenes*

albus of Rosenbach, is the most common microorganism upon the surface of the body, and that "it is very often present in parts of the epidermis deeper than can be reached by any known means of cutaneous disinfection save the application of heat." With reference to this coccus Welch says :

"So far as our observations extend—and already they amount to a large number—this coccus may be regarded as a nearly, if not quite, constant inhabitant of the epidermis. It is now clear why I have proposed to call it the *Staphylococcus epidermidis albus*. It possesses such feeble pyogenic capacity, as is shown by its behavior in wounds as well as by experiments on rabbits, that the designation *Staphylococcus pyogenes albus* does not seem appropriate. Still, I am not inclined to insist too much upon this point, as very probably this coccus, which has hitherto been unquestionably identified by Bossowski and others with the ordinary *Staphylococcus pyogenes albus* of Rosenbach, is an attenuated or modified form of the latter organism, although, as already mentioned, it presents some points of difference from the classical description of the white pyogenic coccus."

According to Welch, this coccus differs from *Staphylococcus pyogenes aureus* not only in color, but also in the fact that it liquefies gelatin more slowly, does not so quickly cause coagulation of milk, and is far less virulent when injected into the circulation of rabbits. It has been shown by the researches of Bossowski and of Welch that this coccus is very frequently present in aseptic wounds, and that usually it does not materially interfere with the healing of wounds, although sometimes it appears to cause suppuration along the drainage tube, and it is the usual cause of "stitch abscess." Bossowski, in fifty cases of wounds treated antiseptically, obtained bacteria from the discharges in forty, and in twenty-six of these cases he found *Staphylococcus pyogenes albus*; *Staphylococcus aureus* was found nine times, *Streptococcus pyogenes* in two, and various non-pathogenic bacteria in eight. In forty-five laparotomy wounds examined by Ghrisky and Robb, in which strict antiseptic precautions had been observed, bacteria were found in thirty-one, and in nineteen of this number *Staphylococcus albus* was present. *Staphylococcus aureus* in five, *Bacillus coli communis* in six, and *Streptococcus pyogenes* in three.

3. STAPHYLOCOCCUS PYOGENES CITREUS.

Isolated by Passet (1885) from the pus of acute abscesses. In thirty-three cases examined it was found associated with *Staphylococcus albus* in two and with *Staphylococcus albus* and *Streptococcus pyogenes* in one.

In its *morphology* this coccus is identical with the two preceding species, from which it is distinguished by the formation of a lemon-yellow pigment, instead of a golden or orange-yellow as in *Staphylococcus aureus*. The pigment is only formed in the presence of oxygen. This coccus is said by Fränkel to liquefy gelatin more slowly than the previously described species—*Staphylococcus aureus* and *Staphylococcus albus*.

As to its *pathogenic properties* we have no definite information. It is included among the pyogenic bacteria because of its occasional presence in

the pus of acute abscesses, although it has heretofore only been found in association with other microorganisms.

4. MICROCOCCUS PYOGENES TENUIS.

Obtained by Rosenbach (1884) from pus in three cases out of thirty examined.

Morphology.—Micrococci, somewhat irregular in size, but larger than *Staphylococcus albus*, and seldom associated in masses. Frequently the individual cocci present the appearance of consisting of two deeply stained masses separated from each other by a paler interspace. Cultures upon surface of nutrient agar form a very thin, transparent layer of about 1 millimetre in breadth along the line of inoculation; this resembles a layer of varnish.

Pathogenesis undetermined.

5. STREPTOCOCCUS PYOGENES.

Synonyms.—*Micrococcus* of erysipelas (Fehleisen); *Streptococcus erysipelatos*; *Streptococcus* of pus; *Streptococcus longus* (Von Lingelsheim).

Obtained by Fehleisen from the skin involved in cases of erysipelas (1883), and by Rosenbach (1884) and Passet (1885) from the pus of acute abscesses. The characters of the "*streptococcus* of erysipelas" of Fehleisen and the "*Streptococcus pyogenes*" of Rosenbach and Passet are generally admitted to be identical, although some bacteriologists still describe them separately and cultures from the two sources are still retained in bacteriological laboratories under the names originally given them.

Rosenbach found *Streptococcus pyogenes* alone in fifteen cases and associated with staphylococci in five cases, out of thirty-three cases examined of acute pus formation. Passet, in thirty-three similar cases, obtained the streptococcus alone in eight and associated with staphylococci in two. Subsequent researches show that the micrococcus is frequently, if not constantly, present in puerperal metritis; that it is the most frequent microorganism associated with ulcerative endocarditis; that it is frequently present in diphtheritic false membranes, and especially in those cases of diphtheritic inflammation which are secondary to scarlet fever and measles (Prudden). Numerous investigations made by bacteriologists during the past twenty years indicate that this is a very important and widely distributed pathogenic microorganism. It has also been frequently found upon exposed mucous surfaces—mouth, nose, vagina—of healthy individuals.

According to the recent researches (1891) of Von Lingelsheim, *Streptococcus pyogenes* differs from *Streptococcus erysipelatos* in being pathogenic both for mice and rabbits, while the latter is pathogenic for rabbits only. The author named, as a result of extended

carefully conducted comparative studies, arrives at the following conclusions :

"According to my observations, there are two great groups among the streptococci. These cannot be distinguished one from the other in cultures in highly albuminous media (pus, blood serum), but present constant differences when cultivated in bouillon. The decisive characteristics in this medium are : macroscopic, the cloudiness of the medium ; microscopic, the length of the chains. The two groups are with difficulty distinguished in agar cultures ; more easily in gelatin, in which the streptococcus which forms short chains causes a slight liquefaction, while the *Streptococcus longus* does not. Upon potato *Streptococcus brevis* alone shows a visible growth. . . . We see here a group of streptococci which we separate from the others, because of their microscopic and cultural differences, under the name of *Streptococcus brevis*, which is also distinguished by having no pathogenic action upon the animals usually experimented upon. We recognize, on the other hand, the streptococci which we have grouped together as *Streptococcus longus* as all pathogenic and about in equal degree for a certain species of animal (rabbits) ; but by experiments upon other species (mice) we arrive at the conclusion that there must also be differences between these streptococci. It appears that the streptococci which are distinguished by their high degree of pathogenic power upon mice are also those which are distinguished in bouillon cultures by the formation of conglomerate masses. We find among these also one which is distinguished by especial virulence for mice, and that this one is distinguished in cultures by its scanty growth upon ox serum."

Von Lingelsheim gives the following classification of the streptococci :

STREPTOCOCCI.

Not pathogenic.	Pathogenic.
<i>Streptococcus brevis</i> .	<i>Streptococcus longus</i> .
Pathogenic for mice and rabbits. (a) <i>Streptococcus murisepticus</i> . (b) <i>Streptococcus pyogenes</i> .	Pathogenic for rabbits. <i>Streptococcus erysipalatos</i> .

Morphology.—Spherical cocci, from $0.4\ \mu$ to $1\ \mu$ in diameter, but varying considerably in dimensions in different cultures, and even in a single chain. Multiply by binary division, in one direction only, forming chains, in which the elements are commonly associated in pairs. Under certain circumstances, instead of forming chains, a culture may contain only, or chiefly, diplococci ; but usually chains containing from four to twenty or more elements are formed, and these are frequently associated in tangled masses. Occasionally one or more cells in a chain greatly exceed their fellows in size, and some bacteriologists suppose that these cells serve as reproductive spores—arthrospores—but this has not been definitely proven.

Stains readily with the aniline colors and by Gram's method.



FIG. 82.—Pus containing streptococci. $\times 800$. (Flügge.)

Biological Characters.—Grows readily in various liquid and solid culture media, including all of those usually employed in bacteriological researches. The most favorable temperature for its development is from 30° to 37° C., but it multiplies freely at the ordinary room temperature—16° to 18° C.

Streptococcus pyogenes is a *facultative anaërobic*, grows both in the presence and absence of oxygen.



FIG. 88.—*Streptococcus* of erysipelas in nutrient gelatin; stick culture at end of four days at 16°–18° C. (Baumgarten).

does not liquefy gelatin, and in gelatin stick cultures it grows along the line of puncture, forming numerous small, spherical, transparent, whitish colonies, which are closely crowded together at the upper portion of the line of growth, and often distinctly separated from each other below; upon the surface there is often no growth, or a scanty development may occur about the point of entrance of the inoculating needle. The minute colonies along the line of puncture are already visible at the end of twenty-four hours in cultures kept in an incubating oven at 30° to 35° C., and at the end of three or four days they have reached their full development, forming a semi-opaque, whitish granular column, upon the margins of which the separate colonies are seen projecting into the gelatin. On gelatin plates very small, transparent colonies are developed, which upon the surface spread out to form a flat, transparent disc of about one-half millimetre. Under a low magnifying power these colonies are seen to be slightly granular and have a yellowish color. At a later date they become darker and less transparent,

and the margin may show irregular projections made up of tangled masses of cocci in chains. The characters of growth in nutrient agar and in jellified blood serum are similar to those in gelatin, and on agar plates colonies are formed similar to those above described, except that they are somewhat smaller and more transparent. Fehleisen and De Simone state that the erysipelas coccus may develop upon the surface of cooked potato, but most authorities—Flügge, C. Fränkel, Passet, Baumgarten—agree that no growth occurs upon potato. Milk is a favorable medium for the growth of this micrococcus, and the casein is coagulated by it. A slightly acid reaction of the culture medium does not prevent its development. The *thermal death-point*, as determined by the writer, is between 52° and 54° C., the time of exposure being ten minutes. According

to De Simone, a temperature of 39.5° to 41° C. maintained for two days is fatal to this micrococcus.

Manfredi and Traversa have injected filtered cultures into frogs, guinea-pigs, and rabbits for the purpose of ascertaining if any soluble toxic substance is produced during the growth of *Streptococcus pyogenes*. They report that in some cases convulsions and in others paralysis resulted from these injections.

Von Lingelsheim has recently (1891) reported the following results obtained in an extended series of experiments made to determine the germicidal power of various chemical agents as tested upon this microorganism—time of exposure two hours: Hydrochloric acid 1:250, sulphuric acid 1:250, caustic soda 1:130, ammonia 1:25, mercuric chloride 1:2,500, sulphate of copper 1:200, chloride of iron 1:500, terchloride of iodine 1:750, peroxide of hydrogen 1:50, carboic acid 1:300, cresol 1:250, lysol 1:300, creolin 1:130, naphthylamin 1:125, malachite green 1:3,000, pyoktanin 1:700.



FIG. 84.—Section from margin of an erysipelatos inflammation, showing streptococci in lymph spaces. From a photograph by Koch. $\times 900$.

Pathogenesis.—When inoculated into the cornea of rabbits *Streptococcus pyogenes* gives rise to keratitis. Inoculations into the ear of the same animal usually give rise to a localized erysipelatos inflammation accompanied by an elevation of temperature in the inoculated ear; at the end of thirty-six to forty-eight hours the inflamed area, which has well-defined margins and a bright-red color, extends from the point of inoculation along the course of the veins to the root of the ear. This appearance passes away in the course of a few days and the animal recovers. Subcutaneous injections into mice or rabbits are usually without result, and the last-named animal also withstands injections of considerable quantities into the general circulation through a vein. When, however, the animal has previously been weakened by the injection of toxic substances the streptococcus may multiply in its body and cause its death (Flügge).

Fehleisen has inoculated cultures, obtained in the first instance from the skin of patients with erysipelas, into patients in hospital suffering from lupus and carcinoma, and has obtained positive results, a typical erysipelatos inflammation having developed

around the point of inoculation after a period of incubation of fifteen to sixty hours. This was attended with chilly sensations and an elevation of temperature. Persons who had recently recovered from an attack of erysipelas proved to be immune.

Sections made from the ear of an inoculated rabbit, or of skin taken from the affected area in erysipelas in man, show the streptococcus in considerable numbers in the lymph channels, but not in the blood vessels. They are more numerous, according to Koch and to Kiehn, upon the margins of the erysipelatous area, and may even be seen in the lymph channels a little beyond the red margin which marks the line of progress of the infection.

The researches of Weichselbaum and others show that *Streptococcus pyogenes* is the infecting microorganism in a certain proportion of the cases of *ulcerative endocarditis*. The author has found it in four cases out of fifteen examined, and in two cases of *endocarditis verrucosa* out of thirteen. In a previously reported series of sixteen cases (fourteen of ulcerative endocarditis and two of *verrucosa*) the streptococcus was found in six.

In *diphtheritic false membranes* this streptococcus is very commonly present, and in certain cases attended with a diphtheritic infection, in which the *Bacillus diphtheriæ* has not been found by competent bacteriologists, it seems probable that *Streptococcus pyogenes* is the pathogenic microorganism responsible for the local inflammation and its results. Thus in a series of twenty-four cases studied by Prudden in 1889 the bacillus of Löffler was not found, "but a streptococcus apparently identical with *Streptococcus pyogenes* was found in twenty-two." Chantemesse and Widal have also reported cases in which a fibrinous exudate resembling that of diphtheria was associated with a streptococcus. "These forms of so-called diphtheria are most commonly associated with scarlatina and measles, erysipelas, and phlegmonous inflammation, or occur in individuals exposed to these diseases; but whether exclusively under these conditions has not yet established" (Prudden).

Löffler has described under the name of *Streptococcus artabrorum* a micrococcus obtained by him from the affected mucous membrane in cases of diphtheria, and which he believes to be accidentally present and without any etiological import in this disease. In its characters it closely resembles *Streptococcus pyogenes* and perhaps a variety of this widely distributed species. Its characters are described by Flügge as follows:

"Cultivated in nutrient gelatin, it forms at the end of three days small transparent, light-gray drops, upon the margin of which, under the microscope, the cocci in twisted chains may be observed. As many as one

dred elements may be found in a single chain, and some of these are distinguished by their size; occasionally whole chains are made up of these large cocci, and when closely observed some of these may present indications of division transversely to the axis of the chain. Subcutaneous inoculation of cultures into mice results in the death of a considerable number of these animals—more than half; and the streptococci are found in the spleen and other organs. Inoculation into the ear of rabbits causes an erysipelatous inflammation. When injected into the circulation of these animals through a vein joint affections are developed in from four to six days, and a purulent accumulation occurs in which the streptococci are found. In two rabbits inoculated in the same way with a culture of the streptococcus of erysipelas, Löffler has observed a similar result."

Recent researches indicate that infection by *Streptococcus pyogenes* through the endometrium is the usual cause of *puerperal fever*. Thus Clivio and Monti demonstrated its presence in five cases of puerperal peritonitis. Czerniewski found it in the lochia of a large number (thirty-five out of eighty-one) of women suffering from puerperal fever, but in the lochia of fifty-seven healthy puerperal women he was only able to find it once. In ten fatal cases he found it in every instance, both in the lochial discharge during life and in the organs after death. Widal carefully studied a series of sixteen cases and arrived at the conclusion that this was the infecting microorganism in all. Bumm and other observers have given similar evidence. Eiselsberg and Emmerich have succeeded in demonstrating the presence of the streptococcus in hospital wards containing cases of erysipelas. That puerperal fever may result from infection through the finger of the accoucheur, when he has previously been in contact with cases of erysipelas, has long been taught, and, in view of the facts above recorded, is not difficult to understand. But in view of the fact that the streptococcus of pus has been found in vaginal mucus and in the buccal and nasal secretions of healthy persons, it may appear strange that cases of puerperal fever not traceable to infection from erysipelas or from preceding cases do not occur more frequently. This is probably largely due to an attenuation of the pathogenic power of the streptococcus when it leads a saprophytic existence. Widal asserts that, when cultivated in artificial media for a few weeks, the cultures no longer have their original virulence, and Bumm has made the same observation. On the other hand, in "*streptococcus-peritonitis*" occurring as a result of puerperal infection Bumm states that the thin, bright-yellow, odorless fluid contained in the cavity of the abdomen is extremely virulent; a very slight trace, a fragment of a drop, injected into the abdominal cavity of a rabbit, is sufficient within twenty-four hours to cause a general septic inflammation with a bloody serous exudation, quickly terminating in the death of the animal; injected subcutaneously it gives rise to an enormous phlegmon which also

quickly proves fatal. But cultures of *Streptococcus pyogenes*, and it has been carried through successive generations in artificial media injected beneath the skin of a rabbit, usually produce no result, at most an abscess of moderate dimensions.

It seems probable that the micrococcus isolated by Flügge from necrotic foci in the spleen of a case of leucocythæmia, and described by him under the name of *Streptococcus pyogenes malignus*, was simply a very pathogenic variety of the streptococcus of pus. He was not able to differentiate it from *Streptococcus pyogenes* by its morphology or growth in culture media, but it proved far more pathogenic when tested upon animals. Mice inoculated subcutaneously with a minute quantity of a pure culture died, without excitation, in three to five days. A large abscess was formed at the point of inoculation, and the blood of the animal contained numerous cocci in pairs and chains. Rabbits inoculated in the ear showed at first the same local appearances as result from inoculations with streptococcus of pus and of erysipelas, but after two or three days symptoms of general infection were developed, and death occurred at the end of three or four days. At the autopsy the cocci were found in the blood, and frequently there were purulent collections in the joints containing the same microorganism. Krause has also described a streptococcus which only differs from *Streptococcus pyogenes* of Rosenbach and Passet by the greater virulence manifested by its cultures.

The fact that pathogenic bacteria may attain an intensified degree of virulence by cultivation in the bodies of susceptible animals was demonstrated by Davaine many years ago, and is fully established by the experiments of Pasteur and others. It is true of the anthrax bacillus, of the writer's *Micrococcus Pasteuri*, and of other well-known pathogenic microorganisms. The reverse of this—attenuation of virulence as a result of cultivation in artificial media—is also well established for several pathogenic species. Now it appears that the attenuated streptococcus is far less likely to give rise to erysipelas or to puerperal infection than is the same microorganism as obtained from a case of one or the other of these infectious diseases. The same is probably true also of *Staphylococcus aureus* and other facultative parasites which are found as saprophytes upon the surface of the body and upon exposed mucous membranes in healthy persons. And it is not improbable that attenuated varieties of these micrococci which find their way into open wounds or into the uterine cavity shortly after parturition, if they escape destruction by the sanguineous discharge, acquire increased pathogenic power from their multiplication in it, as a result of which they are able to invade the living tissues. But it appears probable that

infection through open wounds does not depend alone upon the potency of the pathogenic micrococci present in them, but also upon the absorption of chemical poisons produced by septic (putrefactive) bacteria, which weaken the vital resisting power of the tissues. Gottstein, as a result of experiments made by him, is of the opinion that the resorption of broken-down red blood corpuscles favors infection by pathogenic bacteria present in wounds; and he has shown that the injection into animals of certain toxic substances which destroy the red corpuscles in the circulation makes them susceptible to the pathogenic action of certain bacteria which are harmless for them under ordinary circumstances. Thus a guinea-pig, an animal which is immune against the bacillus of fowl cholera, succumbed to an inoculation made after first injecting subcutaneously 0.06 gramme of hydracetic acid dissolved in alcohol. At the autopsy hæmorrhagic exudations were found in the serous cavities, hæmorrhagic infarctions in the lungs, and quantities of the bacillus injected were found in the blood and in fluid from the cavity of the abdomen.

In man the ever-present pus cocci are more likely to invade the tissues, forming furuncles, carbuncles, and pustular skin eruptions, or erysipelatous and phlegmonous inflammations, when the standard of health is reduced from any cause, and especially when by absorption or retention various toxic organic products are present in the body in excess. It is thus that we would explain the liability to these local infections, as complications or sequelæ of various specific infectious diseases, in the victims of chronic alcoholism, in those exposed to septic emanations from sewers, etc., and probably in many cases from the absorption of toxic products formed in the alimentary canal as a result of the ingestion of improper food, or of abnormal fermentative changes in the contents of the intestine, or from constipation.

The Pus Cocci in Inflammations of Mucous Membranes.—To what extent the pus cocci are responsible for inducing and maintaining non-specific inflammations of mucous membranes has not been determined; but having demonstrated the pyogenic properties of these cocci, their presence in the purulent discharges from inflamed mucous membranes can scarcely be considered as unimportant, notwithstanding the fact that they are also frequently found in secretions from healthy mucous surfaces. They are likewise found upon the skin of healthy persons, and yet we have unimpeachable experimental evidence that they may produce a local inflammation, attended with pus formation, when injected subcutaneously, or even when freely applied to the uninjured surface.

In *otitis media* Levy and Schrader obtained *Staphylococcus albus* in pure cultures in three cases out of ten in which paracentesis was performed, and in two others it was present in association with

other microorganisms. In eighteen cases of otitis media in children Netter found *Staphylococcus aureus* six times and *Staphylococcus pyogenes* thirteen times. Scheibe, in eleven cases in which perforation had not yet taken place, found *Staphylococcus aureus* two and various other microorganisms in the remaining cases. In *Staphylococcus aureus* was not present in any. Habermann found *Staphylococcus aureus* associated with other bacteria in a single case of purulent otitis media. In a series of eight cases occurring as a sequel to influenza Scheibe obtained *Streptococcus pyogenes* in two, "*Streptococcus pneumoniae*" in two, *Staphylococcus aureus* in one, *Streptococcus pyogenes* and *Staphylococcus albus* together in two, and *Streptococcus pyogenes* in association with an undescribed microorganism in one. In all of these cases a slender bacillus was also present. It was shown by microscopical examination, which did not grow in the culture media employed. Bordoni-Uffreduzzi and Grazioplene have tabulated the results obtained by various bacteriologists. They have examined pus obtained through the previously intact tympanic membrane. In thirty-two cases of this character the microorganism most frequently found was *diplococcus pneumoniae* (*Micrococcus pneumoniae crouposæ* of the present writer), which was present in pure culture in thirteen and associated with *Staphylococcus aureus* in one, with *Staphylococcus albus* in one, and with *Streptococcus pyogenes* in one. In the other sixteen cases the pyogenic cocci were present in all but two, in which bacilli were found—*Bacillus anthracis* in one, a non-liquefying bacillus in one. In twenty-seven cases in which the pus was withdrawn from one to thirty days after perforation or spontaneous rupture of the membrane, the pyogenic cocci were present in twenty and *diplococcus pneumoniae* in seven.

In acute *nasal catarrh* Paulsen found *Staphylococcus aureus* in seven cases out of twenty-four examined, and E. Fränkel in twenty-four; but it must be remembered that Von Besser has shown that the micrococcus is frequently present in the secretions from the healthy nasal mucous membrane, and we have experimental evidence that the pus organisms, when introduced into the conjunctival sacs of rabbits (Widmark), do not give rise to catarrhal inflammation. On the other hand, Widmark found that when inoculated into the conjunctival sacs of rabbits an intense *conjunctivitis* resulted, together with keratitis and perforation of the cornea in fifteen per cent of the cases. The same author in his bacteriological researches obtained the pyogenic cocci from the circumscribed abscesses of blepharitis, while in inflammation of the lacrymal sac *Streptococcus pyogenes* was usually present.

Shougolowicz, in the bacteriological examination of twenty-six cases of *trachoma*, found *Staphylococcus albus* in twelve, *Staphylo-*

aureus in nine, *Staphylococcus citreus* in three, and *Staphylococcus cereus albus* in three. These pus organisms were in a number of the cases associated with other well-known saprophytes, and in seven cases a short bacillus not previously described was found. That various bacilli are found in the conjunctival sac of healthy eyes and in different forms of conjunctivitis has been shown by Fick, whose results do not correspond in this respect with those of Gifford, who found almost exclusively micrococci. Whatever may be the final conclusion as to the rôle of the pus cocci heretofore described in the etiology of acute or chronic conjunctivitis, there can be no doubt of the power of the "gonococcus" to induce a virulent inflammation of the conjunctivæ when introduced into healthy eyes.

6. MICROCOCCUS GONORRHOÆÆ

Synonym.—Gonococcus (Neisser).

Discovered by Neisser (1879) in gonorrhœal pus and described by him under the name of "Gonococcus." Cultivated by Bumm (1885), and infective virulence proved by inoculation into man. Constantly present in virulent gonorrhœal discharges, for the most part in the interior of the pus cells or attached to the surface of epithelial cells.

Morphology.—Micrococci, usually joined in pairs or in groups of four, in which the elements are flattened — "biscuit-shaped." The flattened surfaces face each other and are separated, in stained preparations, by an unstained interspace. The diameter of an associated pair of cells varies from 0.8 to 1.6μ in the long diameter—average about 1.25μ —and from 0.6 to 0.8μ in the line of the interspace between the biscuit-shaped elements, which sometimes present a slight concavity of the flattened surfaces. Multiplication occurs alternately in two planes, and as a result of this groups of four are frequently observed. But diplococci are more numerous and are considered as the characteristic mode of grouping. Single, spherical, undivided cells are rarely seen.

It must be remembered that the morphology of this micrococcus as above described does not suffice to distinguish it, for Bumm has shown that "the biscuit form is not at all specific for the gonococcus, but is shared with it by a number of microorganisms, which consist of two hemispherical elements with the flattened surfaces facing each



FIG. 85.—a, gonococci from a pure culture, \times about 1,000; b, gonococci in pus cells and epithelial cell from case of gonorrhœal ophthalmia; c, form and mode of division of gonococci—schematic. (Bumm.)

other and separated by a cleft, and some of these correspond in their morphology, in every detail, with the gonococcus."

Stains quickly with the basic aniline colors, especially with methyl violet, gentian violet, and fuchsin; not so quickly with methylene blue, which is, however, one of the most satisfactory staining agents for demonstrating its presence in pus. Beautiful double-stained preparations may be made from gonorrhœal pus, spread upon a cover glass and "fixed," *secundum artem*, by the use of methylene blue and eosin. Does not stain by Gram's method—*i.e.*, the cocci are decolorized, after having been stained with an aniline color, by being immersed in the iodine solution employed in Gram's method of staining. But this character cannot be depended upon alone for establishing the diagnosis, for Bumm has shown that



FIG. 86.—"Gonococcus" in gonorrhœal pus. From a photomicrograph by Fränkel and Pfeiffer $\times 1,000$.

other diplococci are occasionally found in gonorrhœal pus which do not stain by this method. It serves to distinguish them, however, from the common pus cocci heretofore described—*Staphylococcus aureus*, *Staphylococcus albus*, *Staphylococcus citreus*—which retain their color when treated in the same way. A more trustworthy diagnostic character is that these biscuit-shaped diplococci are found within the pus cells, sometimes one or two pairs only, but more frequently in considerable numbers, and occasionally in such numbers as to completely fill the cell. No similar picture is presented by pus from any other source, with the exception of that from a form of "puerperal cystitis" which Bumm has described. But in this the diplococci contained in the pus cells were to be distinguished by the fact that they retained their color when treated by Gram's method. Owing

to the difficulty of cultivating this micrococcus, and the importance, under certain circumstances, of not making a mistake in its diagnosis, these characters are of exceptional value.

Biological Characters.—The “gonococcus” does not grow in flesh infusions, in nutrient gelatin or agar, but it may be cultivated upon blood serum, and, according to Bumm, grows more readily upon human blood serum than upon that of the lower animals. This he obtained for his experiments from the placenta of a recently delivered woman by passing two ligatures around the cord before separating the child from its placental attachment, and dividing it between them. But even upon blood serum obtained in this way it is not a simple matter to obtain a pure culture. When other micrococci are present, even in small numbers, they take the precedence and overgrow the surface of the culture medium before the gonococcus has made any visible growth. It is therefore necessary to start a culture with pus containing this micrococcus only and in considerable numbers. And the pus should not be spread out in a thin layer, but should be distributed upon the surface in little drops or masses, in which the development commences. A temperature of 30° to 34° C. is most favorable for the development of this micrococcus, and Bumm recommends the transfer to fresh culture material in from eighteen to twenty-four hours. The cultures thrive best in a moist atmosphere, and it is well to place the tubes containing them in a large glass jar partly filled with distilled water and having a tightly fitting cover. The growth under the most favorable conditions is slow, and frequently no development occurs when pus containing numerous gonococci is placed upon blood serum in an incubating oven; or after a slight multiplication development ceases and the cocci undergo degenerative changes and quickly disappear.

Cultures upon the surface of blood serum form a very thin, often scarcely visible layer, with a smooth, moist, shining surface, and by reflected light a grayish-yellow color. The growth at the end of twenty-four hours may extend for a distance of a millimetre along the line of inoculation, but at the end of two or three days no further development occurs and the cocci soon lose their vitality. This micrococcus, then, is *aërobic*. Whether it may also be a facultative anaërobic has not been definitely determined, but it does not grow along the line of puncture when stick cultures are made in blood serum. Its rapid and abundant multiplication in gonorrhœal infection of mucous membranes, and the difficulties attending its cultivation in artificial media, show that the gonococcus is a *strict parasite*.

Lestikow and Löffler, prior to the publication of Bumm's important monograph, had reported successful results in cultivating the gonococcus upon a mixture of blood serum and gelatin. Bockhart

has since recommended a mixture of nutrient agar (two parts), liquefied at a temperature of 50° C., with blood serum (two to three parts) at 20° C. By quickly mixing with this a little pus containing the gonococcus he was able to obtain colonies upon plate culture made by pouring the liquid medium upon sterile glass plates in the usual manner.

Development does not occur below 25° or above 38° C. The writer has shown that a temperature of 60° C. maintained for five minutes destroys the infective virulence of gonorrhœal pus.

Pathogenesis.—That the gonococcus is the cause of the specific inflammation and purulent discharge characteristic of gonorrhœa is now generally admitted upon the experimental evidence obtained by Bumm. Having succeeded in obtaining it in pure cultures from gonorrhœal pus, he made successful inoculations in the healthy urethra in two cases—once with a third culture and once with a culture which had been transferred through twenty successive generations.



FIG. 87.—Gonorrhœal conjunctivitis, second day of sickness; section through the mucous membrane of upper eyelid; invasion of the epithelial layer by gonococci. (Bumm.)

In both cases a typical gonorrhœa developed as a result of the inoculation.

Schrötter and Winkler (1890) report their success in cultivating the gonococcus upon albumin from the egg of the pewee—"Kibitz". In the culture oven at 38° C. a thin, transparent, whitish layer was already visible at the end of six hours and rapidly extended; growth was less abundant at the end of three days, and had entirely ceased by the fifth day. Attempts to cultivate the same microorganism in albumin from hens' eggs gave a negative result.

Aufuso (1891) has cultivated the gonococcus in fluid obtained from the knee joint in a case of chronic synovitis, but failed to cultivate it in the fluid of ascites. A culture of the twelfth generation made upon the culture medium mentioned, solidified by heat, was introduced into the urethra of a healthy man and gave rise to the characteristic attack of gonorrhœa.

The mucous membranes in man which are subject to gonorrhœal infection are those of the urethra, the conjunctiva, the cervix uteri,

and the vagina in children—the vagina in adults is not involved. Inoculations of gonorrhœal pus into the vagina or conjunctival sac of the lower animals—dogs, rabbits, horses, apes—are without result.

The very numerous researches which have been made by competent bacteriologists show that the gonococcus is constantly present in gonorrhœal discharges, and in view of the facts above stated its etiological import appears to be fully established. Bumm has studied the development of blennorrhœa neonatorum, and has shown that soon after infection the presence of gonococci may be demonstrated in the superficial epithelial cells of the mucous membrane and between them; that they soon penetrate to the deeper layers, and that by the end of forty-eight hours the entire epithelial layer is invaded by the diplococci, which penetrate by way of the connecting material—“*Kittsubstanz*”—between the cells. They also multiply in the superficial layers of connective tissue and give rise to an inflammatory reaction, which is shown by an abundant escape of leucocytes from the dilated capillary network. The penetration of the gonococci to the deeper layers of the mucous membrane of the urethra, and even to the corpus cavernosum, was observed by Bockhart in a case studied by him in which death occurred during an acute attack of gonorrhœa. But Bumm concludes from his researches that this is not usual, and that the invasion is commonly limited to the superficial layers of the mucous membrane.

Staphylococcus pyogenes aureus is not infrequently associated with the gonococcus in late gonorrhœal discharges, and the abscesses which occasionally develop as a complication of gonorrhœa, in the prostate, the inguinal glands, or around the urethra, are probably due to its presence, which has been demonstrated in the pus from such abscesses in a number of cases. The same is true of the joint affections and endocarditis which sometimes occur in the course of an attack of gonorrhœa. Although some authors have claimed to find the gonococcus in these so-called metastatic gonorrhœal inflammations, the evidence is not satisfactory, and it seems probable that the *Staphylococcus aureus* is the usual microorganism concerned in these affections.

V.

BACTERIA IN CROUPOUS PNEUMONIA.

THE following account of "The Etiology of Croupous Pneumonia" is from a paper read by the writer at the annual meeting of the Medical Society of the State of New York, at Albany, N. Y. January 6th, 1889:

Acute pneumonia is now generally regarded by the leading clinical authorities and pathologists in this country and in Europe as a specific infectious disease. Green, in his article on "Inflammation of the Lung," Quain's "Dictionary of Medicine," says: "It is maintained by some observers that, like the specific fevers, it is due to a specific cause. Pneumonia whilst differing from these fevers in not being contagious, resembles them in the typical character of its clinical phenomena and, to a less extent, in the local lesion. The changes in the lung occurring in pneumonia cannot be induced by artificial injury of the organ, and it must therefore be assumed that there is something special in the inflammatory process."

This "something special" has been demonstrated by recent research, and it is the object of the present paper to give a historical account of the development of our knowledge with reference to this specific infectious disease, and of the experimental evidence upon which the claim is founded that the microorganism referred to bears an etiological relation to the disease in question.

Evidently, if pneumonia is a specific infectious disease, the microorganism which causes it is widely distributed, and the development of the disease depends rather upon secondary predisposing and exciting causes than upon the accidental introduction of the specific agent.

It cannot be maintained that the disease, as a general rule, is transmitted from individual to individual—i.e., by personal contagion. Clinical experience is entirely opposed to this view, although we have ample evidence that it may occur as an epidemic among individuals who are exposed to the same conditions of environment—as in jails, barracks, etc. Thus at Upsala, Sweden, an epidemic of pneumonia occurred in 1847 in the prison during which sixty-nine of the prisoners were attacked. And again in 1867, during a period of six months (December, 1866, to May, 1867), a similar epidemic was observed in the same prison—sixty-two cases. Other prison epidemics recorded are those at Frankfort in 1875 (seventy-five cases) and in 1876 (ninety-eight cases); at Maringen in 1875 (eighty-three cases) and in 1878 (fifty-eight cases); at the prison of D'Ansberg in 1880 (one hundred and sixty-one cases, with forty-six deaths, in a period of five months).

Again, we have numerous records of village epidemics and of epidemics confined to single houses. In outbreaks of this character, as in epidemics of typhoid fever, of cholera, and of yellow fever, there is a succession of cases occurring at different intervals, but it does not follow that the cases bear any direct relation to each other. On the contrary, everything indicates that, as in the diseases mentioned, in the presence of the in-

agent common predisposing causes relating to the environment, acting upon persons having various degrees of resisting power, induce attacks at various intervals; or it may be that in the presence of the specific cause and predisposing influences an exciting cause, such as exposure to cold, alcoholic excess, etc., is the immediate factor in the development of an attack.

Without stopping to discuss further the facts relating to the epidemic prevalence of the disease under consideration, I call attention to the well-established fact that pneumonia prevails over a wide area of the inhabited surface of the earth, and that by far the larger number of cases occur independently of any recognized connection with previous cases, and often under circumstances in which such connection can be very positively excluded. And, on the other hand, the direct transmission of the disease by the sick to those most closely associated with them, as nurses, etc., if it occurs at all, is evidently a rare exception to the general rule.

We must then conclude, as stated at the outset, that if pneumonia is a specific infectious disease the microorganism which causes it is widely distributed. As a matter of fact, the pathogenic micrococcus which, from the evidence now at hand, appears to be the specific etiological agent in acute pneumonia has been found in the buccal secretions of healthy individuals in various parts of the world—in America, in France, in Italy, and in Germany, and no doubt more extended researches will show that it is extremely common.

This statement may appear at the outset to make the view that the micrococcus in question is the cause of croupous pneumonia quite untenable. For, it may be asked, how is it that the individuals who have this microorganism in their buccal secretions escape an attack of pneumonia? In the present state of our knowledge this question no longer presents any serious difficulties. We know, for example, that the pus organisms—*Staphylococcus pyogenes aureus*, *albus*, and *citreus*—are very frequently found in the buccal secretions and on the surface of the body of healthy individuals, and that, although these micrococci are recognized as the cause of furuncles and of all sorts of acute abscesses, they only give rise to the formation of such abscesses under certain special conditions relating to the general health of the individual, or to a traumatism by which their introduction to vulnerable parts is effected. Again, the tetanus bacillus is a widely distributed microorganism which has been found in the earth, and especially in rich loam, in various parts of the globe. But the hands of farmers and gardeners are constantly soiled with such earth without their contracting tetanus. In this instance it has long been recognized that a traumatism is an essential factor in the chain of events which leads to the development of tetanus, and now we believe, on satisfactory experimental evidence, that it is not the traumatism *per se*, or the injury to the nerves, or exposure to cold, which in certain cases gives rise to this infectious malady, but that the result depends upon the introduction of a specific infectious agent at the time the wound was received or subsequently—the tetanus bacillus of Nicolaier.

In the case of the tubercle bacillus, also, it is extremely probable, in the light of our present knowledge, that this bacillus, in a living condition, not infrequently finds a lodgment in the mouth, upon the Schneiderian mucous membrane, or in the larger bronchial tubes of most individuals who live in populous communities. Here also the infectious agent is only one factor, although an essential one, in the production of the infectious disease. It must be introduced to the vulnerable location, and must find a favorable nidus in the tissues invaded. We have good reason to believe that in this, as well as in other infectious diseases, there are wide differences, inherited or acquired, in the susceptibility of the tissues to invasion when the infectious agent has been introduced to a favorable location.

In a paper read before the Pathological Society of Philadelphia in April, 1885, in discussing the relation of my *Micrococcus Pasteuri* to croupous pneumonia, I say: "It seems extremely probable that this micrococcus is concerned in the etiology of croupous pneumonia, and that the infectious

nature of this disease is due to its presence in the fibrinous exudate into pulmonary alveoli.

"But this cannot be considered as definitely established by the experiments which have thus far been made upon the lower animals. The constant¹ presence of this micrococcus in the buccal secretions of healthy persons indicates that some other factor is required for the development of attack of pneumonia; and it seems probable that this other factor acts by reducing the vital resisting power of the pulmonary tissues, and thus making them vulnerable to the attacks of the microbe. This supposition enables us to account for the development of the numerous cases of pneumonia which cannot be traced to infection from without. The germ being always present, auto-infection is liable to occur when, from alcoholism, sewer poisoning, crowd poisoning, or any other depressing agency, the vitality of the tissues is reduced below the resisting point. We may suppose, also, a reflex vaso-motor paralysis, affecting a single lobe of the lung, for example, and induced by exposure to cold, may so reduce the resisting power of the pulmonary tissue as to permit this micrococcus to produce its characteristic effects.

"Again, we may suppose that a person whose vital resisting power is reduced by any of the causes mentioned may be attacked by pneumonia from external infection with material containing a pathogenic variety of this micrococcus having a potency, permanent or acquired, greater than that possessed by the same organism in normal buccal secretions."

This is the theory by which I have attempted to explain the etiological rôle of this micrococcus in croupous pneumonia. Let us now consider the principal facts which have led to a belief in its causal relation to this disease.

Friedländer, in 1882, observed, in eight fatal cases of pneumonia in which he made autopsies, microorganisms, having an oval form, in the exudate of the pulmonary alveoli; they were in pairs or in short chains. Without firming that this microorganism is the cause of pneumonia, Friedländer seems to have considered it extremely probable that it bore an etiological relation to this disease.

During the same year Leyden and Gunther announced at a meeting of the Medical Society of Berlin (November 20th, 1882) that they had found the same micrococcus in the fibrinous exudate of pneumonia, obtained through the thoracic walls by means of a Pravaz syringe. At the same time Gunther stated that the elliptical cocci, in specimens stained with gentian violet, were surrounded with a colorless capsule.

The following year Matrui published his observations. In sixteen cases he had found an elongated coccus in the fibrinous exudate of pneumonia, always having a very transparent capsule. He had also encountered the same microorganism in the sputa of patients with other diseases, but not so abundantly as in pneumonia.

On November 19th, 1883, Friedländer communicated to the Medical Society of Berlin the results of his culture and inoculation experiments. The "pneumococcus" was characterized by the presence of a capsule which, he says, "always takes the form of the microorganism; if this is round the capsule is round; if it is elliptical the capsule is an ellipse." This capsule, however, was only found in preparations made from the blood of an inoculated animal or from the fibrinous exudate into the alveoli; in cultures it was no longer seen. The cultures in flesh-peptone gelatin presented a nodule-shaped growth which was believed to be characteristic. Growth was rapid in a variety of culture media at the ordinary room temperature (65° to 75° F.), and in a gelatin culture medium no liquefaction occurred.

The following results were obtained by Friedländer in his inoculation experiments: In one series of experiments the "pneumococci," mixed with distilled water, were injected through the thoracic walls into the lungs. Nine rabbits inoculated in this way gave an entirely negative result.

¹I should have said frequent instead of "constant presence."

out of eleven guinea-pigs are said to have succumbed and to have presented the lesions of pneumonia. All of the mice injected died within twenty-four hours, and at the autopsy the lungs were found to be congested and to present foci of red hepatization. In a second series of experiments upon mice they were made to inhale a spray containing the pneumococci in suspension. Several of these animals died and are said to have presented a typical pneumonia. The "pneumococcus," surrounded by its characteristic capsule, was found in the lungs, the spleen, the blood, and the liquid contained in the pleural cavity.

Upon this evidence Friedländer's "pneumococcus," which is now usually described as a bacillus, was very generally accepted as the specific cause of fibrinous pneumonia, and cultures were distributed throughout the laboratories of Europe bearing the label, "Pneumococcus of Friedländer." For some time after the publication of Friedländer's paper all observations made with reference to the presence of oval cocci or of encapsulated cocci in the fibrinous exudate of pneumonia were supposed to confirm his discovery. But now we know that there is another oval coccus which is far more frequently present in the exudate of acute pneumonia, which also presents the appearance of being surrounded by a transparent capsule—less pronounced, however, than that of Friedländer's bacillus—but which is entirely distinct from that of Friedländer and is probably the true pneumococcus. I shall give the distinctive characters of this microorganism later.

At the same time that Friedländer was pursuing his researches in Berlin, Talamon, a French physician, was engaged in similar researches in the laboratory of the Hôtel-Dieu. His results were communicated to the Anatomical Society of Paris on November 30th, 1883, a few days after Friedländer's communication to the Medical Society of Berlin (Germain Séé).

"Talamon did not describe his microbe as having a capsule; according to him, the pneumonia-coccus is characterized by its form. When seen in the fibrinous exudate the microbe has an elliptical form, like a grain of wheat. Cultivated in a liquid medium—an alkaline solution of extract of beef—it is elongated and attenuated, and presents the appearance of a grain of barley. On account of this appearance Talamon has proposed to call it the *lanceolate coccus*. This organism is encountered in the pneumonic exudate obtained after death, or drawn during life by means of a Pravaz syringe from the hepatized portions of the lung. Once only, out of twenty-five cases, it was found in the blood of a patient at the moment of death."

Talamon's inoculation experiments in dogs and guinea-pigs gave a negative result, but out of twenty rabbits injected through the walls of the thorax into the lungs eight showed the characteristic lesions of fibrinous pneumonia. Prof. Séé says, with reference to the evidence in the case of these rabbits as compared with that obtained by Friedländer in his mice: "The rather brief description of the lesions obtained by Friedländer in the mice inoculated by him leaves some doubt in the mind; for the presence of foci (*noyaux*) of induration in congested lungs is not sufficient to characterize fibrinous pneumonia. But the lungs of the rabbits presented by Talamon to the Anatomical Society in support of his communication leave no room for discussion. As he observed, it was not at all a question of foci of congestion, or of broncho-pneumonia, such as one observes habitually in rabbits which die of septicæmia, but a veritable lobar fibrinous pneumonia with pleurisy and pericarditis of the same nature. The naked-eye examination, as well as the microscope, showed no difference in the lesions produced in the rabbit and the pneumonia of man."

On another page Prof. Séé says: "Afanassiew repeated in the laboratory of Prof. Cornil the experiments of Friedländer and of Talamon; by the culture in peptonized gelatin of the pneumonic exudate taken from the cadaver he obtained two species of organisms, round micrococci of large and small dimensions, and oval cocci which corresponded to the microbes described by the two authors" (Friedländer and Talamon) "whose researches we have just reviewed." This quotation indicates that Prof. Séé did not question the

identity of the oval or "lanceolate" coccus found by Talamon in pneumonic exudate, and which in his experiments produced typical pneumonia in rabbits, and the so-called "pneumococcus" of Friedländer, which, according to his account, gave a negative result when injected into rabbits, but caused pneumonia in mice when injected directly into the lungs. Prof. Séguin was not alone in making this inference, which has turned out to be a mistake. The identity of the oval cocci, which had now been seen in the pneumonic exudate by numerous observers, with the microorganism which Friedländer had isolated and cultivated from material obtained post mortem from hepatized lungs, was generally admitted; and all of the observations relating to the presence of oval cocci, having a more or less distinct capsule in the exudate of fibrinous pneumonia, were supposed to give support to the alleged discovery of Friedländer. Now we know that the oval coccus frequently found in such material is not that of Friedländer, but that it is identical with a coccus first observed by the writer in September, 1885, in the blood of rabbits injected with his own saliva and subsequently (1886) named by him *Micrococcus Pasteuri*.

This was, without doubt, the coccus which produced pneumonia in Talamon's experiments upon rabbits; and we must give him the credit of having first experimentally demonstrated the fact that fibrinous pneumonia may be induced by the introduction of this microorganism into the parenchyma of the lung in these animals.

Salvioli, whose experiments were also made in 1884, had a uniformly fatal result from the injection of pneumonic sputum into rabbits (four). He also observed the oval coccus in the material injected, and in the blood of the animals which succumbed to his injections, but did not recognize the identity of this coccus with that which my own experiments and those of Pasteur, Vulpian, and others had shown to be present in normal human saliva and to induce a fatal form of septicæmia in rabbits. On the other hand, he also appears to have taken it for granted that the oval micrococcus encountered by him, and which, under certain circumstances, was surrounded by a transparent capsule, was the "pneumococcus" of Friedländer. Klein appears to have made the same mistaken inference. This is shown by the following quotation from his paper published in 1885:

"In seeking to ascertain what might be the relation between the so-called pneumococci and croupous pneumonia, I have made extensive examinations of the lungs and blood of persons dead of the disease, and also of the sputum of living patients at various stages of their illness. . . . In some of the vesicles, though few and far between, there were present undoubtedly capsulated cocci spoken of by Friedländer and others as pneumococci. . . . As regards the living patients, if we examine typical sputum of croupous pneumonia we find, besides numerous red blood discs and white blood corpuscles, also a few epithelial cells, and in the general gelatinous mass a number of microorganisms, chiefly belonging to the species micrococci.

"These are, as far as size and arrangement go, of two principal types: (a) Oval micrococci about 0.001 millimetre in length, occurring isolated, more commonly as dumbbells and slightly curved chains of four, six, even eight elements. . . . But in all these micrococci the elements are distinctly surrounded by a hyaline zone which, in stained preparations, can be made out as an unstained halo, though in some stained specimens it assumes a tint that is fainter than that of the micrococcus itself; this corresponds to the capsule of Friedländer, and for this reason he called them capsule micrococci."

In a footnote to the paper from which I have quoted Klein says:

"While this paper is passing through the press I receive from Dr. Steiner, of Baltimore, a paper in which he conclusively proves that the micrococci of human saliva, which produce in some instances septicæmia on inoculation into rabbits, are identical with the pneumococci of Friedländer, Salvioli, and others."

My own experiments with pneumonic sputum were made in January

1885, and led me to the identification of the oval coccus found in this material with the coccus found in my own saliva (by inoculations into rabbits) in September, 1880, and subsequently studied by me in an extended series of experiments made during the following years, 1880-84.

But, at the same time, I fell into the error of inference, previously made by Prof. Sée, by Salvioli, and others, and assumed that the "pneumococcus" which Friedländer had obtained from the same source was the same, although I found it difficult to reconcile the experimental data, inasmuch as he had obtained uniformly negative results in his inoculations into rabbits. To explain this discrepancy I suggested that Friedländer's pneumococcus was probably a variety having a different degree of pathogenic power.

This supposition seemed to find support in the fact, which I had previously observed, that my *Micrococcus Pasteuri* became attenuated, as to its pathogenic power, when the cultures were kept for some time; and that there seemed, from the experimental evidence before me, to be different pathogenic varieties in the buccal secretions of different individuals. At this time I had not seen a culture of Friedländer's bacillus. Later, in the autumn of 1885, when I made its acquaintance in Dr. Koch's laboratory, I recognized my mistake and hastened to correct the error.¹

For a detailed account of my experiments with pneumonic exudate I must refer to my paper published in the "Transactions of the Pathological Society of Philadelphia" (vol. xii.) and in the *American Journal of the Medical Sciences* (July, 1885).

With reference to my conclusion that the oval coccus of Talamon and of Salvioli was identical with my *Micrococcus Pasteuri*, I may say that this conclusion has been sustained by the subsequent investigations of Fränkel, Weichselbaum, Bordoni-Uffreduzzi, Netter, Gamelía, and others.

Fränkel's first paper relating to the presence of this microorganism in pneumonic exudate was published in 1885.

Having ascertained that his own saliva contained this oval micrococcus, he was induced to make an extended and interesting series of experiments which led him to the conclusion that this microorganism is far more constantly present in the exudate of fibrinous pneumonia than is the so-called "pneumococcus" of Friedländer. He says:

"Finally, as regards the relative frequency of the two hitherto investigated microorganisms in cases of pneumonia, no positive statement can yet be made. Nevertheless I am inclined to regard the lancet-shaped pneumococcus, which is identical with the microbe of sputum septicæmia, as the more frequent, and the usual infectious agent of pneumonia, on the ground that this organism is so much more frequently found in the sputum of pneumonic patients than in that of healthy individuals. This conclusion is further supported by the fact that it has not hitherto been possible to isolate, directly from the rusty sputum, Friedländer's bacillus."

The extended researches of Weichselbaum, published in 1886, give strong support to the view that this coccus is the usual infectious agent in croupous pneumonia. He examined, in all, the exudate in one hundred and twenty-nine cases of pneumonia.

In ninety-four of these cases the micrococcus in question, called by Weichselbaum "*diplococcus pneumoniae*," was obtained (fifty-four times in cultures); in twenty-one cases he obtained a streptococcus, and in nine only was the bacillus of Friedländer encountered.

Wolf, whose studies were made in Weichselbaum's laboratory, reported the result of his researches in 1887. He found the "*diplococcus pneumoniae*" in sixty-six out of seventy cases of croupous pneumonia examined, and the "pneumococcus of Friedländer" in three cases.

Netter, whose paper was published in November, 1887, found *Micrococcus*

¹ See my paper published in the *American Journal of the Medical Sciences* for July, 1886.

Pasteuri in seventy-five per cent of his cases of pneumonia, and in the sputum of convalescents from this disease its presence was verified in sixty per cent of the cases by inoculation experiments in rabbits. He makes the interesting observation that the sputum of recent convalescents is less virulent for rabbits than that collected at a later period.

Gameléia, who has recently published in the *Annales* of the Pasteur Institute an important paper upon the etiology of fibrinous pneumonia, verified the presence of *Micrococcus Pasteuri* in twelve fatal cases in which he collected material post mortem. He states that in a series of forty consecutive cases Dr. Goldenberg, whose experiments were made in his laboratory, found this micrococcus in every case by inoculation experiments in rabbits or in mice. According to Gameléia, inoculations in mice are more reliable than those made in rabbits, as the mouse is the more susceptible animal. He says: "The author, Weichselbaum, who has made the most extended research upon the etiology of pneumonia, used in his researches the method of cultivation upon gelatin. We must adopt the opinion of Baumgarten, who does not accord any decided value to the negative results of Weichselbaum with reference to the constant presence of *Streptococcus Pasteuri*. Netter, who adopted the method of inoculating the pneumonic sputum into rabbits, and who only found the microbe of Pasteur in seventy-five per cent of his cases, was wrong, in our opinion, in making use of an animal which is too resistant to determine the presence of small quantities of virus. This opinion is confirmed by the fact that Netter rendered some rabbits refractory by his inoculations with material in which he had not found the specific microbe.

"*En résumé*, taking our stand upon the positive results which we have always obtained, as well as upon the superiority of the method of research (inoculations in mice) which we have adopted, we believe ourselves authorized to conclude that fibrinous pneumonia is always dependent upon the microbe of Pasteur."

Fränkel, Weichselbaum, and other recent authors, while maintaining that *Micrococcus Pasteuri* is the most frequent etiological agent in the production of pneumonia, have been disposed to admit that in a certain proportion of the cases the bacillus of Friedländer, and possibly other microorganisms, may bear the same relation to the pneumonic process. Gameléia, on the other hand, believes that the bacillus of Friedländer is a simple saprophyte, the occasional presence of which in pneumonic exudate is without etiological import. He remarks as follows:

"We may be brief as regards the second objection made against the etiological unity of fibrinous pneumonia, viz., with reference to the etiological rights of the microbe of Friedländer. This microbe is found in normal saliva, it is a true saprophyte, and may at times invade the diseased or dead lung. Weichselbaum only found it in seven per cent of his cases, and almost always associated with other microbes, for he only encountered it pure in three cases. As to the researches of the authors who preceded Fränkel, it is sure that the microbe which they often found in sections of diseased lungs, and which they called the microbe of Friedländer, was in fact the microbe of Pasteur, since it was colored by the method of Gram, which decolorizes the bacillus of Friedländer. Many of the positive results, then, which have been reported relative to the last-mentioned microorganism, ought to be put to the account of the other."

This opinion the present writer has entertained since his researches made in 1885.

The experimental evidence offered by Gameléia in favor of the etiological rôle of this micrococcus is most important.

It will be remembered that Talamon produced typical pneumonia in eight rabbits, in 1883, by inoculating them through the thoracic walls with pneumonic exudate. Gameléia says:

"The number of my rabbits in which pneumonia was induced is about two hundred."

The writer found in his experiments, made in 1881, that in making a series of inoculations in rabbits the virus increased in virulence, and that, on the other hand, the micrococcus lost its virulence when the cultures were kept for some time. This fact has been verified by the subsequent researches of Fränkel and of Gameléia. The last-named author has shown that to induce pneumonia in very susceptible animals, like the rabbit, an attenuated variety of the microbe should be injected, for the most virulent cultures quickly cause death by septicæmia. As he expresses it: "Animals which are too susceptible, like the rabbit and the mouse, do not have pneumonia, because they do not offer a local reaction; the virus is generalized in their bodies and they die of an acute septicæmia."

On the other hand, Gameléia has shown that "animals which are but little susceptible to the pneumonic virus offer a local resistance which gives rise to very pronounced reactionary phenomena (extended fibrino-granular cedema), and consequently they present, as a result of intrapulmonary infection, a typical fibrinous pneumonia. Such animals are the dog and the sheep."

In his experiments upon these animals Gameléia obtained the following results:

The sheep was found to survive subcutaneous inoculations, unless very large doses (five cubic centimetres) of the most potent virus were administered. But intrapulmonary inoculation was always followed by typical fibrinous pneumonia, which in the majority of cases proved fatal.

The microbe was rarely found in the blood, and successive inoculations from one sheep to another were not successful. Death occurred, after an intrapulmonary inoculation, on the third, fourth, or fifth day. The pneumonia produced was lobar, and was attended with an extensive fibrinous exudation in which the coccus was found in great abundance. In all, fifty sheep were experimented upon.

The writer found in his experiments, made in 1881, that the dog resists inoculations with this coccus. Gameléia also obtained negative results when moderate doses were injected beneath the skin, but states that "intrathoracic infection always causes a frank, fibrinous pneumonia which is rarely fatal; recovery usually occurs in from ten to fifteen days, after the animal has passed through all the stages of red and gray hepatization which characterizes this affection in man." Twelve dogs were experimented upon.

This micrococcus, then, which in very susceptible animals (mouse, rabbit) invades the blood and quickly causes death by septicæmia, when injected through the thoracic walls in less susceptible animals (dog, sheep), or in an attenuated form in the rabbit, gives rise to the local lesions which characterize fibrinous pneumonia.

Man comes in the category of slightly susceptible animals, as is shown by the comparatively small mortality from pneumonia, and the fact that the micrococcus found in the exudate into the pulmonary alveoli does not invade the blood, unless in rare instances. We may therefore agree with Gameléia in the following statement:

"It is clear that the results obtained in the dog and the sheep, animals which have but a slight susceptibility, are most applicable to human pathology."

In my paper read before the Pathological Society of Philadelphia in April, 1885, from which I have already quoted, I say: "It seems extremely probable that this micrococcus is concerned in the etiology of croupous pneumonia. . . . But this cannot be considered as definitely established by the experiments which have thus far been made upon the lower animals."

The experiments of Gameléia go far toward settling this question in a definite manner, and, considered in connection with those of Talamon and Salvioli, and the extended researches of Fränkel, Weichselbaum, and Netter, leave but little doubt that this is the true infectious agent in acute lobar pneumonia.

7. BACILLUS OF FRIEDLÄNDER.

Synonyms.—Pneumococcus (Friedländer); Bacillus pneumon (Flügge).

Obtained by Friedländer and Frobenius in pure cultures (1881) from the exudate into the pulmonary alveoli in cases of croupous pneumonia. Subsequent researches show that it is only present in a small proportion of the cases—nine times in one hundred and twenty-nine cases examined by Weichselbaum, three times in seven cases examined by Wolf.

Morphology.—Short rods with rounded ends, often so short



FIG. 88.—Bacillus of Friedländer; a, from a culture; b, from blood of mouse, showing capsule. (Flügge.)

to resemble micrococci, especially in very recent cultures; commonly united in pairs or chains of four, and under certain circumstances surrounded by a transparent capsule. The gelatinous envelope—called capsule—is not seen in preparations made from cultures in artificial media, but is very prominent in properly stained preparations from the blood of an inoculated animal.

It often has a diameter equal to or greater than that of the enclosed cell, and appears to consist of a substance resembling mucin, which is soluble in water and dilute alcohol. Where several cells are united in a chain they may all be enclosed in a common envelope, or each may have its own capsule. This capsule is not peculiar to Friedländer's bacillus, as was at first supposed, but is found in other bacilli and also in the writer's *Micrococcus Pasteuri*.

Friedländer's bacillus stains readily with the aniline colors, but is decolorized by the iodine solution used in Gram's method. In preparations from the blood of an inoculated animal, stained by aniline color, the capsule appears as an unstained envelope surrounding the stained cell, but by special treatment the capsule may also be stained. Friedländer's method is as follows: The section or cover-glass preparation is placed for twenty-four hours in a solution of gentian violet and acetic acid, containing fifty parts of a concentrated alcoholic solution of gentian violet, one hundred parts of distilled water, and ten parts of acetic acid. The stained preparation is washed for a minute or two in a one-per-cent solution of acetic acid, dehydrated with alcohol, cleared up with oil of cloves or cedar, and mounted in balsam. The bacillus is quickly stained in dried cover-glass preparations by immersion in aniline-water-gentian-violet solution (two or three minutes). The stained preparation should be decolorized by placing it in absolute alcohol for half a minute, and then washed in distilled water.

Biological Characters.—This bacillus does not, so far as is known, form reproductive spores; it is *non-motile* and *does not liquefy gelatin*. It is *aërobic* and a *facultative anaërobic*. In gelatin stick cultures it presents the “nail-shaped” growth first described by Friedländer, which is not, however, peculiar to this bacillus. The head of the nail is formed by the development around the point of entrance of the inoculating needle of a rounded, white mass having a smooth, shining surface, and its stem by the growth along the line of puncture. This consists of closely crowded, opaque, white, spherical colonies. Gas bubbles sometimes develop in gelatin cultures, and in old cultures the gelatin about the line of growth acquires a yellowish-brown color. The growth in nutrient agar resembles that in gelatin. Upon the surface of blood serum abundant grayish-white, viscid masses are developed. Upon potato the growth is abundant, quickly covering the entire surface with a thick, yellowish-white, glistening layer which often contains gas bubbles when the temperature is favorable. Colonies in gelatin plates appear at the end of twenty-four hours as small, white spheres, which increase rapidly in size, and upon the surface form rounded, smooth, glistening, white masses of considerable size. Under the microscope the colonies present a somewhat irregular outline and a slightly granular appearance. Growth occurs at comparatively low temperatures— 16° to 20° C.—but is more rapid in the incubating oven. The thermal death-point, as determined by the writer, is about 56° C. In the ordinary culture media it retains its vitality for a long time, and may grow when transplanted to fresh culture material after having been preserved for a year or more. At a temperature of 40° C. development ceases.



FIG. 89.—Friedländer's bacillus; stick culture in gelatin; end of four days at 16° – 18° C. (Baumgarten.)

Pathogenesis.—In Friedländer's experiments the bacillus from pure cultures, suspended in water, was injected through the thoracic wall into the right lung of dogs, rabbits, guinea-pigs, and mice. Rabbits proved to be immune; one dog out of five, six guinea-pigs out of eleven, and all of the mice (thirty-two) succumbed to the inoculation. At the autopsy the pleural cavities were found to contain a sero-purulent fluid; the lungs were intensely congested, contained but little air, and in places showed limited areas of red infiltration; the spleen was considerably enlarged; the bacillus was

found in great numbers in the lungs, the fluid in the pleural cavities, and in the blood obtained from the general circulation or from the various organs of the body. Similar appearances presented themselves in the case of the guinea-pigs which succumbed to the inoculation.

These results show that the bacillus under consideration is pathogenic for mice and for guinea-pigs, but they are by no means sufficient to prove that it is capable of producing a genuine croupous pneumonia in man, and it is still uncertain whether its occasional presence in the exudate into the pulmonary alveoli in cases of this disease has any etiological importance.

8. MICROCOCCUS PNEUMONIÆ CROUPOSÆ.

Synonyms.—*Micrococcus Pasteuri* (Sternberg); *Micrococcus* of sputum septicæmia (Fränkel); *Diplococcus pneumoniae* (Weichselbaum); *Bacillus septicus sputigenus* (Flügge); *Bacillus salivarius septicus* (Biondi); Lancet-shaped micrococcus (Talamon); *Streptococcus lanceolatus Pasteuri* (Gameléia).

Discovered by the present writer in the blood of rabbits inoculated subcutaneously with his own saliva in September, 1880; by Pasteur in the blood of rabbits inoculated with the saliva of a child which died of hydrophobia in one of the hospitals of Paris in December, 1880; identified with the micrococcus in the rusty sputum of pneumonia, by comparative inoculation and culture experiments, by the writer in 1885 (paper published in the *American Journal of the Medical Sciences*, July 1st, 1885). Proved to be the cause of croupous pneumonia in man by the researches of Talamon, Salvioli, Sternberg, Fränkel, Weichselbaum, Netter, Gameléia, and others.

The Presence of Micrococcus Pasteuri in the Salivary Secretions of Healthy Individuals.—In September, 1880, while engaged in investigations relating to the etiology of the malarial fevers, I injected a little of my own saliva beneath the skin of two rabbits as a control experiment. To my surprise the animals died, and I found in their blood a multitude of oval microorganisms, united for the most part in pairs, or in chains of three or four elements. These experiments are recorded in my paper entitled “Experimental Investigations Relating to the Etiology of the Malarial Fevers,” published in the Report of the National Board of Health for 1881, pp. 74, 75.

Following up my experiments made in New Orleans (in September, 1880), in Philadelphia (January, 1881), and in Baltimore (March, 1881), I obtained the following results:

“The saliva of four students, residents of Baltimore (in March), gave negative results; eleven rabbits injected with the saliva of six individuals in Philadelphia (in January) gave eight deaths and three

negative results; but in the fatal cases a less degree of virulence was shown in six by a more prolonged period between the date of injection and the date of death. This was three days in one, four days in four, and seven days in one."

In a paper published in the *Journal of the Royal Microscopical Society* (June, 1886) I say :

"My own earlier experiments showed that there is a difference in the pathogenic potency of the saliva of different individuals, and I have since learned that the saliva of the same individual may differ in this respect at different times. Thus during the past three years injections of my own saliva have not infrequently failed to cause a fatal result, and in fatal cases death is apt to occur after a somewhat longer interval, seventy-two hours or more; whereas in my earlier experiments the animals infallibly died within forty-eight hours."

The presence of my *Micrococcus Pasteuri* was demonstrated in the blood of the rabbits which succumbed to the inoculations.

Claxton, in a series of experiments made in Philadelphia in 1882, injected the saliva of seven individuals into eighteen rabbits. Five of these died within five days, and nine at a later period.

Fränkel, whose first publication was made in 1885, discovered the presence of this micrococcus in his own salivary secretions in 1883, and has since made extended and important researches with reference to it. The saliva of five healthy individuals and the sputa of patients suffering from other diseases than pneumonia, injected into eighteen rabbits, induced fatal "sputum septicæmia" in three only. When he commenced his experiments his saliva was uniformly fatal to rabbits, but a year later it was without effect.

Wolf injected the saliva of twelve healthy individuals, and of three patients with catarrhal bronchitis, into rabbits, and induced "sputum septicæmia" in three.

Netter examined the saliva of one hundred and sixty-five healthy persons, by inoculation experiments in rabbits, and demonstrated the presence of this micrococcus in fifteen per cent of the number.

Vignal, in his recent elaborate paper upon the microorganisms of the mouth, says :

"Last year I encountered this microbe continually in my mouth during a period of two months, then it disappeared, and I did not find it again until April of this year, and then only for fifteen days, when it again disappeared without appreciable cause."

The Presence of Micrococcus Pneumoniæ Crouposæ in Pneumonic Sputum.—Talamon, in 1883, demonstrated the presence of this micrococcus in pneumonic sputum, described its morphological characters, and produced typical croupous pneumonia in rabbits by in-

jecting material containing it into the lungs through the thoracic walls.

Salvioli, in 1884, demonstrated its presence in pneumonic sputum by injections into rabbits.

In 1885 the writer made a similar demonstration, and by comparative experiments showed that the micrococcus present in the blood of rabbits inoculated with the rusty sputum of pneumonia was identical with that which he had discovered in 1880 in rabbits inoculated with his own saliva.

The same year (1885) A. Fränkel made a similar demonstration, and published a paper containing valuable additions to our knowledge relating to the biological characters of this microorganism (first publication appeared July 13th, 1885).

In 1886 Weichselbaum published the results of his extended researches relating to the presence of this micrococcus in the fibrinous exudate of croupous pneumonia. He obtained it in ninety-four cases (fifty-four times in cultures) out of one hundred and twenty-nine cases examined.

Wolf (1887) found it in sixty-six cases out of seventy examined.

Netter (1887) in seventy-five per cent of his cases, and in the sputum of convalescents from pneumonia in sixty per cent of the cases examined, by inoculations into rabbits.

Gameléia (1887) in twelve fatal cases of pneumonia in which he collected material from the lungs at the post-mortem examination.

Goldenberg, whose researches were made in Gameléia's laboratory, found it in pneumonic sputum in forty consecutive cases, by inoculations into rabbits and mice.

The Presence of Micrococcus Pneumoniæ Crouposæ in Meningitis.—Numerous bacteriologists have reported finding diplococci in the pus of meningitis, and frequently the microorganisms have been fully identified as "diplococcus pneumoniæ." Thus Netter (1889), in a résumé of the results of researches made by him in twenty-five cases of purulent meningitis, reports as follows:

Thirteen cases were examined microscopically, by cultures, and by inoculations into susceptible animals; six cases by microscopical examination and experiments on animals; and the remainder only by microscopical examination. Four of the cases were complicated with purulent otitis, six with pneumonia, three with ulcerative endocarditis. The "pneumococcus" was found in sixteen of the twenty-five cases; in four *Streptococcus pyogenes* was present; in two *Diplococcus intracellularis meningitidis* of Weichselbaum; in one Friedländer's bacillus; in one Newmann and Schäffer's motile bacillus; in one a small curved bacillus.

In forty-five cases collected from the literature of the subject by

Netter this micrococcus was present in twenty-seven, *Streptococcus pyogenes* in six, and the *Diplococcus intracellularis meningitidis* of Weichselbaum in ten.

Monti (1889), in four cases of cerebro-spinal meningitis, demonstrated the presence of the same micrococcus. In three of his cases pneumonia was also present. In two *Staphylococcus pyogenes aureus* was associated with the "*diplococcus pneumoniae*."

Micrococcus Pneumoniae Crouposæ in Ulcerative Endocarditis.—Weichselbaum, in a series of twenty-nine cases examined (1888), found "*diplococcus pneumoniae*" in seven.

Micrococcus Pneumoniae Crouposæ in Acute Abscesses.—In a case of parotitis occurring as a complication of croupous pneumonia this micrococcus was obtained from the pus in pure cultures by Testi (1889); and in another case in which, as a complication of pneumonia, there developed a purulent pleuritis, abscess of the parotid on both sides, and multiple subcutaneous abscesses, the pus from all of the sources named contained the "*diplococcus*" in great numbers, as

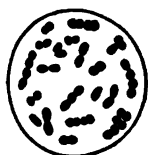


FIG. 90.

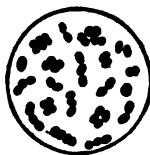


FIG. 91.

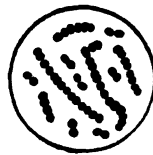


FIG. 92.

FIG. 90.—*Micrococcus pneumoniae crouposæ* from blood of rabbit inoculated with normal human saliva (Dr. S.). $\times 1,000$.

FIG. 91.—*Micrococcus pneumoniae crouposæ* from blood of rabbit inoculated subcutaneously with fresh pneumonic sputum from a patient in the seventh day of the disease. $\times 1,000$.

FIG. 92.—Surface culture of *Micrococcus pneumoniae crouposæ*, on nutrient agar, showing the development of long chains. $\times 1,000$.¹

shown not only by microscopical examination but by inoculation into rabbits.

In a case of tonsillitis resulting in the formation of an abscess Gabbi (1889) obtained the same coccus in pure cultures.

In *otitis media* this micrococcus has been found in a considerable number of cases in the pus obtained by paracentesis of the tympanic membrane, and quite frequently in pure cultures—by Zaufal (1889) in six cases; Levy and Schrader (1889) in three out of ten cases in which paracentesis was performed; by Netter (1889) in five out of eighteen cases occurring in children.

Monti (1889) and Belfanti (1889) report cases of arthritis of the wrist joint, occurring as a complication of pneumonia, in which this micrococcus was obtained in pure cultures. Ortmann and Samter

¹The above figures are from Dr. Sternberg's paper published in the American Journal of the Medical Sciences for July and October, 1885.

(1889), in a case of purulent inflammation of the shoulder joint following pneumonia and pleurisy, obtained the "diplococcus pneumoniae" in pure cultures.

Morphology.—Spherical or oval cocci, usually united in pairs in chains consisting of three or four elements. Longer chains, containing ten or more elements, are sometimes formed, especially in cultures upon the surface of nutrient agar, and it may therefore be regarded as a streptococcus. As observed in the blood of inoculated animals it is usually in pairs consisting of oval or lance-oval elements which are surrounded by a transparent capsule. Owing to the elongated form of the cocci when in active growth, it has been regarded by some authors as a bacillus; but in cultures in liquid media, where development by binary division has ceased, the cells are spherical or nearly so, and in cultures on the surface of nutrient agar the individual cells more nearly approach a spherical form than in the blood of an inoculated animal. The "lanceolate" form was first referred to by Talamon, who described it as having the form of a grain of wheat or even still more elongated like a grain of barley, as seen in the fibrinous exudate of croupous pneumonia. The transparent material surrounding the cells—so-called capsule—is best seen in stained preparations from the fibrinous exudate of croupous pneumonia or from the blood of an inoculated animal. It appears as an unstained marginal band surrounding the elliptical cells, and varies greatly in its extent in different preparations. This capsule probably consists of a substance resembling mucin, and, being soluble in water, its extent depends partly upon the methods employed in preparing specimens for microscopical examination. It is occasionally seen in



FIG. 93.—*Micrococcus pneumoniae crouposæ*, showing capsule, attached to pus cells from exudate in pleural cavity of inoculated rabbit. (Salvioli.)

stained preparations from the surface of cultures on blood serum; and in drop cultures examined under the microscope, by using a small diaphragm, it may be seen to surround the cocci as a scarcely visible halo.

This micrococcus stains readily with aniline colors; and also by Gram's method, which constitutes an important character distinguishing it from Friedländer's bacillus, with which it has often been confounded on account of the morphological resemblance of the two microorganisms.

Biological Characters.—Grows in the presence of oxygen—*aërobic*—but is also a *facultative anaërobic*. Like other micrococci, it has no spontaneous movements. It grows in a variety of culture media when they have a slightly alkaline reaction, but does not develop in a medium which contains the slightest trace of

acid. Nor will it grow at the ordinary room temperature. Scanty development may occur at a temperature of 22° to 24° C., but a temperature of 35° to 37° C. is most favorable for its growth, which is very rapid in a suitable liquid medium. In an infusion made from the flesh of a chicken or a rabbit it multiplies, in the incubating oven, with remarkable rapidity; at the end of six to twelve hours after inoculation the previously transparent fluid will be found to present a slight cloudiness and to be filled throughout with the cocci in pairs and short chains. It does not produce a milky opacity in liquid media, like the pus cocci, for example, but the fluid becomes slightly clouded; multiplication ceases at the end of about forty-eight hours or less, and the liquid medium again becomes transparent as a result of the subsidence of the cocci to the bottom of the receptacle.

It may be cultivated in flesh-peptone-gelatin, containing fifteen per cent of gelatin, at a temperature of 24° C., or in liquefied gelatin (ten per cent) in the incubating oven. In gelatin (fifteen per cent) stick cultures small white colonies develop all along the line of puncture, and in gelatin plates small, spherical, slightly granular, whitish colonies are formed: the *gelatin is not liquefied*. In agar plates extremely minute colonies are developed in the course of forty-eight hours, which resemble little, transparent drops of fluid, and under the microscope some of these are observed to have a compact, finely granular central portion surrounded by a paler, transparent, finely granular marginal zone. Upon the surface of nutrient agar or coagulated blood serum development occurs in the form of minute, transparent, jelly-like drops, which form a thin layer along the line of inoculation in "streak cultures"; and in agar stick cultures the growth along the line of puncture is rather scanty, almost homogeneous, and semi-transparent. Upon potato no development occurs, even in the incubating oven. Milk is a favorable culture medium, and the casein is coagulated as a result of its presence.

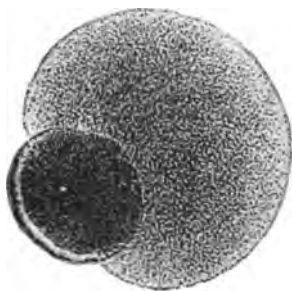


FIG. 94.—Single colony of *Micrococcus pneumoniae crouposæ* upon agar plate, twenty-four hours old. $\times 100$. (Fränkel and Pfeiffer.)

It ceases to grow on solid media at about 40° C., and in favorable liquid media at 42° C. Its thermal death-point, as determined by the writer, is 52° C., the time of exposure being ten minutes. It loses its vitality in cultures in a comparatively short time—four or five days on agar—and is very sensitive to the action of germicidal agents. Its pathogenic power also undergoes attenuation very

quickly when it is cultivated in artificial media, but may be rendered innocuous by passing it through the bodies of susceptible animals. Attenuation of virulence may also be effected by exposing bouillon culture to a temperature of 42° C. for twenty-four hours, or by five hours exposure to a temperature of 41° C.

Emmerich has recently (1891) reported to the Congress of Hygiene and Demography in London the results of experiments made by him relating to immunity in rabbits and mice. Rabbits were rendered immune by the intravenous injection of a very much diluted but virulent culture of the micrococcus. The flesh of these rabbits was rubbed up into a fine paste, and the juices obtained by compressing it in a clean, sterilized cloth. This bloody juice was exposed for twelve hours at a temperature of 10° C., and then sterilized by passing it through a Pasteur filter. Some of this juice was injected into a rabbit, which with twenty-five others was then made to breathe in a spire an atmosphere charged with a spray of a bouillon culture of the micrococcus. As a result of this all of the rabbits died except one which had previously been injected with the immunizing juice. In a similar experiment upon mice six of these animals, which had previously been injected with the immunizing juice, survived the injection of a full dose of a virulent culture, while a control mouse not previously injected with the juice, promptly died after receiving the same quantity of the virulent culture.

The writer in 1881, in experiments made to determine the effect of various disinfectants, as tested upon this micrococcus, obtained experimental evidence that its virulence is attenuated by the use of certain antiseptic agents. Commenting upon the results of these experiments in my chapter on "Attenuation of Virus," in *Journal of Bacteriology* (1884), I say :

"Sodium hyposulphite and alcohol were the chemical reagents which produced the result noted in these experiments ; but it seems probable that a variety of antiseptic substances will be found to be equally effective when used in the proper proportion. Subsequent experiments have shown that neither of these agents is capable of destroying the vitality of the micrococcus in the proportion used (one per cent of sodium hyposulphite or one part of ninety-five-per-cent alcohol to three parts of virus), but both have a restraining influence upon the development of this micrococcus in culture fluids."

The following results were obtained by the writer in his experiments (1881 and 1883) to determine the germicidal and antiseptic value of the agents named, as tested upon this micrococcus.

Alcohol.—A twenty-four-per-cent solution was effective in destroying bouillon cultures in two hours.

Boric Acid.—A saturated solution failed to destroy vital bouillon cultures in two hours' exposure, but 1 : 400 restrained development.

Carbolic Acid.—A one-per-cent solution destroys vitality in two hours, and 1 : 500 restrains development.

Cupric Sulphate destroys the virulence of the coccus in the blood of a rabbit in the proportion of 1 : 400 in half an hour.

Ferric Sulphate failed to destroy vitality in a saturated solution, but restrained development in the proportion of 1 : 200.

Hydrochloric Acid destroys the virulence of the blood of a rabbit containing this micrococcus in the proportion of 1 : 200.

Iodine, in aqueous solution with potassium iodide, destroys vitality in the proportion of 1 : 1,000 and prevents development in 1 : 4,000.

Mercuric Chloride.—One part in forty thousand prevents the development of this micrococcus, and 1 : 20,000 was found to destroy vitality in two hours.

Nitric Acid.—One part in four hundred destroyed the virulence of rabbit's blood containing this micrococcus.

Caustic Potash.—A two-per-cent solution destroyed vitality in two hours.

Potassium Permanganate.—A two-per-cent solution destroyed the virulence of rabbit's blood containing this coccus.

Salicylic Acid, dissolved by the addition of sodium bichlorate.—A solution of 1 : 400 prevented development.

Sulphuric Acid.—One part in two hundred destroys vitality, and 1 : 800 prevents development.

In a recent paper by Bordoni-Uffreduzzi relating to the resisting power of pneumonic virus for desiccation and light, the following results are given: Pneumonic sputum attached to cloths, when dried in the air and exposed to diffuse daylight, retained its virulence, as shown by injections in rabbits, for a period of nineteen days in one series of experiments and for fifty-five days in another. Exposed to direct sunlight the same material retained its virulence after twelve hours' exposure. Cultures have far less resistance, and the protection afforded by the dried albuminous material in which the micrococci were embedded, in the experiments referred to, probably accounts for the virulence being retained so long a time.

Recently (1892) Kruse and Pansini have published an elaborate paper giving an account of their researches relating to "*diplococcus pneumoniae*" and allied streptococci. We give below a summary statement of their results :

Many varieties were obtained by the observers named in their cultures from various sources—from the lungs of individuals dead from pneumonia, from pleuritic exudate, from pneumonic sputa, from bronchitic sputa, from the saliva of healthy persons, from the secretion in a case of subacute nasal catarrh, from the urine of a patient with nephritis.

Pure cultures were obtained by the use of agar plates or by inoculations into rabbits. In all about thirty varieties were obtained and cultivated

through many successive generations. As a rule, the different varieties which at first were seen to have the form of diplococci, when cultivated a length of time in artificial media presented the form of streptococci. The elements which at first were lancet-shaped showed a tendency to become spherical.

The more virulent varieties usually presented the form of diplococci with lancet-shaped elements, or of short chains. A variety which presented long chains could be pronounced, in advance of the experiments on virulence, to possess comparatively little virulence. When by inoculations in rabbits the virulence of such a variety was restored, the tendency to form long chains was less pronounced.

Although, as a rule, no development occurs at 20° C., certain varieties were obtained which, after long cultivation in artificial media, showed decided growth at 18° C.

Decided differences were shown by the cultures from various sources regarding their growth in milk. Out of eighty-four cultures from various sources eleven did not produce coagulation. As a rule, cultures which produced coagulation of milk were virulent for rabbits, and when such cultures were lost their virulence they usually lost at the same time the power of coagulating milk. Virulent cultures die out sooner than those which have been attenuated by continuous cultivation in artificial media; the first, on the face of agar, usually fail to grow at the end of a week, while the attenuated cultures may survive for three weeks or more.

Pathogenesis.—This micrococcus is very pathogenic for man and for rabbits, less so for guinea-pigs. The injection of a minute quantity—0.2 cubic centimetre or less—of a virulent culture beneath the skin of a rabbit or a mouse usually results in the death of the animal in from twenty-four to forty-eight hours. The following is from the writer's first published paper (1881), and refers to the pathological appearances in rabbits :

"The course of the disease and the post-mortem appearances indicate that it is a form of septicæmia. Immediately after the injection there is a rise in temperature, which in a few hours may reach 2° to 3° C. (3.6° to 5.4° F.). The temperature subsequently falls, and shortly before death is often 1° or 2° degrees below the normal. There is loss of appetite and marked depression after twenty-four hours, and the animal commonly dies during the night or early in the morning of the second day after the injection. Death occurs still more quickly when the blood from a rabbit recently deceased is injected. Not infrequently convulsions immediately precede death.

"The most marked pathological appearance is a diffuse inflammation of the subcutaneous tissue, extending in all directions from the point of injection, but especially to the dependent portions of the body. Occasionally there is a little pus near the puncture, but usually death occurs before the pus reaches the point of producing pus. The subcutaneous connective tissue contains a quantity of bloody serum, which possesses virulent properties. The blood which contains a multitude of micrococci. There is usually more or less inflammatory adhesion of the integument to the subjacent tissues. The skin is sometimes dark-colored and gorged with blood, but more frequently of a lighter color than normal and contains much fat. The spleen is usually normal in appearance or enlarged and dark-colored. Changes in the internal organs are more marked in those cases which are of the longest duration.

"The blood commonly contains an immense number of micrococci, often joined in pairs and having a diameter of about 0.5 μ . These are usually blood drawn from superficial veins, from arteries, and from the cavity of the heart immediately after death, and in a few cases their presence

verified during life. Observations thus far made, however, indicate that it is only during the last hours of life that these parasites multiply in the circulating fluid, and in a certain proportion of the cases a careful search has failed to reveal their presence in the blood in post-mortem examinations made immediately after the death of the animal."

In animals which are not examined until some hours after death a considerable increase in the number of micrococci occurs post mortem. The fact that this micrococcus varies very much as to its pathogenic power, as a result of conditions relating to the medium in which it develops, was insisted upon in my first published paper, and has been fully established by later researches (Fränkel, Gameléia). Susceptible animals inoculated with attenuated cultures acquire an immunity against virulent cultures.

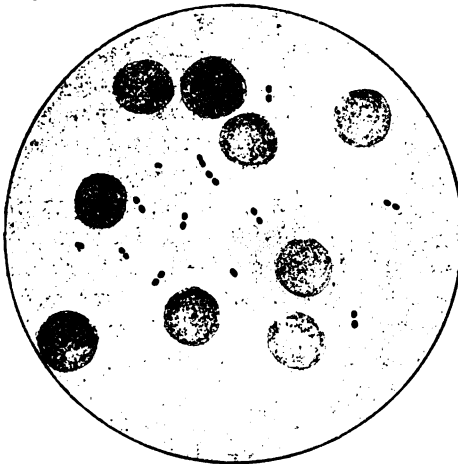


FIG. 85.—*Micrococcus pneumoniae croupose* in blood of rabbit inoculated with pneumonic sputum. $\times 1,000$.

In dogs subcutaneous injections usually give a negative result, or at most a small abscess forms at the point of inoculation. In a single experiment, however, the writer has seen a fatal result in a dog from the injection of one cubic centimetre of bloody serum from the subcutaneous connective tissue of a rabbit recently dead. This shows the intense virulence of the micrococcus when cultivated in the body of this animal. Pneumonia never results from subcutaneous injections into susceptible animals, but injections made through the thoracic walls into the substance of the lung may induce a typical fibrinous pneumonia. This was first demonstrated by Talamon (1883), who injected the fibrinous exudate of croupous pneumonia, obtained after death, or drawn during life by means of a Pravaz syringe from the hepatized portions of the lung, into the lungs of

rabbits. According to Sée, eight out of twenty animals experimented upon exhibited "a veritable lobar, fibrinous pneumonia, with pleurisy and pericarditis of the same nature." Gameléia has also induced pneumonia in a large number of rabbits, and also in the dog and the sheep, by injections directly into the pulmonary tissue. Sheep were found to survive subcutaneous inoculations, unless very large doses (five cubic centimetres) of the most potent virus were injected. But intrapulmonary inoculations invariably induced a typical fibrinous pneumonia which usually proved fatal. In dogs similar injections gave rise to a "frank, fibrinous pneumonia which rarely proved fatal, recovery usually occurring in from ten to fifteen days, after the animal had passed through the stages of red and gray hepatization characteristic of this affection in man."

Monti claims to have produced typical pneumonia in rabbits by injecting cultures of this micrococcus into the trachea.

From the evidence obtained in these experimental inoculations, and that recorded relating to the presence of this micrococcus in the fibrinous exudate of croupous pneumonia, we are justified in concluding that it is the usual cause of this disease, and consequently have described it under the name *Micrococcus pneumoniae crouposæ*. We prefer this to the name commonly employed by German authors—"diplococcus pneumoniae"—because this micrococcus, although commonly seen in pairs, forms numerous short chains of three or four elements in cultures in liquid media, and upon the surface of nutrient agar may grow out into long chains. It would, therefore, more properly be called a streptococcus than a diplococcus.

Recently (1891) G. and F. Klemperer have published an important memoir relating to the pathogenic action of this micrococcus. They succeeded in conferring immunity upon susceptible animals by inoculating them with filtered cultures of the micrococcus, and in some instances this immunity had a duration of six months. A curious fact developed in their researches was that the potency of the substance contained in the filtered cultures was increased by subjecting these to a temperature of 41° to 42° C. for three or four days, or to a higher temperature (60° C.) for an hour or two. When injected into a vein after being subjected to such a temperature immunity was complete at the end of three or four days; but the same material not so heated required larger doses and a considerably longer time (fourteen days) to confer immunity upon a susceptible animal. The unwarmed material caused a considerable elevation of temperature, lasting for some days. The authors mentioned conclude from their investigations that the toxic substance present in cultures of *Micrococcus pneumoniae crouposæ* is a proteid substance, which they propose to call pneumotoxin. The substance produced

in the body of an immune animal, as a result of protective inoculations, upon which the immunity of these animals depends, is also a proteid, which they call anti-pneumotoxin. This they isolated from the blood serum of immune animals. By experiment they were able to demonstrate that the blood serum containing this protective proteid, when injected into other animals, rendered them immune ; and also that it arrested the progress of the infectious malady induced by inoculating susceptible animals with virulent cultures of the micrococcus. When injected into the circulation of an infected animal its curative action was manifested by a considerable reduction of the body temperature.

VI.

PATHOGENIC MICROCOCCI NOT DESCRIBED SECTIONS IV. AND V.

9. DIPLOCOCCUS INTRACELLULARIS MENINGITIDIS

DISCOVERED by Weichselbaum (1887) in the exudate of spinal meningitis (six cases), for the most part within the cells.

Morphology.—Micrococci, usually united in pairs, in groups of four, or in little masses; sometimes solitary and larger (being upon the point of dividing). Distinguished by their position in the interior of pus cells in the exudate, in this respect resembling the gonococcus.

Stain best with Löffler's alkaline solution of methylene blue. Do not retain their color when treated with iodine solution (method).

Biological Characters.—This micrococcus does not grow at room temperature, but upon nutrient agar an abundant development occurs in the incubating oven. Upon the surface of agar a luxuriant, viscid growth, which by reflected light is gray, by transmitted light grayish-white; along the line of puncture occurs only near the surface, indicating that this micrococcus does not grow in the absence of oxygen. Upon plates made from agar (one per cent) and gelatin (two per cent) very small colonies are formed in the interior of the mass, and larger ones, of a yellowish-brown color, on the surface. The former, under the microscope, may be round or slightly irregular, finely granular, and of a yellowish-brown color. The superficial colonies have a yellowish-brown nucleus, surrounded by a more transparent zone. The growth on coagulated blood serum is very scanty, as is that in bouillon. Growth occurs upon potato. This micrococcus quickly loses its power of reproduction in artificial cultures—within six days—and must be transplanted to fresh material at short intervals—two days.

Pathogenesis.—Mice are especially susceptible, and usually die within forty-eight hours after inoculation. Also pathogenic to guinea-pigs, rabbits, and dogs.

10. STAPHYLOCOCCUS SALIVARIUS PYOGENES.

Obtained by Biondi (1887) from an inoculation abscess in a guinea-pig injected subcutaneously with saliva from a child suffering from scarlatina anginosa.

Morphology.—Spherical cocci, 0.3 to 0.5 μ in diameter, usually solitary in the pus of abscesses or in irregular agglomerations.

Stains best by Gram's method.

Biological Characters.—Grows at a comparatively low temperature (12° to 14° C.), and more rapidly in the incubating oven. In gelatin stick cultures, at the room temperature, growth occurs along the line of puncture, and at the end of eight days liquefaction commences in the form of a funnel, at the bottom of which little, white, shining masses accumulate, while at the surface of the liquefied gelatin a white, viscid layer forms. In gelatin plates spherical, well-defined, opalescent, whitish colonies are formed, which cause a tardy liquefaction of the surrounding gelatin. Upon agar-agar the growth is rapid, in the form of a thick layer along the line of inoculation in streak cultures, which has a breadth of about one millimetre at the end of twenty-four hours in the incubating oven, and presents an orange-yellow color at the centre, fading out to white at the margins. The yellow color is not by any means as pronounced as in similar cultures of *Staphylococcus pyogenes aureus*, and liquefaction of gelatin is much slower.

Pathogenesis.—Produces a local abscess when inoculated into dogs, rabbits, guinea-pigs, or mice. When injected into the anterior chamber of the eye of rabbits, hypopyon, followed by spontaneous perforation of the cornea, resulted. Injected into the circulation of a guinea-pig (0.2 to 0.4 cubic centimetre) it gave rise to general infection, and death at the end of eight to ten days.

11. MICROCOCCUS OF PROGRESSIVE TISSUE NECROSIS IN MICE.

Obtained by Koch (1879) from mice inoculated subcutaneously with putrid blood.

Morphology.—Round cells, 0.5 μ in diameter, united in chains, or at times in crowded masses.

Biological Characters not given.

Pathogenesis.—Causes necrosis of the tissues in the vicinity of the point of inoculation in mice, which extends rapidly and causes the death of the animal in about three days. The blood and internal organs remain free from micrococci. (Possibly a very pathogenic variety of *Streptococcus pyogenes*?)

12. MICROCOCCUS OF PROGRESSIVE ABSCESS FORMATION IN RABBITS.

Obtained by Koch (1879) from rabbits inoculated with putrid blood.

Morphology.—Minute cocci, about 0.15 μ in diameter, usually associated in thick, cloud-like zoöglæa masses.

Biological Characters not given.

Pathogenesis.—In rabbits an extensive abscess forms in the vicinity of the point of inoculation, and the animal dies in about twelve days. The walls of the abscess are formed of a thin layer of micrococci associated in zoöglæa masses; the interior contains finely granular, cheesy material, in which the cocci appear to have degenerated and perished. The contents of the abscess injected into other rabbits produce a similar result. The micrococcus does not invade the blood.



FIG. 96.—Micrococcus of progressive tissue necrosis in mice; section of the ear; a, cartilage cells; b, streptococci. (Koch.)

13. MICROCOCCUS OF PYÆMIA IN RABBITS.



FIG. 97. — Micrococcus of pyæmia in rabbits, in capillary from the cortical portion of the kidney. $\times 700$. (Koch.)

Obtained by Koch (1879) in rabbits inoculated subcutaneously with putrefying flesh infusion.

Morphology.—Round cells, 0.25μ in diameter, solitary or in pairs, which usually surround blood corpuscles in a characteristic manner.

Biological Characters not given.

Pathogenesis.—When injected subcutaneously in rabbits the blood is invaded and death results from general infection. At the autopsy a violent infiltration is found at the point of inoculation, there is peritonitis, and metastatic abscesses are found in the lungs and liver. Numerous cocci, closely surrounding the blood corpuscles, are found in the capillaries of the various organs, and in the blood of the heart, etc. Two or three drops of blood from the heart of a rabbit recently died and injected into another animal of the same kind caused its death in about forty hours.

14. MICROCOCCUS OF SEPTICÆMIA IN RABBITS.

Obtained by Koch (1879) from rabbits inoculated subcutaneously with putrefying flesh infusion.

Morphology.—Oval cells, having a long diameter of 0.8 to 1.0μ .

Biological Characters not given.

Pathogenesis.—Produces general infection and death in rabbits and mice. At the autopsy slight oedema is observed at the point of inoculation, the spleen is greatly enlarged; no peritonitis and no embolic processes are observed, such as characterize the pathogenic action of the last-described species (Fig. 13); nor do the cocci accumulate around the red blood corpuscles. They are found in the capillaries of the various organs in masses, and especially in the glomeruli of the kidneys.

15. MICROCOCCUS SALIVARIUS SEPTICUS.

Obtained by Biondi (1887) from the saliva of a case of puerperal sepsis, by inoculations into animals.

Morphology.—Spherical or slightly oval cocci, which, when in rapid multiplication, show slight lateral protrusions.

Biological Characters.—Grows in nutrient gelatin or agar at a temperature of 18° to 20° C., and more rapidly in the incubating oven. It liquefies gelatin. In gelatin plates forms spherical, grayish-white colonies, which may acquire a dark color. In gelatin stick cultures grows along the line of puncture in the form of a column made up of crowded white colonies. Very scanty growth on potato.

Stains with all the aniline colors and by Gram's method.

Pathogenesis.—Produces general infection and death in from four to seven days when inoculated into mice, guinea-pigs, or rabbits. The cocci are found in great numbers, often assembled in masses, in the capillaries of the various organs, but no evidence of inflammatory reaction of the tissues can be observed.

16. MICROCOCCUS SUBFLAVUS (Flügge).

Synonym.—Yellowish-white diplococcus (Bumm).

Obtained by Bumm (1885) from the lochial discharge of puerperal women and from vaginal mucus. Has also been obtained from the urine of

of vesical catarrh, and in the vesicles of pemphigus; also by Fränkel in the vaginal secretion of children suffering from colpitis not of gonorrhoeal origin.

Morphology.—Diplococci, associated in biscuit-shaped pairs, separated by a cleft, and closely resembling the gonococcus of Neisser. Cells from 0.5 to 1.5 μ in diameter.

Stains with the aniline colors and by *Gram's method*—by which character it may be distinguished from the micrococcus of gonorrhoea.

Biological Characters.—Grows at the room temperature upon the surface of nutrient gelatin; small, grayish-white colonies appear along the line of inoculation at the end of twenty-four hours, which later form a confluent layer, first of a pale yellow and finally of an ochreous color. In the course of a few days liquefaction of the gelatin commences in the vicinity of the growth. Coagulated blood serum is also liquefied by this micrococcus.

Pathogenesis.—Inoculations upon mucous membranes susceptible to gonorrhoeal infection are without result. But by injecting the diplococcus from pure cultures, in suspension in distilled water, beneath the skin in man, Bumm obtained as a result local abscess formation—abscesses varying in size from that of a pigeon's egg to that of a man's fist. The diplococcus was present in great numbers in the pus of these abscesses.

17. MICROCOCCUS OF TRACHOMA (?).

Obtained by Sattler (1885) from the contents of the trachomatous follicles in cases of Egyptian ophthalmia; also by Michel (1886), who has given a more exact description of this micrococcus, and has made inoculation experiments which he believes establish its etiological relation to the form of ophthalmia with which it is associated (?).

Morphology.—Very small, biscuit shaped micrococci, in pairs—diplococci—separated by a very narrow dividing line. (This description would apply to some of the more common pus cocci, e.g., *Staphylococcus pyogenes aureus*, which have also been shown to consist of two hemispherical halves separated by a narrow line of division.)

Biological Characters.—Grows slowly upon nutrient gelatin at the room temperature, and does not liquefy this medium, upon the surface of which a grayish-white, broadly extended, glistening layer is formed, which later has a yellowish tint and tulip-shaped margins. Spherical colonies are formed along the line of puncture, which are arranged in a linear series, like a chaplet. In blood serum it grows along the line of puncture as a white, band-like stripe, which subsequently spreads out in the form of white clouds. The growth is more rapid upon nutrient agar or blood serum in the incubating oven. The development upon potato is very scanty. The cultures are viscid, drawing out into long threads when touched with a platinum needle. This micrococcus does not grow in the absence of oxygen—aerobic.

Stains by the aniline colors and by *Gram's method*.

Pathogenesis.—Not pathogenic for rabbits when injected subcutaneously or into the anterior chamber of the eye; but, according to Sattler and to Michel, when inoculated by puncture into the conjunctivæ in man it causes a follicular inflammation resulting in typical trachoma. But Michel was not able to demonstrate the presence of this micrococcus in all of his cases, and extensive researches made since by Baumgarten and by Kartulis (1887) show that in many cases of trachoma, and even in Egyptian ophthalmia (Kartulis), it cannot be found. According to the last-named author, the virulent ophthalmia which prevails in Egypt is gonorrhoeal in its origin, and he has demonstrated the presence of the gonococcus in a large series of cases. A milder, but infectious, acute catarrhal conjunctivitis is characterized by the presence of a minute bacillus, resembling the bacillus of mouse septicæmia, and found in the pus cells. A third group of chronic cases with trachoma, in the researches of Kartulis, failed to show the presence of Sattler's trachoma coccus or any other microorganisms in the contents of the diseased follicles.

18. MICROCOCCUS TETRAGENUS.

First described by Gaffky (Flügge). Obtained by Koch and Gaffky (1881) from a cavity in the lung in a case of pulmonary tuberculosis. Since found occasionally in normal saliva (three times in fifty persons examined by Biondi), and in the pus of acute abscesses (Steinhaus, Park, Vangel). Rather common in the sputum of tuberculous cases.

Morphology.—Micrococci, having a diameter of about 0.5 μ , which divide in two directions, forming tetrads, which are enclosed in a transparent, jelly-like envelope—especially well developed in the blood and tissues of inoculated animals. In culture the cocci are seen in the various stages of division, as large single

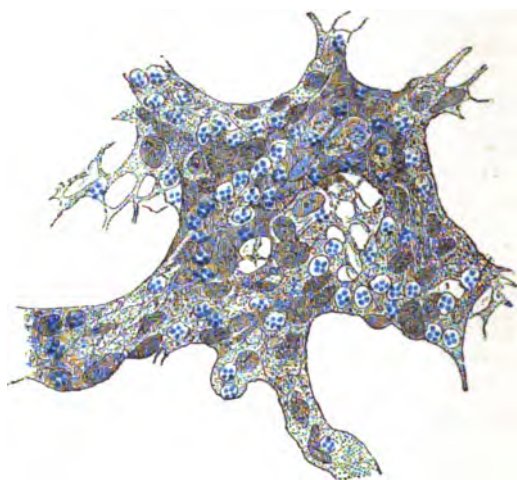


FIG. 98.—*Micrococcus tetragenus*; section of lung of mouse. $\times 800$. (Flügge.)

pairs of oval elements, or groups of four resulting from the reverse division of these latter.

Stains quickly with aniline colors, and in preparations from the blood of an inoculated animal the transparent envelope may also be feebly stained. Stains also by Gram's method.

Biological Characters.—This micrococcus grows, rather slowly, in nutrient gelatin at the ordinary room temperature, without liquefaction of the gelatin. Upon gelatin plates small white colonies are developed in from twenty-four to forty-eight hours, which under the microscope, with a low power, are seen to be spherical or oval-shaped, finely granular, and with a mulberry-like surface. When they come to the surface they form white, elevated, and rather firm masses having a diameter of one to two millimetres. In slant stick cultures a broad and thick white mass forms upon the surface.

and along the line of puncture a series of round, milk-white or yellowish masses form, which usually remain distinct, but may become confluent. Upon the surface of agar the growth is similar to that upon gelatin, or in streak inoculations may consist of a series of spherical, white colonies. Upon cooked potato a thick, viscous layer is formed of milk-white color; the growth upon blood serum is also abundant, especially in the incubating oven. This micrococcus is a facultative anaërobic.

Pathogenesis.—Subcutaneous inoculation of a culture of this micrococcus in minute quantity is fatal to white mice in from two to six days. The animals remain apparently well for the first day or two, then remain quiet and somnolent until death occurs. The cocci are found in comparatively small numbers in the blood of the heart, but are more numerous in the spleen, lungs, liver, and kidneys, from which organs beautiful stained preparations may be made showing the tetrads surrounded by their transparent capsule. Common house mice and field mice are, for the most part, immune, as are the rabbit and the dog. Guinea-pigs sometimes die from general infection, and sometimes a local abscess is the only result of a subcutaneous inoculation.

19. MICROCOCCUS BOTRYOGENUS (Rabe).

Synonyms.—Micrococcus of “myko-desmoids” of the horse; Micrococcus askoformans (Johne); Ascococcus Johni (Cohn).

First described by Bollinger (1870); morphological characters and location in the diseased tissues described by Johni (1884); biological characters determined by Rabe (1886).

Is found in certain diffused or circumscribed growths in the connective tissue of horses—“myko-desmoids.”

Morphology.—Micrococci, having a diameter of 1 to 1.5 μ , usually united in pairs.

In the tissues the cocci are united in colonies of fifty to one hundred μ in diameter, and these are associated in mulberry-like masses visible to the naked eye. The separate colonies are enclosed in a homogeneous, transparent envelope—as in Ascococcus Billrothii. This is not the case, however, in cultures in artificial media.

Stains with the aniline colors.

Biological Characters.—In gelatin plate cultures spherical, sharply defined, silver-gray colonies are developed; later these have a yellowish color and a metallic lustre, and the plate presents the appearance of being powdered with grains of pollen. It gives off a peculiar fruit-like odor, reminding one of the odor of strawberries. In gelatin stick cultures growth occurs along the line of puncture as a pale grayish-white line, which later becomes milk-white; an air

bubble forms near the surface of the gelatin; very slight liquefaction occurs in the immediate vicinity of the line of growth, and after a time the grayish-white thread sinks into an irregular mass, lying at the bottom of the puncture. Upon nutrient agar scarcely any development occurs. Upon potato the growth is abundant, in the form of a pale-yellow, circular layer, and the culture gives off the peculiar odor above described.

Pathogenesis.—When inoculated into guinea-pigs general infection and death result. In sheep and goats it produces a local inflammatory oedema and sometimes necrosis of the tissues. In horses inoculated subcutaneously an inflammatory oedema first occurs, followed at the end of from four to six weeks by the development of new growths in the connective tissue, resembling the tumors found in cases of the disease in the animal from which the micrococcus in question was first cultivated. These tumors contain characteristic mulberry-like conglomerations of colonies made up of the coccus.

20. MICROCOCCUS OF MANFREDI.

Synonym.—Micrococcus of progressive granuloma formation.

Obtained by Manfredi (1886) from the sputum of two cases of croupous pneumonia following measles.

Morphology.—Oval micrococci, having a diameter of 0.6 to 1.0 μ and from 1.0 to 1.5 μ in length; usually associated in pairs, and occasionally in short chains containing three or four elements.

Stains with the aniline colors and by Gram's method.

Biological Characters.—Aërobic; does not liquefy gelatin. Upon gelatin plates forms small, spherical colonies, at first grayish-white, which spread out upon the surface as thin, transparent plates, which by transmitted light have a bluish, by reflected light a pearl-gray color. Later these become thicker and have a pearly lustre. Under the microscope (forty to fifty diameters) the colonies are seen to be slightly granular and the margins have an irregular outline. In gelatin stick cultures a scanty growth occurs along the line of puncture, and a rather thin and limited growth about the point of inoculation. Upon blood serum a thin, greenish-yellow layer, which has irregular margins and a slightly granular, shining surface, is developed. The growth upon potato, at 37° C., is scanty, and consists of a very thin, moist layer, which has a yellowish color and is slightly granular. Growth occurs in favorable media—bouillon, gelatin—at temperatures of 18° to 48° C., but ceases at a temperature of 48° to 50° C.

Pathogenesis.—Pathogenic for dogs, rabbits, guinea-pigs, mice, and birds. In mammals the principal pathological appearance resulting from infection consists in the formation of "granulation tu-

mors" in the parenchymatous organs. These vary in size from that of a millet seed to that of a pea, and undergo caseation. They contain the micrococcus and are infectious. Mammals die in from nine to fifteen days; birds in from one to three or four, and without the formation of the characteristic granuloma, but with general infection of the blood. Cultures which have been kept for several months retain their pathogenic power.

21. MICROCOCCUS OF BOVINE MASTITIS (Kitt).

Obtained by Kitt (1885) from the udder of cows suffering from mastitis and giving milk mixed with pus.

Morphology.—Micrococci, having a diameter of 0.2 to 0.5 μ , solitary, united in pairs, in irregular groups, and occasionally in chains.

Stains with the aniline colors.

Biological Characters.—Does not liquefy gelatin. Upon gelatin plates forms spherical, translucent, glistening colonies, the size of a hemp seed to that of a pin's head; in gelatin stick cultures a nail-shaped growth occurs, the mass at the point of puncture being opaque and of a white color. Upon potato, colonies are quickly developed which have a grayish-white or dirty yellow color, and after a few days have a shining, wax-like appearance. Grows rapidly in milk, causing an acid reaction; in six hours in the incubating oven the milk is pervaded by the micrococcus, or in twelve hours at 20° C.

Pathogenesis.—Injection of pure cultures, suspended in distilled water, into the mammary glands of cows, produces typical, acute, purulent mastitis (Kitt). The micrococcus produced the same result after having been cultivated in artificial media for a year. Subcutaneous inoculations in cows, pigs, guinea-pigs, rabbits, and mice were without result. Injections into the mammary gland of goats were also without effect.

22. MICROCOCCUS OF BOVINE PNEUMONIA (?).

Isolated by Poels and Nolen (1886) from the lungs of cattle suffering from "Lungenseuche" (infectious pleuro-pneumonia of cattle).

Morphology.—Micrococci, varying considerably in size—average diameter 0.9 μ ; solitary, in pairs, or in chains containing several elements; surrounded by a transparent capsule, which stains with difficulty.

Stains with all the aniline colors, and with difficulty by Gram's method.

Biological Characters.—Does not liquefy gelatin, and grows like the bacillus of Friedländer in gelatin stick cultures (nail-shaped growth). In gelatin plates the colonies are spherical, white, and have a very faint yellowish tinge. Grows more rapidly on agar in the incubating oven, and upon potato in the form of a very pale-yellowish layer. Is destroyed by a temperature of 66° C. maintained for fifteen minutes.

Pathogenesis.—Pure cultures injected into the lungs of dogs, rabbits, and guinea-pigs are said to give rise to pneumonic inflammation, and similar results were obtained by injection into the trachea of dogs and by inhalation experiments. Injection of a pure culture into the lungs of a cow caused extensive pneumonic changes; but these did not entirely correspond with the appearances found in the lungs of cattle suffering from infectious pneumonia. Cattle inoculated with a pure culture, by means of a sterilized lancet, did not fall sick, but are believed by Poels and Nolen to have been protected from the disease by such inoculations.

The specific relation of the micrococcus above described to the disease with which it was associated, in the researches of the authors mentioned, has not been established by subsequent investigations.

23. STREPTOCOCCUS SEPTICUS (Flügge).

Found by Nicolaier and by Guarneri in unclean soil during research made in Flügge's laboratory in Göttingen.

Morphology.—Cannot be distinguished from *Streptococcus pyogenes*, does not so constantly form chains, being found in the tissues of inoculated animals, for the most part in pairs.

Biological Characters.—Grows more slowly than *Streptococcus pyogenes* in gelatin plates very minute colonies first appear at the end of three or four days, or along the line of puncture in gelatin stick cultures after five days. Does not liquefy gelatin.

Pathogenesis.—Is very pathogenic for mice and for rabbits, causing death from general infection in two or three days.

24. STREPTOCOCCUS BOMBYCIS.

Synonym.—*Microzyma bombycis* (Béchamp).

Found in the bodies of infected silkworms suffering from *la flac* (maladie des morts-plats). Etiological relation established by Pasteur.

Morphology.—Oval cells, not exceeding $1.5\ \mu$ in diameter, in pairs or short chains.

Biological Characters.—Not determined with precision.

Pathogenesis.—The infected silkworm ceases to eat, becomes weak, dies. Its body is soft and diffuent, and at the end of twenty-four to thirty hours is filled with a dark brown fluid and with gas.

25. NOSEMA BOMBYCIS.

Synonyms.—*Micrococcus ovatus*; *Panhistophyton ovatum*.

Found in the blood and all of the organs of silkworms infected with pébrine (Fleckenkrankheit).

First observed by Cornalia. Etiological relation established by Pasteur.

Morphology.—Shining, oval cells, three to four μ long and two μ wide, solitary, in pairs, or in irregular groups.

Biological Characters.—Not determined with precision.

Pathogenesis.—Dark spots appear upon the skin of infected silkworms which lose their appetite, become slender and feeble, and soon die. Oval corpuscles are found in all of the organs, and also in the eggs of butterflies hatched from infected larvae. Some authors are of the opinion that the oval corpuscles found in this disease do not belong to the *Nosema*, but to an entirely different class of microorganisms—the *Psorosphaera* (Metschnikoff).

26. MICROCOCCUS OF HEYDENREICH.

Synonyms.—*Micrococcus of Biskra* button—Fr. "clou de Biskra"; "Pendesche Geschwür."

Found by Heydenreich (1888) in pus and serous fluid obtained from tumors and ulcers in the Oriental skin affection known as Biskra button.

Morphology.—Diplococci, from 0.86 to $1\ \mu$ in length, surrounded by a capsule; sometimes associated to form tetrads.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* at 20°C ., at the end of forty-eight hours growth occurs along the line of puncture in the form of small, crowded colonies, which produce a grayish-white line; upon the surface a thin, circular layer of a yellowish color is developed. At the end of three to four days liquefaction commences near the surface, where a funnel is formed which extends until about the fourteenth day, when the gelatin is completely liquefied. Upon the surface

of agar, at 37° C., a grayish-white or yellowish layer is formed at the end of twenty-four hours, which has a varnish-like lustre. Upon *potato*, at 30° to 35° C., at the end of forty-eight hours a white or yellow layer has developed.

Pathogenesis.—According to Heydenreich, inoculations in rabbits, dogs, chickens, horses, and sheep cause a skin affection which is identical with that which characterizes Biskra button in man. When rubbed into the healthy skin of man it also produces the development of abscesses.

27. MICROCOCCUS OF DEMME.

Synonym.—Diplococcus of pemphigus acutus (Demme).

Obtained by Demme (1886) from the contents of the bullæ in a case of pemphigus.

Morphology.—Micrococci of from 0.8 to 1.4 μ in diameter; usually united in pairs resembling the "gonococcus" and having a length of 1.8 to 3 μ , very opaque and not surrounded by a capsule; usually associated in irregular masses.

Biological Characters.—*Aërobic* micrococci. Do not grow at the room temperature. Upon *agar plates*, at 37° C., at the end of thirty-six to forty-eight hours milk-white, spherical colonies, which project above the surface, are developed; later club-shaped outgrowths form around the periphery of the colony, giving it the appearance of a rosette, or sometimes of a bunch of grapes. At the end of two weeks the surface is covered with smooth projections and has a cream-like color. In streak cultures upon agar a similar growth occurs along the *impfstrich*, having club-like projections and stalactite-like outgrowths. Growth also occurs upon *potato* at a temperature of 37° C. This micrococcus develops slowly in the incubating oven, and scarcely any growth occurs at a temperature below 32° C.

Pathogenesis.—The injection of a pure culture into the lungs of guinea-pigs gave rise to emaciation and debility and to the formation of foci of broncho-pneumonia, the size of a pea, in the lungs.

28. STREPTOCOCCUS OF MANNEBERG.

Obtained by Manneberg (1888) from the urine in acute cases of Bright's disease.

Morphology.—Micrococci, about 0.9 μ in diameter, solitary, in pairs, or in chains of six to ten elements. Does not differ in morphology from *Streptococcus pyogenes*.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic* micrococcus, which slowly produces a viscid softening of nutrient gelatin. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* forms a white stripe along the line of puncture, which consists of small colonies. At the end of three or four weeks a funnel is formed containing very viscid liquefied gelatin, and at the same time brush-like outgrowths are seen along the line of development. Upon the *surface of agar* the growth resembles that of *Streptococcus pyogenes*, but is somewhat more abundant. Upon *potato*, at 37° C., at the end of four or five days white, drop-like colonies are developed of about 0.5 millimetre in diameter; these become confluent and form a slimy layer. *Milk* becomes strongly acid and coagulates within twelve hours when inoculated with this micrococcus.

Pathogenesis.—Subcutaneous injection of 0.75 to 1 cubic centimetre causes the formation of a local abscess in dogs and rabbits. Intravenous injections produce inflammatory changes in the kidneys; at the end of three or four days the urine contains red blood corpuscles, renal epithelium, blood casts, albumin, and streptococci.

29. MICROCOCCUS ENDOCARDITIDIS RUGATUS (Weichselbaum).

Obtained by Weichselbaum (1890) from the affected cardiac valves in a fatal case of ulcerative endocarditis.

Morphology.—Micrococci, resembling the staphylococci of pus in dimensions and mode of grouping; solitary, in pairs, in groups of four, or in irregular masses.

Biological Characters.—An *aërobic* micrococcus. Does not grow at the room temperature. Upon *agar plates*, at 37° C., at the end of three or four days the superficial colonies consist of a small, brown, central mass surrounded by a granular, semi-transparent, grayish marginal zone; gradually they attain a characteristic wrinkled appearance; the deep colonies, under a low power, are irregular, finely granular, and contain a large central, yellowish-brown nucleus surrounded by a narrow, grayish-brown peripheral zone. In *agar stick cultures* small, spherical colonies are formed upon the surface, which become confluent, forming a grayish-white, wrinkled layer which has a stearin-like lustre and is very viscid; a scanty growth occurs along the line of puncture. Upon *potato*, at 37° C., a scanty development occurs in the form of a small, dry, pale-brown mass. Upon blood serum isolated or confluent, colorless colonies are formed the size of a poppy seed; these are closely adherent to the surface of the culture medium.

Pathogenesis.—When injected subcutaneously into the ear of a rabbit it produces tumefaction and redness; in guinea-pigs, formation of pus. When injected into the circulation of dogs, after injury to the aortic valves, an endocarditis is developed.

30. MICROCOCCUS OF GANGRENOUS MASTITIS IN SHEEP.

Obtained by Nocard (1887) from the milk of sheep suffering from gangrenous mastitis (*mal de pis* or *d'araignée*), a fatal disease which attacks especially sheep which are being milked for the manufacture of cheese, at Roquefort and elsewhere in France.

Morphology.—Micrococci, solitary, in pairs, or in irregular groups, resembling the staphylococci of pus in dimensions and arrangement.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying* micrococcus. Grows at the room temperature in the usual culture media. Upon *gelatin plates*, at the end of forty-eight hours, the colonies are spherical and white in color; under a low power the superficial colonies are circular in outline, homogeneous, and brown in color; they are surrounded by a semi-transparent aureole; liquefaction around the superficial colonies occurs sooner than around those beneath the surface of the gelatin. In *gelatin stick cultures*, at 18° to 20° C., on the second day liquefaction of the gelatin commences near the surface; by the fifth day a pouch of liquefied gelatin has formed, which has the shape of an inverted cone; at the bottom of this an abundant deposit of micrococci is seen, while the liquefied gelatin above is clouded throughout. In *agar stick cultures* development occurs upon the surface as a thick white layer, which gradually extends over the entire surface, and after a time acquires a yellowish tint; development also occurs along the line of puncture. Upon *potato* a thin, viscid, grayish layer is slowly developed; the outline is irregular and the edges thicker than the central portion; the central portion of this layer gradually acquires a yellow color, while the periphery remains of a dirty-white or grayish color. *Blood serum* is liquefied by this micrococcus.

Pathogenesis.—A few drops of a pure culture injected subcutaneously or into the mammary gland of sheep cause an extensive inflammatory cedema and the death of the animal in from twenty-four to forty-eight hours. A cubic centimetre injected into the mammary gland of a goat produced no result; the horse, the calf, the pig, the cat, chickens, and guinea-pigs also proved to be immune. Subcutaneous injections in rabbits produce an extensive abscess at the point of inoculation.

31. STREPTOCOCCUS OF MASTITIS IN COWS.

Obtained by Nocard and Mollereau (1887) from the milk of cows suffering from a form of chronic mastitis (mammitte contagieuse).

Morphology.—Spherical or oval cocci, a little less than one μ in diameter, usually united in long chains.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* streptococcus. Grows in the usual culture media at the room temperature. Develops rapidly in milk or in bouillon at a temperature of 16° to 30° C. The milk of a cow suffering from the form of mastitis produced by this micrococcus, when drawn with proper precautions in sterilized test tubes, at the end of twenty-four hours is acid in reaction; the lower two-thirds of the tube is filled with an opaque, dirty-white, homogeneous deposit, and above this is an opalescent, serous fluid of a bluish or dirty-yellow or slightly reddish color, according to the age of the lesion. A drop of this milk examined under the microscope shows the presence of the streptococcus in great numbers. The addition of two to five per cent of glucose or of gly-

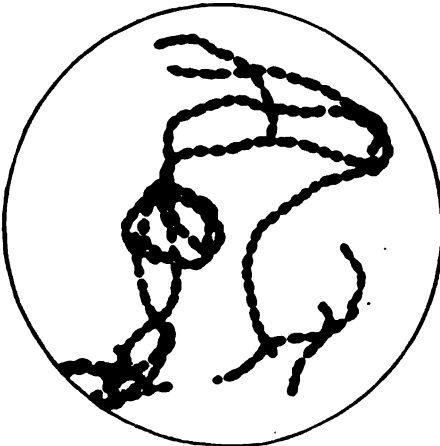


FIG. 99.—Streptococcus of mastitis in cows (Nocard).

cerin to bouillon makes it a more favorable culture medium; the reaction should be neutral or slightly alkaline, as this streptococcus does not grow readily in an acid medium, although it produces an acid reaction in media containing sugar, the acid formed being lactic. In *gelatin stick cultures* the growth upon the surface is scanty, in the form of a thin pellicle around the point of puncture; along the line of inoculation minute, opaque, granular colonies are developed, which, being closely crowded, form a thick line with jagged margins.

In *agar stick cultures* the growth is similar but more abundant. Upon the surface of nutrient gelatin, agar, or blood serum a large number of minute, spherical, semi-transparent colonies are developed along the impfstrich; these have a bluish tint by reflected light; they may become confluent, forming a thin layer with well-defined margins. Upon *gelatin plates*, at 16° to 18° C., colonies are first visible at the end of two or three days; they are spherical and slightly granular, at first transparent and later of a pale-yellow color by transmitted light, which gradually becomes brown. At the end of five or six weeks the colonies are still quite small, well defined, and opaque.

Pathogenesis.—Pure cultures injected into the mammary gland of cows and goats gave rise to a mastitis resembling in its development that from

which the streptococcus was obtained in the first instance. Injections into the cavity of the abdomen or into a vein, of one cubic centimetre of a pure culture, gave a negative result in dogs, cats, rabbits, and guinea-pigs.

32. DIPLOCOCCUS OF PNEUMONIA IN HORSES.

Obtained by Schütz (1887) from the lungs of horses affected with pneumonia.

Morphology.—Oval cocci, usually in pairs, surrounded by a homogeneous, transparent capsule.

Does not stain by Gram's method.

Biological Characters.—An *aërobic, non-liquefying* micrococcus. Grows at the room temperature. Upon *gelatin plates* forms small, spherical, white colonies.

In *gelatin stick cultures* grows along the line of puncture as small, white, separate colonies, which grow larger without becoming confluent. Upon the surface of *agar* small transparent drops are developed along the *impristich*.

Pathogenesis.—The injection of a pure culture into the lung of a horse produces pneumonia and causes its death in eight or nine days. Pathogenic for rabbits, guinea-pigs, and mice.

33. STREPTOCOCCUS CORYZÆ CONTAGIOSÆ EQUORUM.

Obtained by Shütz (1888) from pus from the lymphatic glands involved in horses suffering from the disease known in Germany as *Druse des Pferdes*.

Morphology.—Oval cocci, in pairs, in chains containing three or four elements, or in long chaplets.

Stains with the usual aniline colors—very intensely with Weigert's or Ehrlich's solution.

Biological Characters.—An *aërobic* and *facultative anaërobic* micrococcus. Grows slowly at the room temperature, more rapidly at 37° C. Upon *gelatin plates* at the end of three to five days minute colonies become visible; these never exceed the size of a pin's head. In *gelatin stick cultures* growth upon the surface is scanty or absent; along the line of puncture minute colonies are developed in rows. Upon *agar plates*, at 37° C., at the end of twenty-four hours lentil-shaped colonies are developed the size of a pin's head; under a low power the superficial colonies are seen to have a well-defined, opaque nucleus surrounded by a grayish, transparent marginal zone, which represents a half-fluid, slimy growth which does not extend after the third day and later disappears entirely; the deep colonies are at first well-defined, and later surrounded by wing-like outgrowths. Upon *blood serum*, at 37° C., yellowish, transparent drops are first developed; these become confluent and form a viscid and tolerably thick layer; this later becomes dry and iridescent.

Pathogenesis.—Pathogenic for horses and for mice, producing in these animals an abscess at the point of inoculation, and metastatic abscesses in the neighboring lymphatic glands. Not pathogenic for rabbits, guinea-pigs, or pigeons.

34. HÆMATOCOCCUS BOVIS (Babes).

Obtained by Babes (1889) from the blood and various organs of cattle which had died of an epidemic malady (in Roumania) characterized by hæmoglobinuria. The cocci are found in the blood in great numbers, for the most part enclosed in the red corpuscles.

Morphology.—Biscuit-shaped cocci united in pairs; sometimes oblong in form, isolated or united in groups; the free cocci are surrounded by a pale-yellowish, shining aureole of 0.5 to 1 μ in diameter.

Stains best with Löffler's solution of methylene blue; does not stain by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows very slowly at the room temperature—not below 20° C. In the incubating oven grows in the usual culture media. In *gelatin stick cultures* a scanty development of small, white colonies occurs along the line of puncture. Upon the *surface of agar* small, transparent drops are developed along the impfstrich. Upon *potato*, at 37° C. a thin, broad, yellowish, shining layer is developed in the course of a few days—scarcely visible. Upon *blood serum* small, moist, transparent colonies are developed.

Pathogenesis.—Pathogenic for rabbits and rats, which die in from six to ten days after inoculation with a pure culture; the spleen is found to be enlarged, the lungs hyperæmic, and a bloody serum is found in the cavity of the abdomen; the cocci are present in the blood in considerable numbers, but are rarely seen in the red corpuscles. Inoculations in oxen, horses, goats, sheep, guinea-pigs, and birds were without effect.

35. MICROCOCCUS GINGIVÆ PYOGENES.

Obtained by Miller (1889) from the mouth of a man suffering from alveolar abscess.

Morphology.—Large cocci of irregular dimensions, solitary or in pairs.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows at the room temperature in the usual media. Upon *gelatin plates* it forms spherical, well-defined colonies, which under a low power are at first slightly colored and later opaque. In *gelatin stick cultures* an abundant development occurs both upon the surface and along the line of puncture. Upon the surface of agar a tolerably thick, grayish growth occurs along the impfstrich, which has a purplish tint by transmitted light.

Pathogenesis.—Subcutaneous injections in mice produce a local abscess and necrosis of the skin, followed sometimes by death. Injections into the cavity of the abdomen produced peritonitis and death in from twelve to twenty-four hours.

36. PSEUDODIPLOCOCCUS PNEUMONIÆ.

Obtained by Bonome (1888) from the sero-fibrinous exudate in an autopsy of an individual who died of cerebro-spinal meningitis.

Morphology.—Oval cocci, in pairs or in chains of five or six elements, often surrounded by a transparent capsule; not to be distinguished from *Micrococcus pneumoniae crouposæ*.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature (*Micrococcus pneumoniae crouposæ* does not grow at the room temperature). In *gelatin stick cultures* very small colonies are developed along the line of puncture at the end of twenty-four to twenty-eight hours. Upon the *surface of agar* a rather scanty, moist layer is developed along the impfstrich. Upon *potato* a thin, scarcely visible layer is developed. In *bouillon* the development is abundant; the culture medium acquires a very acid reaction and gives off a strong odor like that of perspiration.

Pathogenesis.—Pathogenic for mice, guinea-pigs, and rabbits, in which animals it produces fatal septicæmia; the spleen is not enlarged, as is the case in animals inoculated with *Micrococcus pneumoniae crouposæ*.

37. STREPTOCOCCUS SEPTICUS LIQUEFACIENS.

Obtained by Babes (1889) from the blood and various organs of a child which died of septicæmia following scarlatina.

Morphology.—Micrococci, about 0.3 to $0.4\ \mu$ in diameter, in pairs or short chains in which the elements are loosely connected.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic, liquefying* micrococcus. Grows in the usual culture media at the room temperature. In *gelatin sticks* at the end of twenty-four hours a thin, granular, whitish stripe is seen along the line of puncture, while the surface seems somewhat depressed. Liquefaction of the gelatin occurs in funnel form; the liquefied gelatin is slightly clouded, and upon the walls of the funnel peculiar, flat, wheel-shaped, jagged colonies are seen. Upon the *surface of agar*, at 36°C , small, white, twin, shining, transparent colonies are developed, which measure a diameter of two to three millimetres. Upon *blood serum* a scarcely perceptible granular layer is developed.

Pathogenesis.—Subcutaneous injections in mice and rabbits produce a local inflammation with œdema, and death occurs in about six days. Streptococci are found in large numbers in the effused serum, in the lungs and in the spleen. After being cultivated for some time in artificial media the cultures lose their pathogenic power.

38. MICROCOCCUS OF KIRCHNER.

Obtained by Kirchner (1890) from the bronchial secretions (in sputum) of patients suffering from epidemic influenza—soldiers in garrison at Tientsin.

Morphology.—Spherical cocci, usually associated in pairs, and surrounded by a capsule. Distinguished from *Micrococcus pneumoniae* crouping by being smaller, quite spherical, and the elements in a pair being more easily separated from each other. Found in the bronchial secretion in pairs, or associated in groups; occasionally seen in chains.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* micrococcus; does not grow in peptone-gelatin at the room temperature. Upon *agar plates*, at 37°C , small, grayish-white, transparent, spherical colonies are developed. In later form round, grayish-white plaques. In *agar stick cultures* the abundant development occurs upon the surface, extending to the walls of the test tube; growth also occurs along the line of puncture.

Pathogenesis.—Not pathogenic for rabbits or for white mice. A guinea-pig which received one cubic centimetre of a bouillon culture in the abdominal cavity died at the end of twenty-four hours; the spleen was not enlarged, the lungs hyperæmic; the micrococci were found in the blood and in other organs. Another guinea-pig, which received one cubic centimetre of bouillon culture in the cavity of the abdomen, recovered after a slight inflammation.

39. MICROCOCCUS NO. II. OF FISCHEL.

Obtained by Fischel (1891) from the blood of two cases of influenza.

Morphology.—Micrococci of from 1 to $1.25\ \mu$ in diameter, in pairs, sometimes in chains.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* minute colonies, visible only under the microscope, are developed at the end of three days. In *gelatin stick cultures* abundant milk-white growth occurs along the line of puncture, and liquefaction of the gelatin commences at the end of four days; this process is slow. Upon *agar plates*, at 37°C ., superficial colonies are developed, resembling a drop of milk. Upon *potato*, at 37°C ., at the end of eight days a thin, shining layer of a yellowish-white color, and about one centimetre broad, is developed; no growth upon potato at the room temperature. Growth occurs in liquid blood serum or in milk. In sterilized whey the micrococcus is said by Fischel to lose its vitality in eight hours.

Pathogenesis.—Pathogenic for dogs and for horses. Intravenous injection of three to four cubic centimetres in dogs is said to produce symptoms resembling those of distemper in this animal, viz., increased temperature, catarrhal conjunctivitis, in some cases keratitis, and in some a mucous discharge from the preputial sac. The micrococcus was not found in the blood of the dogs inoculated by intravenous injection, later than the fourth day.

40. STREPTOCOCCUS OF BONOME.

Obtained by Bonome (1890) from the exudations of the cerebro-spinal meninges and from hæmorrhagic extravasations in the lungs in cases of epidemic cerebro-spinal meningitis.

This streptococcus is said by Bonome to be distinguished from previously known streptococci by the following characters: It does not grow readily in artificial culture media, and soon loses its pathogenic power when preserved in a desiccated condition or cultivated through a few successive generations. It differs from the "pneumococcus" and "meningococcus" by the ball-shaped appearance of its colonies on agar plates, and in the fact that it does not grow upon blood serum; also by the difficulty experienced in carrying it through five or six generations in artificial media.

Pathogenesis.—In white mice and in rabbits a fibrinous inflammation and death result from inoculations with a pure culture, the symptoms resembling those produced by similar inoculations with *Micrococcus pneumoniae crouposæ*. It does not produce septicæmia in white mice, but in rabbits the cocci are found in the blood in chains surrounded by a capsule. In guinea-pigs and dogs a local fibrinous inflammation results from inoculations, and the streptococcus is found in the gelatinous exudate at the point of inoculation. It is distinguished from the streptococcus of erysipelas by its failure to grow in gelatin or in blood serum, and by the appearance of its colonies on agar plates.

41. MICROCOCCUS OF ALMQUIST.

Obtained by Almquist (1891) from the bullæ of pemphigus neonatorum, in nine children suffering from this disease during an epidemic which occurred at Göteborg.

Morphology.—Micrococci from 0.5 to 1 μ in diameter, usually in pairs.

Stains readily with the aniline colors.

Biological Characters.—An *aërobic*, *liquefying*, *chromogenic* micrococcus. Closely resembles *Staphylococcus pyogenes aureus* in its morphology and growth in culture media. Produces a similar golden-yellow pigment.

Pathogenesis.—According to Almquist, this micrococcus is distinguished from *Staphylococcus pyogenes aureus* by its specific pathogenic power. Two inoculations made from a pure culture, by means of a lancet, upon his own arm gave rise to a development of bullæ like those of pemphigus. The process showed no disposition to extend deeper, but the epidermis was raised by a collection of fluid which was at first transparent and later had a milky opacity. From the contents of these bullæ the same coccus was obtained in pure cultures.

42. STAPHYLOCOCCUS PYOSEPTICUS.

Obtained by Hericourt and Richet (1888) from an abscess in the skin of a dog.

In its morphology and biological characters this micrococcus closely resembles *Staphylococcus pyogenes albus*, and it is probably a pathogenic variety of this common species. But the experiments made by the authors referred to show it to be decidedly more pathogenic for rabbits. Subcutaneous injections of a drop or two of a pure culture caused an extensive inflammatory oedema, and death in from twelve to twenty-four hours.

43. *STREPTOCOCCUS PERNICIOSUS PSITTACORUM*.

Micrococcus of gray parrot disease. Eberth and Wolff have described an infectious disease of gray parrots, which is said to be extremely common among the imported birds. The disease is characterized by the formation of nodules upon the surface and in the interior of various organs, and especially in the liver. Micrococci of medium size are found in these nodules and in the blood from the heart; these are sometimes in chains. Microscopic examination of stained sections shows that these cocci are directly related to the tissue necrosis which characterizes the disease. But the micrococcus has not been cultivated and its biological characters are undetermined.

44. *MICROCOCCUS OF FORBES*.

Forbes (1886) has studied an infectious disease of cabbage caterpillars (*Pieris rapæ*), which appears to be due to a micrococcus found by him in large numbers in the bodies of the infected larvæ. This micrococcus, which resembles the common staphylococci in form, was cultivated in liquid media, and successful inoculation experiments were made.

VII.

THE BACILLUS OF ANTHRAX.

[*Fr.*, CHARBON ; *Ger.*, MILZBRAND.]

ANTHRAX is a fatal infectious disease which prevails extensively among sheep and cattle in various parts of the world, causing heavy losses. In Siberia it constitutes a veritable scourge and is known there as the Siberian plague ; it also prevails to a considerable extent in portions of France, Hungary, Germany, Persia, and India, and local epidemics have occasionally occurred in England, where it is known under the name of splenic fever. It does not prevail in the United States. In infected districts the greatest losses are incurred during the summer season.

In man accidental inoculation may occur among those who come in contact with infected animals, and especially during the removal of the skin and cutting-up of dead animals, when there is any cut or abrasion upon the hands. A malignant pustule is developed as the result of such inoculation, but, as a rule, general infection does not occur, as is the case when inoculations are made into the more susceptible lower animals—rabbit, guinea-pig, mouse. Those who handle the hair, hides, or wool of infected animals are also liable to contract the disease by inoculation through open wounds, or by the inhalation of dust containing spores of the anthrax bacillus. Cases of pulmonic anthrax, known formerly in England as “wool-sorters’ disease,” have been occasionally observed in England and in Germany, and are now recognized as being due to infection through the lungs in the manner indicated.

The French physician Davaine, who had observed the anthrax bacillus in the blood of infected animals in 1850, communicated to the French Academy of Sciences the results of his inoculation experiments in 1863 and 1864, and asserted the etiological relation of the bacillus to the disease with which his investigations showed it to be constantly associated. This conclusion was vigorously contested by conservative opponents, but has been fully established by subsequent investigations, which show that the bacillus, in pure cultures, induces

anthrax in susceptible animals as certainly as does the blood of an animal recently dead from the disease.

Owing to the fact that this was the first pathogenic bacillus cultivated in artificial media, and to the facility with which it grows in various media, it has served more than any other microörganism in researches relating to a variety of questions in pathology, physiology, biology, and public hygiene, some of which are discussed in various sections of this volume.

45. *BACILLUS ANTHRACIS.*

Synonyms.—Milzbrandbacillus, *Ger.*; Bactéridie du charbon, *Fr.*

First observed in the blood of infected animals by Pollender (1847) and by Davaine (1850). Etiological relation affirmed by Pasteur

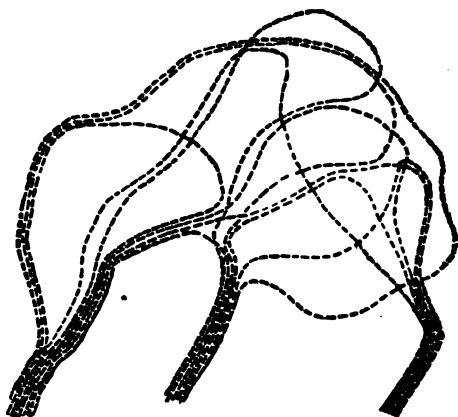


FIG. 100.—*Bacillus anthracis*, from a culture, showing development of long thread-like bundles. $\times 300$. (Klein.)

(1863), and established by the inoculation of pure cultures by Pasteur (1879) and by many other investigators.

Morphology.—Rod-shaped bacteria having a breadth of 1.25μ , and 5 to 20μ in length; or, in suitable culture media, grow out into long, flexible filaments, which are frequently united into twisted, cord-like bundles. These filaments in hanging-drop cultures, before the development of spores, appear to be homogeneous or the protoplasm is clouded and granular, but without distinct segmentation. But in stained preparations the filaments are seen to be made up of a series of rectangular, deeply stained segments. In hanging-drop cultures the ends of the rods appear rounded. In stained preparations from the blood of an infected animal the filaments are seen to present a slight concavity, and a lenticular interspace is formed where two rods come together. The diameter of the

varies considerably in different culture media; and in old cultures irregular forms are frequently seen—"involution forms."

Under favorable conditions endogenous spores are developed in the long filaments which grow out in artificial culture media. These first appear as refractive granules distributed at regular intervals in the segments of the protoplasm, which gradually disappear as the spores are developed; and these are left as oval, highly refractive bodies, held together in a linear series by the cellular envelope, and subsequently set free by its dissolution. The germination of these reproductive bodies results in the formation of rods and spore-bearing filaments like those heretofore described. In this process the spore is first observed to lose its brilliancy, from the absorption of moisture, a prominence occurs at one end of the oval body, and soon the external envelope—exosporium—is ruptured, permitting the softened protoplasmic contents enclosed in the internal spore membrane—endosporium—to escape as a short rod, to which the empty exosporium sometimes remains attached.

The anthrax bacillus stains readily with the aniline colors and also by Gram's method, when not left too long in the decolorizing iodine solution. Löffler's solution of methylene blue is an especially good staining fluid for this as well as for many other bacilli. Bismarck brown is well adapted for specimens which are to be photographed, and also for permanent preparations, as it is less liable to fade than the blue and some other aniline colors.

Biological Characters.—The anthrax bacillus is *aërobic*, but not strictly so, as is shown by the fact that it grows to the bottom of the line of puncture in stick cultures in solid media. It is *non-motile*, and is distinguished by this character from certain common bacilli resembling it in morphology—*Bacillus subtilis*—which were frequently confounded with it in the earlier days of bacteriological investigation.

The anthrax bacillus grows in a variety of nutrient media at a



FIG. 101.—*Bacillus anthracis*, from a culture, showing formation of spores. $\times 1,000$. (Klein.)

temperature of 20° to 38° C. Development ceases at temperature below 12° C. or above 45° C.

This bacillus grows best in neutral or slightly alkaline medium; its development is arrested by a decidedly acid reaction of the culture medium. It may be cultivated in infusions of flesh or of various vegetables, in diluted urine, in milk, etc.

In *gelatin plate cultures* small, white, opaque colonies are developed in from twenty-four to thirty-six hours, which under the microscope are seen to be somewhat irregular in outline and of a pinkish tint; later the colonies spread out upon the surface of the medium, and the darker central portion is surrounded by a brownish wavy filaments, which are associated in tangled bundles. M

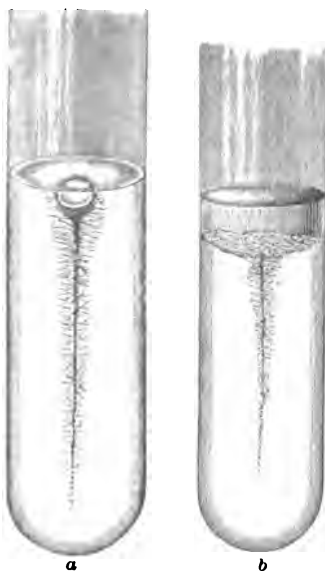


FIG. 102.—Culture of *Bacillus anthracis* in nutrient gelatin: *a*, end of four days; *b*, end of eight days. (Baumgarten.)

like outgrowths from the periphery of the colony may often be seen extending into the surrounding gelatin. At the end of two or three days liquefaction of the gelatin commences, and the central portion is soon surrounded by the liquefied medium, upon the surface of which a white pellicle appears as an irregular white pellicle.

In *stick cultures* growth occurs along the line of puncture as a white, branching thread, from which lateral branches like ramifications extend into the surrounding medium. At the end of two or three days liquefaction of the culture commences near the surface, where development has been most abundant. At first a pasty, white mass is formed, but as liquefaction progresses the upper part of the liquefied gelatin becomes transparent from the subsidence of the motionless bacilli, and these are seen upon the surface of the non-liquefied portion of the medium in the

cloudy, white masses, while below the line of liquefaction the characteristic branching growth may still be seen along the line of puncture.

In *agar plate cultures*, in the incubating oven at 35° to 37° C. colonies are developed within twenty-four hours, which under the microscope are seen to be made up of interlaced filaments, which are very characteristic and beautiful. Upon the surface of nutrient agar a grayish-white layer is formed, which may be removed in ribbons; and in stick cultures in this medium a branching growth is seen, like that in gelatin, but without liquefaction. The ad-

a small quantity of agar to a gelatin medium prevents liquefaction of the gelatin (Flügge).

Upon *blood serum* a rather thick, white layer is formed and liquefaction slowly occurs.

Upon *potato* the growth is abundant as a rather dry, grayish-white layer, of limited extent, having a somewhat rough surface and irregular margins.

Spores are formed only in the free presence of oxygen, as in surface cultures upon potato or nutrient agar, or in shallow cultures in liquid media, and at a temperature of 20° to 35° C. They are not formed during the development of the bacilli in the bodies of living

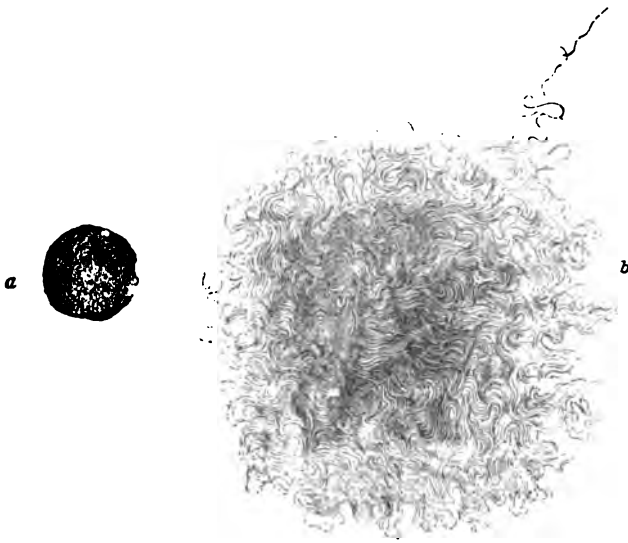


FIG. 103.—Colonies of *Bacillus anthracis* upon gelatin plates: *a*, at end of twenty-four hours; *b*, at end of forty-eight hours. $\times 80$. (Flügge.)

animals, but after the death of the animal the bacillus continues to multiply for a time, and spores may be formed where the fluids containing it come in contact with the air—as, for example, in bloody discharges from the nostrils or from the bowels of the dead animal.

Varieties incapable of spore production have been produced artificially, by several bacteriologists, by cultivating the bacillus under unfavorable conditions. Roux was able to produce a sporeless variety by successive cultivation in media containing a small quantity of carbolic acid—1 : 1,000.

Varieties differing in their pathogenic power may also be produced by cultivation under unfavorable conditions. Thus Pasteur

produced an "attenuated virus" by keeping his cultures for considerable time before replanting them upon fresh soil, and the effect was due to the action of atmospheric oxygen. It is probable that it was rather due to the deleterious action of the products of growth present in the culture media. It has been shown by Chamberlain and Roux that cultivation in the presence of certain chemical substances added to the culture medium—bichromate of potassium 0.01 per cent—causes an attenuation of virulence. The same result occurs when cultures are subjected to a temperature a little below that which is fatal to the bacillus—for eighteen minutes (Chauveau); 42.5° C. for two or three minutes (Koch). Attenuation of pathogenic virulence is also effected by cultivation in the body of a non-susceptible animal, like the frog (Behring, Petruschky); or in the blood of a rat (Behring); by exposure to sunlight (Arloing); and by compressed air (Chauveau).

Anthrax spores may be preserved in a desiccated condition for years without losing their vitality or pathogenic virulence when inoculated into susceptible animals. They also resist a comparatively high temperature. Thus Koch and Wolffhügel found that dry spores exposed in dry air required a temperature of 140° C., maintained for three hours, to insure their destruction. But spores suspended in liquid are destroyed in four minutes by the boiling temperature 100° C. (writer's determination).

The bacilli, in the absence of spores, according to Chauveau, are destroyed in ten minutes by a temperature of 54° C.

For the action of various antiseptic and germicidal agents upon this bacillus we must refer to the sections especially devoted to this subject (Part Second).

Toussaint, by injecting filtered anthrax blood into animals, obtained evidence that it contained some toxic substance which in his experiments gave rise to local inflammation without any noticeable general symptoms. More recent investigations show that a poisonous substance (Hankin) is formed during the growth of the anthrax bacillus, and that cultures containing this *toxalbumin*, in which the bacilli have been removed by filtration through porcelain, produce immunity when injected into susceptible animals, similar to that resulting from inoculations with an attenuated virus. It is probable that the pathogenic power of the anthrax bacillus depends largely upon the presence of this toxalbumin, and that the essential difference between virulent and attenuated varieties depends upon the more abundant production of this toxic substance by the virulent. It has also been shown that virulent cultures produce a larger quantity of acid than those which have been attenuated by any of the agencies above mentioned (Behring).

Martin (1890) has studied the chemical products in filtered cultures of the anthrax bacillus and obtained the following results:

1. Protoalbumose, deutoalbumose, and a trace of peptone. The mixed albumoses were found not to be poisonous except in considerable doses—0.3 grammé injected subcutaneously killed a mouse weighing twenty-two grammes; smaller doses produced a local œdema. A fatal dose caused extensive œdema, coma, and death in twenty-four hours; the spleen was sometimes enlarged. Boiling neutralizes to a considerable extent the toxic power.

2. An alkaloid, soluble in water and in alcohol, but insoluble in benzol, chloroform, or ether. The solutions have a strongly alkaline reaction, and crystalline salts are formed with various acids. This alkaloid is somewhat volatile, and when exposed to light loses to a considerable extent its toxic properties. It produces symptoms similar to those resulting from inoculations with the albumoses, but is more toxic and more prompt in its action. The animal quickly falls into a state of coma; there is extensive œdema about the point of inoculation, and the spleen is usually enlarged. The fatal dose for a mouse weighing twenty-two grammes is from 0.1 to 0.15 grammé; death occurs within two or three hours.

3. In addition to these toxic substances small quantities of leucin and of tyrosin were found in the filtered cultures.

Recently (1892) Petermann has made a series of experiments with filtered cultures of the anthrax bacillus which lead him to the conclusion that "large quantities of a culture in serum from the ox, filtered through porcelain, injected into the veins of a susceptible animal, have a preventive action; but the immunity thus conferred is transitory, not lasting longer than a month or two."

Pathogenesis.—The anthrax bacillus is pathogenic for cattle, sheep, horses, rabbits, guinea-pigs, and mice. White rats, dogs, and frogs are immune, as is also the Algerian race of sheep. The sparrow is susceptible to general infection, but chickens, under normal conditions, are not. Young animals are, as a rule, more susceptible than adults of the same species. Man does not belong among the most susceptible animals, but is subject to local infection as a result of accidental inoculation—malignant pustule—and to pulmonic anthrax from breathing air, containing spores of the anthrax bacillus, during the sorting of wool or hair from infected animals. In animals which have a partial immunity, natural or acquired, as a result of inoculations with attenuated virus, the subcutaneous introduction of virulent cultures may give rise to a limited local inflammatory process, with effusion of bloody serum in which the bacillus is found in considerable numbers; but the blood is not invaded, and the animal, after some slight symptoms of indisposition, recovers. In susceptible

animals injections beneath the skin or into a vein give rise to general infection, and the bacilli multiply rapidly in the circulating fluid. Death occurs in mice within twenty-four hours, and in rabbits, as a rule, in less than forty-eight hours. The blood of the heart and large vessels may be found, in an autopsy made immediately after death, to contain comparatively few bacilli ; but in the capillaries of the various organs, and especially in the greatly enlarged spleen, in the liver, the kidneys, and the lungs, they will be found in great numbers, and well-stained sections of these organs will give an astonishing picture under the microscope, which the student should not fail to see in preparations made by himself. The capillaries in many places will be found stuffed full of bacilli ; or they may even be ruptured as a result of the distention, and the bacilli, together with

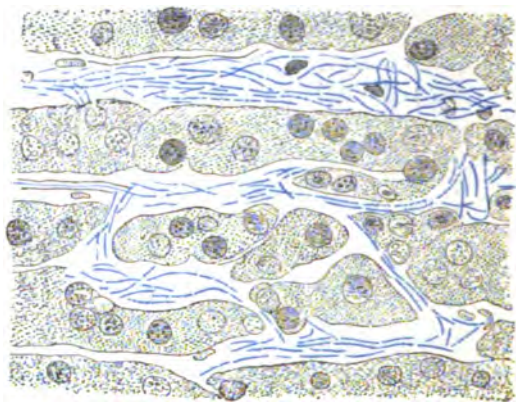


FIG. 104.—*Bacillus anthracis* in liver of mouse. $\times 700$. (Flügge.)

escaped blood corpuscles, will be seen in the surrounding tissues. In the kidneys the glomeruli, especially, appear as if injected with colored threads, and by rupture these may find their way into the uriferous tubules.

These appearances and the general symptoms indicate that the disease produced by the introduction of this bacillus into the bodies of susceptible animals is a genuine septicæmia. As in other forms of septicæmia, the spleen is found to be greatly enlarged ; it has a dark color and is soft and friable. With this exception the organs present no notable changes, although the liver is apt to be somewhat enlarged. In the guinea-pig an extensive inflammatory œdema, extending from the point of inoculation to the most dependent parts of the body, is developed ; the subcutaneous connective tissue is infiltrated with bloody serum and has a gelatinous appearance. This animal comes next to the mouse in susceptibility, and cultures which

are attenuated to such an extent that they will not kill a rabbit or a sheep may still kill a guinea-pig ; or, if not, may kill a mouse. Pasteur has shown that the pathogenic power of the bacillus may be reëstablished by inoculations into susceptible animals, and that an attenuated culture which will not kill an adult guinea-pig may be fatal to a very young animal of this species, and that cultures from the blood of this will have an increased pathogenic virulence.

Very minute quantities of a virulent culture are infallibly fatal to these most susceptible animals, but for rabbits and other less susceptible animals the quantity injected influences the result, and recovery may occur after subcutaneous or intravenous injection of a very small number of bacilli.

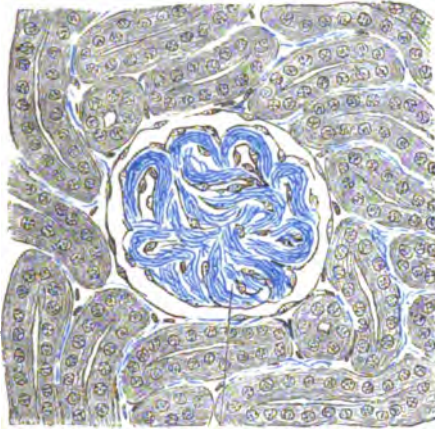


FIG. 105.—*Bacillus anthracis* in kidney of rabbit. $\times 400$. (Baumgarten.)

Infection in cattle and sheep commonly results from the ingestion of spores while grazing in infected pastures. The bacillus itself, in the absence of spores, is destroyed in the stomach. While spores are not formed in the bodies of living animals, their discharges contain the bacillus, and this is able to multiply in them and to form spores upon the surface of the ground when temperature conditions are favorable. It is probable that this is the usual way in which pastures become infected, and that the bloody discharges from the bladder and bowels of animals suffering from the disease furnish a nidus for the external development of these reproductive elements ; as also do the fluids escaping from the bodies of dead animals. And possibly, under specially favorable conditions, the bacillus may lead a saprophytic existence for a considerable time in the superficial layers of the soil.

Buchner has shown by experiment that infection in animals result from respiring air in which anthrax spores are in suspension in the form of dust ; and in man this mode of infection occurs in the so-called wool-sorters' disease.

The question of the passage of the anthrax bacillus from mother to the foetus in pregnant females has received considerable attention. That this may occur is now generally admitted, and appears to be established by the investigations of Strauss and Calkin, Morisani, and others. That it does not always occur is shown, however, by the researches of other bacteriologists, and especially those of Wolff.

VIII.

THE BACILLUS OF TYPHOID FEVER.

RECENT researches support the view that the bacillus described by Eberth in 1880 bears an etiological relation to typhoid fever—*typhus abdominalis* of German authors; and pathologists are disposed to accept this bacillus as the veritable “germ” of typhoid fever, notwithstanding the fact that the final proof that such is the case is still wanting.

This final proof would consist in the production in man or in one of the lower animals of the specific morbid phenomena which characterize the disease in question, by the introduction of pure cultures of the bacillus into the body of a healthy individual. Evidently it is impracticable to make the test upon man, and thus far we have no satisfactory evidence that any one of the lower animals is subject to the disease as it manifests itself in man. The experiments of Fränkel and Simmonds show, however, that this bacillus is pathogenic for the mouse and the rabbit. We shall refer to the experiments of these authors later.

Before the publication of Eberth's first paper Koch had observed this bacillus in sections made from the spleen and liver of typhoid cases, and had made photomicrographs from these sections. His name is, therefore, frequently associated with that of Eberth as one of the discoverers of the typhoid bacillus. Other investigators had no doubt previously observed the same organism, but some of them had improperly described it as a micrococcus. Such a mistake is easily made when the examination is made with a low power; even with a moderately high power the closely crowded colonies look like masses of micrococci, and it is only by focussing carefully upon the scattered organisms on the outer margin of a colony that the oval or rod-like form can be recognized.

Several observers had noted the presence of microorganisms in the lesions of typhoid fever prior to the publication of Eberth's paper, and Browicz in 1875, and Fischel in 1878, had recognized the presence of oval organisms in the spleen which were probably identical with the bacillus of Eberth.

The researches of Gaffky (1884) strongly support the view that

the bacillus under consideration bears a causal relation to fever. Eberth was only successful in finding the bacillus in lymphatic glands or in the spleen in eighteen cases out of which he searched for it. On the other hand, he failed to find it in eleven cases of various nature—partly infectious processes—thirteen cases of tuberculosis in which the lymphatic glands were involved, and in several of which there was ulceration of the mucous membrane of the intestine.

Koch, independently of Eberth and before the publication of his first paper, had found the same bacillus in about half of the cases examined by him, and had pointed out the fact that they were located in the deeper parts of the intestinal mucous membrane, at the limits of necrotic changes, and also in the spleen, where the long, slender bacillus of Klebs was found only in the necrotic portions of the intestinal mucous membrane.

The researches of W. Meyer (1881) gave a larger proportion of successful results. This author confined his attention chiefly to the swollen plaques of Peyer and follicles of the intestine which had not yet undergone ulceration. The short bacillus which had been described by Eberth and Koch was found in sixteen out of twenty cases examined. The observations of this author are in accord with those of Eberth as to the presence of the bacillus in greater abundance in the cases of typhoid which had proved fatal at an early date.

The fact that in these earlier researches the bacilli were not found in a considerable proportion of the cases examined is by no means fatal to the view that they bear an etiological relation to the disease. As Gaffky says in his paper referred to :

“This circumstance admits of two explanations. Either in some cases in which the bacillus has been sought with negative results they may have perished collectively, before the disease process which they had induced had run its course ; or the proof of the presence of the bacilli was wanting only on account of the technical difficulties which attend the finding of isolated colonies.”

Gaffky's own researches indicate that the latter explanation is the correct one.

In twenty-eight cases examined by this author characteristic colonies of the bacillus were found in all but two. In one of these cases one hundred and forty-six sections from the spleen, liver, and mesenteric glands were examined without finding a single colony, and in the other a like result attended the examination of sixty-two sections from the spleen and twenty-one sections from the liver. In the first of these cases, however, numerous colonies were found in recent ulcerations of the intestinal mucous membrane, deeply located in that portion of the tissue which was still intact. These recent ulcers were in the

borhood of old ulcers and are supposed to have indicated a relapse of the specific process. In the second case the negative result is thought by Gaffky to have been not at all surprising, as the patient died at the end of the fourth week of sickness, not directly from the typhoid process, but as a result of perforation of the intestine.

Gaffky has further shown that in those cases in which colonies are not found in the spleen, or in which they are extremely rare, the presence of the bacillus may be demonstrated by cultivation; and that, when proper precautions are taken, pure cultures of the bacillus may always be obtained from the spleen of a typhoid case. Hein has been able to demonstrate the presence of the bacillus and to start pure cultures from material drawn from the spleen of a living patient by means of a hypodermatic syringe. Philopowicz has reported his success in obtaining cultures of the bacillus by the same method.

The fact that a failure to demonstrate the presence of microorganisms by a microscopic examination cannot be taken as proof of their absence from an organ, is well illustrated by a case (No. 18) in which the bacillus was obtained by Gaffky from the spleen and also from the liver, in pure cultures; whereas in cover-glass preparations made from the same spleen he failed to find a single rod, and more than one hundred sections of the spleen were examined before he found a colony.

To obtain pure cultures from the spleen Gaffky first carefully washes the organ with a solution of mercuric chloride, 1:1,000. A long incision is then made through the capsule with a knife sterilized by heat. A second incision is made in this with a second sterilized knife, and a third knife is used to make a still deeper incision in the same track. By this means the danger of conveying organisms from the surface to the interior of the organ is avoided. From the bottom of this incision a little of the soft splenic tissue is taken up on a sterilized platinum needle, and this is plunged into the solid culture medium, or drawn along the surface of the same, or added to liquefied gelatin and poured upon a glass plate. The colonies develop, in an incubating oven, in the course of twenty-four to forty-eight hours.

Gaffky has also shown that the bacillus is present in the liver, in the mesenteric glands, and, in a certain proportion of cases at least, in the kidneys, in which it was found in three cases out of seven.

The appearance of the colonies in stained sections of the spleen is shown in Figs. 106 and 107. Two colonies are seen in Fig. 106 (at *a, a*) as they appear under a low power—about sixty diameters. In Fig. 107 one of the colonies is seen more highly magnified—about five hundred diameters.

Fränkel and Simmonds have demonstrated that the bacilli multi-

ply in the spleen after death, and that numerous colonies were found in portions of the organ which have been kept for twenty-four to forty-eight hours before they were placed in alcohol. Other pieces from the same spleen placed in alcohol soon after the death of the patient show but few colonies or none at all.

This observation does not in any way weaken the evidence of the etiological rôle of the bacillus, but simply shows that dead animal matter is a suitable nidus for the typhoid germ—a fact which has been repeatedly demonstrated by epidemiologists and inculcated upon by sanitarians.

The authors last referred to confirm Gaffky as regards the constant presence of the bacillus in the spleen. In twenty-nine cases they obtained it by plate cultures twenty-five times, and in four that in the four cases attended with a negative result this re-

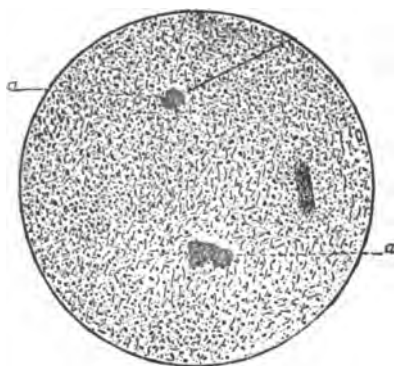


FIG. 106.



FIG. 107.

not at all surprising, inasmuch as the typhoid process had advanced and death resulted from complications.

Gaffky did not succeed in obtaining cultures from the blood of typhoid-fever patients, and concludes from his researches that if bacilli are present in the circulating fluid it must be in very small numbers. He remarks that possibly the result would be different if the blood were drawn directly from a vein instead of from the capillaries of the skin. Fränkel and Simmonds also report that glycerine to which blood drawn from the forefinger of typical cases had been added, remained sterile when poured upon plates in the usual manner—Koch's method. The blood was obtained from six different individuals, all in an early stage of the disease—the second or third week. A similar experiment made with blood obtained from a mortem, from the large veins or from the heart, also gave a negative result in every instance save one. In the exceptional case a

colony developed upon the plate. In view of these results we are inclined to attribute the successful attempts reported by some of the earlier experimenters (Letzerich, Almquist, Maragliano) to accidental contamination and imperfect methods of research. The more recent work of Tayon does not inspire any greater confidence. This author obtained cultures in bouillon by inoculating it with blood drawn from a typhoid patient, and found that these were fatal, *in a few hours*, to guinea-pigs, when injected into the peritoneal cavity. The lesions observed are said to have resembled those of typhoid fever—congestion and tumefaction of Peyer's plaques and of the mesenteric glands, congestion of the liver, the kidneys, etc.

The presence of the bacillus of Eberth in the alvine evacuations of typhoid patients has been demonstrated by Pfeiffer and by Fränkel and Simmonds. This demonstration is evidently not an easy matter, for while the bacilli are probably always present in some portion of the intestine during the progress of the disease, it does not follow that they are present in every portion of the intestinal contents. As only a very small amount of material is used in making plate cultures, and as there are at all times a multitude of bacteria of various species in the smallest portion of fæcal matter, it is not to be expected that the typhoid bacillus will be found upon every plate. Fränkel and Simmonds made eleven attempts to obtain the bacillus by the plate method, using three plates each time, as is customary with those who adhere strictly to the directions of the master, and were successful in obtaining the bacillus in three instances—in two in great numbers and in the third in a very limited number of colonies.

The numerous attempts which have been made to communicate typhoid fever to the lower animals have given a negative result in every instance. Murchison, in 1867, fed typhoid-fever discharges to swine, and Klein has made numerous experiments of the same kind upon apes, dogs, cats, guinea-pigs, rabbits, and white mice, without result. Birch-Hirschfeld, in 1874, by feeding large quantities of typhoid stools to rabbits, produced in some of them symptoms which in some respects resembled those of typhoid; but these experiments were repeated by Bahrdrf upon ten rabbits with an entirely negative result. Von Motschukoffsky met with no better success in his attempts to induce the disease by injecting blood from typhoid patients into apes, rabbits, dogs, and cats. Walder also experimented with fresh and with putrid discharges from typhoid patients, and with blood taken from the body after death, feeding this material to calves, dogs, cats, rabbits, and fowls, without obtaining any positive results. Klebs has also made numerous experiments of a similar nature, and in a single instance found in a rabbit, which died

forty-seven hours after receiving a subcutaneous injection of culture fluid containing his "typhoid bacillus," pathological lesions resembling those of typhoid.

Eberth and Gaffky very properly decline to attach any importance to this solitary case, in which, as the first-named writer remarks, a different explanation is possible, and the possibility of intestinal mycosis not typhoid in its nature must be considered.

Gaffky has also made numerous attempts to induce symptoms in animals by means of pure cultures of Eberth's bacillus given with their food or injected into the peritoneal cavity or subcutaneously. The first experiments were made upon five Java monkeys. For a considerable time these animals were fed daily with pure cultures containing spores. The temperature of the animals was taken twice daily. The result was entirely negative. No better results attended the experiments upon rabbits (16), guinea-pigs (13), rats (7), house mice (11), field mice (4), pigeons (2), one hen and one duck.

Cornil and Babes report a similar negative result from pure cultures of the typhoid bacillus injected into the peritoneal cavity and into the duodenum in rabbits and guinea-pigs.

Fränkel and Simmonds have made an extended series of experiments upon guinea-pigs, rabbits, and mice, and have shown that pure cultures of the bacillus of Eberth injected into the lower mentioned animals—mice and rabbits—may induce death, and that the bacillus may again be obtained in pure cultures from their bodies. It is not claimed that the animals suffer an attack of typhoid fever as the result of these injections, but that their death is due to the introduction into their bodies of the typhoid bacillus, and that the bacillus is thereby proved to be pathogenic.

The failure to produce the characteristic lesions of typhoid fever in lower animals is evidently not opposed to the view that this bacillus is the specific cause of such lesions in man. Fränkel and Simmonds quote from Koch in support of this statement, as follows :

"In my opinion it is not at all necessary, when we experiment upon lower animals, to obtain precisely the same symptoms as in man. In support of my opinion I may refer to the infectious diseases which we are able to produce experimentally in the lower animals. Anthrax runs a very different course in animals and in man; tuberculosis does not present itself in precisely the same manner in one species of animals as in another. Phthisis, as it is called in man, we cannot, in general, produce in animals; and, nevertheless, we cannot assert that the animals experimented upon do not suffer from tuberculosis, and that the conclusions which we draw from such experiments are not perfectly correct."

In Fränkel and Simmonds' experiments a considerable quantity of material was used, and the injections were, for the most part, made into the peritoneal cavity in mice, or into the circulatory system in guinea-pigs.

through a vein in rabbits. The influence of quantity of material used is especially shown in the case of the mice, and the question arises whether the pathogenic power of the bacillus for these animals does not depend upon the simultaneous injection of the ptomaine developed in cultures as a result of the vital activity of the organism. Thus we read that mouse No. 4 resisted an injection of a dilute solution of culture No. 1, but succumbed to a more concentrated solution—one-fifth of a Pravaz syringe. Mouse No. 5 was not killed by the injection of one-third of a syringe of a dilute solution, but subsequently died from the injection of one-third of a syringe of a concentrated solution. Mouse No. 16, injected October 10th with half of a syringe of a very diluted culture, did not die. The injection was repeated on the 17th of October with half a syringe of a concentrated solution, with fatal result.

In all, thirty-five mice were injected, with a fatal result in twenty-seven cases. In rabbits the injections were commonly made in the large vein of the ear, and the quantity of material injected was considerably greater—from one-third the contents of a hypodermatic syringe to two syringefuls. In some instances death occurred within a few hours, in others on the following day or after an interval of two or three days. It is noticeable that the results differ very greatly as to the date of death and the relative quantity of material required to produce a fatal result. This probably depends to some extent upon the size of the animal, and perhaps partly upon individual differences in resisting power.

The experiments, considered together, show that the typhoid bacillus is not pathogenic for these animals in the same sense as is the anthrax bacillus or the bacillus of rabbit septicæmia. These organisms introduced beneath the skin or into the circulation in the smallest amount infallibly produce death, and at the expiration of a period of time which is tolerably uniform.

In all, seventy-nine experiments upon rabbits were made, with the following result: Five injections into the intestine, five into the subcutaneous connective tissue, one into the lung, and two inhalation experiments, *all without result*; twenty injections into the peritoneal cavity furnished two, and forty-six injections into the vein of the ear twenty positive results—*i.e.*, were fatal to the animal.

In the fatal cases the bacilli were proved to be present in the spleen by culture experiments and by microscopical examination of properly stained sections. The colonies were identical in appearance with those found in the spleen of cases of typhoid in man. Colonies were found in the spleens of the rabbits experimented upon exactly as in the human subject—sometimes in the trabeculæ, sometimes in the Malpighian bodies, sometimes free in the splenic pulp.

Brieger has made some very interesting researches with reference to the chemical substances which are produced as a result of physiological processes attending the growth of this bacillus.

Having obtained a culture from the spleen of a typhoid patient and assured himself by comparison with a pure culture given by Koch that he was dealing with the right organism, Brieger planted the bacillus in a culture solution containing grape sugar and salts—*Nährsalzen*—in which it thrived admirably. Such a solution at 30° C. became clouded at the end of twenty-four hours, and gave off an evident odor of ethyl alcohol, which increased from day to day. In addition to ethyl alcohol small quantities of the volatile fatty acids were produced—among them acetic acid. Lactic acid was formed from the grape sugar. The bacillus grew still better in buminous culture fluids. It did not in these give rise to the production of sulphuretted hydrogen or of any of the volatile products of putrefactive decomposition, such as indol and phenol. There was no gas formation in such cultures, even after standing for several weeks, but a slight odor, resembling that of whey, was given off from the cultures. Repeatedly, but not in every case, Brieger succeeded in obtaining from such cultures a very deliquescent product. This was obtained in only very small quantities, when the cultures had remained in the incubating oven for a month. The physiological properties of this base have convinced Brieger that it is a new ptomaine. In guinea-pigs this ptomaine produced a salivary flow of saliva and frequent respiration. Later the animals lost control of their extremities and of the muscles of the trunk; they lay upon their side, but when placed upon their legs were able to move forward a little; they, however, soon fell again and remained motionless upon their side. The pupils gradually became widely dilated and failed to respond to light; the flow of saliva became more profuse; no convulsions occurred. Little by little the pulsations of the heart and the breathing became more frequent. During the course of these symptoms the animals had frequent liquid discharges. Death occurred in from twenty-four to forty-eight hours. At mortem examination showed the heart to be contracted in systole, the lungs to be hyperæmic, the intestine contracted and pale.

The experimental evidence which we have presented, considered in connection with established facts relating to the propagation of typhoid fever, seems to the writer to be convincing as regards the etiological rôle of this bacillus.

No other organism has been found, after the most careful search in the deeper portions of the intestinal glands involved in this disease or in the internal organs; on the other hand, this bacillus has been demonstrated to be constantly present. It is undoubtedly present

during the lifetime of the patients, and is found in greater abundance in those cases which terminate fatally at an early date. It is not a putrefactive organism, and is not developed in the tissues post mortem, although it has been shown by Fränkel and Simmonds that it multiplies rapidly in the spleen after death, up to the time that putrefactive decomposition commences. This is quite in accord with what we should *a priori* have expected, in view of the facts relating to the propagation of typhoid fever. These facts indicate that the disease in question is due to a microorganism which is capable of multiplication external to the human body in a variety of organic media, at comparatively low temperatures, and that it is widely distributed. From the endemic prevalence of the disease over vast areas of the earth's surface we may infer that it is induced by a hardy microorganism. Eberth's bacillus complies with all of these conditions.

There are numerous facts which indicate that the development of an attack of typhoid and the severity of the symptoms depend to some extent upon the quantity of the infectious material introduced into the alimentary canal. Milk or water which has been infected directly by the discharges of typhoid patients is especially dangerous, and there is reason to believe that repeated or concentrated doses of such infectious material may be effective when a single draught of the contaminated fluid, or a greater degree of dilution, would be innocuous.

Again, we have evidence that the typhoid germs may become effective as a result of certain favorable circumstances relating to the individual or to his environment. Those agencies which reduce the vital resisting power of the tissues, and especially exposure to the emanations from putrefying material, to sewer gas, to vitiated air in overcrowded and ill-ventilated apartments, etc., are recognized as favorable to the development of typhoid fever where the specific cause is present. All these facts seem to accord with the experimental evidence which indicates that the pathogenic power of the bacillus of Eberth depends upon the formation of a poisonous ptomaine rather than upon a special facility for multiplying in the tissues of a living animal. Indeed, it seems quite probable that its power to invade living animal tissues depends upon the toxic action of this ptomaine; or, it may be, of other ptomaines produced under certain circumstances in the body in excess or introduced from without. Such toxic agents may serve, when the specific germ is introduced into the intestine in comparatively small numbers, to give it the mastery over the vital resisting power of the tissues subject to invasion, and thus to induce an attack of the disease.¹

¹ The above account of researches relating to the etiology of typhoid fever is from

46. BACILLUS TYPHI ABDOMINALIS.

Synonyms.—*Bacillus typhosus* ; *Typhus bacillus*.

Eberth (1880 and 1881) demonstrated the presence of this in the spleen and diseased glands of the intestine in typhus. Gaffky (1884) first obtained it in pure cultures from source and determined its principal biological characters.

It is found, in the form of small, scattered colonies, in the liver, the glands of the mesentery, the diseased intestines and in smaller numbers in the kidneys, in fatal cases of typhus. It has also been obtained, by puncture, from the spleen during the alvine discharges of the sick, and rarely from the blood. It is not found in the blood of the general circulation, unless, in rare cases and in small numbers.



FIG. 108.

FIG. 108.—*Bacillus typhi abdominalis*, from single gelatin colony. $\times 1,000$. From micrograph. (Fränkel and Pfeiffer.)



FIG. 109.

FIG. 109.—*Bacillus typhi abdominalis*, from single gelatin colony. $\times 1,000$. From micrograph. (Sternberg.)

Morphology.—Bacilli, usually one to three μ in length and 0.5 to 0.8 μ broad, with rounded ends ; may also grow out in long threads, especially upon the surface of cooked potato. The dimensions of the rods differ considerably in different media. Small oval refractive granules are often seen at the extremities of the rods, especially in potato cultures kept in the incubating oven ; these are not reproductive spores, as was at first supposed. The bacteria have numerous flagella arranged around the periphery of the cells, from five to twenty, but many short rods have but one flagellum. This paper was read by the writer at the annual meeting of the Association of Physicians, Washington, D. C., June 18th, 1886.

terminal flagellum. These flagella are spiral in form, about $0.1\ \mu$ in thickness, and from three to five times as long as the rods (Babes).

In stained preparations unstained "vacuoles" may often be seen at the margins of the rods, either along the sides or at the ends; these appear to be due to a retraction of the protoplasm from the cell membrane.

The typhoid bacillus *stains* with the aniline colors, but more slowly than many other bacteria, and easily parts with its color when treated with decolorizing agents—*e.g.*, iodine solution as employed in Gram's method. Löffler's solution of methylene blue is an excellent staining agent for this bacillus, but permanent preparations fade out after a time; fuchsin, gentian violet, or Bismarck brown, in aqueous solution, may also be used. The flagella may be demonstrated by Löffler's method of staining (p. 32).



FIG. 110.—*Bacillus typhi abdominalis*, stained by Löffler's method, showing flagella. $\times 1,000$. From a photomicrograph by Fränkel and Pfeiffer.

To stain the bacillus in sections of the spleen, etc., it is best to leave these in Löffler's methylene blue solution or in the carbol-fuchsin solution of Ziehl for twelve hours or more; or the aniline-fuchsin solution may be used. The sections should be washed in distilled water only, when Ziehl's solution is used, or with a very dilute solution of acetic acid when Ehrlich's tubercle stain is employed (Baumgarten).

Biological Characters.—The typhoid bacillus is a *motile*, *aëro-bic*, *non-liquefying* bacillus, which grows readily in a variety of culture media at the "room temperature." Although it grows most abundantly in the presence of free oxygen, it may also develop in its absence, and is consequently a *facultative anaërobic*.

In *gelatin plate cultures* small, white colonies are developed at the end of thirty-six to forty-eight hours, which under the microscope are seen to be somewhat irregular in outline and of a spherical, oval, or long-oval form; these have by transmitted light a slightly granular appearance and a yellowish-brown color. At the end of three or four days the colonies upon the surface of the gelatin form a grayish-white layer of one to two millimetres in diameter, with more or less irregular margins, and, when developed from deep colonies, with an opaque central nucleus. These colonies, by transmitted light, have a yellowish-brown color towards the centre, where they are thickest,



FIG. 111.—Single colony of *Bacillus typhi abdominalis*, in nutrient gelatin. ($\times 1$) From a photograph by Roux.

while the margins are colorless and transparent; the surface is commonly marked with a network of lines and furrows. Stick cultures in ten-per-cent gelatin, at 18° to 20° C., at the end of three days show upon the surface a whitish, semi-transparent layer, with sharply defined margins and irregular outline, which has a shining, pearly lustre; and along the line of puncture a grayish-white growth, made up of crowded colonies, which are larger and more distinct at the bottom of the line of growth. Upon *nutrient agar*, at a temperature of 35° to 37° C., the growth is more rapid and forms a whitish, semi-transparent layer. The cultures give off a faint putrefactive odor. The growth upon blood serum is rather scanty, in the form of transparent, shining patches along the line of inoculation.

The typhoid bacillus develops abundantly in *milk*, in which fluid it produces an acid reaction; it also grows in various vegetable infusions and in bouillon.

None of the above characters of growth are distinctive, as certain common bacilli found in normal fæces present a very similar appearance when cultivated in the same media.

The growth of this bacillus upon *potato* is an important character, as was first pointed out by Gaffky. In the incubating oven at the end of forty-eight hours,



FIG. 112.—*Bacillus typhi abdominalis*; stick culture in nutrient gelatin, eighth day at 16° - 20° C. (Baumgarten.)

or at the room temperature in three or four days, the surface of the potato has a moist, shining appearance, but there is no visible growth such as is produced by many other bacteria upon this medium. A simple inspection would lead to the belief that no growth had occurred; but if with a platinum needle a little material is scraped from any portion of the shining surface and a stained preparation is made from it, numerous bacilli will be seen, some of which are likely to be in the form of quite long threads, while others are short and have rounded extremities. This "invisible growth" has been shown by the researches of Buchner and others to be most characteristic upon potatoes having a decidedly acid reaction, as is usually the case. When cultivated upon potatoes having an alkaline reaction a thin, visible film of a yellowish-brown color and of limited extent may be developed. Inasmuch as several common and widely distributed bacteria closely resemble the typhoid bacillus in form and in their growth in nutrient gelatin, this character of invisible growth upon potato is very important for its differentiation, especially as the common bacilli referred to—*Bacillus coli communis*, bacillus of *Emmerich*—produce a very distinct and rather thick, yellowish-white mass upon the surface of potato. But recent researches show that this invisible growth, although not a common character, does not belong exclusively to the typhoid bacillus (Babes).

This bacillus in its development in culture media produces acids—according to Brieger small quantities of volatile fat acids, and, in presence of grape sugar, lactic acid. It also grows readily in a decidedly acid medium, and this character has been employed as a test for differentiating it from other similar bacilli; but some of these also grow in a decidedly acid medium, and too much reliance cannot be placed upon this test.

Brieger has shown that indol is not produced in cultures of the typhoid bacillus, and Kitasato has proposed to use the indol test for differentiating this from other similar bacilli which are said, as a rule, to give the indol reaction. This test consists in the addition to ten cubic centimetres of a bouillon culture which has been in the incubating oven for twenty-four hours, of one cubic centimetre of a solution of sodium nitrite (0.02 gramme to one hundred cubic centimetres of distilled water), together with a few drops of concentrated sulphuric acid. If indol is present a red color is developed.

None of the above-mentioned tests are entirely reliable, but, taken together with the morphological and biological characters above described, they may enable the bacteriological expert to give a tolerably confident opinion as to the presence of this bacillus in a water supply suspected of contamination, etc. And when a bacillus having these characters is obtained in a pure culture from the spleen of a typhoid

cadaver the student may be very sure that he has the typhoid bacillus. But in the presence of various similar bacilli, as in faeces, very comparative researches will be required to determine in a manner that a non-liquefying bacillus obtained in pure culture by the plate method is really the one now under consideration. Especially so as the cultures of the typhoid bacillus in the same media may differ considerably at different times, and a number of others are known which resemble it so closely that it is still uncertain whether they are to be considered as varieties of the typhoid bacillus or as distinct species. Thus Babes, in an extended research, found in the organs of typhoid cases, associated with the true typhoid bacillus, other bacilli or varieties very closely resembling it. He has described three varieties (?), obtained by him from other sources, which could only be differentiated from the true typhoid bacillus by a very careful comparison of cultures made side by side in different media.

Cassedebat, also, in an extended examination of the river water at Marseilles with reference to the presence of the typhoid bacillus, found three species which very closely resembled it, but which, on careful comparison were shown to present slight but constant differences in their biological characters. He was not able to distinguish the true typhoid bacillus, and his researches, together with those of other and other recent investigators, make it appear probable that numerous mistakes have been made by bacteriologists who have reported the finding of the typhoid bacillus in river and well water, in faeces, and who have depended mainly upon the character of its growth upon potato in making their diagnosis. Cassedebat found that all three of his pseudo-typhoid bacilli corresponded in their growth upon potato with the bacillus of Eberth. They also corresponded in their growth on gelatin, agar-agar, and blood agar, which, as heretofore remarked, has no characteristic features. All gave a negative indol reaction. Like the typhoid bacillus, they grew in milk without causing coagulation of the casein, but they produced an alkaline reaction in this fluid, while the true typhoid responded with the typhoid bacillus in producing a decided acid reaction. Differences were also observed in bouillon cultures, in bouillon and milk to which various aniline colors had been added, as recommended by Holz.

Whether the typhoid bacillus, as obtained from the spleen of a typhoid cadaver, is in truth specifically distinct from these pseudo-typhoid bacilli, or whether they are all varieties of the same species, differing from modifications in their biological characters acquired during their continuous development under different conditions, is a unsettled question. But, in view of the experimental evidence

available, there is nothing improbable in the supposition that they are simply varieties, and that, as the result of a saprophytic mode of life, this bacillus may undergo more or less permanent modifications.

In the writer's experiments (1887) the thermal death-point of the typhoid bacillus was found to be 56° C., the time of exposure being ten minutes; and potato cultures containing the refractive granules described by Gaffky as spores were found to be infallibly destroyed by a temperature of 60° C. This result has been confirmed by Buchner (1888) and by Janowsky (1890), and the inference seems justified that these granules are not reproductive bodies, as was at first believed; for spores are distinguished by their great resistance to heat and other destructive agencies. According to Buchner, the bacilli containing these refractive granules are even less resistant than fresh cultures in which they are not present, and he is disposed to look upon them as representing a degeneration of the protoplasm of the cells. They do not stain by the methods which are successful in staining the spores of other bacilli, and, in short, present none of the characters which distinguish spores, except the form and high refractive power.

The typhoid bacillus retains its vitality for many months in cultures; the writer has preserved bouillon cultures for more than a year in hermetically sealed tubes, and has found that development promptly occurred in nutrient gelatin inoculated from these. Dried upon a cover glass, it may grow in a suitable medium after having been preserved for eight to ten weeks (Pfuhl). When added to sterilized distilled water it may retain its vitality for more than four weeks (Bolton), (forty days Cassedebat), and in sterilized sea-water for ten days (De Giaksa). Added to putrefying fæces it may preserve its vitality for several months (Uffleemann), in typhoid stools for three months (Karlinski), and in earth upon which bouillon cultures had been poured for five and one-half months (Grancher and Deschamps).

In hanging-drop cultures this bacillus may be seen to exhibit very active movements, the shorter rods rapidly crossing the field with a darting or to-and-fro, progressive motion, while longer filaments move in a serpentine manner.

In addition to the volatile fat acids which, according to Brieger, are formed in small amounts in cultures of the typhoid bacillus, and to lactic acid formed in solutions containing grape sugar, a basic substance possessing toxic properties has been isolated by the chemist named—his *typhotoxine* ($C_7H_7NO_2$). Brieger supposes that other basic substances are likewise formed, but believes this to be the specific product to which the pathogenic action of the bacillus is due. It is a strongly alkaline base, which produces in mice and guinea-pigs salivation, paralysis, dilated pupils, diarrhoea, and death.

Numerous experiments have been made to determine the effect of various germicidal agents required to destroy the vitality of the bacillus, and the action of antiseptics in restraining its development. For the results of these experiments the reader is referred to the sections in Part Second relating to the action of antiseptics and disinfectants.

Pathogenesis.—The very numerous experiments which have been made on the lower animals have not been successful in producing in any one of them a typical typhoid process. Nor is this surprising in view of the fact that, so far as is known, no one of them is susceptible to contract the disease, as man does, by the use of infected water.

The experiments of Fränkel and Simmonds show that when considerable quantities of a pure culture of this bacillus are injected

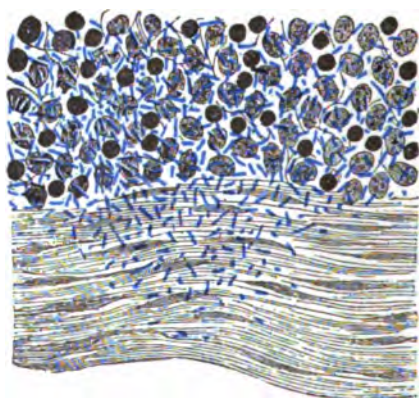


FIG. 113.—Section through wall of intestine, showing invasion by typhoid bacilli (Baumgarten.)

the circulation of rabbits through the ear vein, or into the peritoneal cavity of mice, a certain proportion of the inoculated animals die, usually within forty-eight hours, and that the bacillus may be recovered from the various organs, although it is not present in the blood. But death does not always occur from intravenous injections, and subcutaneous or intraperitoneal injections in rabbits are often without result. Subcutaneous injections in mice proved to be successful in ten cases out of sixteen inoculated by A. Fränkel. Seitz, by following Koch's method—*i. e.*, by rendering the contents of the stomach alkaline, and arresting intestinal peristalsis by the administration of opium—obtained a fatal result, in a majority of the guinea-pigs experimented upon, from the introduction of ten cubic centimeters of bouillon culture into the stomach through a pharyngeal cannula. We may remark, with reference to these results, that while the fact that cultures of the typhoid bacillus have a certain pathogenic

they also show that the animals experimented upon frequently recovered after comparatively large doses, and that the typhoid bacillus is not pathogenic in the same sense as are those microorganisms which, when introduced into the body of a susceptible animal in very minute amount, give rise to general infection and death. On the other hand, a fatal result depends upon the quantity of the culture introduced in the first instance, rather than upon the multiplication of the bacillus in the body of the inoculated animal. This view is confirmed by the experiments of Sirotinin, which show not only that a fatal result depends upon the quantity injected, but also that a similar result follows the injection of cultures which have been sterilized by heat or filtration. The pathogenic action, then, depends upon the presence of toxic substances produced during the growth of the bacillus in artificial culture media. The researches of Brieger, heretofore referred to, show the presence in such cultures of a toxic ptomaine—his typhotoxine—to which the pathogenic potency of these cultures appears to be due. White mice and guinea-pigs usually die in from twenty-four to forty-eight hours when inoculated in the cavity of the abdomen with a virulent culture of the typhoid bacillus—0.1 cubic centimetre to 0.5 cubic centimetre of a bouillon culture three days old. According to Kitasato, the virulence of cultures from different cases of typhoid fever varies considerably.

Detection of the Typhoid Bacillus in Water.—The generally recognized fact that typhoid fever is usually contracted by drinking water contaminated by the typhoid bacillus has led to numerous researches having for their object the discovery of a reliable method of detecting this bacillus when present in water in comparatively small numbers in association with the ordinary water bacilli. The use of Koch's plate method, as commonly employed, will not suffice, because the water bacilli present grow more rapidly and cause liquefaction of the gelatin before visible colonies of the typhoid bacillus are formed; and, owing to the relatively small number of typhoid bacilli, these are likely to escape detection. The aim of bacteriologists has, therefore, been to restrain the growth of these common water bacilli by some agent which does not at the same time prevent the development of the typhoid bacillus. Chantemesse and Widal were the first to propose the use of carbolic acid for this purpose. They recommended the addition of 0.25 per cent of this agent to nutrient gelatin; but, according to Kitasato, the development of the typhoid bacillus is restrained by an amount exceeding 0.20 per cent.

Holz prepares an acid medium by adding gelatin (ten per cent) to the juice of raw potatoes, and asserts that while the typhoid bacillus grows luxuriantly in this medium, many other bacilli fail to develop

in it. The test is said to be still more reliable if 0.05 per cent of carbolic acid is added to the "potato-gelatin." According to the addition of more than 0.1 per cent of carbolic acid to nutrient broth prevents the free development of the typhoid bacillus.

Thoinot has claimed to be able to obtain the typhoid bacillus in mixed cultures—as, for example, from fæces—by suspending a small amount of material containing it for several hours in a solution containing 0.25 per cent of carbolic acid. While other bacteria are destroyed, the typhoid bacillus is said to survive such exposure.

The method of Parietti has recently been tested in a new way by Kamen, and proved to be satisfactory for the detection of the typhoid bacillus in water which was supposed to be the cause of a local epidemic of the disease. The following solution is used:

Carbolic acid,	5	grams
Hydrochloric acid (pure),	4	"
Distilled water,	100	"

Several test tubes, each of which contains ten cubic centimeters of neutral, sterilized bouillon, are used in the experiment. Three to nine drops of the acid solution are added to each tube, and the tubes are then placed in an incubating oven for twenty-four hours to ascertain whether they are still sterile after this exposure. If the bouillon remains clear, from one to ten drops of the acid solution and water are added to each tube and they are returned to the incubating oven. If at the end of twenty-four hours the bouillon is cloudy, this is due, according to Parietti, to the presence of the typhoid bacillus, which is then to be obtained in pure culture by the plate method.

The following method, recently suggested by Hazen and Foote, has been tested with favorable results by Foote. This method depends upon the fact that most of the common water bacilli grow at a temperature of 40° C., whereas this is a favorable temperature for the development of the typhoid bacillus. A small quantity of the suspected water is added to liquefied nutrient agar tubes, and plates are made. These are placed in an incubator at 40° C., and the typhoid bacillus, if present, will develop within two or three days. At the ordinary room temperature more numerous water bacilli would develop upon the same medium abundantly that it would be difficult to recognize colonies of the typhoid bacillus.

Theobald Smith, in a recent paper (*Centralbl. f. Bakt.* Bd. xii., page 367), claims that the typhoid bacillus may be distinguished from other similar bacilli (*Bacillus coli communis*, *B. coli dysenteriae*, *B. coli cholerae*, etc.) by the fact that it does not produce gas.

PLATE V
STERNBERG'S BACTERIOLOGY

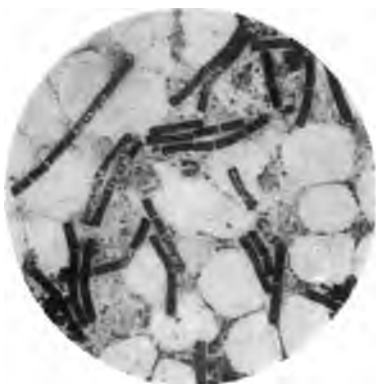


Fig. 1.

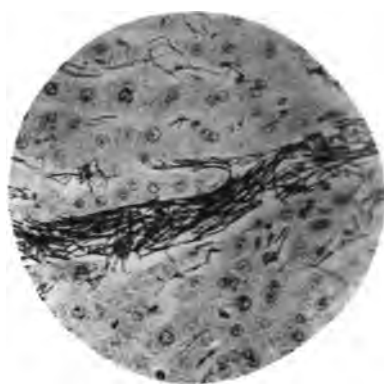


Fig. 2.



Fig. 3.

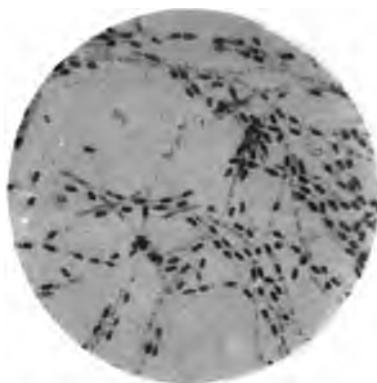


Fig. 4.



Fig. 5.

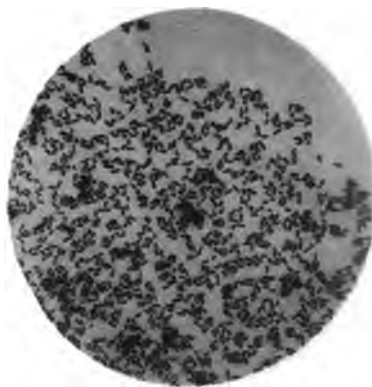


Fig. 6.

media containing sugar—grape sugar, cane sugar, or milk sugar. The medium recommended by Smith for making this test is a peptone-bouillon containing two per cent of grape sugar and made slightly alkaline with carbonate of soda. The liquid becomes clouded throughout at the end of twenty-four hours, but not a trace of gas is developed even after several days. On the other hand, the colon bacillus and other bacilli which closely resemble the typhoid bacillus cause an abundant development of gas in this medium.

PLATE V.

PATHOGENIC BACTERIA.

FIG. 1.—*Bacillus anthracis* from cellular tissue of inoculated mouse. Stained with gentian violet. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

FIG. 2.—*Bacillus anthracis* in section of liver of inoculated rabbit. Stained with Bismarck brown. $\times 250$. Photomicrograph by Sternberg.

FIG. 3.—*Micrococcus gonorrhœæ* in gonorrhœal pus. Stained with gentian violet. $\times 1,000$. Photomicrograph by gaslight (Sternberg).

FIG. 4.—Anthrax spores from a bouillon culture. Double-stained preparation—with carbol-fuchsin and methylene blue. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

FIG. 5.—*Spirillum cholerae Asiaticæ* from a culture upon starched linen at end of twenty-four hours. Stained with fuchsin. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

FIG. 6.—*Bacillus diphtheriæ* from colony upon an agar plate, twenty-four hours old. Stained with Löffler's solution of methylene blue. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

IX.

BACTERIA IN DIPHTHERIA.

DIPHTHERIA is generally recognized by physicians as a specific infectious disease, and, owing to its wide prevalence and fatal character, a precise knowledge of its etiology is of the greatest importance. Until, as a result of recent researches, this was determined, pathologists were in doubt as to whether diphtheria should be considered as primarily a local infection, or whether the local manifestations were secondary to a general systemic infection. But this question appears now to be definitely settled in favor of the former view. We have to-day a very precise knowledge of the specific infecting agent, and have evidence that it produces during its growth a very potent toxic substance, the absorption of which from the seat of local infection accounts in a satisfactory manner for the general symptoms of the disease, which are due to toxæmia and not to an invasion of the blood and tissues by the pathogenic microorganism producing it.

Numerous researches by competent bacteriologists have failed to demonstrate the presence of bacteria in the blood of patients suffering from diphtheria, but a variety of microorganisms have been obtained in cultures from diphtheritic pseudo-membranes, and may be demonstrated by the microscopical examination of stained preparations. Among these are the well-known pus organisms, and especially the *Streptococcus pyogenes*, which appears to be very commonly present, and is perhaps the active agent in the production of certain forms of pseudo-diphtheria. But the malignant, specific diphtheria, so well known in this country and in Europe, has been demonstrated by the recent researches of bacteriologists to be due to a bacillus first recognized by Klebs in stained preparations of diphtheritic false membranes (1883), and cultivated and described by Löffler in 1884. In his first publication Löffler did not claim to have fully demonstrated the etiological relation of this bacillus, but this appears to be fully established by subsequent researches.

In his first research Löffler studied twenty-five cases, and in the greater number of them found in stained preparations the bacillus previously described by Klebs. From six of these cases he

obtained it in pure cultures, and by inoculations in pigeons, chickens, rabbits, and guinea-pigs proved that it gave rise to a diphtheritic inflammation when inoculated into the mucous membrane of the trachea, conjunctiva, pharynx, or vagina. In a second communication Löffler reported his success in finding the same bacillus in ten additional cases, and also that he had isolated from the same source a non-pathogenic bacillus which resembled it very closely. This pseudo-diphtheria bacillus has since been found by other bacteriologists (Von Hoffmann, Roux and Yersin), and it is uncertain whether it is to be considered a distinct species, or a non-pathogenic variety of the diphtheria bacillus as maintained by Roux and Yersin. But its occasional presence does not invalidate the very positive experimental evidence relating to the specific pathogenic power of the true diphtheria bacillus.

Löffler has recently (1890) reviewed the evidence upon which this bacillus is now generally conceded by bacteriologists to be the specific infectious agent in true diphtheria. The following are the principal points in the demonstration :

FIRST.—*It is found in all undoubted cases of diphtheria.* In support of this we have the results of researches made by Löffler, Wyszokowitsch, D'Espine, Von Hoffmann, Ortmann, Roux and Yersin, Kolisko and Paltauf, Zarinko and Sørensen, who in nearly every case have demonstrated without difficulty the presence of this bacillus. On the other hand, Prudden failed to find it in a series of twenty-four cases studied by him ; but his own account of these cases indicates that they were not cases of true diphtheria. He says in a subsequent communication :

"In view of the doubt existing among practitioners as to whether all forms of pseudo-membranous inflammation should be called diphtheria or not, and with the purpose of making a wholly objective study, the writer distinctly stated at the outset of that paper that all the fatal cases of extensive pseudo-membranous laryngitis, as well as pharyngitis, should in his study be considered as cases of diphtheria. This left the question as to the propriety of establishing separate groups of pseudo-membranous inflammation open and free from bias. It was distinctly stated, however, that sixteen out of the twenty-four cases occurred in a large asylum, in which measles and scarlet fever were prevalent during the period in which these studies were under way. Five other cases in another asylum were exposed to similar conditions."

In a subsequent series of "twelve cases of fatal pseudo-membranous inflammation occurring in two children's asylums, in which for many months there had been no scarlatina and no measles, and in which there was no complicating suppurative inflammation and no erysipelas," Prudden (1890) obtained Löffler's bacillus in cultures from eleven, and he says :

"We are now, it would seem, justified, as it did not appear to the writer

that we were two years ago, owing to the large number of important researches which have been made in the interim, in saying that the name diphtheria, or at least primary diphtheria, should be applied, and exclusively applied, to that acute infectious disease, usually associated with a pseudo-membranous inflammation of the mucous membranes, which is primarily caused by the bacillus called *Bacillus diphtheriæ* of Löffler."

In this country Welch and Abbott (1891) have also demonstrated the presence of the Klebs-Löffler bacillus in a series of eight cases occurring in Baltimore, and have proved its specific pathogenic power by inoculations in animals.

With reference to the question as to how long after convalescence is established the diphtheria bacillus may be present in the throat of an infected person, Löffler has made the following research (1890). In a typical case a bacteriological examination was made daily from the commencement until fourteen days after its termination. Fever disappeared on the fifth day, and the exudation had all disappeared on the sixteenth day. Up to this time the bacillus was daily obtained in cultures, and subsequently nearly every day up to the twenty-fifth—that is, for three weeks after the febrile symptoms had disappeared. Roux and Yersin have also obtained the bacillus in cultures from mucus scraped from the throats of convalescents several days after the disappearance of all evidence of the disease.

SECOND. *The Klebs-Löffler bacillus is found only in diphtheria.*—In his earlier researches Löffler obtained the bacillus in a single instance from the mouth of a healthy child, and this fact led him to hesitate in announcing it as his conviction that it was the true cause of diphtheria. But in extended researches made subsequently he has not again succeeded in finding it, except in association with diphtheria, and admits now that he may have been mistaken as to the identity of the bacillus found. This seems not improbable in view of the fact that very similar bacilli have been found by various bacteriologists. Thus Von Hoffmann obtained a very similar but non-pathogenic bacillus from the mucus of chronic nasal catarrh and from healthy mucous membranes; Babes from cases of trachoma, Neisser from ulcers, Zarinko from the surface of various mucous membranes. But all of these were shown to present certain differences in their biological characters by which they could be differentiated from the true diphtheria bacillus.

Welch and Abbott in their comparative studies did not find the Löffler bacillus, "or any bacillus that an experienced bacteriologist would be likely to confound with it." They examined mucus from the throats of healthy children, from those suffering from simple inflammation of the tonsils and pharynx, and from four cases of so-called follicular tonsillitis. As a result of their investigations they agree with Löffler, and with Roux and Yersin, as to "the great prac-

tical value, for diagnostic purposes, of a bacteriological examination of cover-glass specimens and by cultures " of cases in which there is any doubt of the true character of the disease. They say further :

"The only species of bacteria which we have found constantly in the cases of diphtheria has been the Löffler bacillus. Two other species have been present in many cases, viz., the well-known streptococcus, which grows in much smaller colonies and less rapidly than the Löffler bacillus, and a short, oval, often slightly pointed bacillus, growing in long chains running parallel to each other. There are often marked irregularities in shape and especially in size of this bacillus, even of individuals in the same chain. The colonies of this bacillus are grayish-white, moist, larger than those of the streptococcus, but smaller than those of the Löffler bacillus."

THIRD. *As shown by Löffler's earlier researches, pure cultures of this bacillus induce characteristic diphtheritic inflammation when inoculated into the mucous membranes of certain lower animals. Roux and Yersin have also shown that local paralysis is likely to occur in inoculated animals, as is the case in diphtheria in man. In speaking of their inoculations into the trachea in rabbits these investigators say :*

"The affection which is thus induced in the rabbit resembles croup in man. The difficulty which the animal experiences in breathing; the noise made by the air in passing through the obstructed trachea; the aspect of the trachea, which is congested and covered with false membranes; the cedematous swelling of the tissues and glands of the neck, make the resemblance absolutely remarkable."

Welch and Abbott give the following account of the results of inoculations into the trachea in kittens :

"A half-grown kitten is inoculated into the trachea with one platinum loop from a pure culture of the Löffler bacillus on glycerin-agar, eleven days old, derived from Case IV. For the inoculation a small median incision was made over the trachea, in which a hole just large enough to admit the platinum loop was made. The culture was rubbed over the mucosa of the trachea for an extent about three centimetres in length, and in this process sufficient force was used to abrade the mucous membrane. On the day following the inoculation no special alteration in the animal was observed, but on the morning of the second day it was found very weak. In the course of this day it became so weak as to lie completely motionless, apparently unconscious, with very feeble, shallow respiration; several times it was thought to be dead, but on careful examination proved still to be breathing feebly. It was found dead on the morning of the third day. At the autopsy the wound was found gaping and covered with a grayish, adherent, necrotic, distinctly diphtheritic layer. For a considerable distance around the wound the subcutaneous tissues were very cedematous, the edema extending from the lower jaw down over the sternum, and to the sides of the neck, and along the anterior extremities. The lymphatic glands at the angle of the jaw were markedly swollen and reddened. The mucous membrane of the trachea, beginning at the larynx and extending down for six centimetres, was covered with a tolerably firm, grayish-white, loosely attached pseudo-membrane, in all respects identical with the croupous membranes observed in the same situation in cases of human diphtheria."

47. BACILLUS DIPHThERIE.

First observed by Klebs (1883) in diphtheritic false membranes. Isolated in pure cultures and pathogenic power demonstrated by Löffler (1884).

Found in diphtheritic pseudo-membranes, and especially in the deeper portions, intermingled with numerous cellular elements; while the superficial layers of the membrane commonly contain but few cells or bacilli, or are invaded by other species, especially by *Streptococcus pyogenes*. The bacilli are not found in the affected mucous membrane, or in sections from the internal organs in fatal cases of this disease.

Morphology.—Rods, straight or slightly curved, with rounded



FIG. 114. — *Bacillus diphtherie*, from a culture upon blood serum. From a photomicrograph. $\times 1,000$. (Fränkel and Pfeiffer.)

ends, having a diameter of 0.5 to 0.8 μ , and from 2 to 3 μ in length. Irregular forms are very common, and, indeed, are characteristic of this bacillus. In the same culture, and especially in an unfavorable culture medium, very great differences in form and dimensions may be observed; one or both ends may appear swollen, or the central portion may be notably thicker than the extremities, or the rod may be made up of irregular spherical or oval segments. Multiplication occurs by fission only, and the bacilli do not grow out into filaments.

In unstained preparations certain portions of the rod, and especially the extremities, are observed to be more highly refractive than the remaining portion; and in stained preparations these portions are seen to be most deeply colored. The diphtheria bacillus may be *stained* by the use of Löffler's alkaline solution of methylene blue, but is not so readily stained with some of the other aniline colors commonly employed. It stains also by Gram's method. For the demonstration of the bacillus in sections of diphtheritic membrane "nothing can surpass in brilliancy and sharp differentiation sections stained doubly by the modified Weigert's fibrin stain and picro-carmin" (Welch and Abbott).

Biological Characters.—The diphtheria bacillus is *aërobic*, *non-motile*, and *non-liquefying*; it does not form spores. It grows most freely in the presence of oxygen, but is also a *facultative anaërobic*.

Development occurs in various culture media at a temperature of from 20° to 42° C., the most favorable temperature being about 35° C.

It grows readily in *nutrient gelatin* having a slightly alkaline reaction, in *nutrient agar*, *glycerin-agar*, or in *alkaline bouillon*, but the most favorable medium appears to be that first recommended by Löffler—viz., a mixture of three parts of blood serum with one part of bouillon, containing one per cent of peptone, one per cent of grape sugar, and 0.5 per cent of sodium chloride. This mixture is sterilized and solidified at a low temperature, as is usual with blood serum. Upon this the development is so rapid in the incubating oven that, at the end of twenty-four hours, the large, round, elevated colonies, of a grayish-white color and moist appearance, may be easily recognized, while other associated bacteria will, as a rule, not yet have developed colonies large enough to interfere with the recognition of these.



FIG. 115.—Colonies of *Bacillus diphtheriae* in nutrient agar, end of twenty-four hours. $\times 10$. (Fränkel and Pfeiffer.)

Upon *nutrient agar plates* the deep-lying colonies, when magnified about eighty diameters, appear as round or oval, coarsely granular discs, with rather ill-defined margins, or, when several colonies are in juxtaposition, as figures of irregular form. The superficial colonies are grayish-yellow in color, have an irregular, not well-defined outline and a rough, almost reticulated surface. The growth upon *glycerin-agar* is very similar. The first inoculations in a plain nutrient agar tube often give a comparatively feeble growth, which becomes more abundant in subsequent inoculations in the same medium. In stick cultures in *glycerin*—or *plain*—agar, growth occurs to the bottom of the line of inoculation, and also upon the surface, but is not at all characteristic. The same may be said with reference to cultures in *nutrient gelatin*. Plate cultures in this medium containing fifteen per cent of gelatin, at 24° C., give rather small colonies, which are white by reflected light and under the microscope are seen as yellowish-brown, opaque discs, having a more or less irregular outline and a granular structure. In *alkaline bouillon* the growth is sometimes in the form of small, whitish masses along the sides and bottom of the tube, but at others a diffusely clouded growth occurs in this medium; after standing for some time in the incubating oven a thin, white pellicle may form upon the surface of the bouillon. The reaction of the bouillon becomes at first acid, but later it has an alkaline reaction (Welch). With reference to the growth on *potato*, authors have differed, probably because the growth is scarcely visible; upon this point we quote from Welch and Abbott :

"Our experience has been that the *Bacillus diphtheriæ* grows on ordinary steamed potato without any preliminary treatment, but that the growth is usually entirely invisible or is indicated by a dry, thin glaze after several days. Doubtless the invisible character of the growth has led most observers into the error of supposing that no growth existed, whereas the microscopical examination reveals a tolerably abundant growth, which on the first potato is often feeble than on succeeding ones. Irregular forms are particularly numerous in potato cultures, and in general the rods are thicker than on other media. In twenty-four hours, at a temperature of 35° C., microscopical examination shows distinct growth. We have cultivated the bacillus for many generations on potato."

Milk is a favorable medium for the growth of this bacillus, and, as it grows at a comparatively low temperature (20° C.), it is evident that this fluid may become a medium for conveying the bacillus from an infected source to the throats of previously healthy children.

Cultures of the diphtheria bacillus may retain their vitality for several months, and when dried upon silk threads for several weeks colonies are still developed in a suitable medium—in the room from three to four weeks, in an exsiccator five to ten, and in one instance fourteen weeks. In dried diphtheritic membrane, preserved in small fragments, the bacillus retained its vitality for nine weeks, and in larger fragments for twelve to fourteen weeks.

The thermal death-point, as determined by Welch and Abbott, is 58° C., the time of exposure being ten minutes. Löffler had previously found that it did not survive exposure for half an hour to 60° C. With reference to the action of germicidal and antiseptic agents, we refer to the sections in Part Second relating to this subject.

Pathogenesis.—In view of the evidence heretofore recorded, it may be considered as demonstrated that this bacillus gives rise to the morbid phenomena which characterize the fatal disease in man known as diphtheria.

We have already referred to the effects of inoculations into the trachea in rabbits and cats, which give rise to a characteristic diphtheritic inflammation, with general toxæmia and death from the absorption of soluble toxic products formed at the seat of local infection. This inference as to the cause of death seems justified by the fact that the pathogenic bacillus does not invade the blood and tissues, and is supported by additional experimental evidence which we give below. Subcutaneous inoculations in guinea-pigs of a small quantity of a pure culture of the bacillus (0.1 to 0.5 cubic centimetre of a bouillon culture) cause death in from one to four or five days. The usual changes observed at the autopsy are "an extensive local œdema with more or less hyperæmia and ecchymosis at the site of inoculation, frequently swollen and reddened lymphatic glands, increased serous fluid in the peritoneum, pleura, and pericardium, enlarged and hæmorrhagic suprarenal capsules, occasionally

slightly swollen spleen, sometimes fatty degenerations in the liver, kidney, and myocardium. We have always found the Löffler bacilli at the seat of inoculation, most abundant in a grayish-white, fibrino-purulent exudate present at the point of inoculation, and becoming fewer at a distance from this, so that the more remote parts of the cedematous fluid do not contain any bacilli" (Welch and Abbott). The authors quoted agree with Löffler and others in stating that the bacillus is only found at the point of inoculation. In all cases their cultures from the blood and from the various organs gave a negative result.

Rabbits are not so susceptible and may recover after the subcutaneous inoculation of very small doses, but usually die in from four to twenty days when two to four cubic centimetres of a bouillon culture have been introduced beneath the skin. In these animals also there is an extensive local cedema, enlargement of the neighboring lymphatic glands, and a fatty degeneration of the liver. Roux and Yersin have shown that in these animals, when death does not ensue too quickly, paralysis of the posterior extremities frequently occurs, thus completing the experimental proof of the specific pathogenic power of pure cultures of this bacillus.

Similar symptoms are produced in pigeons by the subcutaneous inoculation of 0.5 cubic centimetre or more, but they commonly recover when the quantity is reduced to 0.2 cubic centimetre (Roux and Yersin).

The rat and the mouse have a remarkable immunity from the effects of this poison. Thus, according to Roux and Yersin, a dose of two cubic centimetres, which would kill in sixty hours a rabbit weighing three kilogrammes, is without effect upon a mouse which weighs only ten grammes.

Old cultures are somewhat less virulent than fresh ones, but when replanted in a fresh culture medium they manifest their original virulence. Thus a culture upon blood serum which was five months old was found by Roux and Yersin to kill a guinea-pig in five days, but when replanted it killed a second animal of the same species in twenty-four hours.

Evidently a microorganism which destroys the life of a susceptible animal when injected beneath its skin in small quantity, and which nevertheless is only found in the vicinity of the point of inoculation, must owe its pathogenic power to the formation of some potent toxic substance, which being absorbed gives rise to toxæmia and death. This inference in the case of the diphtheria bacillus is fully sustained by the results of recent experimental investigations. Roux and Yersin (1888) first demonstrated the pathogenic power of cultures which had been filtered through porous porcelain. Old

cultures were found by these experimenters to contain more toxic substance than recent ones, and to cause the death of a pig in the dose of two cubic centimetres in less than two hours. The filtered cultures produced in these animals the effects as those containing the bacilli—local oedema, hæmorrhagic congestion of the organs, effusion into the pleural cavity. What larger doses were fatal to rabbits, and a few drops subcutaneously sufficed to kill a small bird within a few hours. In their second paper (1889) the authors mentioned state that so the reaction of a culture in bouillon is acid its toxic power comparatively slight, but that in old cultures the reaction is alkaline and in these the toxic potency is greatly augmented. With a new culture, filtered after having been kept for thirty days, a one-eighth of a cubic centimetre, injected subcutaneously, sufficed to kill a guinea-pig; and in larger amounts it proved to be fatal to dogs when injected directly into the circulation through a jugular vein.

The same authors, in discussing the nature of the poison in filtered cultures, infer that it is related to the diastases, and that its toxic potency is very much reduced by exposure to a comparatively low temperature—58° C. for two hours—and completely destroyed by the boiling temperature—100° for twenty minutes. It was found to be insoluble in alcohol, and the precipitate obtained by adding alcohol to an old culture proved to contain the toxic substance. Löffler also has obtained, by adding five volumes of alcohol to one of a pure culture, a white precipitate, soluble in water, which killed rabbits in the dose of 0.1 to 0.2 gramme when injected beneath the skin of these animals. It gave rise to a local inflammation and necrosis of the skin in the vicinity of the point of inoculation, and to hyperæmia of the internal organs. This deadly *toxine* is supposed to be an albuminoid substance, but its exact chemical composition has not yet been determined.

Brieger and Fränkel have succeeded in rendering guinea-pigs immune against virulent cultures of the diphtheria bacillus by exposing bouillon cultures three weeks old, which had been sterilized by exposure for an hour to 60° to 70° C., into the subcutaneous tissue (ten to twenty cubic centimetres). At first the susceptibility of the animal is rather increased than diminished, but at the end of three weeks immunity is said to be complete. Fränkel is of the opinion that immunity results from the introduction of a substance which is not identical with the toxic product to which the cultures owe their pathogenic power. This latter is destroyed by a temperature of 60° C., while the substance which gives immunity is still active in the cultures after exposure to a temperature of 60° to 70°, as is shown by the protective results of inoculations made with such cultures.

The recent researches of Behring show that the blood of immune animals contains some substance which neutralizes the toxic product contained in virulent cultures of the diphtheria bacillus. This effect is said to be produced when blood from such an animal is added to a filtered culture without the body, as well as when the culture is injected into the living animal. This remarkable fact, if fully confirmed by further investigations, opens up a new field of experimental research, and may lead to important results in the therapeutics of this and other infectious diseases.

According to Roux and Yersin, "attenuated varieties" of the diphtheria bacillus may be obtained by cultivating it at a temperature of 39.5° to 40° C. in a current of air; and these authors suggest that a similar attenuation of pathogenic power may occur in the fauces of convalescents from the disease, and that possibly the similar non-pathogenic bacilli which have been described by various investigators have originated in this way from the true diphtheria bacillus. These authors further state, in favor of this view, that from diphtheritic false membrane, preserved by them in a desiccated condition for five months, they obtained numerous colonies of the bacillus in question, but that the cultures were destitute of pathogenic virulence. They say:

"It is then possible, by commencing with a virulent bacillus of diphtheria, to obtain artificially a bacillus without virulence, quite similar to the attenuated bacilli which may be obtained from a benign diphtheritic angina, or even from the mouth of certain persons in good health. This microbe, obtained artificially, resembles completely the pseudo-diphtheritic bacillus; like it, it grows more abundantly at a low temperature; it renders bouillon more rapidly alkaline; it grows with difficulty in the absence of oxygen."

48. PSEUDO-DIPHTHERITIC BACILLUS.

Löffler, Von Hoffmann, and others have reported finding bacilli which closely resemble the *Bacillus diphtheriæ*, but which differ from it chiefly in being non-pathogenic. The following account we take from the latest paper upon the subject by Roux and Yersin (troisième mémoire, 1890).

Found by Roux and Yersin in mucus from the pharynx and tonsils of children—from forty-five children in Paris hospitals, suffering from various affections, not diphtheritic, fifteen times; from fifty-nine healthy children in a village school on the seaboard, twenty-six times. Of six children with a simple angina but two furnished cultures of this bacillus, while it was obtained in five out of seven cases of measles.

Its characters are given as follows :

"The colonies of the pseudo-diphtheritic bacillus, cultivated serum, are identical with the true diphtheria bacillus. At a temperature of 33° to 35° multiplication is rapid, and it continues at the ordinary temperature, although slowly. Under the microscope the appearance of which forms these colonies is the same as that of *Bacillus diphtheriae*, stains readily with Löffler's solution of methylene blue, and in Gram's method. Sometimes it colors uniformly, at others it appears granular. It grows in alkaline bouillon, giving a deposit upon the bottom of the vessel containing the culture, and in this medium often presents various forms, pear-shaped, or club-shaped. It is destroyed in a liquid medium at a temperature of 58° C. maintained for ten minutes. All of these characters are common to the pseudo-diphtheritic bacillus and the true *Bacillus diphtheriae*. As a difference between them we may note that the pseudo-diphtheritic bacillus is often shorter in colonies grown upon blood serum; that in bouillon they are more abundant; that they continue at a temperature of 22°, at which the true bacillus grows very slowly. When we make a comparison of cultures in bouillon they become acid and then alkaline; a change occurs much sooner in the case of the pseudo-diphtheritic bacillus. Like the true bacillus, the pseudo-diphtheritic grows in a vacuum abundantly than the other.

"Inoculations into animals of cultures of this bacillus have not caused their death; but we may remark that in some experiments a not very marked inflammation has been produced in guinea-pigs at the point of inoculation, while in others there has been no local lesion. The most marked cedema results from cultures obtained from cases of measles.

"Do the facts which we have reported explain the question which we are now asking? Can we conclude that there is a relation between the two bacilli? On the one side, the presence of the pseudo-diphtheritic bacillus in the throats of healthy persons, and of those who have anginas manifestly not dangerous, seems to be opposed to the idea of a relationship between them. On the other hand, when we consider that the non-virulent bacillus is very abundant in fatal diphtheria, that it is more abundant in benign diphtheria, that it comes more common in severe cases as they progress towards recovery, and finally, that they are more numerous in persons who have recovered from diphtheria than in healthy persons, it is difficult to accept the idea that the two microbes are entirely distinct. The morphological differences which have been referred to are so slight that they prove nothing. The only difference between the organisms can only be distinguished by their action upon animals. The difference of virulence does not at all correspond with the difference of form. As regards the form and the aspect of cultures, the true diphtheria bacilli differ less than virulent anthrax differs from a vegetative anthrax bacillus, which, however, originate from the same source. Besides, the sharp distinction which we make between the virulent and non-virulent bacilli is arbitrary; it depends upon the susceptibility of the animals. If we inoculate animals still more susceptible, there are pseudo-diphtheritic bacilli which we must class as virulent; and if, on the contrary, we substitute rabbits for guinea pigs in our experiments, there are virulent bacilli which we must call pseudo-diphtheritic. In our experiments we do not simply encounter bacilli which are very virulent and bacilli which are non-virulent; between these two extremes there are bacilli of every degree of virulence."

Abbott has recently (1891) published the result of his experiments with reference to the presence of the pseudo-diphtheritic bacillus in benign throat affections. He made a bacteriological study of the throats of three patients, nine of whom were suffering from acute pharyngitis, and fourteen from acute follicular tonsillitis, eight from ordinary

nasal catarrh, two from simple enlarged tonsils, fifteen from chronic pharyngitis, one from subacute laryngitis, one from chronic laryngitis, one from rhinitis, and two from an affection of the tonsils and pharynx. In forty-nine cases nothing of particular interest was observed. A variety of microorganisms were isolated, and of these the pyogenic micrococci were the most common.

In four cases microorganisms were found which resembled the *Bacillus diphtheriæ* of Löffler in their morphology and growth in culture media, but which proved not to be pathogenic. Abbott says: "The single point of distinction that can be made out between the organisms obtained from Cases I., III., and IV. and the true bacillus of diphtheria is in the absence of pathogenic properties from the former, whereas in addition to this point of distinction the organism from Case II. gives, as has been stated, a decided and distinct growth upon the surface of sterilized potato."

49. *BACILLUS DIPHTHERIÆ COLUMBRARUM.*

Described by Löffler (1884), who obtained it from diphtheritic pseudo-membranes in the mouths of pigeons dead from an infectious form of diphtheria which prevails in some parts of Germany among these birds and among chickens.

Reddened patches first appear upon the mucous membrane of the mouth and fauces, and these are covered later with a rather thick, yellowish layer of fibrinous exudate. In pigeons the back part of the tongue, the fauces, and the corners of the mouth are especially affected; in chickens the tongue, the gums, the nares, the larynx, and the conjunctival mucous membrane. The disease is especially fatal among chickens, the young fowls and those of choice varieties being most susceptible. It is attended at the outset by fever, and usually proves fatal within two or three weeks, but may last for several months.

Morphology.—Short bacilli with rounded ends, usually associated in irregular masses, and resembling the bacilli of rabbit septicæmia (fowl cholera), but a little longer and not quite so broad. In sections from the liver they are seen in irregular groups in the interior of the vessels.

Biological Characters.—An *aërobic, non-motile, non-liquefying* bacillus.

Grows in *nutrient gelatin* in the form of spherical, white colonies along the line of puncture, and upon the surface as a whitish layer. Under the microscope the colonies in gelatin plates have a yellowish-brown color and a slightly granular surface. Upon *blood serum* the growth consists of a semi-transparent, grayish-white layer. Upon *potato* a thin layer is formed having a grayish tint.

Pathogenesis.—Pigeons inoculated with a pure culture in the mucous membrane of the mouth are affected exactly as are those which acquire the disease naturally. Subcutaneous inoculations in pigeons give rise to an inflammation resulting in local necrotic changes. Pathogenic for rabbits and for mice. Subcutaneous injections in mice give rise to a fatal result in about five days. The bacillus is found in the blood and in the various organs, in the interior of the vessels, and sometimes in the interior of the leucocytes; they are especially numerous in the liver. The lungs are dotted with red spots, the spleen is greatly enlarged, and the liver has a marbled appearance from the presence of numerous irregular white masses scattered through the pale-red parenchyma of the organ. These white masses are seen, in sections, to consist of necrotic liver tissue, in the centre of which the bacilli

are found in great numbers, in the interior of the vessels. This ap-
is so characteristic that Löffler considers inoculations in mice to be a
reliable method of establishing the identity of the bacillus. Not pa-
for chickens, guinea-pigs, rats, or dogs.

There seems to be some doubt whether the form of diphtheria vi-
vails among pigeons, and which Löffler has shown to be due to the
above described, is identical with the diphtheria of chickens. Diph-
man has been supposed by some authors to be identical with the
prevails among fowls, and possibly this may be the case under cer-
cumstances. But the evidence seems to be convincing that the
infectious diphtheria of fowls which is peculiar to them, and which
ordinary circumstances, is not communicated to man.

50. BACILLUS DIPHThERIE VITULORUM.

Described by Löffler (1884) and obtained by him from the pseu-
branous exudation in the mouths of calves suffering from an infectio-
of diphtheria. The disease is characterized by the appearance of
patches upon the mucous membrane of the cheeks, the gums, the
and sometimes of the larynx and nares of infected animals. There
lowish discharge from the nose, an abundant flow of saliva, occas-
tacks of coughing, and diarrhoea. Death may occur at the end of
five days, but usually the animal survives for several weeks. Dip-
patches similar to those in the mouth are also found in the large
and scattered abscesses in the lungs.

Löffler, in a series of seven cases examined, obtained from the de-
tions of the pseudo-membranous deposit a long bacillus which appe-
the cause of the disease.

Morphology.—Bacilli, five to six times as long as broad, usually
long filaments. The diameter of the rods is about half that of the
of malignant oedema.

Biological Characters.—Attempts to cultivate this bacillus in
gelatin, blood serum from sheep, and various other media were unsu-
But when fragments of tissue containing the bacillus were placed
serum from the calf a whitish border, consisting of the long bacilli,
veloped. These could not, however, be made to grow when trans-
fresh blood serum.

Pathogenesis.—Mice inoculated subcutaneously with the fre-
theritic exudation died in from seven to thirty days. The autopsy
an extensive infiltration of the entire walls of the abdomen, which of-
trated the peritoneal cavity and enveloped the liver, the kidneys,
intestine in a yellowish exudate. The bacillus was found in this
and by inoculating a little of it into another animal of the same
similar result was obtained. Not pathogenic for rabbits or guinea-p

51. BACILLUS OF INTESTINAL DIPHThERIA IN RABBIT

Described by Ribbert (1887) and obtained by him from the organ-
bits which succumbed to an affection characterized by a diphtheritic
mation of the mucous membrane of the intestine. The autopsy reve-
swelling of the mesenteric glands and minute necrotic foci in the l-
spleen.

Morphology.—Bacilli with slightly rounded ends, from three t-
long and 1 to 1.4 μ in diameter; often united in pairs or in filame-
taining several elements.

Stains with the aniline colors, but not so readily in sections
other microorganisms. Ribbert recommends staining with anilin-
fuchsin solution, washing in water, then placing the sections in me-
blue solution, and decolorizing in alcohol. Does not stain by
method.

Biological Characters.—An *aërobic, non-liquefying* (non-motile) bacillus. Upon *gelatin plates* semi-transparent, grayish colonies are formed, which later have a brownish color; the surface of these is finely granular and of a pearly lustre. In stick cultures in nutrient gelatin the growth along the line of puncture is very scanty. On *potato* a flat, whitish layer is formed, which extends slowly over the surface. Grows best at a temperature of 30° to 35° C.

Pathogenesis.—Pure cultures injected into the peritoneal cavity or subcutaneously in rabbits caused the death of these animals in from three to fourteen days, according to the quantity injected. At the autopsy necrotic foci are found in the liver and spleen, and the mesenteric glands are enlarged, but the intestine presents a healthy appearance. But when cultures are introduced into the alimentary canal the characteristic diphtheritic inflammation of the mucous membrane of the intestine is induced. This result was obtained both by direct injection into the lumen of the intestine and by injecting cultures into the mouth.

X.

BACTERIA IN INFLUENZA.

A NUMBER of bacteriologists have made careful researches on the recent extended epidemic of influenza, and quite recently a bacillus has been discovered, both by Pfeiffer and by C. Berlin, which there is good reason to believe is the specific cause of this disease. Before describing this we shall refer briefly to other researches.

Babes has described no less than seventeen distinct species of bacteria isolated by him, principally from nasal or bronchial mucus. Among them a considerable number closely resemble *Streptococcus pyogenes* and *Micrococcus pneumoniae crouposæ*. No one form was found with sufficient constancy to justify the inference that it was the specific cause of the disease.

Klebs, in examining blood drawn from the fingers of patients with influenza, observed an enormous number of small, actively motile, highly refractive bodies, which in their size, form, and movements corresponded with similar bodies previously observed by him in the blood of patients with pernicious anaemia, but which were far more numerous. These were believed by Klebs to be flagellate infusoria ("flagellata"). The observations of other bacteriologists have not thus far confirmed those of Klebs regarding the presence of microorganisms of this class in the blood of patients with influenza.

Kowalski, who made bacteriological researches in sixteen cases of influenza, was able to find microorganisms of any kind in the blood, examined either fresh and in dried preparations. In his cultures from the nasal and bronchial secretions of the sick he obtained in five cases *Staphylococcus pyogenes aureus*, in four *Staphylococcus pyogenes albus*, in two *Micrococcus pneumoniae*, in two *Streptococcus pyogenes*, in two *Staphylococcus citreus*, in one Friedländer's bacillus, in one *Staphylococcus albus*, in one *Staphylococcus cereus flavus*. In addition to these he found three species not previously described. One of these he found in two cases; this grew upon the surface of agar as small, transparent colonies, did not grow upon potato, in sterilized milk, or in bouillon; it was arranged in pairs or in chains, and is designated by Kowalski "*Gonococcus*."

Prior, in a bacteriological study of fifty-three cases, twenty-nine of which were without complication and twenty-four complicated by pneumonia, found in the sputum of uncomplicated cases, as the most abundant microorganism at the outset of the attack, *Micrococcus pneumoniae crouposæ*; next to this came *Staphylococcus pyogenes aureus* and *Micrococcus pyogenes*; when the acme of the attack was past the two species named quickly diminished in numbers, while streptococci were found in longer time. In cases of croupous pneumonia following influenza "*Micrococcus pneumoniae*" was constantly found in great numbers.

Fischel (1891) obtained in cultures from the blood of two cases

ferent micrococci, one of which was pathogenic for dogs and horses and gave rise to symptoms in these animals resembling those of influenza (see Micrococcus No. II. of Fischel, No. 39, page 324).

Kirchner (1891) found constantly in the sputum of recent cases a diplococcus enclosed in a jelly-like capsule, which differed in its biological and pathological characters from *Micrococcus pneumoniae crouposa* (see Micrococcus of Kirchner, No. 38, page 324).

52. BACILLUS OF INFLUENZA.

Discovered by Pfeiffer (1892) in the purulent bronchial secretion, and by Canon in the blood of patients suffering from epidemic influenza. Pfeiffer found the bacillus in thirty-one cases examined by him, and in uncomplicated cases it was present in the purulent bronchial secretion in immense numbers and in a pure culture. Canon, whose independent observations were published at the same time, examined the blood of twenty influenza patients in stained preparations, and found the same bacillus in nearly all of them. His method of demonstrating it is as follows:

The blood is spread upon clean glass covers in the usual way. After the preparations are thoroughly dry they are placed in absolute alcohol for five minutes. They are then transferred to the following staining solution (Czenzynke's): concentrated aqueous solution of methylene blue, forty grammes; one-half-per-cent solution of eosin (dissolved in seventy-per-cent alcohol), twenty grammes; distilled water, forty grammes. The cover glasses immersed in this staining solution are placed in an incubating oven at 37° C. for from three to six hours, after which they are washed with water, dried, and mounted in balsam. In successful preparations the red blood corpuscles are stained red by the eosin, and the leucocytes blue. The bacillus is seen in these as a short rod, often resembling a diplococcus. It is sometimes seen in large numbers, but usually only a few rods are seen after a long search—four to twenty in a single preparation. In six cases it was found in numerous aggregations containing from five to fifty bacilli each. In these cases the blood was drawn during a fall of temperature or shortly after.

Morphology.—Very small bacilli, having about the same diameter as the bacillus of mouse septicæmia, but only half as long. Solitary or united in chains of three or four elements.

Stains with difficulty with the basic aniline dyes—best with dilute Ziehl's solution, or Löffler's methylene blue solution, with heat. The two ends of the bacilli are most deeply stained, causing them to resemble diplococci. Pfeiffer says: "I am inclined to believe that some of the earlier observers also saw the bacilli described by me, but that, misled by their peculiar behavior with regard to staining agents, they described them as diplococci or streptococci." Do not stain by Gram's method.

Biological Characters.—An *aërobic, non-motile* bacillus. not grow in nutrient gelatin at the room temperature. Spore formation not observed. Upon the surface of *glycerin-agar* in the incubating oven very small, transparent, drop-like colonies are developed at the end of twenty-four hours. These can only be recognized by the aid of a lens. "A remarkable point about them is that the colonies always remain separate from each other, and do not, as all species known to me do, join together and form a continuous mass. This feature is so characteristic that the influenza bacilli can be thereby with certainty distinguished from other bacteria" (Kitasato). On 1.5 per cent sugar-agar the colonies appear as extremely small droplets, clear as water, often only recognizable with a lens (Pfeiffer).

In *bouillon* a scanty development occurs, and at the end of twenty-four hours small, white particles are seen upon the surface, which subsequently sink to the bottom, forming a white, woolly deposit while the bouillon above remains transparent. This bacillus does not grow at temperatures below 28° C.

Canon has obtained colonies, resembling those described by Kitasato, in cultures from the blood of influenza patients. His cultures were made upon glycerin-agar in Petri's dishes. Ten or twelve drops of blood from a puncture made in the finger of the patient, after sterilization of the surface, were allowed to fall upon the agar medium and this was placed in the incubating oven. As the number of bacilli in the blood is small, a considerable quantity is used. Colonies are visible at the end of twenty-four to forty-eight hours.

The influenza bacillus is quickly destroyed by desiccation. A pure culture diluted with water and dried is destroyed with loss of vitality in twenty hours; in dried sputum the vitality is retained somewhat longer, but no growth occurs after forty hours. The thermal death-point is 60° C. with five minutes' exposure (Pfeiffer and Beck).

Pathogenesis.—Pfeiffer infers that this is the specific cause of influenza in man for the following reasons:

1. They were found in all uncomplicated cases of influenza, whether examined in the characteristic purulent bronchial secretion, or in absolutely pure cultures. They were frequently situated in the protoplasm of the pus corpuscles; in fatal cases they were found to have penetrated from the bronchial tubes into the peribronchitis tissue, and even to the surface of the pleura, where in two cases they were found in pure cultures in the purulent exudation.

2. They were only found in cases of influenza. Numerous control experiments proved their absence in ordinary bronchitis, diphtheria, etc.

PLATE VI
STERNBERG'S BACTERIOLOGY.

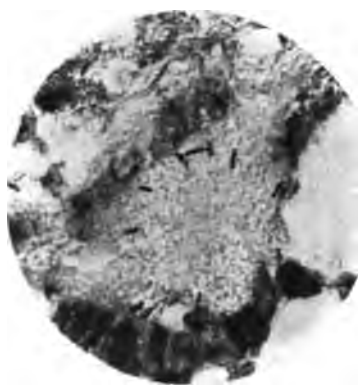


Fig. 1.

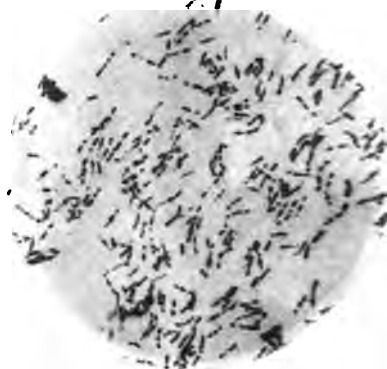


Fig. 2.



Fig. 3.

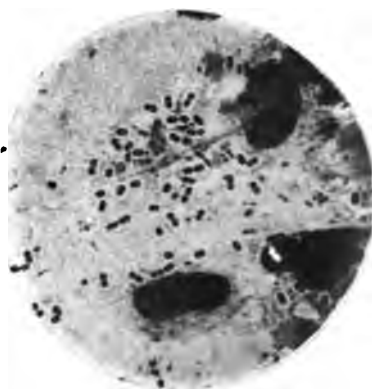


Fig. 4.



Fig. 5.

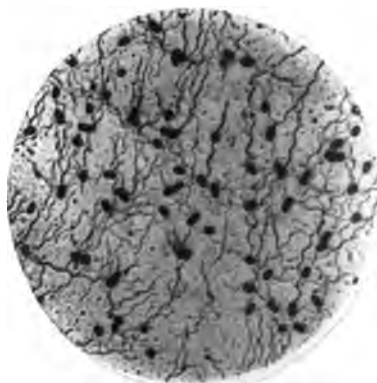


Fig. 6.

3. The presence of the bacilli corresponded with the course of the disease, and they disappeared with the cessation of the purulent bronchial secretion.

In his preliminary report of his investigations Pfeiffer says :

“Numerous inoculation experiments were made on apes, rabbits, guinea-pigs, rats, pigeons, and mice. Only in apes and rabbits could positive results be obtained. The other species of animals showed themselves refractory to influenza.”

PLATE VI.

PATHOGENIC BACTERIA.

FIG. 1.—*Bacillus tuberculosis* in giant cell. $\times 1,000$. Photomicrograph made at the Army Medical Museum, Washington, by Gray.

FIG. 2.—*Bacillus tuberculosis* from a culture on glycerin-agar. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

FIG. 3.—*Bacillus tetani* from an agar culture. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

FIG. 4.—*Micrococcus pneumoniae crouposæ* in sputum of a patient with pneumonia. $\times 1,000$. Stained by Gram's method. Photomicrograph by Fränkel and Pfeiffer.

FIG. 5.—*Bacillus septicæmiæ hæmorrhagicæ* (“bacillus of fowl cholera”) in blood from the heart of an inoculated pigeon. $\times 1,000$. Photomicrograph by Fränkel and Pfeiffer.

FIG. 6.—*Bacillus* of hog cholera, showing flagella. Stained by Löffler's method. $\times 1,000$. Photomicrograph made at the Army Medical Museum, Washington, by Gray.

XI.

BACILLI IN CHRONIC INFECTIOUS DISEASES.

IN tuberculosis, leprosy, glanders, and syphilis we have a group of infectious diseases which present many points of resemblance. All run a chronic course; all may be communicated to susceptible animals by inoculation; in all, the lymphatic glands in the vicinity of the point of inoculation become enlarged, and new growths, consisting of various cellular elements of a low grade of vitality, are developed in the tissues which are the point of predilection for each; in all, these new growths show a tendency to degenerative changes, as a result of which abscesses, caseous masses, or open ulcers are formed.

In two of the diseases in this group—tuberculosis and glanders—the infectious agent has been obtained in pure cultures and its specific pathogenic power demonstrated by inoculations in susceptible animals; in one—leprosy—there is but little doubt that the bacillus constantly found in the new growths characteristic of the disease bears an etiological relation to it, although this has not been demonstrated, the bacillus not having as yet been cultivated in artificial media. The evidence with reference to the parasitic nature of the fourth disease mentioned as belonging to this group—syphilis—is still unsatisfactory, but there is every reason to believe that it will also eventually be proved to be due to a parasitic microorganism.

The announcement of the discovery of the tubercle bacillus was made by Koch, in March, 1882, at a meeting of the Physiological Society of Berlin. At the same time satisfactory experimental evidence was presented as to its etiological relation to tuberculosis in man and in the susceptible lower animals, and its principal biological characters were given.

This achievement, the result of patient and intelligent scientific investigation, will always rank as one of the most important in the history of medicine. The previous demonstration by Villemin (1865)—confirmed by Cohnheim (1877) and others—that tuberculosis might be induced in healthy animals by inoculations of tuberculous material, had paved the way for this great discovery, and advanced

pathologists were quite prepared to accept it. The more conservative have since been obliged to yield to the experimental evidence, which has received confirmation in all parts of the world. To-day it is generally recognized that tuberculosis is a specific infectious disease due to the tubercle bacillus.

As evidence of the thorough nature of Koch's personal researches in advance of his first public announcement, we give the following résumé of his investigations :

In nineteen cases of miliary tuberculosis the bacilli were found in the tubercular nodules in every instance ; also in twenty-nine cases of pulmonary phthisis, in the sputum, in fresh cheesy masses, and in the interior of recently formed cavities ; in tuberculous ulcers of the tongue, tuberculosis of the uterus, testicles, etc. ; in twenty-one cases of tuberculous—scrofulous—lymphatic glands ; in thirteen cases of tuberculous joints ; in ten cases of tubercular bone affections ; in four cases of lupus ; in seventeen cases of Perlsucht in cattle. His experimental inoculations were made upon two hundred and seventy-three guinea-pigs, one hundred and five rabbits, forty-four field mice, twenty-eight white mice, nineteen rats, thirteen cats, and upon dogs, pigeons, chickens, etc. Very extensive comparative researches were also made, which convinced him that the bacillus which he had been able to demonstrate in tuberculous sputum and tissues by a special mode of staining was not to be found in the sputa of healthy persons, or of those suffering from non-tubercular pulmonary affections, or in organs and tissues involved in morbid processes of a different nature.

53. BACILLUS TUBERCULOSIS.

Discovered by Koch (first public announcement of discovery March 24th, 1882). The bacilli are found in the sputum of persons suffering from pulmonary or laryngeal tuberculosis, either free or in the interior of pus cells ; in miliary tubercles and fresh caseous masses, in the lungs or elsewhere ; in recent tuberculous cavities in the lungs ; in tuberculous glands, joints, bones, and skin affections (lupus) ; in the lungs of cattle suffering from pulmonary tuberculosis—Perlsucht ; and in tubercular nodules generally in animals which are infected naturally or by experimental inoculations.

In the giant cells of tubercular growths they have a peculiar and characteristic position, being found, as a rule, upon the side of the cell opposite to the nuclei, which are crowded together in a crescentic arrangement at the opposite pole of the cell. Sometimes a single bacillus will be found in this position, or there may be several. Again, numerous bacilli may be found in giant cells in which the nuclei are distributed around the periphery. They are more numer-

ous in tuberculous growths of recent origin, and often cannot be demonstrated, by microscopical examination, in caseous material from the centre of older nodules. But such material, when inoculated into susceptible animals, gives rise to tuberculosis, and the usual inference is that it contains spores of the tubercle bacillus.

Morphology.—The tubercle bacilli are rods with rounded ends, of from 1.5 to 3.5 μ in length, and are commonly slightly curved or bent at an angle; the diameter is about 0.2 μ . In stained preparations unstained portions are frequently seen, which are generally believed to be spores, but this is by no means certain. From two to six of these unstained spaces may often be seen in a single rod, and owing to this alternation of stained and unstained portions the bacilli may, under a low power, be mistaken for chains of micrococci. The rods are usually solitary, but may be united in pairs, or in short chains containing three or four elements. In old cultures irregular



FIG. 116. — *Bacillus tuberculosis*.
× 1,000. From a photomicrograph.

forms may be observed, the rods being sometimes swollen at one extremity, or presenting the appearance of having a lateral bud-like projection—involution forms.

The *staining* characters of this bacillus are extremely important for its differentiation and recognition in preparations of sputum, etc. Unlike most microorganisms of the same class, it does not readily take up the aniline colors, and when stained it is not easily decolorized, even by the use of strong acids. The failure to observe it in tuberculous material, prior to Koch's discovery, was no doubt due to the fact that it does not stain in the usual aqueous solutions of the aniline dyes. Koch first recognized it in preparations placed in a staining fluid to which an alkali had been added—solution of methylene blue with caustic potash; but this method was not very satisfactory, and he promptly adopted the method devised by Ehrlich, which consists essentially in the use of a solution of an aniline color—fuchsin or methyl violet—in a saturated aqueous solution of aniline oil, and decolorization with a solution of a mineral acid—nitric acid one part to three parts of water.

The original method of Ehrlich gives very satisfactory results, but various modifications have since been proposed, some of which are advantageous. The carbol-fuchsin solution of Ziehl is now largely employed; it has the advantage of prompt action and of

keeping well. The staining is effected more quickly if heat is applied. The tubercle bacilli stain by Gram's method, but this is not to be recommended for general use, owing to the fact that the protoplasm of the rods is frequently contracted into a series of spherical, stained bodies, which might easily be mistaken for micrococci.

The *examination of sputum* for the presence of the tubercle bacillus is recognized as a most important procedure for the early diagnosis of pulmonary tuberculosis. It is attended with no special difficulties, and every physician should be acquainted with the technique.

The patient should be directed to expectorate into a clean, wide-mouthed bottle or glass-covered jar the material coughed up from the lungs, and especially, in recent cases, that which is coughed up upon first rising in the morning. This should be placed in the physician's hands as promptly as possible; although a delay of some days does not vitiate the result, and the tubercle bacilli may still be demonstrated after the sputum has undergone putrefaction. It is well to pour the specimen into a clean, shallow vessel having a blackened bottom—a Petri's dish placed upon a piece of dead-black paper will answer very well. In tuberculous sputum small, lenticular masses of a yellowish color may usually be observed, and one of these should be selected for microscopical examination, by picking it up with a platinum needle and freeing it as far as possible from the tenacious mucus in which it is embedded. If such masses are not recognized take any purulent-looking material present in the specimen, whether it be in small specks distributed through the mucus, or in larger masses. A little of the selected material should be placed in the centre of a clean cover glass and another thin glass cover placed over it. By pressure and a to-and-fro motion the material is crushed and distributed as evenly as possible; the glasses are then separated by a sliding motion. The film is permitted to dry by exposure in the air. When dry the cover glass, held in forceps, is passed three times through the flame of an alcohol lamp or Bunsen burner to fix the albuminous coating. Too much heat causes the film to turn brown and ruins the preparation. The staining fluid (Ziehl's carbol-fuchsin) may then be poured upon the cover glass, or this may be floated upon the surface of the fluid contained in a shallow watch glass. Heat is now applied by bringing the cover glass over a flame and holding it there until steam begins to be given off from the surface of the staining fluid; it is then withdrawn and again

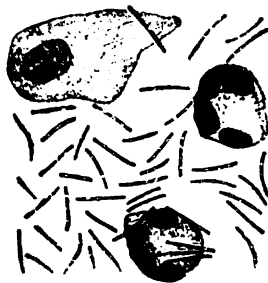


FIG. 117.—*Bacillus tuberculosis* in sputum. $\times 1,000$. (Baumgarten.)

gently heated at intervals for a minute or two. The cover glass is then washed in water, and the film will be seen to have a uniform deep-red color. The next step consists in decolorization in the acid solution (twenty-five-per-cent solution of nitric or of sulphuric acid). The cover glass is gently moved about in this solution for a few seconds, and the color will be seen to quickly fade to a greenish tint. The object is to remove all color from the cells and the albuminous background, so that the bacilli, which retain their color in presence of the acid, may be clearly seen. The preparation is next washed in dilute alcohol (sixty per cent) to remove the fuchsin which has been set free by the acid. If decolorization was not carried far enough the film will be seen to still have a red color, especially in places where it is thickest, when it is removed from the dilute alcohol and washed out in water. In this case it will be necessary to return it to the acid solution and again wash it in the dilute alcohol and in water. It may now be placed in a solution of methylene blue or of vesuvin for a contrast stain. The tubercle bacilli are distinguished by the fact that they retain the red color imparted to them in the fuchsin solution, while other bacteria present, having been decolorized in the acid solution, take the contrast stain and appear blue or brown, according to the color used. The double-stained preparation, after a final washing in water, may be examined at once, or dried and mounted in balsam for permanent preservation.

Of the various other methods which have been proposed, that of Fränkel, as modified by Gabbett, appears to be the most useful. This consists in staining as above directed with Ziehl's carbol-fuchsin solution, and in then placing the cover glass directly in a second solution which contains both the acid for decolorizing and the contrast stain. This second solution contains twenty parts of nitric acid, thirty parts of alcohol, fifty parts of water, and sufficient methylene blue to make a saturated solution (one to two parts in one hundred). After remaining in this solution for a minute or two the cover glass is washed in water, and upon microscopical examination the tubercle bacilli, if present, will be seen as red rods which strongly contrast with the blue background.

The methods recommended for cover-glass preparations may also be used for staining the tubercle bacillus in thin sections of tuberculous tissues, except that it is best not to employ heat. The sections may be left for an hour in the carbol-fuchsin solution, or for twelve hours in the Ehrlich-Weigert tubercle stain—eleven cubic centimetres of saturated alcoholic solution of methyl violet, ten cubic centimetres of absolute alcohol, one hundred cubic centimetres of aniline water. They should then be decolorized by placing them for

about half a minute in dilute nitric acid (ten per cent); then wash out color in sixty-per-cent alcohol; counter-stain for two or three minutes in a saturated aqueous solution of methylene blue; dehydrate with absolute alcohol or with aniline oil; clear up in oil of cedar, and mount in xylol balsam. If the aniline-water-methyl-violet solution has been used for staining the bacilli a saturated solution of vesuvin may be used as a contrast stain.

Biological Characters.—A *parasitic, aërobic, non-motile* bacillus, which grows only at a temperature of about 37° C. Is also a *facultative anaërobic* (Fränkel).

The question as to *spore formation* has not been definitely determined. It has been generally assumed that the unstained spaces which are frequently seen in the bacilli are spores; and the fact that

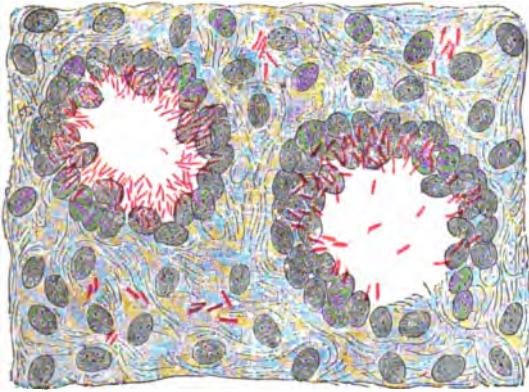


FIG. 118.—Section through a tuberculous nodule in the lung of a cow, showing two giant cells containing tubercle bacilli. $\times 950$. (Baumgarten.)

caseous material in which a microscopical examination has failed to demonstrate the presence of bacilli may produce tuberculosis, with bacilli, when inoculated into guinea-pigs, has been explained upon the supposition that this material contained spores. But a few bacilli present in such caseous material might easily escape detection. As pointed out by Fränkel, the oval spaces in stained specimens have not the sharply defined outlines of spores. Moreover, the bacilli, when examined in unstained preparations, do not contain corresponding refractive bodies, recognizable as spores. And when the bacilli are stained by Gram's method the protoplasm is often contracted in the form of little, spherical stained masses, while the unstained spaces are larger and no longer have the oval form presented in rods stained by Ehrlich's method. The great resisting power of the bacillus to heat and to desiccation has been supposed to be due to the presence

of spores. But, so far as resistance to heat is concerned, this is not so great as was at one time believed. Schill and Fischer (1884), assuming that the tubercle bacillus forms spores, made quite a number of experiments to determine its thermal death-point. They subjected sputum containing the bacillus to a temperature of 100° C., and tested the destruction of vitality by inoculations into guinea-pigs. Exposure to steam at a temperature of 100° C. for two to five minutes was effective in every experiment, with one exception. One guinea-pig died tuberculous after having been inoculated with sputum exposed to this temperature for two minutes. This result was assumed to show that the bacillus would survive lower temperatures, but it is evident that additional experiments were required to establish this fact. In 1887 the writer made a few similar experiments at a lower temperature, and guinea-pigs inoculated with tuberculous sputum exposed for ten minutes to a temperature of 90° , 80° , and 60° C. failed to become tuberculous, while another guinea-pig, inoculated with the same material after exposure to a temperature of 50° C. for ten minutes, died tuberculous. These results correspond with those subsequently (1888) reported by Yersin, who tested the thermal death-point of this bacillus by the culture method. This author assumes that the bacilli form spores, but states as a result of his experiments that "at the end of ten days bacilli heated for ten minutes at 55° C. gave a culture in glycerin-bouillon; those heated to 60° , at the end of twenty-two days; while those heated to 70° and above failed to grow in every instance. This experiment, repeated a great number of times, always gave the same result. The tubercle bacilli then resist a temperature of 60° C. for ten minutes, and it is to be remarked that the resistance of spores to heat appears to be no greater than that of the bacilli themselves." Yersin remarks in a footnote that "the spores which served for these experiments did not appear as more or less irregular granules taking the coloring matter strongly, but as veritable spores with sharply defined outlines, to the number of one or two in a bacillus, or three at the outside. These spores are particularly clear in cultures upon glycerin-agar several weeks old."

It may be that bacteriologists have been mistaken in the inference that all spores possess a greater resisting power for heat than that exhibited by bacilli in the absence of spores. That this is true as regards anthrax spores and many others, the thermal death-point of which has been determined by exact experiments, does not prove that it is true for all. And it is known that there are wide differences in the resisting power both of the spores of different species and in the vegetating cells. To admit that the tubercle bacillus or the typhoid bacillus, etc., may form spores which have no greater

resisting power against heat than the bacilli themselves, would therefore simply be an admission that some bacteriologists had made a mistaken inference based upon incomplete data. In view of the facts stated we can simply repeat what was said at the outset, viz., the question as to spore formation has not been definitely determined.

The tubercle bacillus is a *strict parasite*, and its biological characters are such that it could scarcely find natural conditions, outside of the bodies of living animals, favorable for its multiplication. It therefore does not grow as a saprophyte under ordinary circumstances. But it has been noted by Roux and Nocard that when it has been cultivated for a time in artificial media containing glycerin it may grow in a plain bouillon of veal or chicken, in which media it fails to develop when introduced directly from a culture originating from the body of an infected animal. This would indicate the possibility of its acquiring the ability to grow as a saprophyte; and we can scarcely doubt that at some time in the past it was a true saprophyte. The experiments of Nuttall indicate that the bacillus may multiply, under favorable temperature conditions, in tuberculous sputum outside of the body. And it is extremely probable that multiplication occurs in the muco-purulent secretion which accumulates in pulmonary cavities in phthisical patients. In these cavities its development may, in a certain sense, be regarded as saprophytic, as it feeds upon non-living organic material.

Koch first succeeded in cultivating this bacillus upon coagulated *blood serum*, prepared as directed in Section VIII., Part First, of the present volume. Roux and Nocard have since shown (1888) that it grows very well on nutrient agar to which glycerin has been added (six to eight per cent), and also in veal broth containing five per cent of glycerin. It is difficult to obtain pure cultures from tuberculous sputum, on account of the presence of other bacteria which grow much more rapidly and take full possession of the medium before the tubercle bacillus has had time to form visible colonies. For this reason it is best to first inoculate a guinea-pig with the tuberculous sputum and to obtain cultures from it after tuberculous infection has fully developed. The inoculated animals usually die at the end of three or four weeks. It is best to kill one which gives evidence of being tuberculous, and to remove one or more nodules from the lungs through an opening made in the chest walls. The greatest care will be required to prevent contamination by other common microorganisms. The instruments used must be sterilized by heat, and the skin over the anterior thoracic wall carefully turned back; then, after again sterilizing knives and scissors, cut an opening into the chest cavity, draw out the root of the lung, and take up with

slender sterilized forceps, or with a strong platinum loop, one or more well-defined tubercular nodules. These may be conveyed directly to the surface of the solid culture medium and then broken up and rubbed over the surface as thoroughly as possible; or they may first be crushed between two sterilized glass slides, and then transferred with the platinum loop and thoroughly rubbed into the surface of the culture medium.

This breaking-up of the tuberculous nodules and distribution of the bacilli upon the surface of the culture medium is essential for the success of the experiment. Instead of using the tubercular nodules in the lungs, an enlarged lymphatic gland from the axilla or elsewhere may be used, as first recommended by Koch. This is to be crushed in the same way; and it will be best to inoculate a number of tubes at the same time, as accidental contamination or failure to develop is very liable to occur in a certain number. Owing to the liability of the blood serum to become too dry for the development of the bacillus, it is best to keep the cultures in a moist atmosphere, or to prevent evaporation by applying a rubber cap over the open end of the test tube. This should be sterilized in a solution of mercuric chloride (1 : 1,000); and the end of the cotton plug should be burned off just before applying it, for the purpose of destroying the spores of mould fungi, which in a dry atmosphere would be harmless, but under the rubber cap are likely to sprout and to send their mycelium through the cotton plug to the interior of the tube, thus destroying the culture.

Upon coagulated blood serum the growth first becomes visible at the end of ten to fourteen days (at 37° C.), and at the end of three



FIG. 119.—Tubercle bacilli from surface of culture upon blood serum. $\times 500$. (Koch.)

weeks a very distinct and characteristic development has occurred. The first appearance is that of dry-looking, grayish-white points and scales, which are without lustre, and are sometimes united to form a thin, irregular, membranous-looking layer. Under the microscope, with an amplification of eighty diameters, the early, thin surface growth upon blood serum presents a characteristic appearance. The bacilli, arranged in parallel rows, form variously curved figures, of which we may obtain impressions by carefully applying a dry cover glass to the surface. Upon staining the preparation in the usual way the same arrangement of the bacilli which adhered to the thin glass cover will be pre-

served. The growth is more abundant in subsequent cultures, which have been kept up in Koch's laboratory from his original pure cultures up to the present time; in these the bacillus still pre-

serves its characters of form and growth, and its specific pathogenic power.

Pastor (1892) has succeeded in obtaining pure cultures of the tubercle bacillus from sputum by the following ingenious method: After proving by microscopic examination that the sputum of a tuberculous individual contains numerous bacilli, he has the patient cleanse his mouth as thoroughly as possible with sterilized water, and then expectorate some material, coughed up from the lungs, into a sterilized test tube. By shaking with sterilized water a fine emulsion is made, and this is filtered through fine gauze. The filtrate, which is nearly transparent, contains numerous tubercle bacilli. A few drops of the emulsion are now added to liquefied gelatin in a test tube, and a plate is made in the usual way. This is kept for three or four days at the room temperature, during which time the common mouth bacteria capable of growth form visible colonies. By means of a hand lens a place is now selected in which no colonies are seen, and a bit of gelatin is excised with a sterilized knife. This piece is transferred to the surface of blood serum or glycerin-agar, and placed in the incubating oven, where in due time colonies of the tubercle bacillus will usually be found to develop.

Another method of accomplishing the same result has recently been described by Kitasato. This is a method devised by Koch some time since and successfully employed in his laboratory. The morning expectoration of a tuberculous patient, raised from the lungs by coughing, is received in a Petri's dish. A bit of sputum, such as comes from the tuberculous cavity in the lungs of such a patient, is now isolated with sterilized instruments and carefully washed in at least ten successive portions of sterilized water. By this procedure the bacteria accidentally attached to the viscid mass of sputum during its passage through the mouth are washed away. In the last bath the mass is torn apart and a small portion from the interior is used to make a microscopic preparation, the examination of which shows whether only tubercle bacilli are present. If this be the case cultures upon glycerin-agar are started from material obtained from the interior of the same mass. The colonies obtained in this way appear in about two weeks as round, white, opaque, moist, and shining masses. Kitasato's researches show that the greater portion of the tubercle bacilli in sputum obtained in this way, and in the contents of lung cavities, are incapable of development, although this fact cannot be recognized by a microscopic examination of stained specimens.

On account of the greater facility of preparing and sterilizing *glycerin-agar*, and the more rapid and abundant development upon this medium, it is now usually employed in preference to blood

serum. The growth at the end of fourteen days is more abundant than upon blood serum at the end of several weeks. When numerous bacilli have been distributed over the surface of the culture medium a rather uniform, thick, white layer, which subsequently acquires a

yellowish tint, is developed; when the bacilli are few in number or are associated in scattered groups separate colonies are developed, which acquire considerable thickness and have more or less irregular outlines; they are white at first, then yellowish-white. Fränkel describes the tubercle bacillus as a facultative anaërobic, and it would appear that it must be able to grow in situations where it can obtain very little oxygen from its development in the interior of tuberculous nodules, lymphatic glands, etc. But in stick cultures in glycerin-agar development only occurs near the surface, and not at all in the deeper portion of the medium. In view of its abundant growth on the surface it is difficult to understand this failure to grow along the line of puncture, if it is in truth a facultative anaërobic.



FIG. 120.—Culture of tubercle bacillus upon glycerin-agar. Photograph by Roux.

In peptonized veal broth containing five per cent of glycerin the bacillus develops at first in the form of little flocculi, which accumulate at the bottom of the flask and which by agitation are easily broken up. At the end of two or three weeks the bottom of the flask is covered with similar flocculi, which form an abundant deposit.

Pawlowski and others report success in cultivating the tubercle bacillus upon the surface of cooked *potato* enclosed in a test tube after the method of Bolton and Roux. The open end of the tube is hermetically sealed in a flame after the bacilli have been planted upon the obliquely-cut surface of the potato; this prevents drying. According to Pawlowski, better results are obtained if the surface of the potato is moistened with a five-per-cent solution of glycerin. The growth is said to be seen at the end of about twelve days as grayish, dry-looking flakes; at the end of three or four weeks it forms a dry, smooth, whitish layer, and no further development occurs.

The range of temperature at which this bacillus will grow is very restricted; 37° C. is usually given as the most favorable point,

but Roux and Nocard say that the most favorable temperature appears to be 39° , and that development is slower at 37° .

The experiments of Koch, Schill and Fischer, and others show that the bacilli retain their vitality in desiccated sputum for several months (nine to ten months—De Toma); but they are said to undergo a gradual diminution in pathogenic virulence, which is more rapid when the desiccated material is kept at a temperature of 30° to 40° C. In the experiments of Cadéac and Malet portions of the lung from a tuberculous cow, dried and pulverized, produced tuberculosis in guinea-pigs at the end of one hundred and two days. They retain their vitality for a considerable time in putrefying material (forty-three days—Schill and Fischer; one hundred and twenty days—Cadéac and Malet). The resisting power of this bacillus against germicidal agents is also greater than that of certain other pathogenic microorganisms, but not so great as to justify the inference that it forms spores. It is not destroyed by the gastric juice in the stomach, as is shown by successful infection experiments in susceptible animals, by mixing cultures of the bacillus with their food (Baumgarten, Fischer), and also by experiments with an artificially prepared gastric juice (Falk). They are destroyed, in sputum, in twenty hours by a three-per-cent solution of carbolic acid, even when they present the appearance usually ascribed to the presence of spores (Cavagnis); also by absolute alcohol, a saturated aqueous solution of salicylic acid, saturated aniline water, etc. (Schill and Fischer). The more recent experiments of Yersin upon pure cultures of the bacillus gave the following results: "Tubercle bacilli, containing spores, were killed by a five-per-cent solution of carbolic acid in thirty seconds, by one-per-cent in one minute; absolute alcohol, five minutes; iodoform-ether, one per cent, five minutes; ether, ten minutes; mercuric chloride, 1:1,000 solution, ten minutes; thymol, three hours; salicylic acid, 2.5 per cent, six hours.

The tubercle bacillus appears to be especially susceptible to the action of light. In his address before the Tenth International Medical Congress (Berlin, 1890) Koch says that when exposed to direct sunlight the tubercle bacillus is killed in from a few minutes to several hours, according to the thickness of the layer; it is also destroyed by diffuse daylight in from five to seven days when placed near a window. This fact has an important hygienic bearing, especially in view of the fact that the tubercle bacillus is not readily killed by desiccation, putrefaction of the material containing it, etc. Tuberculous sputum expectorated upon sidewalks, etc., being exposed to the action of direct sunlight, will in many cases be disinfected by this agent by the time complete desiccation has occurred—i.e., before it is in a condition to be carried into the air as dust.

Sawizky has recently (1891) made a series of experiments to determine the length of time during which dried tuberculous sputum retains its virulence. He arrives at the conclusion that virulence is not suddenly but gradually lost, and that in an ordinary dwelling room dried sputum retains its specific infectious power for two and one-half months.

Metschnikoff states that when kept at a temperature of 42° C. for some time the tubercle bacillus undergoes a notable diminution in its pathogenic power, and that when kept at a temperature of 43° to 44° it after a time only induces a local abscess when injected subcutaneously into guinea-pigs. The experiments of Löte also indicate that an "attenuation of virulence" has occurred in the cultures preserved in Koch's laboratory, originating in 1882 from the lungs of a tuberculous ape. The author named made experiments with cultures from this source (ninetieth to ninety-fifth successive culture), and at the same time with a culture obtained from Roux, of Pasteur's laboratory. Rabbits inoculated with cultures from the last-mentioned source developed a hectic fever at the end of two weeks, and died tuberculous at the end of twenty-one to thirty-nine days. Twelve rabbits were inoculated with the cultures from Koch's laboratory; the injections were made either subcutaneously, into a vein, into the pleural cavity, or into the cavity of the abdomen. No elevation of temperature occurred in any of the animals, and they were found at the end of a month to have increased in weight. At the end of six weeks one of them was killed and tubercular nodules were found in various organs. The remaining animals were killed at the end of one hundred and forty-four to one hundred and forty-eight days. The two inoculated subcutaneously presented no sign of general tuberculosis, but a small yellow nodule containing bacilli was found at the point of inoculation. Those inoculated by injection into a vein showed one or two nodules in the lungs containing a few bacilli. In Koch's original experiments rabbits were killed by intravenous inoculation of his cultures in from thirteen to thirty-one days. That this attenuation of virulence depends upon a diminished production of a toxic product to which the bacillus owes its pathogenic power appears to be very certain, in view of the fact that the late cultures in a series have a more vigorous and abundant development than the more pathogenic cultures obtained directly from the animal body.

The discovery by Koch of a *toxine* in cultures of this bacillus, which is soluble in glycerin, and which in very minute doses produces febrile reaction and other decided symptoms when injected subcutaneously into tuberculous animals, must rank as one of the first

importance in scientific medicine, whatever the final verdict may be as to its therapeutic value in tubercular diseases in man.

The toxic substance contained in Koch's glycerin extract from cultures of the tubercle bacillus, now generally known under the name of tuberculin, is soluble in water, insoluble in alcohol, and passes readily through dialyzing membranes. It is not destroyed by the boiling temperature. According to the chemical examination of Jolles, the "lymph" contains fifty per cent of water and does not contain alkaloids or cyanogen compounds. It contains albuminates, which are thrown down as a voluminous white precipitate by tannic acid, and are redissolved by hot water containing sodium chloride and very diluted potash solution. The elementary analysis gave N 5.90 per cent, C 35.19 per cent, and H 7.02 per cent. The results obtained are believed to show that the active substance present in the lymph is a toxalbumin. In experiments made with Koch's lymph in Pasteur's laboratory by Bardach, a very decided elevation of temperature was produced in tuberculous guinea-pigs by the subcutaneous injection of 0.1 gramme, and a fatal result by the injection of 0.2 to 0.5 gramme. In man a decided febrile reaction is produced in tuberculous patients by very much smaller doses—0.001 cubic centimetre.

Hammerschlag, in his chemical researches, found that the tubercle bacillus yields a larger proportion of substances soluble in alcohol and ether than any other bacilli tested (twenty-seven per cent). The alcoholic extract contains fat, lecithin, and a toxic substance which produces convulsions in rabbits and guinea-pigs. The portion insoluble in alcohol and ether contains cellulose and an albuminoid substance. No ptomaines were found, but a toxalbumin was isolated, which caused an elevation of temperature in rabbits of 1° to 2° C., lasting for a day or two.

Hunter reports the following results of his chemical examination of tuberculin. It contains—

1. Albumoses, chiefly protoalbumose and deutoalbumose, along with heteroalbumose, and occasionally a trace of dysalbumose.

2. Alkaloidal substances, two of which can be obtained in the form of the platinum compounds of their hydrochlorate salts.

3. Extractives, small in quantity and of unrecognized nature.

4. Mucin.

5. Inorganic salts.

6. Glycerin and coloring matter.

The following conclusions are reached with reference to its toxic properties :

1. Tuberculin owes its activity, not to one principle, but to at least three, and probably more, different substances.

2. Its action in producing local inflammation, fever, and general constitutional disturbance is not a simple but an extremely complex one.

3. Its active ingredients are of the nature of albumoses, alkaloidal substances, and extractives. The action of these is in certain instances antagonistic.

4. Its remedial and inflammatory actions are connected with the presence of certain of its albumoses, while its fever producing properties are chiefly associated with substances of non-albuminous nature.

5. The albumoses are not lost by dialysis; the latter are. By the adoption of suitable methods it is thus possible to remove the substances which cause the fever, while retaining those which are beneficial in their action.

6. The fever produced by tuberculin is thus absolutely unessential to its remedial action.

In a recent communication (October, 1891) Koch has given a full account of his method of preparing crude tuberculin, and also the process by which he obtains from this a tuberculin which appears to be pure, or nearly so. To obtain considerable quantities of the crude product the tubercle bacillus is cultivated in an infusion of calves' flesh or of beef extract to which one per cent of peptone and four to five per cent of glycerin have been added. This culture liquid must be made slightly alkaline, and it is placed in flasks with a flat bottom, which should not be more than half-filled—thirty to fifty cubic centimetres. The inoculation is made upon the surface with small masses from a culture upon blood serum or glycerin-agar. By accident Koch discovered that these masses floating upon the surface give rise to an abundant development and to the formation of a tolerably thick and dry white layer, which finally covers the entire surface. At the end of six to eight weeks development ceases and the layer after a time sinks to the bottom, breaking up meanwhile into fragments. These cultures, after their purity has been tested by a microscopical examination, are poured into a suitable vessel and evaporated to one-tenth the original volume over a water bath. The liquid is then filtered through porcelain. The crude tuberculin obtained by this process contains from forty to fifty per cent of glycerin, and consequently is not a suitable medium for the development of saprophytic bacteria, if they should by accident be introduced into it. It keeps well and preserves its activity indefinitely.

From this crude tuberculin Koch has obtained a white precipitate, with sixty-per-cent alcohol, which has the active properties of the crude tuberculin as originally prepared. This is fatal to tuberculous guinea-pigs in doses of two to ten milligrammes. It is soluble in water and in glycerin, and has the chemical reactions of an albuminous body. In preparing it one and a half volumes of absolute alcohol are added to one volume of the crude tuberculin, and, after stirring it to secure uniform admixture, this is put aside for twenty-four hours. At the end of this time a flocculent deposit will be seen at the bottom of the vessel. The fluid above this is carefully poured off, and an equal quantity of sixty-per-cent alcohol poured into the vessel

for the purpose of washing the precipitate. This is again allowed to settle and the procedure is repeated three or four times, after which the precipitate is washed with absolute alcohol. It is then placed upon a filter and dried in a vacuum exsiccator.

An analysis of this purified tuberculin by Proskauer gave 18.46 per cent of ash, consisting almost entirely of potassium and magnesium phosphate. The elementary analysis gave 48.13 per cent of carbon, 7.06 per cent of hydrogen, 14.46 per cent of nitrogen, and 1.17 per cent of sulphur.

Recently (1892) Tizzoni and Cattani have presented some experimental evidence which indicates that injections of Koch's tuberculin into guinea-pigs may produce in these animals a certain degree of immunity against tuberculosis; and that this immunity depends upon the presence of an anti-tuberculin formed in the body of the partially immune animal.

Numerous experiments made by veterinary surgeons upon tuberculous cows show that the injection of Koch's tuberculin in these animals, in doses of thirty to forty centigrammes, produces a rise of temperature of from 1° to 3° C. The febrile reaction usually occurs in from twelve to fifteen hours after the injection. Its duration and intensity do not depend upon the extent of the tuberculous lesions, but is even more marked when these are slight than in advanced cases. In non-tuberculous animals no reaction occurs, and the experiments made justify the suspicion that tuberculosis exists if an elevation in temperature of a degree or more occurs as a result of the subcutaneous injection of the dose mentioned.

When the number of tubercle bacilli in sputum is comparatively small they may easily escape observation. Methods have therefore been suggested for finding them under these circumstances. Ribbert (1886) proposed the addition to the sputum of a two-per-cent solution of caustic potash, and boiling the mixture. The tenacious mucus is dissolved, and when the mixture is placed in a conical glass vessel the bacilli are deposited at the bottom and may easily be found in the sediment after removing the supernatant fluid. The same object is accomplished by Stroschein (1889) by the addition to sputum of three times its volume of a saturated solution of borax and boracic acid in water.

A method of *estimating the number* of bacilli in sputum has recently been proposed by Nuttall, which appears to give sufficiently accurate results and to be useful in judging of the progress of a case or of the results of treatment. For the details of this method we must refer to the author's paper (Johns Hopkins Hospital *Bulletin*, vol. xi., No. 13, 1891). It consists essentially in first making the sputum fluid by the addition of a solution of caustic potash; in

then shaking it thoroughly in a bottle containing sterilized gravel or pounded glass ; in carefully measuring the total quantity of fluid, and in dropping upon glass slides uniform drops by means of a graduated pipette ; in spreading these uniformly by means of a platinum needle and a turn table ; in covering the dried film with a film of blood serum, and coagulating this by heat ; and, finally, in staining and counting the bacilli in a series of slides from the same specimen, and from the average number found in a single drop estimating the total number in the sputum for twenty-four hours.

Pathogenesis.—Man, cattle, and monkeys are most subject to contract the disease naturally, and it may be communicated by inoculation to many of the lower animals—guinea-pigs, field mice, rab-

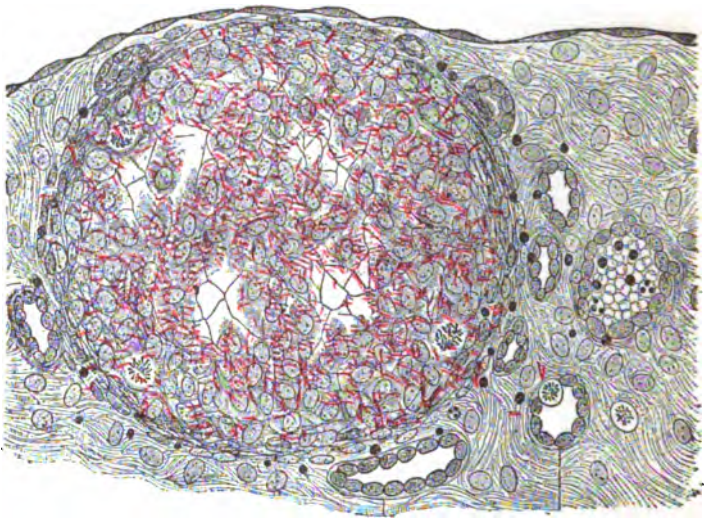


FIG. 131.—Limited epithelioid celled tubercle of the iris. $\times 950$. (Baumgarten.)

bits, and cats are among the most susceptible animals ; and in larger doses dogs, rats, white mice, and fowls may also be infected.

When tuberculous sputum is introduced beneath the skin of a guinea-pig the nearest lymphatic glands are found to be swollen at the end of two or three weeks, at the same time there is a thickening of the tissues about the point of inoculation ; later a dry crust forms over the local tuberculous tumefaction, and beneath this is a flattened ulcer covered with cheesy material. The animals become emaciated and show difficulty in breathing, and usually succumb to general tuberculosis, especially involving the lungs, within four to eight weeks. Injections of tuberculous sputum, or of pure cultures of the

bacillus, into the peritoneal cavity give rise to extensive tuberculosis of the liver, spleen, and lungs, and to death, as a rule, within three or four weeks. Rabbits are less susceptible to subcutaneous injections, but die within seventeen to twenty days when virulent—recent—cultures are injected into the circulation. As a result of such an inoculation the animal rapidly loses flesh and has a decided elevation of temperature, commencing at the end of the first week and increasing considerably during the last days of life. At the autopsy the spleen and liver are found to be greatly enlarged, but they do not contain any tubercles that can be recognized by the naked eye (Yersin). They contain, however, great numbers of tubercle bacilli, both free and in the cells. Injections of a small quantity of a pure culture into the anterior chamber of the rabbit's eye cause first iris-tuberculosis, followed by swelling and caseation of the nearest lymph glands, and finally general infection and death; when larger quantities are injected general tuberculosis is quickly developed. The influence of quantity—number of bacilli—is also shown in subcutaneous, intravenous, or intraperitoneal injections into guinea-pigs and rabbits (Hirschberger, Gebhardt, Wyssokowitsch). Thus rabbits which received less than one hundred and fifty bacilli, in sputum, in the experiments of Wyssokowitsch, did not develop tuberculosis; and in guinea-pigs the smaller the number injected the more protracted the course of the disease was found to be.

Tuberculosis in man no doubt results, in a large proportion of the cases, from the respiration, by a susceptible individual, of air containing the tubercle bacillus in suspension in a desiccated condition. As already stated, it has been demonstrated by experiment that the bacillus retains its vitality in desiccated sputum for several months. The experiments of Cornet have demonstrated that in the dust of apartments occupied by tuberculous patients tubercle bacilli are very commonly present in sufficient numbers to induce tuberculosis in guinea-pigs inoculated in the peritoneal cavity with such dust, while negative results were obtained from inoculations with dust from other localities. In view of these facts the usual mode of infection is apparent. Infection may also occur through an open wound or abrasion of the skin, as in the small, circumscribed tumors which sometimes develop upon the hands of pathologists as a result of handling tuberculous tissues. A few instances of accidental inoculation through wounds made by glass or earthen vessels containing tuberculous sputum have also been recorded. A more common mode of infection, especially in children, is probably by way of the intestinal glands, from the ingestion of the milk of tuberculous cows. That infection may occur by way of the intestine has been proved by experiments upon rabbits, which develop tuberculosis when fed upon

tuberculous sputum. And that the tubercle bacillus is frequently, if not usually, present in the milk of tuberculous cows has been proved by the experiments of Bollinger, Hirschberger, Ernst, and others. In Hirschberger's investigations milk from tuberculous cows induced tuberculosis in guinea-pigs, when injected subcutaneously or into the peritoneal cavity, in fifty-five per cent of the cases studied (twenty). The conclusion is reached that the milk may contain tubercle bacilli even when the udder of the cow is not involved. Ernst also, from an examination of the milk from thirty-six tuberculous cows in which the udder was apparently not involved, found the tubercle bacillus by microscopical examination in five per cent of the samples examined (one hundred and fourteen).

The prevalence of tuberculosis among cattle is shown by numerous investigations, and especially by the official inspections of slaughtered animals made in Germany. Thus in Saxony, in the year 1889, of 611,511 cattle examined 6,135 were found to be tuberculous (about one per cent); in Berlin, 1887-1888, out of 130,733 animals slaughtered 4,300 were found to be tuberculous (3.2 per cent). In view of the facts stated the great mortality from tubercular diseases among children, many of whom are removed from other probable sources of infection, is not difficult to understand, and the practical and simple method of preventing infection in this way, afforded by the sterilization (by heat) of milk used as food for infants, must commend itself to all.

54. BACILLUS TUBERCULOSIS GALLINARUM.

The researches of Maffucci (1889) and of Cadiot, Gilbert, and Roger (1890) show that the bacillus obtained from spontaneous tuberculosis in chickens, although closely resembling the bacillus of human tuberculosis, is not identical with it, varying especially in its pathogenic power. This view is sustained by the observations of Koch, who says in his address before the Tenth International Medical Congress (Berlin, 1890):

"The care which it is necessary to exercise in judging of the characters which serve to differentiate bacteria, even those which are well known, I have learned in the case of the tubercle bacillus. This species is so definitely characterized by its staining reactions, its growth in pure cultures, and its pathogenic qualities, and indeed by each of these characters, that it seems impossible to confound it with other species. Nevertheless in this case also one should not rely upon a single one of the characters mentioned for determining the species, but should follow the safe rule that all available characters should be considered, and the identity of a certain bacterium should only be regarded as demonstrated when it has been shown to correspond in all of these particulars. When I made my first researches with reference to the tubercle bacillus I was controlled by this rule, and tested tubercle bacilli from various sources, not only with reference to their staining reactions, but also with reference to their growth in culture media and

pathogenic characters. Only in the tuberculosis of chickens I was not able to apply this rule, as at that time it was not possible for me to obtain fresh material from which to make pure cultures. As, however, all other forms of tuberculosis had given identical bacilli, and the bacilli of chicken tuberculosis in their appearance and behavior towards the aniline colors entirely corresponded with these, I believed myself justified in assuming their identity, notwithstanding the incompleteness of the research. Later I received pure cultures from various sources, which apparently originated from tubercle bacilli, but in several regards differed from these; especially in the fact that inoculation experiments, made by experienced and reliable investigators, led to dissimilar results, which it was necessary to regard as unexplained contradictions. At first I believed that these differences depended upon changes such as are frequently observed in pathogenic bacteria, when these are cultivated in pure cultures outside of the body for a long time under more or less unfavorable conditions. In order to solve the riddle I attempted by various influences to change the common tubercle bacilli into the presumed variety referred to. They were cultivated for several months at so high a temperature that only a scanty growth was obtained; in other experiments still higher temperatures were allowed to act repeatedly for so long a time that the cultures were brought as nearly as possible to the point of killing the bacilli. In a similar way I subjected the cultures to the action of chemical agents, of light, or absence of moisture; they were cultivated for many generations in association with other bacteria; inoculated successively in animals having but a slight susceptibility. But, in spite of all these attempts, only slight variations were obtained in their characters—far less than other pathogenic bacteria undergo under similar circumstances. It appears, therefore, that the tubercle bacilli retain their characters with special obstinacy; this is in accord with the fact that pure cultures which have now been cultivated by me in test tubes for more than nine years, without in the meantime having been in a living body, are still entirely unchanged with the exception of a slight diminution of virulence. . . . It happened about a year ago that I received a living chicken which was suffering from tuberculosis, and I used this opportunity to make cultures directly from the diseased organs of this animal, which previously I had not been able to do. When the cultures grew I saw to my surprise that they had precisely the appearance and all of the characters possessed by the enigmatical cultures resembling those of the genuine tubercle bacillus. Later I learned that these also originated from tuberculosis in fowls, but, upon the assumption that all forms of tuberculosis are identical, had been considered genuine tubercle bacilli. A verification of my observations I find in the recently published researches of Prof. Maffucci with reference to tuberculosis of fowls."

According to Maffucci, adult chickens are refractory against the action of the *Bacillus tuberculosis* from man, and there are slight morphological and biological differences in the bacilli from the two sources.

Recently Cadiot, Gilbert, and Roger have made a series of experiments with the bacillus of tuberculosis in fowls. They found the bacilli to be very numerous in the livers of chickens suffering from spontaneous tuberculosis, and inoculated with material from this source six chickens, five rabbits, and twelve guinea-pigs. The chickens, when inoculated in the cavity of the abdomen or by injection into a vein, died in from forty-one to ninety-three days from general tuberculosis. Four of the rabbits died of general tuberculosis, presenting the same appearance as that following inoculation with

bacilli from human tuberculosis. Of the guinea-pigs, which were inoculated in the cavity of the abdomen, eleven remained in good health and one only died of general tuberculosis. These experiments show a decided difference in the pathogenic properties of tubercle bacilli from the two sources, for the guinea-pig is especially susceptible to tuberculosis as a result of similar inoculations with bacilli from human tuberculosis. We must therefore conclude that the bacillus found in spontaneous tuberculosis in fowls is a distinct variety of *Bacillus tuberculosis*. Whether this variety would cause tuberculosis in man, if introduced into susceptible subjects, has not been determined; and, as pointed out by Koch, this question can only be answered in the affirmative if it should be obtained in pure cultures from cases of human tuberculosis.

Since the above was written Maffucci has published (1892) an elaborate memoir upon tuberculosis of fowls. His conclusions are stated as follows:

"The bacillus of tuberculosis in fowls is distinguished from that of tuberculosis in mammals by the following points of difference:

"1. It does not induce tuberculosis in guinea-pigs, and seldom causes general tuberculosis in rabbits.

"2. The cultures in various media have a different appearance from those of the *Bacillus tuberculosis* of mammals.

"3. The temperature at which it develops varies between 35° and 45° C., and the thermal death-point is 70° C.

"4. At 45° to 50° C. the cultures show long, thick, and branched forms.

"5. The bacillus retains its vegetative and pathogenic power at the end of two years.

"6. This bacillus produces a substance which is toxic for guinea-pigs and is but slightly toxic for grown fowls.

"7. The tuberculosis produced in fowls by this bacillus is without giant cells."

55. *BACILLUS LEPRÆ*.

Discovered by Hansen (1879), chiefly in the interior of the peculiar round or oval cells found in leprous tubercles. Discovery confirmed by Neisser (1879) and by many subsequent observers.

While found chiefly in the leprous tubercles of the skin and mucous membranes, the bacilli have also been found in the lymphatic glands, the liver, the spleen, the testicles, and, in the anæsthetic form of the disease, in the thickened portions of nerves involved in the leprous process. Some observers have also reported finding them in the blood, but this appears to be quite exceptional. In the leprous cells they are commonly found in great numbers, and they may also be seen in the lymph spaces outside of these cells. They are not found in the epidermal layer of the skin, but, according to Babes, they may penetrate the hair follicles.

Morphology.—The bacillus of leprosy resembles the tubercle bacillus in form, but is of more uniform length and not so frequently

bent or curved. The rods have pointed ends; and in stained preparations unstained spaces, similar to those observed in the tubercle bacillus and generally assumed to be spores, are to be seen, although not quite so distinctly as in the latter. The bacilli are said by Flügge to be from four to six μ in length and less than one μ in width—probably considerably less, for the same author states that the tubercle bacillus has about the diameter of the bacillus of mouse septicæmia, and this is given as 0.2 μ .

This bacillus *stains* readily with the aniline colors and also by Gram's method. Although it differs from the tubercle bacillus in the ease with which it takes up the ordinary aniline colors, it resembles it in retaining its color when subsequently treated with strong solutions of the mineral acids. Double-stained preparations are therefore easily made by first staining sections or cover-

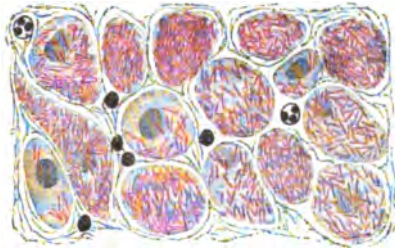


FIG. 122.—Section of a recent lepra nodule of the skin. $\times 950$. (Baumgarten.)

glass preparations in Ziehl's carbol-fuchsin solution or in an aqueous solution of methyl violet, decolorizing in acid, washing in alcohol, and counter-staining with methylene blue—or, if methyl violet was used in the first instance, with vesuvin.

Biological Characters.—The earlier attempts to cultivate this bacillus were without success, but recently Bordoni-Uffreduzzi has obtained from the marrow of the bones of a leper a bacillus which he believes to be the leprosy bacillus, and which he was able to cultivate upon blood serum to which a certain amount of peptone and of glycerin had been added. At first this bacillus only grew with difficulty and in the incubating oven; but after it had been cultivated artificially through a number of generations it is said to have grown upon ordinary nutrient gelatin at the room temperature. The bacillus obtained in this way is said to have retained its color when treated with acids, after having been stained with aniline-fuchsin, corresponding in this respect with the bacillus of leprosy and the tubercle bacillus. But it differed considerably in its morphology from the *Bacillus lepræ* as seen in the tissues of lepers, being considerably thicker, and it was not so promptly stained by the aniline colors as is the bacillus found in the tissues. Moreover, attempts to cultivate the same bacillus from leprosy tubercles of the skin were unsuccessful, as were also inoculation experiments into the anterior chamber of the eye in rabbits. It is therefore a matter of doubt as to whether the bacillus obtained by Bordoni-Uffreduzzi is identical with that present

in such numbers in the cells of the leprous tubercles, to which the name *Bacillus lepræ* has been given. Quite recently the announcement has been made that the "India Leprosy Commission" has succeeded in cultivating the leprosy bacillus in blister serum. Not having seen a detailed account of the experiments of this Commission, the writer is unable to estimate the value of their work and the reliability of the alleged success in cultivating this bacillus.

Some of the earlier observers described the bacillus of leprosy as motile, but this assertion seems to have been based upon some error of observation, and it is now generally agreed that, like the tubercle bacillus, it is without proper movements. The question of spore formation has not been definitely settled. As before remarked, unstained portions, occurring at regular intervals, are seen in the rods in stained preparations; but no satisfactory evidence has been presented to show that these are truly reproductive spores.

Pathogenesis.—The inference that the bacillus above described bears an etiological relation to the disease with which it is associated is based upon the demonstration of its constant presence in leprous tissues—which has now been repeatedly made in various and distant parts of the world—and of its absence from the same tissues involved in different morbid processes. As it has not been obtained in pure cultures, the final proof of such etiological relation is still wanting. We have, however, experimental evidence to show that leprous tissues containing this bacillus are infectious and may reproduce the disease. The experiment has been made upon man by Arning, who inoculated a condemned criminal subcutaneously with fresh leprous tubercles. The experiment was made in the Sandwich Islands, and the man was under observation until his death occurred from leprosy at the end of about five years. The first manifestations of the disease became visible in the vicinity of the point of inoculation several months after the experimental introduction of the infectious material.

Positive results have also been reported in the lower animals by Damsch, by Vossius, and by Melcher and Ortmann. The last-named investigators inoculated rabbits in the anterior chamber of the eye with portions of leprous tubercles excised for the purpose from a leper. The animals died from general infection at the end of several months, and the characteristic tubercles containing the bacillus were distributed through the various organs.

56. BACILLUS MALLEI.

Synonyms.—The bacillus of glanders; *Der Rotzbacillus*, *Ger.*; *Bacille de la morve*, *Fr.*

Discovered by Löffler and Schütz (1882), and proved to be the cause of glanders by the successful inoculation of pure cultures.

Found especially in the recent nodules in animals infected with glanders; also in the same after ulceration, and in the discharge from the nostrils, pus from the specific ulcers, etc.; sometimes in the blood of infected animals (Weichselbaum).

Morphology.—Bacilli with rounded ends, straight or slightly curved, rather shorter and decidedly thicker than the tubercle bacillus; usually solitary, but occasionally united in pairs, or in filaments containing several elements (in potato cultures). In stained preparations unstained or feebly stained spaces are seen in the rods, alternating with the deeply stained protoplasm of the cell. As in the tubercle bacillus, which presents a similar appearance, these spaces have been supposed by some bacteriologists to represent spores; but Löffler believes them to represent rather a degeneration of the protoplasm. Baumgarten and Rosenthal claim to have demonstrated the presence of spores by the use of Neisser's method of staining, but they do not consider it established that the unstained spaces in the rods referred to are of this nature.

The glanders bacillus may be *stained* with aqueous solutions of the aniline colors, but the staining is more intense when the solution



FIG. 123.—*Bacillus mallei*. $\times 1,000$. From a photomicrograph. (Fränkel and Pfeiffer.)

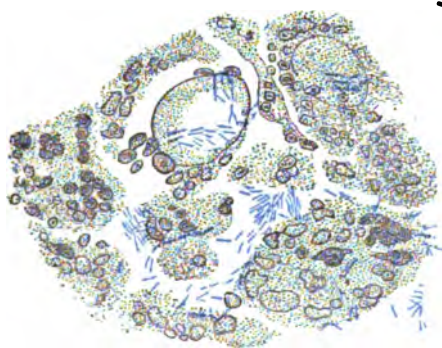


FIG. 124.—Section of a glanders nodule. $\times 700$. (Flügge.)

is made feebly alkaline. Add to three cubic centimetres of a 1:10,000 solution of caustic potash, in a watch glass, one cubic centimetre of a saturated alcoholic solution of an aniline color (methylene blue, gentian violet, or fuchsin); or the aniline-water-fuchsin, or methyl violet solution of Ehrlich may be used, with the addition just before use of an equal quantity of 1:10,000 solution of caustic potash. Löffler recommends that cover-glass preparations be placed in Ehrlich's solution and heated for five minutes; then decolorized in a one-

per-cent solution of acetic acid to which sufficient tropæolin has been added to give it the yellow color of Rhine wine; then quickly washed in distilled water. This bacillus presents the peculiarity of losing very quickly in decolorizing solutions the color imparted to it by the aniline staining solutions. For this reason the staining of the bacillus in sections is attended with some difficulty. Löffler recommends his alkaline methylene-blue solution for staining sections; and for decolorizing, a mixture containing ten cubic centimetres of distilled water, two drops of strong sulphuric acid, and one drop of a five-per-cent solution of oxalic acid. Thin sections should be left in this acid solution about five seconds. The method more recently recommended by Kühne also gives good results in skilful hands (see p. 34).

Biological Characters.—An *aërobic, non-motile, parasitic* bacillus, which may be cultivated in various artificial media at a temperature of 37° C. The lowest temperature at which development occurs (22° C.—Löffler) is a little above that at which nutrient gelatin is liquefied; the highest limit is 43° C. According to Fränkel, the glanders bacillus is a *facultative anaërobic*. Baumgarten and Rosenthal claim to have demonstrated the presence of spores by Neisser's method of staining. Löffler was led to doubt the formation of spores from the results of his experiments upon the thermal death-point of this bacillus, and its comparatively slight resistance to desiccation and destructive chemical agents. He found that exposure for ten minutes to a temperature of 55° C., or for five minutes to a three- to five-per-cent solution of carbolic acid, or for two minutes to a 1 : 5,000 solution of mercuric chloride, was effectual in destroying its vitality. As a rule, the bacilli do not grow after having been preserved in a desiccated condition for a few weeks; and in a moist condition the cultures cannot be preserved longer than three or four months—usually not so long as this (Löffler). The bacillus does not grow in infusions of hay, straw, or horse manure, and it is doubtful whether it finds conditions in nature favorable for its saprophytic existence. It grows, in the incubating oven, in neutral bouillon, in nutrient gelatin, or in nutrient agar, and still better in *glycerin-agar*. Upon the last-mentioned medium it grows, even at the room temperature (Kranzfeld), but better still in the incubating oven, as a pale-white, transparent streak along the line of inoculation, which at the end of six or seven days may have a width of seven to eight millimetres. According to Raskina, nutrient agar made with milk forms an extremely favorable medium, upon which a thick, pale-white layer develops in two or three days, which on the third or fourth day acquires an amber-yellow color, and the deeper layers acquire a brownish-red tint.

The growth upon solidified *blood serum*, in the course of three or

four days at 37° C., consists of yellowish, transparent drops, which later coalesce into a viscid layer, which has a milky appearance from the presence of numerous small crystals (Baumgarten). The growth upon cooked *potato* is especially characteristic. In the incubating oven, at the end of two or three days, a rather thin, yellowish, transparent layer develops, which resembles a thin layer of honey. Later this ceases to be transparent, and the amber color changes, at the end of six to eight days, to a reddish-brown color; and outside of the reddish-brown layer, with more or less irregular outlines, the potato for a short distance acquires a greenish-yellow tint.

Pathogenesis.—Glanders occurs principally among horses and asses, but may be contracted by man from contact with infected animals; it has also been communicated, in one instance with a fatal result, by subcutaneous inoculation, resulting accidentally from the use of an imperfectly sterilized hypodermic syringe which had previously been used for injecting cultures of the bacillus into guinea-pigs. The field mouse and the guinea-pig are especially susceptible to infection by experimental inoculations; the cat and the goat may be infected in the same way. Lions and tigers in menageries are said to have contracted glanders from being fed upon the flesh of infected animals (Baumgarten). Rabbits have but slight susceptibility, and the same is true of sheep and dogs; swine, cattle, white mice, and common house mice are immune.

The etiological relation of the bacillus is fully established by the experiments of Löffler and Schütz, confirmed by other bacteriologists, which show that pure cultures injected into horses, asses, and other susceptible animals, produce genuine glanders. The disease is characterized in the equine genus by the formation of ulcers upon the nasal mucous membrane, which have irregular, thickened margins and secrete a thin, virulent mucus; the submaxillary lymphatic glands become enlarged and form a tumor which is often lobulated; other lymphatic glands become inflamed, and some of them suppurate and open externally, leaving deep, open ulcers; the lungs are also involved and the breathing becomes hurried and irregular. In farcy, which is a more chronic form of the same disease, circumscribed swellings, varying in size from a pea to a hazelnut, appear on different parts of the body, especially where the skin is thinnest; these suppurate and leave angry-looking ulcers with ragged edges, from which there is an abundant purulent discharge. The specific bacillus can easily be obtained in pure cultures from the interior of suppurating nodules and glands which have not yet opened to the surface, and the same material will give successful results when inoculated into susceptible animals. But the discharge from the nostrils or from an open ulcer contains comparatively few bacilli; and as these are

associated with various other bacteria which grow more readily in our culture media, it is not easy to obtain pure cultures, by the plate method, from such material.

In the guinea-pig subcutaneous inoculation is followed in four or five days by tumefaction at the point of inoculation, and after a time a prominent tumor with caseous contents is developed ; ulceration of the skin follows, and a chronic, purulent ulcer with irregular, indurated margins results ; after a time this may cicatrize. Meanwhile the lymphatic glands become involved, and the symptoms of general

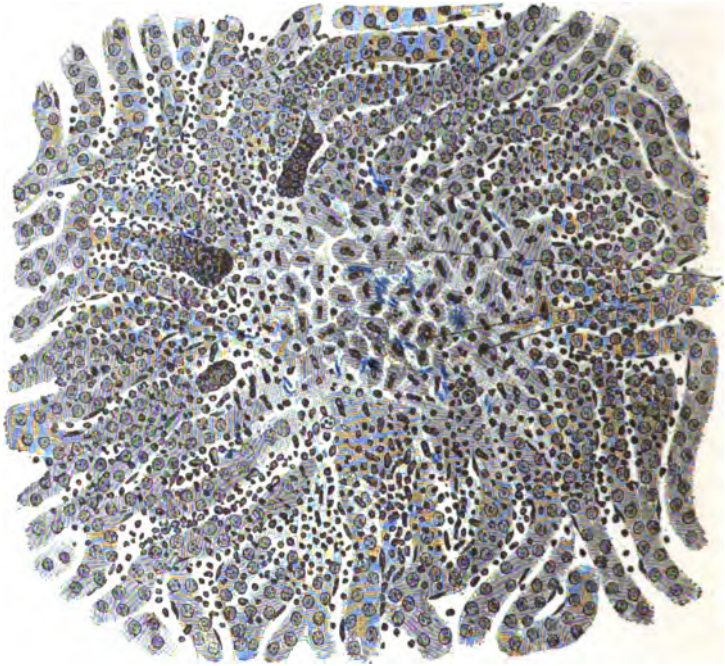


FIG. 125.—Section through a glandular nodule in liver of field mouse. Tissue $\times 250$. Bacilli $\times 500$. (Baumgarten.)

infection are developed at the end of four or five weeks ; the glands suppurate, and in males the testicles are also involved ; finally a diffuse inflammation of the joints occurs, and death results from exhaustion. In the guinea-pig the specific ulcers upon the nasal mucous membrane, which characterize the disease in the horse, are rarely developed to any great extent.

In field mice general infection occurs at once as a result of the subcutaneous injection of a small quantity of a pure culture, and the animal dies at the end of three or four days. Upon post-mortem

examination the principal changes are found in the liver and in the greatly enlarged spleen. Scattered through these organs are minute gray points which are scarcely visible to the naked eye. In the guinea-pig, which succumbs at a later date, these nodules are larger and closely resemble miliary tubercles, both macroscopically and under the microscope, in stained sections of the tissues. Similar nodules are also found in the kidneys and in the lungs; they have a decided tendency to undergo purulent degeneration. The bacilli are found principally in these nodules, of recent formation, and are commonly associated in groups, as if they had been enclosed in the interior of a cell the membranous envelope of which had undergone degeneration and disappeared.

As before remarked, it is not an easy matter to demonstrate the bacillus in sections of the tissues containing these nodules, owing to the facility with which they lose their color in alcohol and other decolorizing agents. For this reason it will be best to dehydrate sections by the use of aniline oil (Weigert's method) or to resort to Kühne's method of staining.

It is also difficult to demonstrate the presence of the bacillus in nodules which have undergone purulent degeneration, in the secretions from the nostrils of horses suffering from glanders, or in the pus from the specific ulcers and suppurating glands; for they are present in comparatively small numbers. But the virulent nature of these discharges is shown by inoculations into guinea-pigs or mice, and it is easier to obtain a pure culture from such virulent material by first inoculating a susceptible animal than directly by the plate method; for the small number of bacilli present, and their association with other bacteria which develop more rapidly in our culture media, make this a very uncertain procedure. For establishing the diagnosis of glanders, therefore, Löffler recommends the inoculation of guinea-pigs with pus from a suppurating gland or ulcer, or the nasal discharge from a suspected animal, rather than a direct attempt to demonstrate the presence of the bacillus by staining and culture methods.

The method proposed by Strauss gives more prompt results. This consists in the intraperitoneal injection of cultures or of the suspected products into the cavity of the abdomen of male guinea-pigs. If the glanders bacillus is present the diagnosis may be made within three or four days from the infectious process established in the testicles. At the end of this time the scrotum is red and shining, the epidermis desquamates, and suppuration occurs, the pus sometimes perforating the integument. This pus is found to contain the glanders bacillus. The animal usually dies in the course of twelve to fifteen days. When the animals are killed three or four days

after the inoculation, the two layers of the tunica vaginalis testis are found to be covered with a purulent exudate containing the glanders bacillus and to be more or less adherent. Even as early as the second day the tunica vaginalis is seen to be covered with granulations.

An attenuation of virulence occurs in cultures which have been kept for some time, and inoculations with such cultures may give a negative result; or, when considerable quantities are injected, may produce a fatal result at a later date than is usual when small amounts of a recent culture are injected into susceptible animals.

Kalning, Preusse, and Pearson have obtained from cultures of the glanders bacillus a glycerin extract similar to the crude tuberculin of Koch—*mallein*. This, when injected into animals suffering from glanders, gives rise to a considerable elevation of temperature, and it has been proposed to use it as a means of diagnosis in cases of suspected infection in animals in which the usual symptoms have not yet manifested themselves. The value of the test has already been demonstrated by the experiments of Heyne, Schilling, and others.

57. BACILLUS OF LUSTGARTEN.

Synonym.—Syphilis bacillus.

Found by Lustgarten (1884) in syphilitic lesions and in secretions of syphilitic ulcers, and believed by him to be the specific infectious agent in this disease. No satisfactory experimental evidence that this is the case has yet been obtained.

Morphology.—Straight or curved bacilli, which bear considerable resem-

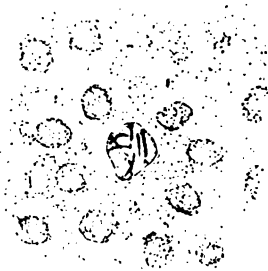


FIG. 126.

FIG. 126.—Migrating cell containing syphilis bacilli. (Lustgarten.)



FIG. 127.

FIG. 127.—Pus from hard chancre containing syphilis bacilli. (Lustgarten.)

blance to tubercle bacilli, but differ from them in the staining reactions. They are usually more or less curved, or bent at a sharp angle, or S-shaped; the ends often present slight knob-like swellings; the length is from three and one-half μ to four and one-half μ , and the diameter is from 0.25 to 0.3 μ . With a high power the contour is seen to be not quite regular, but wavy in outline, and bright, shining spaces in the deeply stained rods may be observed; these, from two to four in a single rod, are believed by Lustgarten

to be spores. The bacilli are not found free in the tissues, but are enclosed in cells of a round-oval or polygonal form, which are said to be about double the size of a white blood corpuscle. The bacilli are not numerous, and very commonly only one or two are found in a single cell, but groups of six or eight may sometimes be seen, especially upon the margins of a syphilitic lesion, and in the tissues in the immediate vicinity of the infiltration, which show but little change or are apparently healthy (Lustgarten).

The presence of these bacilli in syphilitic lesions was demonstrated by Lustgarten by the following *staining method*: The thin sections are placed in the Ehrlich-Weigert gentian-violet solution (one hundred parts aniline water, eleven parts saturated alcoholic solution of gentian violet) for from twelve to twenty-four hours at the room temperature, and two hours in the incubating oven at 40° C. The sections are then thoroughly washed in alcohol and placed for ten seconds in a 1.5-per-cent solution of potassium permanganate; in this solution a precipitate of peroxide of manganese is formed, which adheres to the section; this is dissolved and washed off in a dilute aqueous solution of pure sulphuric acid; the sections are then washed in water, and, if not completely decolorized, are returned for a few seconds to the permanganate solution and again washed off in the acid; it may be necessary to repeat this operation three or four times. Finally the sections are dehydrated and mounted in balsam in the usual manner. Cover-glass preparations are made in the same way, except that, after being taken from the staining solution, they are washed off in water instead of in alcohol.

Another method of staining, recommended by De Giacomini, consists in placing the sections for twenty-four hours in aniline-water-fuchsin solution (cover-glass preparations may be stained in the same solution, hot, in a few minutes), then washing them in water, and decolorizing in a solution of perchloride of iron—first in a dilute and then in a saturated solution.

The method of staining employed by Lustgarten serves to differentiate his bacillus from many other microorganisms, but not from the tubercle bacillus and the bacillus of leprosy, which, as he pointed out, may be stained in the same way. And it has since been shown by Alvarez and Tavel, and by Matterstock, that in smegma from the prepuce or the vulva, bacilli are found which have the same staining reaction and are similar in their morphology to the bacillus of Lustgarten. This by no means proves that the smegma bacilli found under the prepuce of healthy persons are identical with the bacilli found by Lustgarten and others in sections of tissues involved in syphilomata. In the absence of pure cultures and inoculation experiments it is impossible to establish identity, however similar may be the characters referred to. Several well-known pathogenic bacilli resemble quite as closely in these particulars other bacilli which have, nevertheless, been differentiated from them by culture and inoculation experiments. We may mention especially in this connection the bacillus of diphtheria, as obtained from the pseudo-membranous exudation in a genuine case of this disease, and the pseudo diphtheria bacilli found by Roux and Yersin in the fauces of healthy children. On the other hand, since it has been shown that similar bacilli are common in preputial smegma, we cannot attach great importance to the finding of Lustgarten's bacillus in primary syphilitic sores; and it has not been found in sufficient numbers, or with sufficient constancy, by those who have searched for it subsequently to the publication of Lustgarten's investigations, to give strong support to the view that it is the specific infectious agent in syphilis. Baumgarten, who has searched in vain for Lustgarten's bacillus in uncomplicated visceral syphilomata, suggests that the bacilli found occasionally in such lesions were perhaps tubercle bacilli and represented a mixed infection. As the bacillus under consideration has not been obtained in cultures, we have no information as to its biological characters and pathogenesis.

THE SYPHILIS BACILLUS OF EVE AND LINGARD.

Eve and Lingard (1886) report that they have obtained in cultures from

the blood and diseased tissues of syphilitics who have not undergone mercurial treatment, bacilli which in their form and dimensions resemble the tubercle bacilli, but which stain readily by the common aniline colors and by Gram's method, and are not stained by Lustgarten's method. They grow readily upon solidified blood serum, forming a thin, pale-yellow or brownish-yellow layer. Inoculations of pure cultures into apes were without result. The negative results which have attended the culture experiments and microscopical examinations of the blood and diseased tissues, made by many competent bacteriologists in other parts of Europe, make it appear probable that the bacilli described by the English investigators named belong to some saprophytic species, and that they are not usually present in syphilomata or the blood of syphilitic patients.

MICROCOCCHI OF DISSE AND TAGUCHI.

Disse and Taguchi (1886) claim to have obtained from the blood of syphilitics micrococci which they were able to cultivate in artificial media at 20 to 40° C., and which formed on the surface of such media a grayish-white layer consisting of diplococci which are motile and of larger motionless cocci. The diplococci are said to originate from division of the larger cocci. Inoculations into rabbits, dogs, and sheep gave rise to chronic interstitial inflammatory processes in the lungs and liver, to granulomata in various organs, and to fatty degenerative changes in the walls of the arteries, which, in the opinion of the authors named, correspond with the pathological changes produced by syphilitic infection in man. We remark, with reference to the supposed etiological relation of this coccus, that bacteriologists in Europe have not confirmed the authors named as to the presence of this micrococcus in the blood of syphilitics, and that the micrococcus of progressive granuloma formation described by Manfredi produces similar pathological changes in inoculated animals; also that there is no evidence that the animals experimented upon are subject to syphilitic infection.

58. BACILLUS OF RHINOSCLEROMA (?).

First observed by Von Frisch (1882) in the newly formed tubercles of rhinoscleroma. Cultivated by Paltauf and Von Eiselberg (1886).

Rhinoscleroma is a chronic affection of the skin, and especially of the mucous membrane of the nares, which is characterized by the formation of tubercular thickenings of the skin and tumefaction of the nasal mucous membrane, followed sometimes by ulceration. It prevails in Italy, Austria, and to a slight extent in some parts of Germany. Pathologists generally regard it as an infectious process, although this has not been proved.

The bacilli, first described by Von Frisch, appear to be constantly present in the newly formed tubercles. They are commonly found in certain large hyaline cells peculiar to the disease, and may also be observed in the lymphatic vessels or scattered about in the involved tissues.

Morphology.—Short bacilli with rounded ends, usually united in pairs, and surrounded by a gelatinous capsule resembling that of Friedländer's bacillus. According to Eisenberg, the bacilli are two to three times as long as broad, and may grow out into filaments.

These bacilli stain readily with the aniline colors and by Gram's method. The capsule may be demonstrated by the methods usually employed in staining Friedländer's bacillus, or by the following method which is especially recommended by Alvarez: The excised portions of tissue involved in the disease are placed for twenty-four hours in a one-per-cent solution of osmic acid and then in absolute alcohol. When properly hardened thin sections are made; these are stained in a hot solution of aniline-water-methyl-violet for a few minutes, and then decolorized, by Gram's method, in iodine solution.

Biological Characters.—An *aërobic, non-motile, non-liquefying* bacillus (facultative anaërobic?).

In gelatin stick cultures the growth resembles that of Friedländer's bacillus—*i.e.*, a nail-like growth, consisting of densely crowded, opaque colonies along the line of puncture, and a heaped-up, white, glistening mass upon the surface, hemispherical in form and viscous in consistence. Upon *gelatin* plates yellowish-white, spherical colonies are developed within two or three days, which under the microscope are seen to be granular. Upon *potato* a cream-like growth occurs along the line of inoculation, which is white or yellowish-white in color, and in which gas bubbles may be developed. Development is most rapid at a temperature of 35° to 38°, but also occurs at the room temperature.

Pathogenesis.—The etiological relation of this bacillus to the disease with which it is associated has not been established. It is pathogenic for mice and for guinea-pigs, less so for rabbits; in this regard, as in its morphology and growth in various culture media, it bears a close resemblance to Friedländer's bacillus, which is also found not infrequently in the nasal secretions of healthy persons and in those suffering from chronic nasal catarrh or ozæna.

The principal points of difference, as pointed out by Baumgarten, are as follows: The bacillus of rhinoscleroma is usually more decidedly rod-shaped



FIG. 138.—Bacillus of rhinoscleroma in lymphatic vessels of the superficial part of tumor. $\times 1,200$. (Cornall and Babes)

than Friedländer's bacillus, although both may be of so short an oval as to resemble micrococci. The first-mentioned bacillus constantly presents the appearance of being surrounded by a transparent capsule, even in the cultures in artificial media, while Friedländer's bacillus in such media does not usually present this appearance, unless as a result of special treatment. Finally, the bacillus of rhinoscleroma may retain its color, in part at least, when treated by Gram's method, while Friedländer's bacillus is completely decolorized when placed in the iodine solution employed in this method.

Notwithstanding these points of difference, Baumgarten is not entirely satisfied that this bacillus is a distinct species, and several bacteriologists have maintained that it is identical with the bacillus of Friedländer.

59. BACILLUS OF KOUBASOFF.

Obtained by Koubasoff (1889) from new growths in the stomach of a person who died of cancer of the stomach.

Morphology.—Bacilli with round ends, or with one end pointed, two or three times as long as the tubercle bacillus and three or four times as thick.

Stains readily with the aniline colors.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Forms spores in the centre of the rods. Grows in the usual culture media at the room temperature, more rapidly at 36° C.

In stick cultures in *glycerin-gelatin* the growth resembles an inverted stethoscope; at the surface a circular, bluish membrane is formed, which is depressed in the form of a funnel, while along the line of puncture a slender, yellowish, jagged column is developed. Upon *agar*, at 36° C., a bluish-white layer is quickly developed. Upon *potato* the growth resembles that of the typhoid bacillus at first; later a granular membrane is formed; under a low power the granules appear to be formed of intertwined masses of filaments. The growth upon *blood serum* is similar to that upon *agar*.

Pathogenesis.—Subcutaneous injections in guinea-pigs cause their death in one to two weeks, in rabbits in one to two months, in cats and dogs in two months or more. Death occurs in a shorter time in animals which have been fed upon cultures than as a result of subcutaneous injections. The animals become very much emaciated and have paralysis of the sphincter muscles. At the autopsy flat or nodular elevations, which are often ulcerated, are seen here and there upon the mucous membrane of the stomach and intestine; the mesentery, especially of the small intestine, is hyperæmic; the mesenteric glands are swollen, as are also the inguinal glands. In the liver and sometimes in the ovary, uterus, and spleen larger or smaller nodules are seen.

60. BACILLUS OF NOCARD.

Obtained by Nocard (1888) from pus collected from the superficial abscesses in cattle suffering from a chronic infectious disease which prevails especially upon the island of Guadeloupe—known as “*farcin du bœuf*”; *Ger.* “*Wurmkrankheit*.”

Morphology.—A long and slender bacillus, about as thick as the bacillus of rouget (*Bacillus mrisepiticus*); usually seen in tangled masses which consist of an opaque central portion surrounded by long filaments, which apparently give off lateral ramifications. (This description of the morphology gives rise to the suspicion that the microorganism described by Nocard is a microscopic fungus rather than a bacillus.) According to Nocard, the branching is more apparent than real, and is in fact a false dichotomization, such as is seen in the genus *Cladotrix*.

Stains best by Weigert's method; is decolorized by Gram's method. Does not stain readily with most aniline colors.

Biological Characters.—An *aërobic*, *non-motile* bacillus, which does not grow in nutrient gelatin at the room temperature. Grows in the usual culture media at a temperature of 30° to 40° C. Forms small oval spores. Is destroyed in ten minutes by a temperature of 70° C. Upon the *surface of agar* it forms irregular, opaque, yellowish-white colonies, which are thickest at the margin, have a dull, dusty-looking, mammillated surface, and after a time become confluent, forming a thick, wrinkled, membranous layer. Upon *potato* development is rapid in the form of prominent, dry, pale-yellow plaques. In *bouillon* whitish flocculi are formed, most of which fall to the bottom, while some float upon the surface, where they form dry, dusty-looking, rounded pellicles of a dirty-gray color with a greenish reflection.

Pathogenesis.—The guinea-pig is the most susceptible animal. When injected into the peritoneal cavity of one of these animals it produces, in from nine to twenty days, lesions which closely resemble those of miliary tuberculosis. At the autopsy the peritoneum is found to be covered with nodules, in the centre of which the bacillus is found in tangled masses; the liver, spleen, kidneys, and intestine are also studded with pseudo-tubercles, but these are only found in the peritoneal coat and not in the parenchyma of the various organs, or in the organs of the thoracic cavity. Intravenous injections give rise to lesions similar to those of general miliary tuberculosis, the organs generally containing a considerable number of nodules, in the centre of which tufts of bacilli are found. In cattle and sheep similar lesions result from intravenous injections, but without causing the death of the animal. The dog, the cat, the horse, the ass, and the rabbit are immune. Subcutaneous inoculations in guinea-pigs produce an extensive local abscess, followed by a chronic induration of the neighboring lymphatic glands.

XII.

BACILLI WHICH PRODUCE SEPTICÆMIA IN SUSCEPTIBLE ANIMALS.

WHEN, as a result of accidental (natural) or experimental inoculation, a microorganism is introduced into the body of a susceptible animal which is able to multiply in its blood, producing a general infection, we speak of this general blood infection as a *septicæmia*. When pathogenic microorganisms which are unable to multiply in the blood establish themselves in some particular locality in the animal body which is favorable for their growth, and by the formation of toxic products, which are absorbed, give rise to general symptoms of poisoning, we designate the affection *toxæmia*. As examples of this mode of pathogenic action we may mention diphtheria and tetanus. As a rule, the various forms of septicæmia are quickly fatal, and, as the microorganisms to which they are due multiply in the blood of the infected animal, this fluid possesses infectious properties, and, when inoculated in the smallest quantity into another susceptible animal, reproduces the same morbid phenomena. A typical example of this class of diseases is found in anthrax, to which disease a special section has already been devoted (VII.). But in this and other forms of septicæmia subcutaneous inoculations do not, as a rule, result in the immediate invasion of the blood by the parasitic microorganism. Often a local inflammatory process of considerable extent is first induced; and in some cases general infection only occurs a short time before the death of the animal, depending, perhaps, upon a previous toxæmia from the absorption of toxic products developed at the seat of local infection. The pathogenic action, then, in acute forms of septicæmia appears to result, not alone from the presence and multiplication of the pathogenic microorganism in the blood, but also from the toxic action of products evolved during its growth.

Some of the pathogenic bacilli of this class now known to bacteriologists have been discovered by studying the infectious diseases induced by them in lower animals among which these diseases prevail naturally—*i.e.*, independently of human interference. Many

more are known to us from experiments made in pathological laboratories, in testing by inoculations into animals bacteria obtained from various sources, with reference to their pathogenic power. We include in this group only those bacilli which induce fatal septicæmia in susceptible animals when injected into the circulation or subcutaneously in a comparatively small quantity—*e.g.*, less than half a cubic centimetre of a bouillon culture.

61. BACILLUS SEPTICÆMIÆ HÆMORRHAGICÆ.

Synonyms.—Bacillus of fowl cholera; Microbe du choléra des poules (Pasteur); Bacillus cholerae gallinarum (Flügge); Bacillus der Hühnercholera; Bacillus of rabbit septicæmia; Bacillus cuniculicida (Flügge); Bacillus der Kaninchenseptikämie (Koch); Bacillus der Rinderseuche (Kitt); Bacillus der Schweineseuche (Löffler and Schütz); Bacillus der Wildseuche (Hueppe); Bacillus der Büffelseuche (Oreste-Armanni); (Bacterium of Davaine's septicæmia?)

It is now generally admitted by bacteriologists that Koch's bacillus of rabbit septicæmia (1881) is identical with the bacillus ("micrococcus") of fowl cholera previously described by Pasteur (1880). The similar bacilli found in the blood of animals dead from the infectious diseases known in Germany as Wildseuche (Hueppe), Rinderseuche (Kitt), Schweineseuche (Schütz), and Büffelseuche (Oreste-Armanni) appear also to be identical with the bacillus of rabbit septicæmia and fowl cholera. This view is sustained by Hueppe and by Baumgarten, and by the recent comparative researches of Caneva (1891) and of Bunzl-Federn (1891).

This is evidently a widely distributed pathogenic bacillus; it was obtained by Koch from rabbits inoculated with putrefying flesh infusion, by Gaffky from impure river water, and by Pasteur from the blood of fowls suffering from the infectious disease known in France as *choléra des poules*. It is not infrequently found in putrefying blood, and its presence in the salivary secretions of man has occasionally been demonstrated (Baumgarten).

FIG. 129 Bacillus septicæmiæ hæmorrhagiæ in the blood of a rabbit. $\times 950$. (Baumgarten.)

With reference to the American swine plague described by Salmon and Smith, we are informed by Smith, in his most recent publication upon the subject (*Zeitschrift für Hygiene*, Band x., page 493), that cultures of the German Schweineseuche bacillus, received from the Berlin Hygienic Institute, compared with his cultures from infected swine in this country, agreed in all particulars, except that the former were decidedly more pathogenic for swine and for rabbits.

It appears extremely probable that the form of septicæmia studied

by Davaine (1872), which he induced in the first instance by injecting putrid ox blood into rabbits, was due to the same pathogenic bacillus. The writer obtained this bacillus (1887) in Cuba from the blood of rabbits inoculated with liver tissue taken from a yellow-fever cadaver and kept for forty-eight hours in an antiseptic wrapping. The name which we have adopted is that proposed by Hueppe for the form of septicæmia to which it gives rise—"Septikæmia hæmorrhagica."

Morphology.—Short bacilli with rounded ends, from 0.6 to 0.7 μ in diameter and about 1.4 μ long; sometimes united in pairs, or in chains of three or four elements. In stained preparations the extremities are usually stained, while the central portion of the rod remains unstained. This "end staining" causes the rods to present the appearance of diplococci when examined with a comparatively low power, and some of the earlier observers described the microorganism under consideration as a micrococcus. It is quickly stained by the aniline colors usually employed, but loses its color when treated by Gram's method.

Biological Characters.—A *non-motile, aërobic, non-liquefying* bacillus. Does not form spores. Grows in various culture media at the room temperature, but more rapidly at 35° to 37° C.—the lowest temperature at which development occurs is about 13° C. Although this is an aërobic bacillus and a certain amount of oxygen is necessary for its development, it appears to grow better when the amount is somewhat restricted than it does on the surface of nutrient media.

Upon *gelatin plates*, at the end of two or three days, small, white colonies are developed upon or near the surface; these are finely granular and spherical, with a more or less irregular outline, and by transmitted light have a yellowish color; later the central portion of the colonies is of a yellowish-brown color and is surrounded by a transparent peripheral zone. The superficial colonies are commonly smaller than those which develop a little below the surface of the gelatin. In *stick cultures* in nutrient gelatin the growth upon the surface consists of a thin, whitish layer in the vicinity of the point of puncture, having an irregular, jagged outline—sometimes there is no development upon the surface; along the line of puncture the growth consists of rather transparent, discrete or confluent colonies. In *streak cultures* upon nutrient agar, or gelatin, or blood serum the growth is limited to the immediate vicinity of the line of inoculation, and consists of finely granular, semi-transparent colonies, which form a thin, grayish-white layer with irregular, somewhat thickened margins. Upon *potato* no development occurs, as a rule, at the room temperature, but in the in-

cubating oven a rather thin, transparent, grayish-white or yellowish, waxy layer is developed in the course of a few days. According to Bunzl-Federn, the bacillus of fowl cholera and that of rabbit septicæmia grow upon potato, while the bacillus of Wildseuche, Schweineseuche, and Büf-



FIG. 130. — *Bacillus septicæmiæ hæmorrhagicæ*; stick culture in nutrient gelatin, end of four days at 16°-18° C. (Baumgarten.)

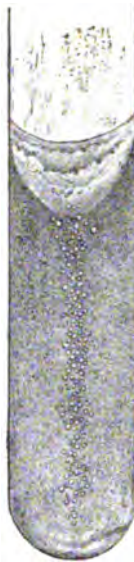


FIG. 131. — *Bacillus* of Schweineseuche; old stick culture in nutrient gelatin. (Schütz.)

felseuche do not. According to Caneva, none of the bacilli of this group grow upon potato. The same author states that the growth in milk is scanty and does not produce coagulation, while Bunzl-Federn finds that the bacillus of fowl cholera and of rabbit septicæmia produce coagulation and the others do not. These differences are not, however, considered by the author last named as sufficient to establish the specific difference of the bacilli from these different sources. He looks upon them rather as varieties of the same species. Bunzl-Federn has also ascertained that when cultivated in a peptone solution all of the bacilli of this group, with the exception of that obtained from the so-called Büffelseuche, give the reaction for phenol and for indol—the bacillus of Büffel-

seuche gives the indol reaction only. Development in *bouillon* is rapid and causes a uniform turbidity of the fluid. Cultures of this bacillus may retain their vitality for three months or more when kept in a moist condition; but the bacillus usually fails to grow after having been kept for a few days in a desiccated condition; according to Hueppe, it may resist desiccation for fourteen days. The *thermal death-point*, as determined by Salmon for the bacillus of fowl cholera, is 56° C., the time of exposure being ten minutes (55° C. with fifteen minutes' exposure—Baumgarten). It is not readily destroyed by putrefaction (Kitt). A solution of mercuric chloride of 1 : 5,000 destroys it in one minute, and a three-per-cent solution of carbolic

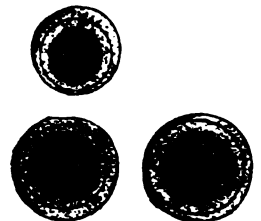


FIG. 132. — *Bacillus* of swine plague; colonies on gelatin plate, end of seven days. $\times 60$. (Smith.)

destroys it in one minute, and a three-per-cent solution of carbolic

acid in six hours (Hueppe). Pasteur (1880) has shown that when cultures of this bacillus (microbe of fowl cholera) in bouillon are kept for some time they gradually lose their pathogenic virulence, and he has ascribed this "attenuation of virulence" to the action of atmospheric oxygen. He also ascertained that the particular degree of virulence manifested by the mother culture after a certain interval could be maintained in successive cultures made at short intervals. He was thus able to cultivate different pathogenic varieties, and to use these in making protective inoculations, by which susceptible animals were preserved from the effects of virulent cultures injected subsequently.

Attenuated cultures recover their virulence when inoculated into very susceptible animals. Thus a culture which would produce a non-fatal and protective attack in a chicken may, according to Pasteur, kill a small bird, like a sparrow; and by successive inoculations from one sparrow to another the original degree of virulence may be restored, so that a minute quantity of a pure culture would be fatal to a chicken.

Pathogenesis.—Pathogenic for chickens, pigeons, pheasants, sparrows, and other small birds, for rabbits and mice, also for swine (Schweineseuche), for cattle (Rinderseuche), and for deer (Wildseuche). Subcutaneous injection of a minute quantity of a virulent culture usually kills chickens within forty-eight hours. Some time before death the fowl falls into a somnolent condition, and, with drooping wings and ruffled feathers, remains standing in one place until it dies. Infection may also occur from the ingestion of food moistened with a culture of the bacillus or soiled with the discharges from the bowels of other infected fowls. At the autopsy the mucous membrane of the small intestine is found to be inflamed and studded with small hæmorrhagic foci, as are also the serous membranes; the spleen is notably enlarged. The bacilli are found in great numbers in the blood, in the various organs, and in the contents of the intestine. In rabbits death commonly occurs in from sixteen to twenty hours, and is often preceded by convulsions. The temperature is elevated at first, but shortly before death it is reduced below the normal. The post-mortem appearances are: swelling of the spleen and lymphatic glands; ecchymoses or diffuse hæmorrhagic infiltrations of the mucous membranes of the digestive and respiratory passages, and in the muscles; and at the point of inoculation a slight amount of inflammatory cedema. The bacilli are found in considerable numbers in the blood within the vessels, or in that which has escaped into the tissues by the rupture of small veins. They are not, however, so numerous as in some other forms of septicæmia—*e.g.*, anthrax, mouse septicæmia—when an examination is made imme-

diately after death ; later the number may be greatly increased as a result of post-mortem multiplication within the vessels. The rabbit is so extremely susceptible to infection by this bacillus that inoculation in the cornea by a slight superficial wound usually gives rise to general infection and death. This animal may also be infected by the ingestion of food contaminated with a culture of the bacillus. It is by this means that Pasteur proposed to destroy the rabbits in Australia, which have multiplied in that country to such an extent as to constitute a veritable pest. Both in fowls and in rabbits the disease may under certain circumstances run a more protracted course—*e.g.*, when they are inoculated with a small quantity of an attenuated culture. In less susceptible animals—guinea-pigs, sheep, dogs, horses



FIG. 132.—Bacillus of Schweineseuche, in blood of rabbit. (Schütz.)

—a local abscess, without general infection, may result from the subcutaneous injection of the bacillus ; but these animals are not entirely immune. In the infectious maladies of swine, cattle, deer, and other large animals to which reference has been made, and which are believed to be due to the same bacillus, the symptoms and pathological appearances do not entirely correspond with those in the rabbit or the fowl ; but the bacillus as obtained from the blood of such animals corresponds in its morphological and biological characters with Pasteur's microbe of fowl cholera and Koch's bacillus of rabbit septicæmia, and pure cultures from the various sources mentioned are equally fatal to rabbits and to fowls. In the larger animals pulmonary and intestinal lesions are developed, and in swine a diffused red color of the skin, similar to that observed in the disease known in Germany as Schweinerothlauf (Fr. rouget), is sometimes seen.

According to Baumgarten, bacilli from Wildseuche or from Rinderseuche inoculated into swine give rise to fatal Schweineseuche, and bacilli from any of these forms of disease, when inoculated into pigeons, produce characteristic fowl cholera; but the bacillus as obtained from Schweineseuche or Wildseuche is not fatal to chickens, and the bacillus from Schweineseuche is fatal to guinea-pigs, which have but slight susceptibility to the bacillus of rabbit septicæmia. Notwithstanding these differences he agrees with Hueppe in the view that the bacilli from the various sources mentioned are specifically identical; although evidently, if this view is adopted, we must admit that varieties exist which differ somewhat in their pathogenic power.

The researches of Smith and of Moore show that "an attenuated variety of bacteria, belonging to the group of swine-plague bacteria and not distinguishable from them, inhabit the mouth and upper air passages of such domesticated animals as cattle, dogs, and cats" (Smith).

62. BACILLUS OF CHOLERA IN DUCKS.

Obtained by Cornil and Toupet (1888) from the blood of ducks, in the Jardin d'Acclimation at Paris, which had died of an epidemic disease characterized by diarrhoea, feebleness, and muscular tremors, and which resulted fatally in two or three days.

Morphology.—Does not differ in its morphology from the bacillus of fowl cholera (*Bacillus septicæmiæ hæmorrhagicæ*); short rods with rounded ends, from 1 to 1.5 μ in length and 0.5 μ broad.

Stains with the usual aniline colors, but not by Gram's method; the ends stain more deeply than the central portion.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature. In its growth in various media, as well as in its morphology, Cornil and Toupet found this bacillus to correspond with the bacillus of fowl cholera. In *gelatin stick cultures* the growth upon the surface consists of a thin, grayish layer, and along the line of puncture as small, semi-transparent, slightly yellowish, spherical colonies. Upon agar, in the incubating oven, at the end of twelve hours small, lentil-shaped, waxy colonies are formed, which later may have a diameter of three to four millimetres. Upon *potato* circular, yellowish colonies are formed, which become confluent and form a somewhat depressed, pale-yellow layer.

Pathogenesis.—According to Cornil and Toupet, this bacillus is pathogenic for ducks, but not for chickens or pigeons, and only kills rabbits when injected in considerable quantity. Ducks die in from one to three days from subcutaneous injections, or by the ingestion of food to which the bacillus has been added.

63. BACILLUS OF HOG CHOLERA (Salmon and Smith).

Synonyms.—Bacillus of swine plague (Billings); Bacillus of swinepest (Selander).

According to Smith, this bacillus was first described by Klein (1884); it was first obtained in pure cultures and its principal characters determined by Salmon and Smith (1885), and has since been

studied in cultures and by experimental inoculations by Selander, Billings, Frosch, Welch, Caneva, Bunzl-Federn, and others.

The bacillus is found in the blood and various organs of hogs which have succumbed to the infectious disease known in this country as hog cholera; and also in the contents of the intestine, from which it may be obtained by inoculations into rabbits, but is not easily isolated by the plate method owing to the large number of other bacteria present (Smith).

Morphology.—Short bacilli with rounded ends, 1.2 to 1.5 μ in length and 0.6 to 0.7 μ in breadth; usually united in pairs.

This bacillus is easily stained by the aniline colors usually employed, but does not retain its color when treated by Gram's method. When the staining agent is allowed to act for a very short time the



FIG. 131.—Bacillus of hog cholera; stained by Löffler's method to show flagella. $\times 1,000$. From a photomicrograph made at the Army Medical Museum. (Gray.)

ends of the rods may be stained while the central portion remains unstained.

Biological Characters.—An *aërobic* (facultative anaërobic), *non-liquefying*, *actively motile* bacillus. In many of its characters this bacillus closely resembles the one last described (*Bacillus septicæmiæ hæmorrhagicæ*), but it is distinguished from it by its active movements, which, according to Smith, may be still observed in cultures which have been kept for weeks or months. Does not form spores. Grows readily in various culture media at the room temperature—more rapidly in the incubating oven. Upon *gelatin plates* colonies are developed in from twenty-four to forty-eight hours. The deep colonies are spherical and homogeneous, and have a brownish color by transmitted light; they seldom exceed one-half millimetre in diameter.

The superficial colonies may attain a diameter of two millimetres; they present no distinctive characters. Upon agar plates the colonies may have a diameter of four millimetres; they have a grayish, transparent appearance and a shining surface. In *gelatin stick cultures* small, yellowish-white colonies are developed along the line of inoculation, which may become confluent; upon the surface a thin, pearly layer is developed about the point of inoculation, which may have a diameter of six millimetres or more. Upon *potato* a straw-yellow layer is developed, which later acquires a darker color. In slightly alkaline *bouillon* a slight cloudiness may be observed at the end of twenty-four hours, and at the end of one or two weeks, if not disturbed, a deposit is seen at the bottom of the tube and a thin, broken film may form upon the surface. The development of this bacillus in *milk* produces a direct solution of the casein without previous coagulation; when a solution of litmus has been added to milk it retains its blue color in presence of this bacillus, while the bacillus previously described causes it to change to red. Neither phenol nor indol is produced in solutions containing peptone (Bunzl-Federn)—another distinguishing character from the *Bacillus septicæmiæ hæmorrhagicæ*. This bacillus may be cultivated in slightly acid media, which after a time acquire an alkaline reaction.

In Smith's experiments this bacillus was found to resist desiccation from nine days to several months, according to the thickness of the layer dried upon the cover glass; bacilli from an agar culture in some experiments failed to grow after seventeen days, and in others still gave cultures after four months. Bouillon cultures are sterilized in four minutes by a temperature of 70° C., in fifteen minutes by 58° C., and in one hour by 54° C. (Smith). Novy has isolated from cultures of the hog-cholera bacillus a toxic basic substance which he calls susotoxin. This was obtained by Brieger's method; it is a yellowish-brown, syrup-like liquid, which, when injected into rats in doses of 0.125 to 0.25 cubic centimetre, causes their death in less than thirty-six hours. He also obtained by precipitation with absolute alcohol, from cultures concentrated in a vacuum at 36° C., a toxalbumin which when dried was in the form of a white powder easily soluble in water. Rats died in three or four hours after receiving subcutaneously a dose of 0.1 to 0.5 gramme.

Pathogenesis.—Pathogenic for swine, rabbits, guinea-pigs, mice, and pigeons.

In certain parts of the United States the disease known as "hog cholera" frequently prevails among swine as a fatal epidemic. It may occur as an acute and quickly fatal septicæmia, or in a more chronic form lasting from two to four weeks or even longer. In the acute form death may occur within twenty-four hours, and hæm-

orrhagic extravasations are found upon the mucous and serous membranes and in the parenchyma of the lungs, kidneys, and lymphatic glands. The spleen is greatly enlarged, soft, and dark in color. In the chronic form of the disease the most notable changes are found in the alimentary canal. These are most constant and characteristic in the cæcum and colon, which may be studded with spherical, hard, necrotic masses or extensive diphtheritic patches. According to Smith, the hæmorrhagic and necrotic form of the disease may exist at the same time in different animals of the same herd. The bacilli are found in all of the organs, and especially in the spleen, where they are associated in irregular colonies similar to those of the typhoid bacillus. Smith has demonstrated their presence in urine taken from the bladder immediately after the death of the animal, and states that the kidneys are almost always involved, as shown by the presence of albumin and tube casts in the urine.

An extremely minute quantity of a bouillon culture injected beneath the skin of a rabbit causes its death in from seven to twelve days; a larger quantity may produce a fatal result in five days; intravenous injections of very small amounts may be fatal within forty-eight hours. After a subcutaneous injection the animal remains in apparent good health for three or four days, after which it loses its appetite and is indisposed to move; several days before death the temperature is suddenly elevated from 2° to 3° C., and it remains high until the fatal termination. At the autopsy the spleen is found to be enlarged and of a dark-red color; the liver is studded with small, yellowish-white, necrotic foci; the kidneys have undergone parenchymatous changes; the heart is fatty; and the intestinal mucous membrane is more or less marked with hæmorrhagic extravasations. The bacilli are found in all of the organs. In house mice the results of experimental inoculations are similar to those in rabbits. Guinea-pigs succumb when inoculated subcutaneously with one-tenth cubic centimetre; pigeons require a still larger dose—about three-quarters of a cubic centimetre. Swine are killed by the intravenous injection of one to two cubic centimetres of a recent bouillon culture, but, as a rule, do not succumb to subcutaneous injections. Cultures recently obtained from diseased animals are more virulent than those which have been propagated for a considerable time in artificial media.

Smith has described a variety of the hog-cholera bacillus obtained during an epidemic in which the disease was of longer duration—about four weeks—than is usual, and in which there was commonly found at the autopsy a diphtheritic inflammation of the mucous membrane of the stomach. This bacillus differed from the typical form by being somewhat larger and in forming considerably larger colonies in gelatin plates—two or three times

as large. It also produced a greater opacity in peptonized bouillon, and in general showed a more vigorous growth in various nutrient media. It differed also in its pathogenic power, as tested upon rabbits, causing death at a later date or not at all; and in fatal cases the swelling of the spleen and necrotic foci in the liver, produced by the first-described species, were absent.

64. BACILLUS OF BELFANTI AND PASCAROLA.

Synonym.—*Impftetanusbacillus*.

Obtained by Belfanti and Pascarola (1888) from the pus of wounds in an individual who succumbed to tetanus.

Morphology.—Bacilli with rounded ends, sometimes so short as to resemble micrococci; resemble the *Bacillus septicæmiæ hæmorrhagicæ* (fowl cholera).

Stains with the usual aniline colors and also by Gram's method. The ends are commonly more deeply stained than the central portion.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* yellowish-gray, finely granular, spherical colonies with smooth outlines are developed. In *gelatin stick cultures*, at 18° to 25° C., at the end of twenty-four hours small, spherical colonies are developed along the line of puncture, which are isolated or closely crowded; upon the surface a rather thin, shining, grayish-white, iridescent, circular layer is formed; gas is given off which has not a disagreeable odor. Upon the surface of *agar* elevated, shining, gray colonies develop along the *impfstrich*, or a gray, shining band is formed which increases in thickness but not in breadth—usually less than one-half centimetre broad. Old cultures give off an acid odor. Upon *blood serum* a thin, white layer is developed along the line of inoculation. Upon *potato* a thin, white, varnish-like layer is formed.

Pathogenesis.—Very pathogenic for rabbits, guinea-pigs, white mice, and sparrows. Not pathogenic for chickens, pigeons, or geese.

65. BACILLUS OF SWINE PLAGUE, MARSEILLES.

Synonyms.—*Bacillus der Schweineseuche*, Marseilles (Rietsch and Jobert); *Bacillus der Frettchenseuche*—ferret disease (Eberth and Schimmelbusch); *Bacillus der Amerikanischen Rinderseuche* (Caneva); *Bacillus of spontaneous rabbit septicæmia* (Eberth).

The recent researches of Caneva and of Bunzl-Federn agree as to the identity of the bacillus obtained by Rietsch and Jobert (1887) from swine attacked with a fatal epidemic disease in Marseilles, and the bacillus found by Eberth and Schimmelbusch (1889) in the blood of ferrets suffering from a fatal form of septicæmia studied by them. The first-named bacteriologist also identifies a bacillus supposed by Billings to be the cause of "Texas fever" in cattle ("*Amerikanische Rinderseuche*") and the bacillus of swine plague (Billings) with the above. Bunzl-Federn obtained cultures of Billings' swine-plague bacillus at two different times. He identifies the one first received with the bacillus now under consideration, and the other with the bacillus of hog cholera (Salmon).¹

¹ The author named says: "With reference to the bacillus of swine plague (Billings), I obtained, as did Caneva, a decided production of acid in the cultures

Morphology.—Bacilli with rounded ends, about twice as long as broad, and one-third smaller than the bacillus of typhoid fever (Eberth and Schimmelbusch). The bacillus of hog cholera is shorter and more slender than the Marseilles bacillus, and the bacillus of Löffler and Schütz (No. 61) is still smaller (Rietsch and Jobert).

In stained preparations the extremities of the rods are usually deeply stained, while the central portion remains unstained—"polar staining." By Löffler's method of staining the presence of flagella may be demonstrated (Frosch).

Stains readily with the aniline dyes usually employed, but does not retain its color when treated by Gram's method.

Biological Characters.—An *aërobic* (facultative anaërobic), *non-liquefying*, *actively motile* bacillus. Grows readily at the room temperature, and is distinguished from the bacillus of septicæmia hæmorrhagica by its active movements and more rapid and abundant development in the various culture media usually employed. It is distinguished from the bacillus of hog cholera (No. 63) by producing phenol and indol in solutions containing peptone, by causing coagulation of milk, and by producing an acid reaction in this fluid. Grows in culture media having an acid reaction.

Rietsch and Jobert give the following account of the characters of growth in various culture media, as compared with the bacillus of hog cholera and the bacillus of Schweineseuche (Löffler, Schütz), No. 61:

Gelatin streak cultures. At the end of twenty-four hours this bacillus had developed considerably, while the growth of the hog-cholera bacillus was scarcely to be discerned with the naked eye, and the bacillus of Schweineseuche did not form a visible growth until the end of forty-eight hours. After several days the bacillus of swine plague (Marseilles) formed an opaque, yellowish-white streak, which, when examined with a low-power lens, had a brown color by transmitted light and a bluish-white color by reflected light. The streak of the Löffler-Schütz bacillus was not so thick and not so opaque, and was made up of small, nearly transparent colonies; the hog-cholera bacillus came between the other two. Upon *blood serum*, *agar*, and *glycerin-agar* the Marseilles bacillus grew more rapidly than the other two, forming a layer which was opaque and of a white color, with bluish and reddish reflections. Upon *potato* it formed a thick, opaque, yellowish layer, while the growth of the hog-cholera bacillus was much thinner and that of the Löffler-Schütz bacillus scarcely to be seen. In *bouillon* the Löffler-Schütz bacillus,

first sent by Billings; but upon testing later cultures received directly from Billings and from other sources, the result was exactly the opposite—viz., a decided production of alkali in milk and identity with the hog-cholera bacillus of Salmon."

at the end of three days at 37° C., had not produced any perceptible cloudiness, while the Marseilles bacillus at the end of twenty-four hours had caused the fluid to be clouded, a film of bacteria had formed upon the surface and a deposit at the bottom of the tube; the hog-cholera bacillus produced a less degree of opacity in the bouillon.

Pathogenesis.—This bacillus is pathogenic for sparrows and other small birds when injected beneath the skin in small amounts, and also for pigeons in a longer time—five to fourteen days. Frosch reports a negative result from subcutaneous injections into rabbits, guinea-pigs, mice, and pigeons, but his cultures appear to have become attenuated, as the recent cultures of Eberth and Schimmelbusch were fatal to pigeons in four out of five experiments. Two rabbits were inoculated subcutaneously by Rietsch and Jobert with half a Pravaz syringeful of a pure culture of the Marseilles bacillus; one of these died on the sixth day and the other survived.

In sparrows, which succumb in from twenty-four to thirty-six hours after receiving a small amount of a pure culture in the breast muscle, the bacillus is present in the blood in large numbers, and a purulent pleuritis and pericarditis is found at the autopsy. In the ferrets from which Eberth and Schimmelbusch obtained their cultures the bacillus was not present in the blood in sufficient numbers to be readily demonstrated by microscopical examination, but it was obtained in pure cultures from the liver, spleen, and lungs. The principal pathological appearances noted were enlargement of the spleen and pneumonia. Caneva reports that the Marseilles bacillus injected into white mice gives rise to an extensive abscess at the point of inoculation, but does not kill adult animals. In a young mouse which succumbed to such an injection the bacilli were not generally distributed in the tissues, but were found as emboli in the smaller capillaries. This bacillus, then, is distinguished from the similar bacilli previously described (Nos. 61 and 63) by its comparatively slight pathogenic power, as well as by its more vigorous growth in culture media, and the other characters heretofore mentioned.

66. BACILLUS SEPTICUS AGRIGENUS.

Obtained by Nicolaier from soil which had been manured.

Morphology.—Resembles the bacillus of fowl cholera and of rabbit septicæmia, of which it is perhaps a variety, but is usually somewhat longer. It also sometimes shows the end-staining characteristic of *Bacillus septicæmiæ hæmorrhagicæ*, but not so constantly and not so sharply defined.

Biological Characters.—An *aërobic*, (*non-liquefying* ?), non-motile bacillus. Does not form spores.

In *gelatin plate cultures* spherical, finely granular colonies are developed having a yellowish-brown central portion, which is separated by a dark ring from a grayish-brown marginal zone; later this difference in color disappears and the colonies become more decidedly granular. In stick cultures the growth consists of a thin layer which is not at all characteristic.

Pathogenesis.—Small quantities of a pure culture injected into the ear vein of a rabbit cause its death in from twenty-four to thirty-six hours; pathogenic also for house mice and for field mice. At the autopsy no notable pathological changes are observed. The bacilli are found in blood from the heart and in the capillaries of the various organs, but are not so numerous as in rabbit septicæmia; they show a special inclination to adhere to the margins of the red blood corpuscles.

67. BACILLUS ERYSIPELATUS SUI.

Synonyms.—Bacillus of hog erysipelas; Bacillus des Schweinerothlauf (Löffler, Schütz); Bacille du rouget du porc (Pasteur); Bacillus of mouse septicæmia; Bacillus murisepticus (Flügge); Bacillus des Mäusesepdikämie (Koch).

The bacillus of mouse septicæmia, first described by Koch (1878), resembles so closely in its morphology, characters of growth, and pathogenic power the bacillus of Schweinerothlauf of Löffler and Schütz (1885) that they can scarcely be considered as distinct species, although, from slight differences which have been observed, they are perhaps entitled to separate consideration as varieties of the same species. Flügge, Eisenberg, Fränkel, and other authors, while

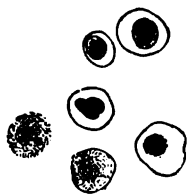


FIG. 135.—Bacillus of mouse septicæmia in leucocytes from blood of mouse. $\times 700$. (Koch.)

recognizing the fact that the bacilli from the two sources closely resemble each other, apparently do not consider them identical and describe them separately. Baumgarten, on the other hand, describes them under one heading and considers it highly probable that they are identical, although he also admits slight differences in the morphological characters and growth in culture media. These differences are, however, no greater than we have in artificially produced varieties of other well-known microorganisms, and we think it best to follow Baumgarten in describing them under a single heading.

Koch first obtained this bacillus by injecting putrefying blood or flesh infusion, during the first days of putrefactive change, beneath the skin of mice. A certain proportion of the animals experimented upon contracted a fatal form of septicæmia, and the bacillus under consideration was found in their blood. The bacillus of Schweinerothlauf was obtained by Löffler and by Schütz from the blood and various organs of swine which had succumbed to the infectious malady known in Germany as rothlauf and in France as rouget.

Morphology.—Extremely minute bacilli, about $1\ \mu$ in length and $0.2\ \mu$ in diameter. The Schweinerothlauf bacilli are described as somewhat thicker and longer by Flügge, by Fränkel, and by Eisenberg, but Baumgarten states that they are somewhat more slender and

on the average shorter than the bacillus of mouse septicæmia. The bacilli are solitary, or in pairs the elements of which are often united at an angle; occasionally a chain of three or four elements may be observed, and in old cultures the bacilli may grow out into short threads which are straight or more or less curved and twisted. Small refractive bodies may sometimes be distinguished in the rods, and these have been supposed by some authors to be spores, but this has not been demonstrated.

This bacillus *stains* readily with the ordinary aniline staining agents and also by Gram's method.

Biological Characters.—A *facultative anaërobie, non-liquefying* bacillus. According to



FIG. 136.—Bacillus of rouget, from a pure culture. $\times 1,000$. From a photomicrograph. (ROUX.)

Schottelius, the rothlauf bacilli are sometimes motile, but Flügge states that other observers have not seen them in active motion. Fränkel says they have the power of voluntary motion. Eisenberg says that the bacillus of mouse septicæmia is motionless, and Fränkel says "they seem to be incapable of voluntary motion." Baumgarten remarks: "Whether the bacilli exhibit voluntary movements has not been determined." Although this bacillus is not strictly anaërobie, it grows better in the absence of oxygen than in its presence. Development occurs in various culture media at the room temperature, but is more rapid in the culture oven. In *gelatin stick cultures* no development occurs upon the surface, but the growth along the line of puncture is very characteristic; this consists of a delicate, cloud-like, radiating growth, which extends, in the course of a few days, almost to the walls of the test tube. The rothlauf bacillus does not extend so rapidly through the gelatin, and the branching, cloud-like growth is not as delicate; Flügge compares it to the brush of bristles used for cleansing test tubes.

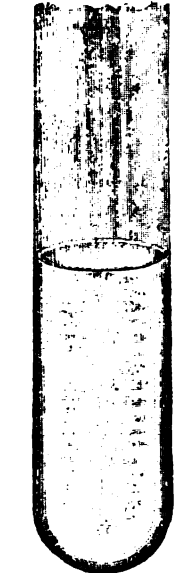


FIG. 137.—Bacillus of mouse septicæmia; culture in nutrient gelatin, end of four days at 18° C. (Baumgarten.)

In old cultures in nutrient gelatin a slight softening of the gelatin occurs along the line of growth, and as a result of

evaporation and desiccation a funnel-shaped cavity is formed in the culture medium in the course of two or three weeks. In *gelatin plates* colonies are developed in the course of two or three days in the deeper layers of the gelatin, but not upon the surface; these are nebulous, grayish-blue, radiating masses, which are so delicate as to be scarcely visible without the aid of a lens or a dark background. Under a low power they appear as branching, feathery masses, which have been compared by Flügge to the radiating growth of "bone corpuscles." In older cultures they coalesce and cause a nebulous opacity of the whole plate, which has a bluish-gray lustre.



FIG. 133.—*Bacillus* of mouse septicæmia; single colony in nutrient gelatin. $\times 80$. (Flügge.)

Upon the surface of nutrient agar or blood serum a very scanty development occurs along the line of inoculation. No growth occurs upon potato. In bouillon the bacilli cause a slight cloudiness at the outset, and later a scanty, grayish-white deposit upon the bottom of the test tube; no film is formed upon the surface.

The thermal death-point of this bacillus, as determined by the writer (1887), is 58°C ., the time of exposure being ten minutes. In the experiments of Bolton it was destroyed in two hours by mercuric chloride solution in the proportion of 1 : 10,000; by carbolic acid and by sulphate of copper in one-per-cent solution. These results are opposed to the view that the minute refractive granules which may sometimes be seen in the interior of the rods are reproductive spores, for all known spores have a much greater resisting power to heat and the chemical agents named.

Pathogenesis.—Pathogenic for swine, rabbits, white mice, house mice, pigeons, and sparrows. Field mice, guinea-pigs, and chickens are immune.

Swine may be infected by the ingestion of food containing the rothlauf bacillus, as has been demonstrated by allowing them to eat the intestine of an animal which had recently succumbed to the disease, and also by the subcutaneous injection of pure cultures. The disease usually terminates fatally within three or four days, and sometimes in less than twenty-four hours. It is characterized by fever, debility, loss of appetite, and by the appearance upon the surface of the body of red patches, which gradually extend and become confluent, producing after a time a uniform dark-red or brown color of the entire surface. The discharges from the bowels frequently contain bloody mucus. At the autopsy, in acute cases, the spleen is notably enlarged, and the liver and kidneys are likely to be more or less swollen, as are also the lymphatic glands, especially those of the mesentery; the gastric and intestinal mucous membranes are

usually inflamed and spotted with hæmorrhagic extravasations ; the serous membranes also may be inflamed, and the cavities of the pleuræ, pericardium, and peritoneum usually contain more or less fluid. The bacilli are found in the blood vessels throughout the body, and are especially numerous in the interior of the leucocytes. Cornevin and Kitt have shown that the contents of the intestine also contain the bacilli in large numbers, and the disease appears to be propagated among swine principally by the contamination of their food with the alvine discharges of diseased animals.

Pigeons are very susceptible to the pathogenic action of this bacillus, and usually die within three or four days after inoculation with a pure culture. Rabbits are not so susceptible, although a certain proportion die from general infection after being inoculated in the ear. The first effect of such an inoculation is to produce an erysipelatous inflammation. When the animal recovers it is subsequently immune.

White mice and house mice are extremely susceptible, but field

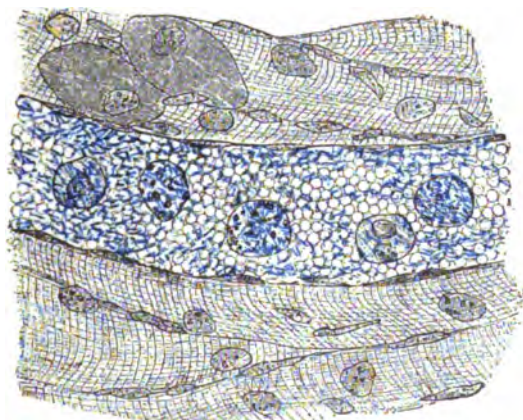


FIG. 139.—Section of diaphragm of a mouse dead from mouse septicæmia, showing bacilli in a capillary blood vessel. (Baumgarten.)

mice are immune. This remarkable fact was first ascertained by Koch by experiments with his bacillus of mouse septicæmia. House mice which have been inoculated with a minute quantity of a pure culture of the rothlauf, or mouse septicæmia, bacillus, die in from forty to sixty hours. The animal is usually found dead in a sitting position, with its back strongly curved, and for many hours before death it remains quietly sitting in the same position ; the eyes are glued together by a sticky secretion from the conjunctival mucous membrane. At the autopsy the spleen is found to be very much enlarged, and there may be a slight amount of œdema at the point of inoculation.

The bacilli are found in the blood vessels generally, and are very numerous in the interior of the leucocytes, which are sometimes completely filled with them.

Pasteur's first studies relating to the etiology of "rouget" were made, in collaboration with Chamberlain, Roux, and Thuillier, in 1882. His description of the microorganism to which he attributed the disease does not correspond with that subsequently isolated by Löffler and by Schütz; but the last-named bacteriologists, and Schottelius also, found the characteristic rothlauf bacillus in cultures from his laboratory which had been prepared for the protective inoculation of swine—"vaccins." Pasteur found, by experimental inoculations of his bacillus of rouget into pigeons, that the virulence of his cultures was increased by successive inoculations through a series of these birds, as shown by the occurrence of death at an earlier date, and also by the fact that blood taken from the last pigeon in a series was more virulent for swine than that from the first or from an infected pig. On the other hand, the virulence was diminished by inoculations into rabbits; and, by passing the bacillus through a series of these animals, a vaccine was obtained which produced a comparatively mild and non-fatal attack in swine. In practice the use of two different vaccines is recommended, a mild—"attenuated"—virus being first inoculated, and, after an interval of twelve days, a second vaccine having greater pathogenic potency. These inoculations have been extensively practised in France, and that immunity from the disease may be secured in this way is well established, having been confirmed in Germany by Schütz, by Lydtin, and by Schottelius. There is, however, some doubt as to the practical value of the method, inasmuch as a certain number of the inoculated animals die, and there appears to be danger that the disease may be spread by the alvine discharges of inoculated animals. In a region where the annual losses from the disease are considerable, and where the soil is, perhaps, thoroughly infected with rothlauf bacilli, protective inoculations probably afford the best security against loss. But under other circumstances the quarantine of infected animals and thorough disinfection of the localities in which cases have occurred will probably prove a better mode of procedure.

68. BACILLUS COPROGENES PARVUS.

Synonym.—Mäuseseptikämieähnlicher Bacillus (Eisenberg).

Obtained by Bienstock from human fæces.

Morphology.—A very minute bacillus, which is but little longer than it is broad, and might easily be mistaken for a micrococcus.

Biological Characters.—Grows very slowly on nutrient gelatin, forming a scarcely visible film along the line of inoculation, which at the end of several weeks is scarcely one millimetre wide. Is not motile.

Pathogenesis.—In white mice an extensive oedema is developed at the

point of inoculation at the end of ten or twelve hours, and the animal dies within thirty-six hours. The bacilli are found in great numbers in the effused serum at the point of inoculation and in comparatively small numbers in the blood. A rabbit inoculated with a pure culture obtained from a mouse died at the end of eight days. The inoculation, which was made in the ear, gave rise to a local erysipelatous inflammation.

69. BACILLUS CAVICIDA.

Synonym.—Brieger's bacillus. Probably identical with *Bacterium coli commune* of Escherich.

Obtained by Brieger (1884) from human faeces.

Morphology.—Small bacilli, about twice as long as broad, which closely resemble the colon bacillus of Escherich (*Bacterium coli commune*).

Biological Characters.—An *aërobie* (facultative anaërobie), *non-liquefying* bacillus.

The growth in gelatin plate cultures is said to be very characteristic, the colonies being "in the form of very beautifully grouped, whitish, concentric rings, which are arranged like the scales upon the back of a turtle." (Eisenberg). The writer has studied cultures of this bacillus brought from the bacteriological laboratories of Germany, side by side with cultures of the *Bacterium coli commune* of Escherich, and has found no appreciable differences in the colonies in gelatin plates, or in the growth in various culture media. Upon *potato* it grows rapidly in the incubating oven, forming a dirty-yellow, moist layer.

Pathogenesis.—This bacillus, as first obtained by Brieger, was characterized by being very pathogenic for guinea-pigs, which were invariably killed, within seventy-two hours, by the subcutaneous injection of a minute quantity of a pure culture. The bacillus was found in great numbers in the blood of animals which succumbed to an experimental inoculation. The writer's experiments with this bacillus, made in 1889, indicate that its pathogenic power had become attenuated, inasmuch as considerable quantities of a pure culture injected into guinea-pigs did not cause the death of the animals—culture used came originally from Germany. Not pathogenic for rabbits or for mice.

70. BACILLUS CAVICIDA HAVANIENSIS.

This bacillus was obtained by the writer from the contents of the intestine of a yellow-fever cadaver, in Havana, 1889, through inoculated guinea-pigs.

Morphology.—A bacillus with rounded ends, from 1.0 to three μ long and about 0.7 μ broad, frequently united in pairs.

Stains readily with the ordinary aniline colors.

Biological Characters.—An *aërobie* and *facultative anaërobie*, *non-liquefying*, *actively motile* bacillus.

In *gelatin stick* cultures the growth upon the surface is very scanty and thin, not extending far from the point of puncture; along the line of puncture are developed small, translucent, pearl-like, spherical colonies, which later become opaque and sometimes granular. In gelatin roll tubes, at the end of twenty-four hours at 22° C., the deep colonies are very small spheres, of a pale straw color; later they become opaque, light brown spheres, or may have a dark central mass surrounded by a transparent zone. The superficial colonies at the end of five days are small, translucent masses of a pale straw color towards the centre, with thin and irregular margins, sometimes with



FIG. 140.—*Bacillus cavicida havaniensis*; from a potato culture. $\times 1,000$. From a photomicrograph. (Sternberg.)

a central light-brown nucleus; at the end of ten days the deep colonies are still quite small, of a brown color, and opaque.

In glycerin-agar roll tubes, at the end of twenty-four hours, the deep colonies are in the form of a biconvex lens, and appear spherical when viewed in face and biconvex when seen from the side; they have a straw color by transmitted light and are bluish-white by reflected light; the superficial colonies are translucent, with a bluish-white lustre.

On *potato*, at 22° C., at the end of forty eight hours there is a thin, dirty-yellow growth of limited extent; at the end of ten days there is a thin, gamboge-yellow layer and little masses of the same color; the growth is quite thin, with irregular outlines, and is confined to the vicinity of the impfstrich.

Grows in nutrient agar containing 0.2 per cent of hydrochloric acid. Thermal death-point 55° C. Grows in *agua coco* without forming gas, and causes this liquid and bouillon to become slightly translucent—not milky.

Pathogenesis.—Pathogenic for guinea-pigs, less so for rabbits. Guinea-pigs inoculated subcutaneously with a few drops of a pure culture die in ten or twelve hours from general infection. There is usually a considerable effusion of bloody serum in the vicinity of the point of inoculation, and the spleen is more or less enlarged.

71. BACILLUS CRASSUS SPUTIGENUS.

Obtained by Kreibohm (1886) from the sputum of two individuals, and once in scrapings from the tongue.

Morphology.—Short, thick bacilli, of oblong form, with rounded corners, often bent or twisted—"sausage-shaped." Immediately after division the bacilli are about one-half longer than they are broad, but before dividing

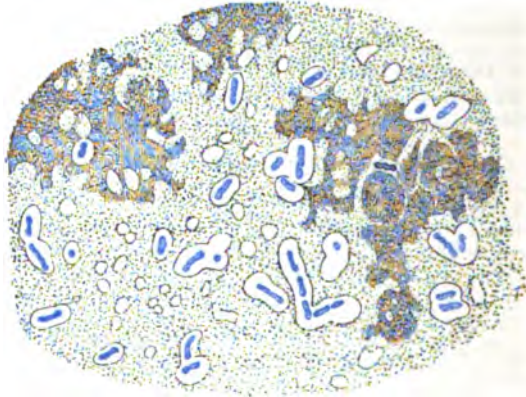


FIG. 141.—*Bacillus crassus sputigenus*, from blood of mouse. $\times 700$. (Flügge.)

again they may attain a length of three to four times the breadth. Irregular forms with swollen ends or uneven contour are frequently seen.

This bacillus is quickly stained by the ordinary aniline colors and also by Gram's method.

Biological Characters.—An *aërobic*, *non-liquefying* (non-motile ?) bacillus. Grows in various culture media at the room temperature—more rapidly in the incubating oven. "Appears to form spores at 35° C." (Flügge).

In *gelatin plates*, at the end of thirty-six hours, grayish-white colonies are developed, which soon reach the surface of the gelatin and spread out as

round, viscid, grayish-white drops, which project considerably above the surface of the culture medium. Under a low magnifying power recent colonies appear as spherical, grayish-brown discs, the surface of which is marked with dark points or lines. The superficial colonies are more transparent, have irregular outlines, and the surface, especially near the margins, is coarsely granular. The development in stick cultures is very rapid and resembles that of Friedländer's bacillus—"nail-shaped" growth. Upon potato the growth is also similar to that of Friedländer's bacillus, and consists of a thick, grayish-white, moist, and shining layer.

Pathogenesis.—Mice inoculated with a small quantity of a pure culture die from acute septicæmia in about forty-eight hours. The bacilli are found in blood from the heart and from the various organs—most numerous in the liver. Rabbits are killed within forty-eight hours by intravenous injection of a small quantity, and the blood contains the bacillus in great numbers. Larger amounts injected into the circulation of rabbits or dogs cause death in a few hours (three to ten), preceded by diarrhoea, and in some instances bloody discharges from the bowels. At the autopsy an acute gastro-enteritis is found.

72. BACILLUS PYOGENES FŒTIDUS.

Obtained by Passet (1885) from an abscess of the anus.

Morphology.—Short bacilli with rounded ends, 1.45μ long and 0.58μ broad; usually associated in pairs or in short chains.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Grows rapidly in the usual culture media at the room temperature. In the interior of the rods, in stained preparations, one or two unstained, spherical places may sometimes be seen, which have been supposed to be spores (?). The independent motion exhibited by this bacillus is not very active. In gelatin plates white colonies are developed at the end of twenty-four hours, which upon the surface spread out as grayish-white plaques, having a diameter sometimes of one centimetre; these are thickest in the centre and of a whitish color; the colonies may become confluent. In *gelatin stick cultures* the growth upon the surface, at the end of twenty-four hours, consists of a thin, grayish-white layer with rather thick, irregular margins; along the line of puncture more or less crowded colonies. Upon *potato* the bacillus forms an abundant, shining, pale-brown layer. The cultures give off a disagreeable putrefactive odor.

According to Eisenberg, mice and guinea-pigs are killed in twenty-four hours by injections beneath the skin or into the cavity of the abdomen, and numerous bacilli are found in the blood.

73. PROTEUS HOMINIS CAPSULATUS.

Obtained by Bordoni-Uffreduzzi (1887) from two cadavers presenting the pathological appearances of the so-called "Haderkrankheit."

Morphology.—Bacilli, varying considerably in dimensions; somewhat thicker than the anthrax bacillus; often swollen in the middle or at the extremities; more or less curved; isolated, united in pairs or in long filaments; in stained preparations from agar cultures or from blood the bacilli are surrounded by a "capsule."

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* (facultative anaërobic?), *non-liquefying, non-motile* bacillus. Formation of spores not observed. Grows in the usual culture media at the room temperature. At a temperature of 15° to 17° C. long filaments are formed, in which the bacilli are surrounded with a capsule; at 22° to 24° C. the bacilli are for the most part isolated, but few filaments being formed; at 32° to 37° C. the bacilli are so short as to resemble micrococci; development ceases at a temperature of 8° and is very slow at 15° C.

This bacillus grows as well in an acid medium as in one which is slightly alkaline. In *gelatin plates*, at the end of eighteen to twenty-four hours, colonies are formed which under a low power are seen to be spherical and to contain a quantity of shining granules; the following day, at a temperature of 15° to 17° C., the colonies may be as large as a pin's head and still

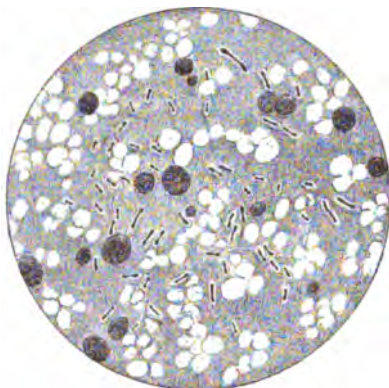


FIG. 149.—*Proteus hominis capsulatus*, from liver of mouse. $\times 1,000$. (Bordoni-Uffreduzzi.)

remain spherical or slightly oval, but the outline is no longer so uniform, and between the shining points in the interior a confused network may be seen; as the colony becomes larger it is raised above the surface of the gelatin, becomes opaque, and has a pearly lustre like that of Friedländer's bacillus. In *gelatin stick cultures* the growth resembles that of Friedländer's bacillus—"nail-shaped growth." Upon the surface of *nutrient agar* a rapidly extending, semi-transparent layer is formed. Upon *potato*, at 15° to 17° C., at the end of twenty-four hours transparent drops are seen in the vicinity of the point of inoculation, and later a moist, shining, colorless layer, of tough consistence, is formed, which gradually extends over the surface. The growth upon *blood serum* resembles that upon nutrient

agar, and the blood serum is not liquefied. In liquid blood serum or in bouillon the bacilli are isolated—not in filaments; they cause a clouding of the liquid, and an abundant deposit accumulates at the bottom of the tube, while a film of bacilli forms upon the surface. The cultures never give off a putrefactive odor.

Pathogenesis.—Pathogenic for dogs and for mice, less so for rabbits and for guinea-pigs. Agar cultures grown in the incubating oven at 32° to 37° C. are more pathogenic than cultures in gelatin at the room temperature. A small quantity of a recent culture injected subcutaneously in mice causes their death in from one to four days, according to the quantity and age of the culture; the recent cultures are most virulent. When the animal lives more than twenty-four hours it has a mucous diarrhoea. At the autopsy the spleen is found to be much enlarged and dark in color; the lymphatic glands are also swollen and hæmorrhagic, the liver and kidneys hyperæmic; in the vicinity of the point of inoculation is a subcutaneous œdema of jelly-like appearance and numerous punctiform hæmorrhages are seen. The bacillus is found in great numbers in the effused serum from the subcutaneous tissues, in the blood, the contents of the intestine, and in the parenchyma of the various organs. When examined at once the bacilli in the subcutaneous œdema and in the lymphatic glands are usually quite short, and even spherical, while in the blood they are somewhat longer and may appear as short filaments with swollen ends, surrounded by a capsule. When the examination is made some time after the death of the animal longer filaments are quite numerous. Rabbits and guinea-pigs are killed by the intravenous injection of comparatively small amounts of a recent culture, but quite large doses are required to produce a fatal result when the injection is made beneath the skin. From two to three cubic centimetres of a recent culture injected into the circulation of a dog give rise to symptoms of toxæmia, and the animal usually dies on the second day. At the autopsy the abdominal organs are found to be hyperæmic, the mucous membrane of the intestine swollen, red in color, and covered with bloody mucus. The bacillus is found in the blood and in the various organs. When smaller doses are injected into a vein (a few drops) the animal, after a few hours, has a mucous diarrhoea and

vomiting, or efforts to vomit. Death usually occurs at the end of two or three days. At the autopsy the spleen is found to be normal, the other organs slightly hyperæmic, and the intestinal mucous membrane in a state of catarrhal inflammation. The bacilli are found in the blood and in the various organs in considerable numbers.

74. *PROTEUS CAPSULATUS SEPTICUS*.

Obtained by Banti (1888) from a case of "acute hæmorrhagic infection."

According to Banti, this is possibly identical with the preceding species—*Proteus hominis capsulatus*—but in some respects more nearly resembles Friedländer's bacillus.

75. *BACILLUS ENTERITIDIS*.

Obtained by Gärtner (1888) from the tissues of a cow which was killed in consequence of an attack characterized by a mucous diarrhoea, and also from the spleen of a man who died twelve hours after eating the flesh of this animal.

Morphology.—Short bacilli, about twice as long as broad, frequently united in pairs; chains of four to six elements are sometimes seen.

Stains with the usual aniline colors, and presents the peculiarity of staining deeply at one end while the remainder of the rod is but slightly stained. When two bacilli are united the deeply stained ends are in apposition.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Spore formation not determined. Grows in the usual culture media at the room temperature. Upon *gelatin plates* pale-gray, superficial colonies are formed at the end of twenty-four hours; under a low power these are seen to be coarsely granular and transparent; the central portion usually presents a greenish color; deep colonies are spherical, indistinctly granular, and of a brownish color; in older colonies a marginal transparent zone is seen which appears to be made up of minute fragments of glass of a pale-brown color. In *gelatin stick cultures* but slight development occurs along the line of puncture; upon the surface a thick, grayish-white layer is formed, which after a time becomes very much wrinkled. Upon the surface of *agar*, at 37° C., at the end of eighteen to twenty hours a grayish-yellow layer has formed. Upon *potato* a moist, shining, yellowish-gray layer is developed. The growth upon *blood serum* is rapid in the form of a gray layer along the line of inoculation.

Pathogenesis.—White mice and house mice usually die in from one to three days when fed with a pure culture of this bacillus. Rabbits and guinea-pigs die in from two to five days from subcutaneous injections—less pathogenic for pigeons and canary birds. Dogs, cats, chickens, and sparrows are immune. A goat died in twenty hours after receiving an intravenous injection of two cubic centimetres of a culture in blood serum. The principal pathological appearance consists in an intense inflammation of the intestinal mucous membrane. The bacilli are found in blood from the heart and also in the contents of the stomach.

76. *BACILLUS OF GROUSE DISEASE*.

Obtained by Klein (1889) from the lungs and liver of grouse which had succumbed to an epidemic disease.

Morphology.—Bacilli with rounded ends, from 0.8 to 1.6 μ long; may also be seen as spherical or oval cells 0.6 μ long and 0.4 μ thick; solitary, in pairs, or in chains of three to four elements.

Stains best with Weigert's solution of methylene blue in aniline water.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the

room temperature—better in the incubating oven. Upon *gelatin plates*, at 20° C., at the end of twenty-four hours small, angular, transparent scales may be seen upon the surface with a low-power lens; at the end of three or four days these form flat, more or less irregular, shining, gray colonies, with thin and often dentate margins; these colonies may become confluent and form a dry, scaly layer which by reflected light has a peculiar, fatty lustre. In *gelatin stick cultures* the superficial growth is in the form of a transparent, dry, grayish layer with dentate margins, not more than three to five millimetres in diameter. Upon *agar*, at 36° to 37° C., a thin, whitish-gray, dry layer is formed.

Pathogenesis.—Pathogenic for mice, for guinea-pigs, for linnets, and for green-finches; less so for sparrows. Chickens, pigeons, and rabbits, according to Klein, are immune. Of eight mice inoculated subcutaneously with one or two drops of a bouillon culture, six died within forty-eight hours and two recovered. Out of eight guinea-pigs inoculated in the same way four died in forty-eight hours and two recovered. At the autopsy the lungs and liver were found to be hyperæmic, the spleen not enlarged. The bacilli were present in large numbers in blood from the heart and in the lungs.

77. BACILLUS GALLINARUM.

Obtained by Klein (1889) from the blood of chickens which succumbed to an epidemic disease resembling "fowl cholera." The bacillus is believed by Klein not to be identical with Pasteur's bacillus of fowl cholera, and is said not to be pathogenic for rabbits, which would seem to differentiate it from this bacillus (*Bacillus septicæmiæ hæmorrhagicæ*).

Morphology.—Bacilli with rounded ends, from 0.8 to 2 μ long and 0.3 to 0.4 μ thick; often in pairs.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature—better in the incubating oven. Upon *gelatin plates* forms grayish-white, superficial colonies, which later present the appearance of flat, homogeneous, whitish discs with thin edges and irregular margins, and by transmitted light have a brownish color. The deep colonies are small and spherical, and have a brownish color by transmitted light. In *gelatin stick cultures* a thin, gray layer with irregular margins and of limited extent forms upon the surface, and a scanty growth occurs along the line of puncture in the form of a grayish-white line. Upon the surface of *agar*, at 37° C., a thin, gray layer with irregular margins has developed at the end of twenty-four hours; later this extends over the entire surface as a thin, grayish-white layer. No growth occurs upon *potato* at 37° C. In *bouillon*, at 37° C., development occurs, with clouding of the bouillon, within twenty-four hours; later a deposit consisting of bacilli is seen at the bottom of the tube, but no film forms upon the surface.

Pathogenesis.—Chickens inoculated subcutaneously with a pure culture die in from twenty-four hours to eight or nine days. Pigeons and rabbits are immune.

78. BACILLUS SMARAGDINUS FÆTIDUS.

Obtained by Reimann (1887) from the nasal secretions in a case of ozaena.

Morphology.—Small, slender, slightly curved bacilli, about half as large as the tubercle bacilli; usually arranged in parallel groups.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying* bacillus. Spore formation not observed. Grows slowly at the room temperature in the usual culture media—more rapidly at 37° C. In *gelatin stick cultures* development occurs along the line of puncture, and at the end of forty-eight hours a slight liquefaction, in form of a funnel, occurs near the

surface; after the eighth day liquefaction progresses more rapidly. About the sixth day a bright-green color is recognized in the upper part of the tube by reflected light. Upon *agar plates*, at 37° C., at the end of forty-eight hours minute colonies are formed, of irregular form, which have a white color with a shade of green; in older colonies the central portion may be finely granular and brownish-yellow in color, while the marginal zone is more transparent; the agar has by reflected light a deep emerald green color. In *agar stick cultures*, at the end of twenty hours, an abundant development has occurred without color; at the end of forty-eight hours the culture medium is of a bright green color throughout; later the color changes to brown. A dirty-yellow layer forms upon the surface of the agar. Upon *potato*, at 37° C., a dark-brown layer forms in the vicinity of the line of inoculation; later this is chocolate brown.

The cultures in gelatin and agar give off a peculiar, penetrating odor similar to that of *jasmin*.

Pathogenesis.—Pathogenic for rabbits when injected into a vein or subcutaneously. Death occurs in from thirty-six to forty-eight hours. At the autopsy hæmorrhagic extravasations are found beneath the pericardium and the pleuræ; abscesses in the lungs and liver. The bacilli are found in the blood and in the various organs in large numbers.

79. BACILLUS PNEUMOSEPTICUS.

Obtained by Babes (1889) from the blood and tissues of an individual who died of septic pneumonia.

Morphology.—Small, straight bacilli about 0.2 μ thick.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* superficial colonies are formed which are flat, irregular in outline, whitish, shining, and semi-transparent; under a low power finger-like offshoots are seen about the periphery. In *gelatin stick cultures* an abundant development occurs along the line of puncture; the colonies give off a strong sperm-like odor. Upon the surface of *agar* small, whitish, flat, shining colonies with ill-defined outlines are formed, which soon become confluent and cover the surface; an abundant white deposit is seen in the condensation water. Upon *potato* a moist, white layer is formed. Upon *blood serum* circular, whitish, transparent colonies are formed along the line of inoculation, which soon coalesce.

Pathogenesis.—Very pathogenic for rabbits, guinea-pigs, and mice when injected subcutaneously in small amount. The animals die in from two to three days without any noticeable local inflammation and with symptoms of septicæmia. The lungs and spleen are found to be hyperæmic. The bacilli are found in the blood free, or sometimes enclosed in the leucocytes; they are only found in small numbers in the capillaries of the internal organs. Cultures gradually lose their virulence when propagated in artificial media.

80. BACILLUS CAPSULATUS.

Obtained by Pfeiffer (1889) from the blood of a guinea-pig which died spontaneously.

Morphology.—Thick bacilli with rounded ends, usually two or three times as long as broad; often united in chains of two or three elements; may grow out into homogeneous filaments. Stained preparations show the bacilli to be enveloped in an oval capsule which may be considerably broader than the bacilli themselves—two to five times as broad; where several bacilli are united they are surrounded by a single capsular envelope.

Stains with the usual aniline colors, but not by Gram's method. In preparations which are deeply stained with hot fuchsin or gentian violet solu-

tion the capsule is so deeply stained that the bacillus is hidden; by careful treatment with a weak solution of acetic acid the capsule may be differentiated as a pale-red or violet envelope surrounding the deeply stained bacilli.



FIG. 148.—*Bacillus capsulatus*, from peritoneal exudate of an inoculated guinea-pig. $\times 1,000$. From a photomicrograph. (Fleffer.)

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non liquefying*, *non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. The cultures in agar or upon potato are very viscid and draw out into long threads when touched with the platinum needle: the blood of an animal killed by inoculation with this bacillus has the same viscid character. Upon *gelatin plates* minute colonies are first visible at the end of twenty-four to thirty-six hours; later the deep colonies are white, oval masses the size of a pin's head; the superficial colonies attain the size of a lentil, and are flattened, hemispherical masses with a porcelain-white color. In *gelatin stick cultures* growth occurs to the bottom of the line of puncture, and on the surface a shining white, circular,

arched mass forms around the point of puncture, resembling the growth of Friedländer's bacillus. Upon the surface of *agar*, at 37°C , at the end of twenty-four hours a thick, soft layer of a pure white color is formed, which is very viscid and resembles the growth of *Micrococcus tetragenus* upon the same medium. Upon *potato* an abundant and viscid, shining, yellowish-white layer is quickly developed.

Pathogenesis.—Pathogenic for white mice and for house mice, which die at the end of two or three days after being inoculated at the root of the tail with a small quantity of a pure culture. Inoculation from mouse to mouse increases the virulence of the cultures. At the autopsy the superficial veins are distended with blood, the inguinal glands enlarged, the spleen considerably enlarged, the liver and kidneys hyperæmic, the intestine pale, the heart distended with blood, which usually is very viscid and is drawn out into threads when touched with the platinum needle. The bacilli are found in the blood and in all of the organs, in the contents of the peritoneum and pleuræ, and in the exudate in the vicinity of the point of inoculation. Pathogenic also for guinea pigs and for pigeons; guinea-pigs are infallibly killed within thirty-six hours by the injection of a single drop of a bouillon culture, twenty-four hours old, into the cavity of the abdomen; the blood contains the bacillus in enormous numbers, as does the viscid fluid found in the peritoneal cavity. Rabbits do not succumb to intraperitoneal or subcutaneous inoculations, but are killed by the intravenous injection of one cubic centimetre of a recent bouillon culture. Putrefactive changes occur very quickly in animals killed by inoculation with this bacillus.

81. *BACILLUS HYDROPHILUS FUSCUS*.

Obtained by Sanarelli (1891) from the lymph of frogs suffering from a fatal infectious disease.

Morphology.—Bacilli with rounded ends, usually from 1 to $3\ \mu$ in length; often short oval; may grow out into filaments of 12 to $20\ \mu$ in length.

Biological Characters.—An *aërobic*, *liquefying*, *motile* bacillus. Grows in the usual culture media at the room temperature. In *gelatin stick cul-*

tures, at 18° to 20° C., liquefaction has already commenced along the line of puncture at the end of twelve hours, and at the end of thirty-six to forty-eight hours half of the gelatin is liquefied in funnel shape; on the third or fourth day the gelatin is completely liquefied, and a thick, white, flocculent deposit is seen at bottom of the tube. In *glycerin-agar*, at 37° C., a slight, bluish, diffuse fluorescence is seen upon the surface at the end of twelve hours, and soon after a luxuriant growth, which soon covers the entire surface, is developed; at the end of twenty-four to thirty-six hours large gas bubbles begin to form in the agar; gradually the fluorescence disappears, the surface growth becomes thicker and has a dirty-gray color which changes later to brownish. *Blood serum* is a favorable medium and is rapidly liquefied by this bacillus. Upon *potato* the growth is most characteristic. At the end of twelve hours a thin, straw-yellow layer is developed along the impfstrich; this gradually becomes yellow, and at the end of four to five days has a brown color, resembling that of the glanders bacillus upon potato.

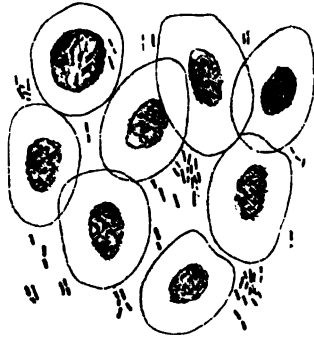


FIG. 144.—*Bacillus hydrophilus fuscus*, in blood of triton. (Sanarelli.)

Pathogenesis.—Pathogenic for frogs, toads, lizards, and other “cold-blooded” animals; also for guinea-pigs, rabbits, dogs, cats, mice, chickens, and pigeons. When a few drops of a bouillon culture are injected into the muscles of the thigh, swelling and redness at the point of inoculation are quickly developed, and death usually occurs in eight to ten hours. The bacilli are found in great numbers in the blood and in all of the organs. Guinea pigs die from general infection within twelve hours after receiving a subcutaneous injection of a small amount of a pure culture; the spleen is enlarged and the liver and spleen hyperæmic; an extensive inflammatory oedema in the vicinity of the inoculation wound is frequently observed; the bacilli are very numerous in the blood and in all the organs. Rabbits die in five to six hours from an intravenous injection. Adult dogs are immune, but new-born dogs (three to four days old) die infallibly, after receiving a subcutaneous injection of a small quantity of a pure culture, in twelve to thirty-six hours. Young cats also succumb to similar inoculations. Chickens and pigeons die within five to seven hours after receiving an intravenous injection, but resist subcutaneous injections.



FIG. 145.—*Bacillus hydrophilus fuscus*; culture in nutrient gelatin, end of six-teen hours. (Sanarelli.)

82. *BACILLUS TENUIS SPUTIGENUS*.

Obtained by Pansini (1890) from sputum.

Morphology.—Short bacilli, usually in pairs and surrounded by a capsule.

Stains by Gram's method.

Biological Characters.—An *aërobie*, *non-liquefying*, *non-motile* bacillus. Grows in nutrient gelatin at the room temperature. Develops abundantly on *potato*. Coagulates *milk* and produces an acid reaction in this medium.

Pathogenesis.—Pathogenic for rabbits and white rats; not for guinea-pigs or for white mice (in small doses).

83. BACILLUS OF LASER.

Obtained by Laser (1892) from mice which succumbed to an epidemic disease in Fränkel's laboratory at Königsberg.

In its characters this bacillus closely resembles the bacillus of swine plague (No. 65), and is perhaps identical with it.

Morphology.—A small bacillus, with rounded ends, about twice as long as broad. Has flagella both at the extremities and sides.

Stains by the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *actively motile* bacillus. Spore formation not observed. Grows either in the incubating oven or at the room temperature. Thermal death-point 65° to 70° C.—ten minutes' exposure. Upon *gelatin plates*, at the end of two days, the deep colonies are spherical, finely granular, and brownish in color; the superficial are transparent, fluely granular, and leaf-like. In *gelatin stick cultures* growth occurs along the entire line of puncture as well as upon the surface. At the end of three days a considerable evolution of gas is usually observed. In *agar* an abundant development is seen at the end of twenty-four hours in the incubating oven; upon the surface a grayish-white, shining layer with dentate margins is formed along the track of the needle. In *bouillon*, at 37° C., development is abundant and rapid; a thin film is formed on the surface at the end of the second day. Upon *potato* a brownish layer is formed at the end of twenty-four hours. In *milk* an acid reaction is produced.

Pathogenesis.—Pathogenic for field mice, guinea-pigs, rabbits, and pigeons. The bacillus is found in the blood and various organs of infected mice. The spleen is found to be greatly enlarged.

84. BACILLUS TYPHI MURIUM (Löffler).

Obtained by Löffler (1889) from mice which died in his laboratory from an epidemic disease due to this bacillus.

Morphology.—Short bacilli, resembling the bacillus of diphtheria in pigeons, and varying considerably in dimensions—like the bacillus of typhoid fever; grows out into flexible filaments.

Stains with the aniline colors—best with Löffler's solution of methylene blue.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not determined. Has flagella around the periphery of the cells, like those of the typhoid bacillus, and exhibits similar active movements. In *gelatin stick cultures*, at the room temperature, growth occurs upon the surface, at the end of forty-eight hours, in the form of a flat, grayish-white, round, semi-transparent mass the size of a pin's head; later the surface colony increases in extent and has more or less irregular margins. In *gelatin plate cultures* the deep colonies are at first round, slightly granular, transparent, and grayish; later they are of a yellowish-brown color and decidedly granular. The superficial colonies are very granular and marked by delicate lines—similar to colonies of the typhoid bacillus. Upon *agar* a grayish-white layer is developed which is not at all characteristic. Upon *potato* a rather thin, whitish layer is formed, and around this the potato acquires a dirty bluish-gray color. In *milk* an abundant development occurs, and a decidedly acid reaction is produced without causing any perceptible change in the appearance of the fluid.

Pathogenesis.—Pathogenic for white mice, which die in from one to two weeks after infection; also to field mice, which succumb to subcutaneous injections of a pure culture, and also, in from eight to twelve days, when fed upon potato cultures or bread moistened with a small quantity of a bouillon culture. Löffler believes that this bacillus may be used for the destruction of field mice in grain fields, inasmuch as they invariably die after ingesting food which has been contaminated with it, and also from eating the bodies

of other mice which have died as a result of infection. House mice are also susceptible. Rabbits, guinea-pigs, pigeons, and chickens were found by Löffler not to be susceptible to infection by feeding.

85. BACILLUS OF CAZAL AND VAILLARD.

Obtained by Cazal and Vaillard (1891) from cheesy nodules upon the peritoneum and in the pancreas of an individual who died in the hospital at Val de Grace.

Morphology.—Bacilli with rounded ends, but little longer than they are broad; solitary, in pairs, or in chains of ten to fifteen or more elements.

Stains with the usual aniline colors, but not by Gram's method; the extremities of the rods are more deeply stained than the central portion—"polar staining."

Biological Characters.—An *aërobie* and *facultative anaërobie*, *liquefying*, *motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature—more rapidly in the incubating oven at 37° C. In *gelatin stick cultures*, at the end of twenty-four hours, a series of punctiform, white colonies is developed along the line of puncture; upon the surface development is more abundant, and at the end of forty-eight hours liquefaction commences; this progresses slowly from above downward, and a white, flocculent deposit accumulates at the bottom of the liquefied gelatin. Upon the surface of *agar*, at the end of twenty-four hours at 37° C., a moist, transparent, opalescent layer is developed, which rapidly extends over the entire surface; later this layer becomes somewhat thicker, whitish, and cream-like in consistence, without losing its transparency. Upon *potato* a thick, prominent, moist, and slightly viscid layer is developed, which at first has a pale-yellow and later a yellowish-brown color. In *bouillon* development is abundant, producing a milky opacity of the liquid; a thick, flocculent deposit accumulates at the bottom of the tube; the reaction of the culture liquid becomes very alkaline. All of the cultures give off a peculiar odor, slightly ammoniacal and resembling that of putrid urine. The cultures retain their vitality for several months—in a closed tube for more than a year. The thermal death-point is 60° C. with fifteen minutes' exposure.

Pathogenesis.—Pathogenic for rabbits and mice, but not for guinea-pigs. In mice death occurs from general infection, at the end of forty-eight to sixty hours, from the subcutaneous injection of one eighth cubic centimetre of a recent bouillon culture. In rabbits injection of one cubic centimetre into the circulation causes the death of the animal in thirty-six to fifty hours. The symptoms induced are a foetid diarrhoea and paralysis of the extremities. When smaller doses are injected (0.5 cubic centimetre) a chronic malady is developed, characterized at the outset by diarrhoea and emaciation, then by the development of tumors which resemble those found in the man from whom the cultures were first obtained. These tumors are for the most part located in the subcutaneous connective tissue; after a time they attain the size of a chestnut and ulcerate, allowing the escape of a semi-fluid, purulent material. The animals usually recover. Similar tumors are developed as a result of subcutaneous injections of one to three cubic centimetres of a recent bouillon culture.

86. BACILLUS OF BABES AND OPRESCU.

Obtained by Babes and Opreescu (1891) from a case of septicæmia hæmorrhagica presenting some resemblance to exanthematic typhus.

Morphology.—In agar cultures the bacilli are from 0.4 to 0.5 μ thick, and are frequently united in pairs; associated with these rod-shaped bacteria are forms which are of a short oval. In gelatin cultures oval forms are more numerous; they have a diameter of 0.3 to 0.4 μ , and often appear to be surrounded by a capsule. In fresh cultures the bacilli are often in form of

a figure 8, and are only stained at the point of contact of the two segments. In potato cultures they are sometimes elongated and swollen at one extremity.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, actively motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—more rapidly at 37° C. In *gelatin stick cultures* yellowish-white colonies are developed along the line of puncture; at the bottom these may have a diameter of one to two millimetres, and they have a brown color. Upon the surface an irregular, lobulated, whitish, translucent, paraffin like layer is developed. At the end of eight days the surface growth consists of large, confluent, transparent plaques, with irregular outlines and crenated, elevated margins; along the line of puncture large, separate, lenticular or spherical colonies are seen; these have a brownish-white color. At the end of two months the surface growth is concentric and still more transparent, while the colonies near the surface have become almost brown. Upon the *surface of agar*, at 37° C., a narrow band is developed along the line of inoculation; above, this is composed of transparent, shining, flat, round colonies having a diameter of one millimetre or more; below, the colonies are confluent and form a transparent, whitish layer. In glycerin-agar development is still more abundant, and may already be perceived at the end of twelve hours. Crystals are seen below the surface in agar cultures and about the superficial colonies in gelatin. Upon *potato* a uniform, thin, grayish, very transparent layer is developed, which sometimes has a brownish-gray tint. At the end of a few days the potato acquires a brownish color. In *bouillon* cloudiness of the medium is apparent at the end of ten hours; twenty-four hours later a whitish precipitate is seen at the bottom of the tube, which is more abundant when the culture medium contains glucose; later a thin pellicle is seen upon the surface and the bouillon acquires a yellowish color.

Pathogenesis.—Recent cultures are pathogenic for rabbits, guinea-pigs, pigeons, and mice, which die from general infection in from two to four days. Old cultures are less virulent.

87. BACILLUS OF LUCET.

Obtained by Lucet (1891) from chickens and turkeys suffering from an infectious form of septicæmia characterized by dysenteric discharges—"Dysenterie epizootique des poules et des dindes."

Resembles *Bacillus gallinarum* of Klein, and is perhaps identical with this microorganism.

Morphology.—Short bacilli, from 1.2 to 1.8 μ long, usually in pairs.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, non-motile* bacillus. Spore formation not observed. Grows slowly in the usual culture media at the room temperature—more rapidly at 37° C.

In *gelatin plates* small, shining, moist, white, circular colonies are developed, which look like little drops of wax; later these increase in size, and especially in thickness, forming hemispherical masses. In *gelatin stick cultures* grayish, punctiform colonies are developed along the line of puncture, and upon the surface a circular, prominent, whitish plaque. Streak cultures upon the surface of gelatin are in the form of a dirty-white or grayish-white, moist streak, with regular margins, limited to the line of inoculation, but increasing in thickness until it breaks loose and slips down the oblique surface of the culture medium. The deposit which collects in this way acquires, as it becomes old, in the deepest portion a reddish color. Upon *agar* it forms a thick, yellowish-white, mucus-like layer with straight or slightly dentate margins. In *bouillon* it produces a decided clouding of the liquid, and an abundant grayish, pulverulent sediment accumulates at the bottom of the tube; the bouillon after a time becomes transparent above this sediment and



Fig. 1.



Fig. 2.



Fig 3.

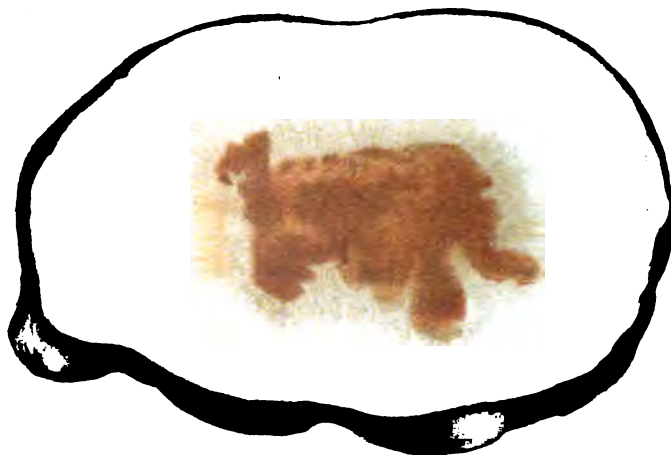


Fig. 4.

BACILLUS OF GLANDERS (LOEFFLER)

is viscid, drawing out into threads. In the absence of oxygen the characters of growth are the same as in its presence. The cultures acquire an alkaline reaction; they are sterilized by exposure for ten minutes to a temperature of 60° C. Does not grow upon *potato*.

Pathogenesis.—Pathogenic for chickens and turkeys. Not pathogenic for pigeons, guinea-pigs, or rabbits when injected subcutaneously or into the peritoneal cavity, but kills rabbits when injected into a vein. In the infected fowls the bacilli are found in small numbers in the blood, more numerous in the kidneys and liver, still more numerous in the spleen, and in enormous numbers in the intestinal mucus, where in acute cases it is found almost in a pure culture. Fowls do not contract the disease as a result of the ingestion of grains soiled with cultures of the bacillus, but become infected when fed with animal food to which a pure culture has been added.

88. CAPSULE BACILLUS OF LOEB.

Obtained from a case of keratomalacia infantum by inoculating culture media with a little of the softened exudate in the cornea.

Morphology.—Resembles *Bacillus capsulatus* of Pfeiffer, but this is said to be somewhat larger and thicker. In the blood of mice, however, both bacilli vary considerably in size, and according to Loeb it was not possible to determine with certainty that one bacillus was, on the average, larger than the other.

In staining reactions, also, no difference was observed—both bacilli stain with the usual aniline colors, and under certain circumstances the centre of the rods is less deeply stained than the extremities.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Grows in the usual culture media at the room temperature. In its growth in culture media it closely resembles *Bacillus capsulatus* of Pfeiffer (No. 80).

Pathogenesis.—Pathogenic for mice and for guinea-pigs, but not for rabbits and pigeons; Pfeiffer's bacillus is pathogenic for these animals.

PLATE VII.

BACILLUS OF GLANDERS.

FIG. 1.—*Bacillus mallei* from the liver of a field mouse, cover-glass preparation. (Löffler.)

FIG. 2.—*Bacillus mallei* from a recent culture upon blood serum. (Löffler.)

FIG. 3.—*Bacillus mallei* in section of spleen of a field mouse dead from glanders. (Löffler.)

FIG. 4.—Culture of glanders bacillus upon cooked potato. (Löffler.)

XIII.

PATHOGENIC AËROBIC BACILLI NOT DESCRIBED IN PREVIOUS SECTIONS.

A CONSIDERABLE number of saprophytic bacilli are pathogenic for small animals when injected into the circulation, or subcutaneously, or into a serous cavity in considerable quantity—one to five cubic centimetres or more—but fail to produce any appreciable effect when introduced into the bodies of these animals in minute doses, and do not multiply in the blood to any considerable extent, although in fatal cases they may usually be recovered in cultures from the blood and tissues. These bacilli are pathogenic by reason of the toxic ptomaines produced by them, or because of local inflammatory processes which they induce, or for both of these reasons combined. Some of them may also, under certain circumstances, multiply in the blood and thus give rise to septicæmia as well as to toxæmia; this is the case, for example, with the “colon bacillus” of Escherich. When injected in considerable quantity into the circulation of a guinea-pig it causes the death of the animal within twenty-four hours, and the bacillus is found in the blood in great numbers; but minute amounts injected into a vein, or larger amounts injected subcutaneously, do not usually produce general infection. It is, therefore, not included among the “bacilli which produce septicæmia in susceptible animals.” There is reason to believe, however, that under certain circumstances this bacillus may have sufficient pathogenic potency to produce a genuine septicæmia in guinea-pigs. Thus the original cultures of Brieger’s bacillus, which appears to be a variety of the colon bacillus, are reported to have produced fatal septicæmia in guinea-pigs when injected subcutaneously in small amounts. A strict division into pathogenic bacilli which produce general blood infection—septicæmia—and those which produce a fatal result owing to the production of toxic chemical substances is not possible; for many pathogenic bacteria produce general infection when injected in comparatively large doses, and at the same time give rise to symptoms of toxæmia; or general infection may occur in animals of one species, and fatal toxæmia without septicæmia in

those of another species. Many of the bacilli described in the present section are common saprophytes, which have been shown by laboratory experiments to be pathogenic for certain animals when introduced into their bodies in a certain amount, which differs greatly for different bacteria and for different species of animals. The experiments of Cheyne and others show how largely the pathogenic power of saprophytic bacteria depends upon the quantity of a culture which is injected, as well as upon the age of the culture and the seat of the inoculation—in the blood, the abdominal cavity, the subcutaneous tissues, or the muscles. And the bacteriologist named has also shown that pathogenic power depends, in some instances at least, upon the combined action of the toxic substances introduced in the first instance and of the living bacteria. Thus Cheyne found that one-tenth of a cubic centimetre of a bouillon culture of *Proteus vulgaris* injected into the dorsal muscles of a rabbit infallibly caused its death within forty-eight hours, but when the dose was reduced to one-fortieth cubic centimetre the animal recovered. But if to this amount (one-fortieth cubic centimetre) he added one cubic centimetre of a sterilized (by heat) culture of the same bacillus instead of diluting with distilled water, and injected the mixture into the dorsal muscles of a rabbit, death occurred in every experiment within forty-eight hours. The sterilized culture injected by itself produced no effect in this dose (one cubic centimetre), and Cheyne believes that the fatal result in these experiments was due to the fact that the toxic products present in the sterilized culture overcame the natural resisting powers of the tissues and enabled the bacillus to multiply over a larger area than would otherwise have been the case. As a result of this, toxic substances were produced in the body of the animal in sufficient quantity to cause general toxæmia and death; whereas the bacilli alone, in the dose mentioned, were not able to invade the tissues in the vicinity of the point of inoculation, and gave rise to a local abscess only. The same explanation is probably true for very many of the saprophytic bacteria which have been shown to possess pathogenic power; and it is probable that many of those which are now classed by bacteriologists as non-pathogenic would prove to be pathogenic in the same way if thoroughly tested upon various species of animals, although it might be necessary to use unusually large doses to accomplish the same result.

89. BACILLUS COLI COMMUNIS.

Synonyms.—*Bacterium coli commune* (Escherich); Colon bacillus of Escherich; Emmerich's bacillus (*Bacillus Neapolitanus*). Probably identical with *Bacillus cavica* (Brieger's bacillus).

Obtained by Emmerich (1885) from the blood, various organs, and the alvine discharges of cholera patients at Naples; by Weisser (1886) from normal and abnormal human fæces, from the air, and from putrefying infusions; by Escherich (1886) from the fæces of healthy children; since shown to be constantly present in the alvine discharges of healthy men, and probably of many of the lower animals. Found by the writer in the blood and various organs of yellow-fever cadavers, in Havana (1888 and 1889).

Numerous varieties have been cultivated by different bacteriologists, which vary in pathogenic power and to some extent in their growth in various culture media; but the differences described are not sufficiently characteristic or constant to justify us in considering them as distinct species.

Morphology.—Differs considerably in its morphology as obtained from different sources and in various culture media. The typical form is that of short rods with rounded ends, from two to three μ in



FIG. 146. — Bacillus coli communis, $\times 1,000$. (Escherich.)

length and 0.4 to 0.6 μ broad; but under certain circumstances the length does not exceed the breadth—about 0.5 μ —and it might be mistaken for a micrococcus; again the prevailing form in a culture is a short oval; filaments of five μ or more in length are often observed in cultures, associated with short rods or oval cells. The bacilli are frequently united in pairs. The presence of spores has not been demonstrated. In unfavorable culture media the bacilli, in stained preparations, may present unstained places, which are supposed by Escherich to be due to degenerative changes in the protoplasm. Under certain circumstances some of the rods in a pure culture have been observed by Escherich to present spherical, unstained portions at one or both extremities, which closely resemble spores, but which he was not able to stain by the methods usually employed for staining spores, and which he is inclined to regard as “involution forms.”

This bacillus *stains* readily with the aniline colors usually employed by bacteriologists, but quickly parts with its color when treated with iodine solution—Gram’s method—or with diluted alcohol.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* bacillus. Sometimes exhibits independent movements, which are not very active. One rod of a pair, in a hanging-drop culture, may advance slowly with a to-and-fro movement, while the other follows as if attached to it by an invisible band (Escherich). The writer’s personal observations lead him to believe that, as a rule, this bacillus does not exhibit independent movements. Does not form spores. Grows in various culture media at the room

temperature—more rapidly in the incubating oven. Grows in a decidedly acid medium.

In *gelatin plates* colonies are developed in from twenty-four to forty-eight hours, which vary considerably in their appearance according to their age, and in different cultures in the same medium. The deep colonies are usually spherical and at first are transparent, homogeneous, and of a pale-straw or amber color by transmitted light; later they frequently have a dark-brown, opaque central portion surrounded by a more transparent peripheral zone; or they may be coarsely granular and opaque; sometimes they have a long-oval or "whetstone" form. The superficial colonies differ still more in appearance; very young colonies by transmitted light often resemble little drops of water or fragments of broken glass; when they have sufficient space for their development they quickly increase in size, and may attain a diameter of three to four centimetres; the central portion is thickest, and is often marked by a spherical nucleus of a dark-brown color when the colony has started below the surface of the gelatin; the margins are thin and transparent, the thickness gradually increasing towards the centre, as does also the color, which by transmitted light varies from light straw color or amber to a dark brown. The outlines of superficial colonies are more or less irregular, and the surface may be marked by ridges, fissures, or concentric rings, or may be granular. The writer has observed colonies resembling a rosette, or a daisy with expanded petals. Escherich speaks of colonies which present star-shaped figures surrounded by concentric rings.

In *gelatin stick cultures* the growth upon the surface is rather dry, and may be quite thin, extending over the entire surface of the gelatin, or it may be thicker with irregular, leaf-like outlines and with superficial incrustations or concentric annular markings. An abundant development occurs all along the line of puncture, which in the deeper portion of the gelatin is made up of more or less closely crowded colonies; these are white by reflected light, and of an amber or light-brown color by transmitted light; later they may become granular and opaque. Frequently a diffused cloudy appearance is observed near the surface of the gelatin, and under certain circumstances branching, moss-like tufts develop at intervals along the line of growth. One or more gas bubbles may often be seen in recent stick cultures in gelatin.

Upon nutrient agar and blood serum, in the incubating oven, an abundant, soft, white layer is quickly developed. Upon *potato* an abundant, soft, shining layer of a brownish-yellow color is developed. The growth upon potato differs considerably, according to the age of the potato. According to Escherich, upon old potatoes there may

be no growth, or it may be scanty and of a white color. In *milk*, at 37° C., an acid reaction and coagulation of the casein are produced at the end of eight or ten days. In the absence of oxygen this bacillus is able to grow in solutions containing grape sugar (Escherich). In bouillon it grows rapidly, producing a milky opacity of the culture liquid. The thermal death-point of Emmerich's bacillus, and of the colon bacillus from fæces, was found by Weisser to be 60° C., the time of exposure being ten minutes. The writer has obtained corresponding results. Weisser found that when the bacilli from a bouillon culture were dried upon thin glass covers they failed to grow



FIG. 147.



FIG. 148.

FIG. 147.—*Bacillus coli communis* in nutrient gelatin containing twenty per cent of gelatin, end of two weeks, showing moss-like tufts along the line of growth. (Sternberg.)

FIG. 148.—A portion of the growth shown in Fig 147, at *a*, magnified about six diameters. From a photograph. (Sternberg.)

after twenty-four hours. These results give confirmation to the view that the bacillus under consideration does not form spores.

Pathogenesis.—Comparatively small amounts of a pure culture of the colon bacillus injected into the circulation of a guinea-pig usually cause the death of the animal in from one to three days, and the bacillus is found in considerable numbers in its blood. But when injected subcutaneously or into the peritoneal cavity of rabbits or guinea-pigs, a fatal termination depends largely on the quantity injected; and although the bacillus may be obtained in cultures from the blood and the parenchyma of the various organs, it is not present

in large numbers, and death appears to be due to toxæmia rather than to septicæmia. Mice are not susceptible to infection by subcutaneous injections. Small quantities injected beneath the skin of guinea-pigs usually produce a local abscess only; larger amounts—two to five cubic centimetres—frequently produce a fatal result, with symptoms and pathological appearances corresponding with those resulting from intravenous injection. These are fever, developed soon after the injection, diarrhoea, and symptoms of collapse appearing shortly before death. At the autopsy the liver and spleen appear normal, or nearly so; the kidneys are congested and may present scattered punctiform ecchymoses (Weisser). According to Escherich, the spleen is often somewhat enlarged. The small intestine is hyperæmic, especially in its upper portion, and the peritoneal layer presents a rosy color; the mucous membrane gives evidence of more or less intense catarrhal inflammation, and contains mucus, often slightly mixed with blood. In rabbits death occurs at a somewhat later date, and diarrhoea is a common symptom. In dogs the subcutaneous injection of a considerable quantity of a pure culture may give rise to an extensive local abscess.

Varieties.—Booker, in his extended studies relating to the bacteria present in the fæces of infants suffering from summer diarrhoea, has isolated seven varieties “which closely resemble *Bacterium coli commune* in morphology and growth in agar, neutral gelatin, and potato, but by means of other tests a distinction can be made between them.” These are described as follows:

BACILLUS *d* OF BOOKER.

“Found in two cases of cholera infantum and the predominating form in one serious case of catarrhal enteritis.

“*Morphology.*—Resembles *Bacterium coli commune*.

“*Growth in Colonies.*—Gelatin: Colonies grow luxuriantly in gelatin, and thrive in acid and sugar gelatin equally as well as in neutral gelatin. In the latter the colonies closely resemble, but are not identical with, the *Bacterium coli commune*. In acid gelatin they differ very much from *Bacterium coli commune*. The colonies spread extensively and are bluish-white with concentric rings. Slightly magnified, they have a large, uniform, yellow central zone surrounded by a border composed of perpendicular threads placed thickly together. Sometimes a series of these rings appear with intervening yellow rings.

“Agar: The colonies are round, spread out, and blue or bluish-white. Slightly magnified, they have a pale-yellow color.

“*Stab Cultures.*—Gelatin: In sugar gelatin the surface growth has a nearly colorless centre surrounded by a thick border with an outer edge of fine, hair-like fringe; the growth along the line of inoculation is fine and delicate. In neutral gelatin the growth is not so luxuriant as on sugar gelatin; on the surface it is thick and white, with a delicate stalk in the depth.

“Agar: Thick white surface growth with a well-developed stalk in the depth.

“Potato: Luxuriant yellow, glistening, moist, and slightly raised surface, with well-defined borders.

"*Action on Milk*.—Coagulated into a gelatinous coagulum in twenty-four hours at 38° C., and into a solid clot in two days.

"*Milk Litmus Reaction*.—Milk colored blue with litmus is changed to light pink in twenty-four hours at 38° C. The pink color gradually fades, and by the second or third day is white or cream color with a thin layer of pink on top. The pink color extends in a few days about one-half down the clot.

"*Temperature*.—Grows best about 38° C.

"*Spores* have not been observed.

"*Gas Production*.—Gas bubbles are produced in milk; not observed on potato."

BACILLUS *e* OF BOOKER.

"Found as the predominating form in two cases of dysentery one of which was fatal and the other a mild case.

"*Morphology*.—Resembles *Bacterium coli commune*.

"*Growth in Colonies*.—Gelatin: The colony growth varies considerably with slight difference in the gelatin. In ten-per-cent neutral gelatin the colonies resemble those of *Bacterium coli commune*. On the second or third day, when the colonies have just broken through the surface and are spread out, it is impossible to distinguish one variety from the other, but as the colonies grow older a difference can generally be recognized. In sugar and acid gelatin the colonies have a clear centre with white border; slightly magnified, a uniform brown centre surrounded by a brown zone composed of fine, needle-like rays perpendicular to the border. After cultivating for a few generations on acid and sugar gelatin the colonies cease to develop, and either grow in very small colonies or do not grow at all. The activity is regained if cultivated on neutral gelatin.

"*Agar*: Colonies are large, round, and have a mother-of-pearl appearance. Slightly magnified, a uniform yellow color.

"*Slab Cultures*.—*Agar*: Luxuriant, nearly colorless surface growth, with well-developed stalk along the line of inoculation in the depth.

"*Potato*: Golden-yellow, glistening, slightly raised surface with well-defined borders.

"*Action on Milk*.—Milk becomes gelatinous in twenty-four hours at 38° C., and in a few days a solid coagulum is formed. Milk colored blue with litmus is reduced to white or cream color in twenty-four to forty-eight hours at 38° C., with a thin layer of pink at the top of the culture. The pink color gradually extends lower in the coagulum.

"*Temperature*.—Thrives best at about 38° C.

"*Spores* have not been observed.

"*Gas Production*.—Occurs in milk, but not seen in potato cultures.

"*Relation to Gelatin*.—Does not liquefy gelatin.

"*Resemblance*.—Resembles *Bacterium coli commune* and *bacillus d*; differing from the former in the character of the colony growth on acid and sugar gelatin, and in ceasing to develop in these media after several generations. It differs from *bacillus d* in this latter respect."

BACILLUS *f* OF BOOKER.

"Found in one case of cholera infantum and one case of catarrhal enteritis.

"*Morphology*.—Resembles *Bacterium coli commune*.

"*Growth in Colonies*.—Gelatin: It is difficult to distinguish the colony growth from the *Bacterium coli commune*. There is often a difference in the colonies planted at the same time and kept under similar conditions, but it is not very marked nor always the same kind of difference. The tendency to concentric rings is greater in this variety. The colonies develop somewhat better on neutral and sugar gelatin than on acid gelatin.

"**Agar:** The colonies are large, round, and bluish-white. Slightly magnified, a light-yellow color.

"**Stab Cultures.**—Gelatin: The culture is spread over the surface and has a mist-like appearance; in the depth along the line of inoculation is a delicate stalk.

"**Agar:** Thick, luxuriant, white surface growth, with a well-developed stalk along the line of inoculation in the depth.

"**Potato:** Bright-yellow, glistening, moist surface with well-defined borders, and but slightly raised above the surrounding potato.

"**Action on Milk and Litmus Reaction.**—Milk is coagulated into a solid clot in twenty-four hours at 38° C. Milk colored blue with litmus is changed to pink in twenty-four hours at 38° C., and in forty-eight hours is reduced to white or cream color with a thin pink layer on top.

"**Gas Production.**—Gas bubbles arise in milk cultures, but they have not been observed on potato cultures.

"**Temperature.**—Grows better at 38° C.

"Spores have not been observed.

"**Relation to Gelatin.**—Does not liquefy gelatin.

"**Resemblance.**—It closely resembles *Bacterium coli commune* and Brieger's bacillus in the character of its growth upon different media, but is readily distinguished from both, as is also Brieger's bacillus from the *Bacterium coli commune*, by the following differential test recently made known by Dr. Mall. Yellow elastic tissue from the ligamentum nuchæ of an ox is cut into fine bits and placed in test tubes containing water with ten-per-cent bouillon and one-per-cent sugar, and sterilized from one and one-half to two hours at a time for three consecutive days. Into this is inoculated two species of bacteria, one of which is the bacterium under observation, the other a bacillus found in garden earth. The latter bacillus is anaërobic, grows in hydrogen, nitrogen, and ordinary illuminating gas, in the bottom of bouillon, in the depth but not on the surface of agar stab cultures, and not at all in gelatin stab cultures. It has a spore in one end making a knob bacillus. Different species of bacteria—*Streptococcus Indicus*, tetragenus, cholera, swine plague, *Bacterium lactis aërogenes*, *Bacterium coli commune*, Brieger's bacillus, and a number of varieties of bacteria which I have isolated from the fæces—were inoculated with the head bacillus into the above-described elastic-tissue tubes. The tubes inoculated with Brieger's bacillus developed a beautiful purple tint, which started as a narrow ring at the top of the culture, gradually extending downward and deepening in color until the whole tube had a dark-purple color. This color reaction began in five to fourteen days, and was constantly present in a large number of tests. Tubes inoculated with bacillus *f* gave a much fainter purple color, which was longer in appearing and never became so dark as with Brieger's bacillus.

"Tubes inoculated with the other species of bacteria above mentioned gave no color change and remained similar to control. Bacillus *f* also shows a slight difference from *Bacterium coli commune* in coagulating milk and reducing litmus more rapidly, and appears to produce more active fermentation in milk. Like Brieger's bacillus, the gelatin colonies more frequently show a concentric arrangement than those of the *Bacterium coli commune*."

BACILLUS *g* OF BOOKER.

"Found in one case of serious gastro-enteric catarrh. It was not in large quantity.

"**Morphology and Biological Characters.**—In morphology, character of growth on agar, gelatin, and potato, it resembles *Bacterium coli commune*.

"**Action on Milk and Litmus Reaction.**—Milk is not coagulated, and milk colored blue with litmus is changed to pink in a few days, and holds this color. These characteristics distinguish it from the *Bacterium coli commune*.

"**Gas Production.**—Not observed in milk or potato cultures.

"**Relation to Gelatin.**—Does not liquefy gelatin."

BACILLUS *h* OF BOOKER.

"Found in one case of mild dysentery, not in large quantity.

"*Morphology*.—Resembles *Bacterium coli commune*.

"*Growth in Colonies*.—Gelatin: In plain neutral gelatin the colonies resemble those of *Bacterium coli commune*. In sugar gelatin the colonies are white and spread extensively. Slightly magnified, they have a round, dark centre surrounded by a yellow, loose zone with an outer white rim; later the whole colony has a uniform yellow color and is not compact.

"Agar: Colonies are white, round, and large. Slightly magnified, they are brownish-yellow.

"*Stab Cultures*.—Nothing characteristic in gelatin and agar.

"Potato culture is yellow, dry, and slightly raised, with well-defined borders.

"*Action on Milk and Litmus Reaction*.—Milk is coagulated into a solid clot in two days at 38° C. Milk colored blue with litmus is changed to pink in twenty-four hours.

"*Gas Production*.—Occurs in milk; not observed on potato.

"*Relation to Gelatin*.—Does not liquefy gelatin."

BACILLUS *k* OF BOOKER.

"Found in two cases of cholera infantum and one of catarrhal enteritis.

"*Morphology*.—Resembles *Bacterium coli commune*.

"*Growth in Colonies*.—Gelatin: In neutral gelatin the colonies cannot be distinguished from those of *Bacterium coli commune*. In acid gelatin the colonies do not spread so extensively as those of *Bacterium coli commune*, and they have a decided concentric arrangement, a wide white centre surrounded by a narrow, transparent blue ring, and outside of this a white border. Slightly magnified, the colonies have an irregular, yellowish-brown centre mottled over with dark spots and surrounded by a light-yellow ring bordered by a brownish-yellow wreath.

"Agar: Colonies are large, round, and bluish-white. Slightly magnified, a light brownish-yellow color.

"*Stab Cultures*.—Gelatin: In sugar gelatin the surface growth is extensive, nearly colorless, and has a rough, misty appearance. In the depth is a delicate growth. In plain neutral gelatin the surface growth is bluish-white, thick, and not so extensively spread; the growth in the depth is also thicker.

"Potato culture is moist, dirty-cream color, has raised surface and defined border.

"*Action on Milk*.—Milk becomes gelatinous in twenty-four hours at 38° C., and a solid clot in two days. Milk colored blue with litmus is changed to pink in twenty-four hours, and reduced to white with a pink layer on top in two days."

BACILLUS *n* OF BOOKER.

"Found in large quantity, but not the predominating form, in one case of chronic gastro-enteric catarrh—extremely emaciated.

"*Morphology*.—Resembles *Bacterium coli commune*.

"*Growth in Colonies*.—Gelatin: In neutral gelatin the colonies are spread out and have a frosty or ground-glass appearance. The centre is blue and border white, but both have the ground-glass appearance. Slightly magnified, the central part is light yellow and the border brown with a rough, furrowed surface. In acid gelatin the white border is wider and the surface is rougher.

"Agar: Colonies are round, blue, or bluish-white, and spread out. Under the microscope they have a light-yellow color.

"*Stab Cultures*.—Gelatin: Has a rough, nearly colorless surface growth, and a thick stalk in the depth along the line of inoculation.

"Agar: Thick white surface growth with well-developed stalk in the depth.

"*Action on Milk and Litmus Reaction.*—Milk remains liquid, and milk colored blue with litmus is changed to pink.

"*Gas Production.*—Not observed in milk or potato cultures.

"*Relation to Gelatin.*—Does not liquefy gelatin.

"*Spores* have not been noticed."

Bacillus Coli Communis in Peritonitis.—The researches of A. Fränkel show that *Bacillus coli communis* may be obtained in pure cultures from the exudate into the peritoneal cavity in a considerable proportion of the cases of peritonitis, and there is good reason for believing that in these cases it was the cause of the inflammatory process. Thirty-one cases were examined by Fränkel, with the following result: Pure cultures of *Bacillus coli communis* were obtained in nine cases; of *Streptococcus* (*pyogenes*?) in seven; of *Bacillus lactis aërogenes* in two; of "*diplococcus pneumoniae*" in one; of *Staphylococcus pyogenes aureus* in one. Of the remaining eleven cases, seven gave mixed cultures, and in three of these *Bacillus coli communis* was the most abundant species. The author referred to has also shown that pure cultures of *Bacillus coli communis* injected into the cavity of the abdomen of rabbits cause a typical peritonitis. The present writer has frequently obtained the same result in experiments made with this bacillus. It would appear, therefore, that the peritonitis which so constantly results from wounds of the intestine is probably due, to a considerable extent, to the introduction of this microorganism from the lumen of the intestine, where it is constantly found, into the peritoneal cavity, where the conditions are favorable for its rapid development.

90. BACILLUS LACTIS AÉROGENES.

Synonym —*Bacillus lactis aërogenes* (Escherich).

Obtained by Escherich (1886) from the contents of the small intestine of children and animals fed upon milk; in smaller numbers from the faeces of milk-fed children, and in one instance from uncooked cow's milk.

Morphology.—Short rods with rounded ends, from 1 to 2 μ in length and from 0.1 to 0.5 μ broad; short oval and spherical forms are also frequently observed, and, under certain circumstances, longer rods—3 μ —may be developed: usually united in pairs, and occasionally in chains containing several elements. In some of the larger cells Escherich has observed unstained spaces, but was not able to obtain any evidence that these represent spores.

This bacillus stains readily with the ordinary aniline colors, but does not retain its color when treated by Gram's method.

Biological Characters.—An *aërobic* (facultative anaërobic), *non-liquefying*, *non-motile* bacillus. Does not form spores. Grows in various culture media at the room temperature—more rapidly in the incubating oven. Upon *gelatin plates*, at the end of twenty-four hours, small white colonies are developed. Upon the surface these form hemispherical, soft, shining masses which, examined under the microscope, are found to be homogeneous



FIG. 149.—*Bacillus lactis aërogenes*. $\times 1,000$. Escherich.)

and opaque, with a whitish lustre by reflected light. The deep colonies are spherical and opaque and attain a considerable size. In gelatin stick cultures the growth resembles that of Friedländer's bacillus—i.e., an abundant growth along the line of puncture and a rounded mass upon the surface, forming a "nail-shaped" growth. In old cultures the upper portion of the gelatin is sometimes clouded, and numerous gas bubbles may form in the gelatin. Upon the surface of nutrient agar an abundant, soft, white layer is developed. Upon old potatoes, in the incubating oven, at the end of twenty-four hours a yellowish-white layer, several millimetres thick, is developed, which is of paste-like consistence and contains about the periphery a considerable number of small gas bubbles; this layer increases in dimensions, has an irregular outline, and larger and more numerous gas bubbles are developed about the periphery, some the size of a pea; later the whole surface of the potato is covered with a creamy, semi-fluid mass filled with gas bubbles. On young potatoes the development is different; a rather luxuriant, thick, white or pale-yellow layer is formed, which is tolerably dry and has irregular margins; the surface is smooth and shining, and a few minute gas bubbles only are formed after several days.

Pathogenesis.—Injections of a considerable quantity of a pure culture into the circulation of rabbits and of guinea-pigs give rise to a fatal result within forty-eight hours.

In his first publication relating to "the bacteria found in the dejecta of infants afflicted with summer diarrhoea," Booker has described a bacillus which he designates by the letter B, which closely resembles *Bacillus lactis aërogenes* and is probably identical with it. He says:

"*Summary of Bacillus B.*—Found nearly constantly in cholera infantum and catarrhal enteritis, and generally the predominating form. It appeared in larger quantities in the more serious cases. It was not found in the dysenteric or healthy faeces. It resembles the description of the *Bacillus lactis aërogenes*, but the resemblance does not appear sufficient to constitute an identity, and, in the absence of a culture of the latter for comparison, it is considered a distinct variety for the following reasons: *Bacillus B* is uniformly larger, its ends are not so sharply rounded, and in all culture media long, thick filaments are seen, and many of the bacilli have the protoplasm gathered in the centre, leaving the poles clear. There is some difference in their colony growth on gelatin, and in gelatin stick cultures *Bacillus B* does not show the nail-form growth with marked end swelling in the depth. In potato cultures the *Bacillus lactis aërogenes* shows a difference between old and new potatoes, while *Bacillus B* does not show any difference.

"*Bacillus B* possesses decided pathogenic properties, which was shown both by hypodermic injections and feeding with milk cultures."

91. BACILLUS C OF BOOKER.

Found by Booker (1889) in a case of cholera infantum.

Morphology.—Resembles *Bacillus lactis aërogenes* of Escherich.

Biological Characters.—Resembles *Bacillus lactis aërogenes*, but differs from it in not coagulating milk; the growth on potato also is more luxuriant and the surface is more thickly covered with gas bubbles.

BACILLI OF JEFFRIES.

Jeffries, in a study of the alvine discharges of children suffering from summer diarrhoea, isolated a number of bacilli resembling *Bacillus coli communis* and *Bacillus lactis aërogenes* of Escherich. He says:

"While Brieger's bacillus and the lactic acid bacillus of Escherich were not found, a whole group of species in growth, form, and general physiology closely resembling them have been isolated. This group is represented by bacilli A, G, J, K, P, S, Z; they seem to form a genus; the form is very

much alike. All are good anaërobic growers; all form gas; all turn milk distinctly acid; and all closely resemble one another in pure cultures. Many would doubtless class these altogether as one species; but if species are to be recognized at all, we must come to recognizing every fixed difference as constituting a species.

"This group occurred—always very abundantly—in eighteen out of the twenty-two cases of summer diarrhoea, and is also well represented among the kittens. They are, however, so much like the harmless forms found by Escherich that they may for the present be laid aside as of no specific significance. They are also almost the only forms tested which failed to produce intestinal troubles in kittens. Excluding these, there is no species, or group of species, left either generally occurring or in sufficient numbers to be regarded as the specific pathogenic plant of summer diarrhoea."

92. BACILLUS ACIDIFORMANS.

Obtained by the writer (1888) from a fragment of yellow-fever liver preserved for forty-eight hours in an antiseptic wrapping; since obtained from

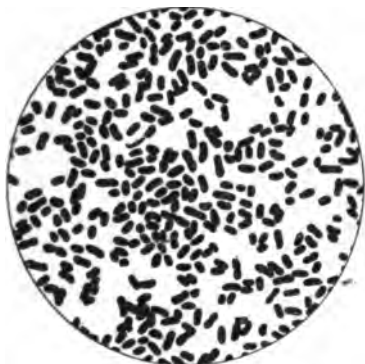


FIG. 150.

FIG. 150.—*Bacillus acidiformans*, from a potato culture. $\times 1,000$. From a photomicrograph (Sternberg.)



FIG. 151.

FIG. 151.—Culture of *Bacillus acidiformans* in nutrient gelatin, end of four days at 22°C . From a photograph. (Sternberg.)

liver preserved in the same way from two comparative autopsies—*i.e.*, not cases of yellow fever.

Morphology.—A short bacillus with rounded corners, sometimes short oval in form; from $1\frac{1}{2}$ to 3μ in length and about 1.2μ in breadth; may grow out into filaments of 5 to 10μ , or more, in length; in some cultures the short oval form predominates.

Stains readily with the aniline colors usually employed, and by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Does not form spores. Grows rapidly at the room temperature in the usual culture media. Grows in decidedly acid media; in culture media containing glycerin or glucose it produces an abundant evolution of carbon dioxide, and a volatile acid is formed.

It does not liquefy gelatin, and in stick cultures grows abundantly both on the surface and along the line of puncture. At the end of twenty-four hours, at 22°C ., a rounded white mass is formed upon the surface, resembling

the growth of Friedländer's bacillus; at the bottom of the line of puncture the separate colonies are spherical, opaque, and pearl-like by reflected light. Gas bubbles are formed in the gelatin. At the end of a week the surface is covered with a thick, white, semi-fluid mass.

In gelatin roll tubes the superficial colonies are translucent or opaque, and circular or somewhat irregular in outline; by reflected light they are slightly iridescent; the deep colonies are spherical, opaque, and homogeneous.

The growth upon the surface of *nutrient agar* is abundant and rapid, of a shining milk-white color, and cream-like in consistence. An abundant development forms along the line of puncture and the culture medium is split up by gas bubbles. In glycerin-agar the evolution of gas is very abundant and the culture medium acquires an intensely acid reaction.

On *potato* the growth is abundant and rapid at a temperature of 20° to 30° C., forming a thick, semi-fluid mass of a milk-white color.

I have not obtained any evidence that this bacillus forms spores; the cultures are sterilized by ten minutes' exposure to a temperature of 160° F.

When cultivated in bouillon to which five per cent of glycerin has been added the culture medium acquires a milky opacity, and there is a copious precipitate, of a viscid consistence, consisting of bacilli; during the period of active development the surface is covered with gas bubbles, as in a saccharine liquid undergoing alcoholic fermentation, and the liquid has a decidedly acid reaction.

Pathogenesis.—Pathogenic for rabbits and for guinea-pigs when injected into the cavity of the abdomen—one to two cubic centimetres of a culture in bouillon. The animal usually dies in less than twenty-four hours. The bacilli are found in the blood in rather small numbers, and are frequently seen in the interior of the leucocytes. The spleen is enlarged, the liver normal, the intestine usually hyperæmic.

93. BACILLUS CUNICULICIDA HAVANIENSIS.

Obtained by the writer (1889) from the contents of the intestine of yellow-fever cadavers, and also from fragments of yellow-fever liver preserved for forty-eight hours in an antiseptic wrapping—my bacillus α , Havana, 1889.

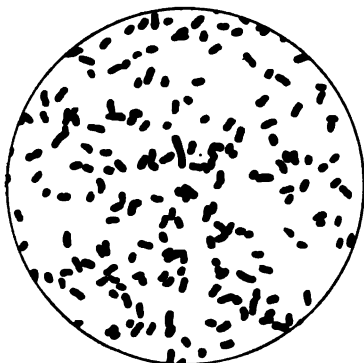


FIG. 152.—*Bacillus cuniculicida* Havaniensis, from a single colony in nutrient gelatin. $\times 1,000$. From a photomicrograph. (Sternberg.)

Morphology.—This bacillus resembles the colon bacillus in form, but is somewhat larger, from 2 to 4 μ in length and from 0.8 to 1 μ in diameter; sometimes associated in pairs; may grow out into short filaments—not common. The ends of the rods are rounded, and under certain circumstances vacuoles are seen at the extremities, especially in potato cultures.

Stains quickly with the aniline colors usually employed, and also by Gram's method.

Biological Characters.—An *aërobie* and *facultative anaërobie*, *non-liquefying* bacillus. Under certain circumstances may exhibit active movements, but is usually motionless.

A very curious thing with reference to this bacillus is that it presented active movements in my cultures made directly from yellow-fever cadavers, but that these movements were not constant, and that since my return to Baltimore I have not, as a rule, observed active movements in cultures from the same stock, which, however, preserved their pathogenic power and other

characters. In Havana these movements were usually not observed in all the bacilli in a field under observation, but one and another would start from a quiescent condition on an active and erratic course; sometimes spinning actively upon its axis, and again shooting across the field as if propelled by a flagellum.

My notes indicate that cultures passed through the guinea-pig are more apt to be motile.

In gelatin stick cultures the growth of bacillus *x* resembles that of the colon bacillus, but the colonies at the bottom of the line of puncture are more opaque and not of a clear amber color like that of colonies of the colon bacillus. Upon the surface the growth is thicker than that of the colon bacillus, and forms a milk-white, soft mass.

The colonies in gelatin Esmarch roll tubes vary considerably at different times. Deep colonies are usually spherical, homogeneous, light brown in color, and more opaque than the similar colonies of the colon bacillus. At the end of a few days the deep colonies become quite opaque, and may be lobate, like a mulberry, or coarsely granular; sometimes the deep colonies have an opaque central portion surrounded by a transparent marginal zone.

In old gelatin roll tubes these deep colonies form opaque white hemi-



FIG. 153.



FIG. 154.

Fig. 153.—*Bacillus cuniculicida Havaniensis*; colonies in gelatin roll tube, third day at 20° C. $\times 6$. From a photograph. (Sternberg.)

FIG. 154.—*Bacillus cuniculicida Havaniensis*; colonies in gelatin roll tube, end of forty-eight hours. $\times 10$. From a photograph. (Sternberg.)

spheres projecting from the surface of the dried culture medium, and little tufts of acicular crystals are sometimes observed to project from the side of such old colonies.

The superficial colonies are circular or irregular in outline, with transparent margins and an opaque central portion, sometimes corrugated. They are finely granular and iridescent by reflected light, and of a milk-white color; by transmitted light they have a brownish color. Young colonies closely resemble those of the colon bacillus. This bacillus grows well at a temperature of 20° C. (68° F.), but more rapidly and luxuriantly at a higher temperature—30° to 35° C.

It grows well in agar cultures, and especially in *glycerin-agar*, in which it produces some gas and an acid reaction. The growth on the surface of *glycerin-agar* cultures is white, cream-like in consistence, and quite abundant.

It grows well in an agar or gelatin medium made acid by the addition of 0.2 per cent (1:500) of hydrochloric acid.

In cocoanut water it multiplies rapidly, producing a milky opacity of the previously transparent fluid, an acid reaction, and an evolution of carbon dioxide.

On *potato* it produces a thick layer, which may cover the entire surface in three or four days, and which has a dirty-white, cream-white, or pinkish-white color and cream-like consistence. The growth upon potato varies at different times, evidently owing to differences in the potato.

When stained preparations are examined with the full light of the Abbe condenser the ends of some of the rods appear to be cut away, leaving a concave extremity; but by using a small diaphragm to obtain definition it will be seen that the cell wall extends beyond the stained portion of the rod and includes what appears to be a vacuole. There is no reason to believe that this appearance is due to the presence of an end spore, for the supposed vacuole is not refractive, as a spore would be, and my experiments on the thermal death-point of this bacillus indicate that it does not form spores. Cultures are sterilized by exposure for ten minutes to a temperature of 160° F. (71.2° C.).

Pathogenesis.—Very pathogenic for rabbits when injected into the cavity of the abdomen. Injections of a small quantity of a pure culture into the ear vein or subcutaneously generally give a negative result. Injections of from one to five cubic centimetres of a culture in bouillon, blood serum, or *agua coco*, into the cavity of the abdomen, frequently prove fatal to rabbits in a few hours—two to six.

The negative results obtained in injecting cultures beneath the skin or into the ear vein of rabbits show that this bacillus does not induce a fatal septicæmia in these animals, and the fatal result when injections are made into the peritoneal cavity does not appear to be due to an invasion of the blood, but rather to the local effect upon the peritoneum, together with the toxic action of the chemical products resulting from its growth.

It is true that I have always been able to recover the bacillus from the liver, or from blood obtained from one of the cavities of the heart, even in animals which succumb within a few hours to an injection made into the cavity of the abdomen. But the direct examination of the blood shows that the bacilli are present in very small numbers, and leads me to believe that the bacillus does not multiply, to any considerable extent at least, in the circulating fluid.

The spleen is not enlarged, as is the case in anthrax, rabbit septicæmia, and other diseases in which the pathogenic microorganism multiplies abundantly in the blood.

On the other hand, there is evidence of local inflammation in the peritoneal cavity. When death occurs within a few hours the peritoneum is more or less hyperæmic and there is a considerable quantity of straw-colored fluid in the cavity of the abdomen. When the animal lives for twenty hours or more there is a decided peritonitis with a fibrinous exudation upon the surface of the liver and intestine. Usually the liver, in animals which die within twenty-four hours, is full of blood, rather soft, and dark in color. In a single instance I found the liver to be of a light color and loaded with fat.

The rapidly fatal effect in those cases in which I have injected two or more cubic centimetres of a culture into the cavity of the abdomen has led me to suppose that death results from the toxic effects of a ptomaine contained in the culture at the time of injection. The symptoms also give support to this supposition. The animal quickly becomes feeble and indisposed to move, and some time before death lies helpless upon its side, breathing regularly, but is too feeble to get up on its feet when disturbed. Death sometimes occurs in convulsions, but more frequently without—apparently from heart failure.

Pathogenic also for guinea-pigs when injected into the cavity of the abdomen, but death does not occur in so short a time—eighteen to twenty hours. Subcutaneous injections of one-half to one cubic centimetre gave a

negative result in eleven out of thirteen guinea-pigs inoculated—two died within twenty-four hours.

94. BACILLUS LEPORIS LETHALIS.

Obtained by Dr. Paul Gibier (1888) from the contents of the intestine of yellow-fever patients; also by the writer from the same source (1888, 1889) in exceptional cases and in comparatively small numbers. Named and described by present writer.

Morphology.—Bacilli with rounded ends, from 1 to 3 μ in length and about 0.5 μ in breadth. The length may vary in the same culture from a short oval to rods which are two or three times as long as broad, or it may grow out into flexible filaments of considerable length. In recent cultures the bacilli are frequently united in pairs.

Stains readily with the aniline colors usually employed. In cultures which are several days old, or in recent cultures when the stained preparation is washed in alcohol, the ends of the rods are commonly more deeply stained than the central portion—"end staining"; and in old cultures some of the bacilli are very faintly stained.

Biological Characters.—An *aërobie*, *liquefying*, *actively motile* bacillus. Does not form spores.

In *gelatin stick cultures*, at the end of twenty-four hours at a temperature of 20° to 22° C., there is an abundant development along the line of puncture and commencing liquefaction at the surface. Later, the liquefaction is funnel-shaped, and there is an opaque white central core along the line of puncture, with liquefied gelatin around it. Liquefaction progresses most rapidly at the surface, and in the course of three or four days the upper portion of the gelatin for a distance of half an inch or more is completely liquefied, and an opaque white mass, composed of bacilli, rests upon the surface of the unliquefied portion.

In *gelatin roll tubes* the young colonies upon the surface are transparent and resemble somewhat small fragments of broken glass; later liquefaction occurs rapidly. Deep colonies in gelatin roll tubes, or at the bottom of stick cultures, are spherical, translucent, and of a pale straw color.

Upon the surface of nutrient agar it grows rapidly, forming a rather thin, translucent, shining, white layer, which covers the entire surface at the end of two or three days at a temperature of 20° C.

Upon *potato* the growth is rapid and thin, covering the entire surface, and is of a pale-yellow color.

This bacillus grows at a comparatively low temperature, and its vitality is not destroyed by exposure for an hour and a half in a freezing mixture at 15° C. below zero (5° F.).

Decided growth occurred in a stick culture in gelatin exposed in Baltimore during the month of January in an attic room. During the twenty-two days of exposure the highest temperature, taken at 9 A.M. each day, was 11° C., and the lowest 2° C. At a temperature of 16° to 20° C. development in a favorable culture medium is rapid.

There is no evidence that this bacillus forms spores; cultures are sterilized by exposure to a temperature of 60° C. for ten minutes.

Coagulated blood serum is liquefied by this bacillus. It retains its vitality for a long time in old cultures, having grown freely when replanted at the end of a year from a hermetically sealed tube containing a pure culture in blood serum.

Pathogenesis.—This bacillus is very pathogenic for rabbits when injected into the cavity of the abdomen in quantities of one cubic centimetre or more; it is less pathogenic for guinea-pigs, and is not pathogenic for white rats when injected subcutaneously. Gelatin cultures seem to possess more intense pathogenic power than bouillon cultures, and cultures from the blood of an animal recently dead as the result of an inoculation are more potent

than those from my original stock which had not been passed through a susceptible animal.

The mode of death in rabbits is quite characteristic. A couple of hours after receiving in the cavity of the abdomen two or three cubic centimetres of a liquefied gelatin culture the animal becomes quiet and indisposed to eat or move about. Soon after it becomes somnolent, the head drooping forward and after a time resting between the front legs, with the nose on the floor of its cage. It can be roused from this condition, and raises its head in an indifferent and stupid way when pushed or shaken, but soon drops off again into a profound sleep. Frequently the animals die in a sitting position, with their nose resting upon the floor of the cage between the front legs. I have not seen this lethargic condition produced by inoculations with any other microorganism. Convulsions sometimes occur at the moment of death.

The time of death depends upon the potency of the culture and its quantity as compared with the size of the animal. From three to four cubic centimetres of a liquefied gelatin culture usually kill a rabbit in from three to seven hours.

The rapidity with which death occurs when a considerable quantity of a liquefied gelatin culture is injected into the cavity of the abdomen, and the somnolence which precedes death, give rise to the supposition that the lethal effect is due to the presence of a toxic chemical substance rather than to a multiplication of the bacillus in the body of the animal. And this view is supported by the fact that animals frequently recover when the dose administered is comparatively small and especially when it is injected subcutaneously.

In all cases in which death occurs, even when but a few hours have elapsed since the inoculation was made, I have recovered the bacillus in cultures made from blood obtained from the heart or the interior of the liver, and, as stated, these cultures appear to have a greater virulence than those not passed through the rabbit.

In sections of the liver and kidney stained with Löffler's solution of methylene blue the bacilli are seen, and are often in rather long-jointed filaments.

95. BACILLUS PYOCYANUS.

Synonyms.—*Bacillus* of green pus ; *Microbe du pus bleu* ; *Bacillen des grünblauen Eiters* ; *Bacterium aeruginosum*.

Obtained by Gessard (1882) from pus having a green or blue color, and since carefully studied by Gessard, Charrin, and others.



FIG. 155. — *Bacillus pyocyaneus*.
(Flügge.) × 700.

This bacillus appears to be a widely distributed saprophyte, which is found occasionally in the purulent discharges from open wounds, and sometimes in perspiration and serous wound secretions (Gessard). The writer obtained it, in one instance, in cultures from the liver of a yellow-fever cadaver (Havana, 1888).

Morphology.—A slender bacillus with rounded ends, somewhat thicker than the *Bacillus murisepticus* and of about the same length (Flügge); frequently united in pairs, or chains of four to six elements; occasionally grows out into filaments.

Biological Characters.—An *aërobic*, *liquefying*, *motile* bacillus. Grows readily in various culture media at the room tempera-

ture—more rapidly in the incubating oven. Is a facultative anaërobic (Fränkel). Does not form spores. The thermal death-point, as determined by the writer, is 56° C., the time of exposure being ten minutes. In *gelatin plate cultures* colonies are quickly developed, which give to the medium a fluorescent green color; at the end of two or three days liquefaction commences around each colony, and usually the gelatin is completely liquefied by the fifth day. Before liquefaction commences the deep colonies, under a low power, appear as spherical, granular masses, with a serrated margin, and have a yellowish-green color; the superficial colonies are quite thin and finely granular; at the centre, where they are thickest, they have a greenish color, which fades out towards the periphery.

In *stick cultures* in nutrient gelatin development is most abundant near the surface, and causes at first liquefaction in the form of a shallow funnel; later the liquefied gelatin is separated from that which is not liquefied by a horizontal plane, and a viscid, yellowish-white mass, composed of bacilli, accumulates upon this surface, which gradually has a lower level as liquefaction progresses; the transparent, liquefied gelatin above is covered with a delicate, yellowish-green film, and the entire medium has a fluorescent green color. Upon nutrient *agar* a rather abundant, moist, greenish-white layer is developed, and the medium acquires a bright green-color, which subsequently changes to olive green. Upon *potato* a viscid or rather dry, yellowish-green or brown layer is formed, and the potato beneath and immediately around the growth has a green color when freely exposed to the air or to the vapors of ammonia. In *milk* the casein is first precipitated and then gradually dissolved, while at the same time ammonia is developed. The *green pigment* is formed only in the presence of oxygen; it is soluble in chloroform and may be obtained from a pure solution in long, blue needles; acids change the blue color to red, and reducing substances to yellow. According to Ledderhose, it is an aromatic compound resembling anthracene, and is not toxic. According to Gessard's latest researches (1890), two different pigments are produced by this bacillus, one of a fluorescent green and the other—pyocyanin—of a blue color. Cultivated in egg albumin the fluorescent green pigment, which changes to brown with time, is alone produced. In bouillon and in media containing peptone or gelatin both pigments are formed, and the pyocyanin may be obtained separately by dissolving it in chloroform. In an alkaline solution of peptone (two per cent) to which five per cent of glycerin has been added the blue pigment alone is formed.

Pathogenesis.—The experiments of Ledderhose, Bouchard, and others show that this bacillus is pathogenic for guinea-pigs and rabbits. Subcutaneous or intraperitoneal injections of recent cultures—

one cubic centimetre or more of a culture in bouillon—usually cause the death of the animal in from twelve to thirty-six hours. An extensive inflammatory œdema and purulent infiltration of the tissues result from subcutaneous inoculations, and a sero-fibrinous or purulent peritonitis is induced by the introduction of the bacillus into the peritoneal cavity. The bacillus is found in the serous or purulent fluid in the subcutaneous tissues or abdominal cavity, and also in the blood and various organs, from which it can be recovered in pure cultures, although not present in great numbers, as is the case in the various forms of septicæmia heretofore described. When smaller amounts are injected subcutaneously the animal usually recovers after the formation of a local abscess, and it is subsequently immune when inoculated with doses which would be fatal to an unprotected animal. Immunity may also be secured by the injection of a considerable quantity of a sterilized culture. Bouchard has also produced immunity in rabbits by injecting into them the filtered urine of other rabbits which had been inoculated with a virulent culture of the bacillus. It has been shown by Bouchard, and by Charrin and Guignard, that in rabbits which have been inoculated with a culture of the anthrax bacillus a fatal result may be prevented by soon after inoculating the same animals with a pure culture of the *Bacillus pyocyaneus*. The experiments of Woodhead and Wood indicate that the antidotal effect is due to chemical products of the growth of the bacillus, and not to an antagonism of the living bacterial cells. They were able to obtain similar results by the injection of sterilized cultures of the *Bacillus pyocyaneus*, made soon after infection with the anthrax bacillus.

96. BACILLUS OF FIOCCA.

Found by Fiocca in the saliva of cats and dogs.

Closely resembles the influenza bacillus of Pfeiffer and of Canon.

Morphology.—Resembles the bacillus of rabbit septicæmia, but is only half as large—from 0.2 to 0.33 μ in breadth. The length is but little greater than the breadth. Usually seen in pairs, closely resembling diplococci. When cultivated on potato it appears to be a micrococcus, but in the blood of infected animals and in bouillon cultures it is seen to be a short bacillus.

Stains with difficulty with the usual aniline colors, but is readily stained by Ehrlich's method or with Ziehl's solution.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows best at 37° C. and does not develop at temperatures below 15° C. In agar plates, at 37° C., small, punctiform colonies are developed at the end of twenty-four hours; these do not increase in size later; under the microscope the deep colonies are seen to be spherical, granular, and dark yellow in color; the superficial colonies are more or less round, with irregular outlines, transparent, slightly granular, and often have a shining nucleus at the centre. Upon gelatin plates the colonies have a similar appearance, but are not visible in less than four or five days. In streak cultures upon the surface of agar small, punctiform colonies are seen along the track of the needle at the end of twenty-four hours, resembling fine dewdrops; the following day

these colonies are a little larger and less transparent; they remain distinct, especially along the margins of the line of growth. Upon *potato* a very thin, transparent layer is developed, which does not change the appearance of the surface of the potato, but slightly increases its resistance to the platinum needle. In *bouillon* small flocculi, suspended in the clear liquid, are developed within twenty-four hours; these subsequently sink to the bottom.

Milk is not coagulated by this bacillus, and no gas is produced in media containing sugar.

Pathogenesis.—Pathogenic for rabbits, guinea-pigs, young rats, and mice, in which animals it produces general infection, and death—in rabbits—at the end of twenty-four hours. The bacillus is found in the blood in great numbers.

97. *PROTEUS VULGARIS*.

Obtained by Hauser (1885) from putrefying animal substances, and since shown to be one of the most common and widely distributed putrefactive bacteria. This and the other species of *Proteus*



FIG. 156.—*Proteus vulgaris*; "swarming islands" from a gelatin culture. $\times 265$. (Hauser.)

described by the same bacteriologist (*Proteus mirabilis*, *Proteus Zenkeri*) have no doubt frequently been encountered by previous observers, and are among the species formerly included under the name "*Bacterium termo*," which was applied to any minute motile bacilli found in putrefying infusions.

Morphology.—Bacilli with rounded ends, about 0.6μ broad, and varying greatly in length, being sometimes short oval, and at others from 1.25 to 3.75μ in length; also grow out into flexible filaments, which may be more or less wavy or spiral in form. The short rods are commonly seen in pairs; they have terminal flagella; involution forms are frequently seen, the most common being spherical bodies about 1.6μ in diameter. In old cultures in bouillon, or in cultures made in meat infusion in the incubating oven, the short oval forms greatly predominate, but in recent cultures in nutrient gelatin fla-

ments of considerable length are encountered in association with shorter rods.

Stains readily with fuchsin or gentian violet—not so well with the brown aniline colors ; does not stain by Gram's method (Cheyne).

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *motile* bacillus. Grows rapidly in the usual culture media at the room temperature.

The growth upon *gelatin plates* (five per cent of gelatin) at the room temperature is very characteristic ; at the end of six or eight hours small depressions in the gelatin are observed, which contain liquefied gelatin and grayish-white masses of bacilli. Under a low power these depressions are seen to be surrounded by a marginal zone consisting of two or three layers, outside of which is a zone of a single layer, from which amoeba-like processes extend upon the surface of the gelatin. These processes are constantly undergoing changes in their form and position, and may become separated from the mother colony, or remain temporarily attached to it by a narrow thread consisting of bacilli ; after a time the entire surface of the gelatin is covered with wandering, amoeba-like colonies ; these rapidly cause liquefaction, which by the end of twenty-four to forty-eight hours has reached a depth of one millimetre or more over the entire surface. The deep colonies also are surrounded by processes projecting into the gelatin, which may be observed to suddenly advance and again to be retracted towards the central zoöglösa-like mass. Liquefaction around the colony rapidly progresses, and actively motile rods and spiral filaments may be seen about the periphery of this liquefied gelatin, while about it is a radiating crown of irregular processes, some of which may be screw-like or corkscrew-formed. In ten-per-cent gelatin the migration of surface colonies, above described, is not observed. In *gelatin stick cultures* liquefaction occurs along the entire line of puncture, and soon the contents of the tube are completely liquefied ; near the surface of the liquefied gelatin the growing bacilli form a grayish-white cloudiness, and at the bottom of the tube an abundant flocculent deposit is formed. Upon the surface of *nutrient agar* a rapidly extending, moist, thin, grayish-white layer is formed. Upon *potato* this bacillus produces a dirty-white, moist layer. The cultures in media containing albumin or gelatin have a putrefactive odor and acquire a strongly alkaline reaction. A temperature of 20° to 24° C. is most favorable for the growth of this bacillus. It is a facultative anaërobic and grows in an atmosphere of hydrogen or of carbon dioxide, although not so rapidly as in the presence of oxygen. The movements are often extremely active and difficult to follow under the microscope ; again they may be quite deliberate, or the bacilli may remain motionless

for a time and again dart off in active motion. The long terminal flagella may sometimes be discerned by means of a good objective and careful manipulation of the light.

Pathogenesis.—Pathogenic for rabbits and for guinea-pigs when injected into the circulation, into the cavity of the abdomen, or subcutaneously in considerable quantity. Cultures in nutrient gelatin are said by Cheyne to be more pathogenic (toxic) than those in bouillon. When injected into the muscles of rabbits a much smaller dose produces a fatal result than when injected subcutaneously. In Cheyne's experiments, made in London (1886), one-tenth cubic centimetre of a liquefied gelatin culture, injected into the dorsal muscles, was invariably fatal in from twenty-four to thirty-six hours; a dose of one-twentieth cubic centimetre, injected in the same way, usually caused death; while one-fortieth cubic centimetre gave rise to an extensive local abscess, and the animals died at the end of six or eight weeks. Doses of less than one-five-hundredth cubic centimetre produced no effect. Cheyne estimates that one cubic centimetre of a culture in nutrient gelatin contains 4,500,000,000 bacilli, and, consequently, that a smaller number than 9,000,000 produced no effect when injected into the muscular tissue of rabbits. Injections into the subcutaneous connective tissues of a dose twice as large as that which invariably proved fatal when injected into the muscles usually caused an extensive abscess, but did not kill the animal; and, after recovery from the effects of such an injection, the rabbit was found to be immune against a similar dose injected into the muscles. Foà and Bonome have succeeded in producing immunity against the effects of virulent cultures of this bacillus by inoculating rabbits with filtered cultures, and also by injecting beneath the skin of these animals a solution of neurin, which they believe to be the principal toxic product present in the cultures.

Proteus Vulgaris in Cholera Infantum.—The extended researches of Booker have led him to the conclusion that this bacillus plays an important part in the production of the morbid symptoms which characterize cholera infantum. *Proteus vulgaris* was found in the alvine discharges in a considerable proportion of the cases examined, but was not found in the fæces of healthy infants. "The prominent symptoms in the cases of cholera infantum in which the proteus bacteria were found were drowsiness, stupor, emaciation and great reduction in flesh, more or less collapse, frequent vomiting and purging, with watery and generally offensive stools."

Another bacillus found by Booker in a considerable number of his cases he has designated by the letter A (No. 103).

98. PROTEUS OF KARLINSKI.

Synonym.—*Bacillus murisepticus pleomorphus* (Karlinski). Probably identical with *Proteus vulgaris* of Hauser.

Obtained by Karlinski (1889) from a fibro-purulent uterine discharge, and from abscesses in the uterus and its appendages in a puerperal woman.

Morphology.—Resembles *Proteus vulgaris* of Hauser in its morphology, and presents various forms under different circumstances relating to the culture medium, the temperature, age of culture, etc.—sometimes as spherical or short oval cells, at others as longer or shorter rods or spiral filaments; usually as bacilli with round ends two and a half times as long as thick, often united in pairs.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation not observed. Grows rapidly in the usual culture media at the room temperature. In *gelatin plate cultures*, at the end of ten hours, small colonies are developed which have well-defined outlines, are oval or whetstone-shaped, of a light-brown color by transmitted light and white by reflected light, with a somewhat darker margin and a smooth surface, sometimes marked by shallow clefts; at the end of twenty hours the colonies commence to have irregular margins, and the surface of the gelatin above them is marked by concentric rings. At the end of thirty hours the colonies have formed a bulb-shaped liquefaction of the gelatin, and delicate, ray-like offshoots are seen around the margin. At the end of two days the bulbous cavities are about one and a half millimetres in diameter and contain a cloudy, grayish-white liquid; they are surrounded by a moist-looking, gray, irregular marginal zone. In *gelatin stick cultures*, at the end of twenty-four hours, a funnel-shaped liquefaction of the gelatin occurs near the surface, and a grayish-white, cloudy mass is developed along the line of puncture; at the end of forty-eight hours a sac-like pouch of liquefied gelatin has formed, and in the course of four or five days the gelatin is entirely liquefied. Upon *agar plates* the colonies are at first oval in form and white by reflected light, or pale brown by transmitted light; at the end of thirty hours the surface becomes wrinkled or folded and is surrounded by radiating, delicately twisted offshoots. Upon the surface of agar a white layer is developed. Upon *potato* a whitish-gray, soft, homogeneous layer, which after standing a long time has a darker color. Upon *blood serum* a thin, grayish-white layer is formed and the serum is rapidly liquefied. Gelatin cultures acquire a strongly alkaline reaction and give off a disagreeable odor resembling that of butyric acid.

Pathogenesis.—White mice inoculated at the root of the tail die in from twenty-two to twenty-four hours; the spleen is greatly enlarged; the bacilli are found in blood from the various organs—less numerous in blood from the heart. Field mice and house mice are less susceptible. Subcutaneous injections in rabbits may give rise to local inflammation and also to general infection. In white rats and guinea-pigs a local abscess may result from a subcutaneous inoculation.

99. PROTEUS MIRABILIS.

Obtained by Hauser (1885) from putrefying animal substances.

Morphology.—Bacilli resembling very closely the preceding species (*Proteus vulgaris*), but presenting more numerous involution forms, which may be spherical, pear-shaped, or spermatozoa-like, etc. The bacilli are about 0.6μ in diameter and vary greatly in length, being sometimes nearly spherical, or forming rods of 2 to 3.75μ in length, or long filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation has not been observed. Grows in the usual culture media at the room temperature. Does not liquefy gelatin as

rapidly as *Proteus vulgaris*. Upon *gelatin plates*, at the end of twelve hours, superficial colonies of two to three millimetres in diameter are formed; under a low power these appear finely granular and brownish in color, and have an irregular outline; outgrowths from the margin extend in various directions and form new colonies, which may be attached for a time by a long and slender thread consisting of bacilli. The movement of these new colonies is not as pronounced as in the case of the preceding species, and

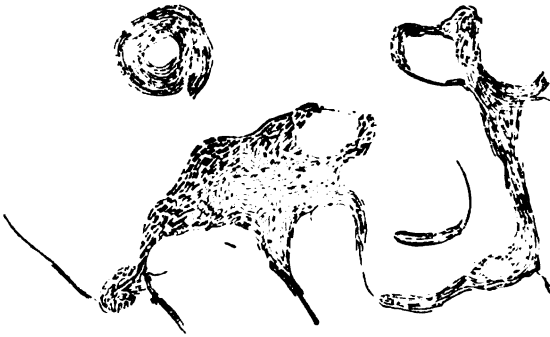


FIG. 157.—"Swarming islands" of *Proteus mirabilis*, from a gelatin culture. $\times 285$. (Hauser.)

they are characterized by the presence of numerous distorted bacilli—involution forms. The deep colonies form spiral zoöglœa masses.

In gelatin stick cultures the whole surface is first covered with threads and islands of bacilli, which after a time form an anastomosing network, and finally a confluent layer which at the end of forty-eight hours is rather thick,



FIG. 157.—Spiral zoöglœa from a culture of *Proteus mirabilis*. $\times 95$. (Hauser.)

with a moist, shining surface and grayish color, and appears to be perforated with numerous small, sieve-like openings. These thinner and transparent places disappear after a time, and at the end of two or three days liquefaction of the gelatin commences; complete liquefaction does not occur until the fifth or sixth day, or even later. Along the line of puncture finely granular colonies are first formed, from which long threads are given off, which form after a short time a tolerably broad zone of threads and spiral zoöglœa masses.

Pathogenesis.—In Hauser's experiments filtered cultures (two to six cubic centimetres), injected into the circulation or into the cavity of the abdomen in rabbits, caused fatal toxæmia.

100. PROTEUS ZENKERI.

Obtained by Hauser (1885) from putrefying animal substances.

Morphology.—Bacilli which vary greatly in length—average about 1.65μ , and about 0.4μ broad.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon the surface of nutrient gelatin a laminated mass forms about the point of puncture, from the periphery of which offshoots are given off, at the extremities of which colonies are formed, as in the case of *Proteus mirabilis*. Gradually a rather thick, grayish-white, opaque layer is formed, which covers the entire surface of the gelatin and is easily detached from it. This species is distinguished from the two preceding by the fact that it does not liquefy gelatin or blood serum and does not give off a decided putrefactive odor when cultivated in these media.

Pathogenesis.—Considerable quantities injected into small animals give rise to local abscesses and to symptoms of toxæmia.

101. PROTEUS SEPTICUS.

Obtained by Babes (1889) from the mucous membrane of the intestine and the various organs of a boy who died of septicæmia.

Morphology.—Bacilli about 0.4μ broad and varying greatly in length; slightly curved rods or flexible filaments, often associated in loose chains.

Stains by the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic*, *liquefying*, *motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. In *gelatin plates* centres of liquefaction are quickly formed and rapidly extend. The spherical, liquefied places have at first a wavy or dentate outline, and are surrounded by a branching, transparent, granular margin which rapidly extends in advance of the liquefaction. In *stick cultures* in nutrient gelatin liquefaction of the entire contents of the tube may take place within twenty-four hours, or a broad, liquefied sac is formed along the line of puncture. Gelatin cultures give off a very disagreeable odor. Upon the surface of *nutrient agar*, at 37°C. , a peculiar, thick network extends over the surface in the course of a few hours. Upon *potato* an elevated, brownish-white, shining layer is formed. *Blood serum* is liquefied by this bacillus.

Pathogenesis.—Pathogenic for mice, less so for rabbits. In mice death occurs in from one to three days after the subcutaneous injection of a small quantity of a pure culture; the bacilli are present in the blood in small numbers.

102. PROTEUS LETHALIS.

Synonym.—*Proteus bei Lungengangrän des Menschen* (Babes).

Obtained by Babes (1889) from the spleen and gangrenous portions of the lung of a man who died of septicæmia.

Morphology.—Short rods with round ends, from 0.8 to 1.5μ thick; often swollen in the middle, like a lemon or a flask; forms short, flexible filaments which also present similar swellings.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Not observed to form spores. Grows in the usual culture media at the room temperature. In *gelatin plates* forms hemispherical, elevated, whitish, translucent colonies, which later send out

coarse branches which ramify over the surface of the gelatin. A similar growth is observed upon the surface of gelatin stick cultures, and an abundant development takes place along the line of puncture. Upon *nutrient agar* a thick, opaque, slightly yellowish layer is formed. Upon *potato* a moist, shining, brownish layer is developed, and the potato acquires a brownish color. Upon *blood serum* the growth is less abundant than on agar; the blood serum is not liquefied. This bacillus grows rapidly at the room temperature; it is destroyed by a temperature of 80° C., and presumably does not form spores.

Pathogenesis.—Recent cultures are very pathogenic for mice and for rabbits, less so for guinea-pigs. The subcutaneous injection of a small quantity of a pure culture kills susceptible animals in two or three days. More or less cedema is found at the point of inoculation. Injections into the rectum of rabbits gave rise to hæmorrhagic enteritis, peritonitis, and death at the end of four days.

103. BACILLUS A OF BOOKER.

Obtained by Booker (1889) from the alvine discharges of children suffering from cholera infantum.

Morphology.—Bacilli with round ends, varying greatly in length, usually three to four μ long and 0.7 μ broad (in recent agar cultures). In older cultures the bacilli are shorter and smaller.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Grows at the room temperature in the usual culture media. In *gelatin plates* colonies are visible at the end of twenty-four hours; under the microscope these are nearly colorless, and liquefaction soon occurs around them. In *gelatin stick cultures* complete liquefaction occurs in three or four days. Upon *agar* a colorless layer covering the entire surface is developed in three or four days, and an abundant development occurs along the line of puncture. Agar colonies have a bluish look, and are surrounded by an indistinct halo which shades off gradually into the surrounding agar; under a low power the colonies are light-brown and the borders indistinct; the surface has a delicate, wavy appearance. Upon *potato* the growth is luxuriant and of a dirty-brown color. Blood serum is liquefied by this bacillus.

Milk is coagulated into a gelatinous mass having an alkaline reaction; later the coagulum is dissolved.

Pathogenesis.—Mice and guinea-pigs fed with cultures in milk die in from one to eight days.

104. BACILLUS ENDOCARDITIDIS GRISEUS.

Obtained by Weichselbaum (1888) from the affected valves in a case of endocarditis recurrens ulcerosa.

Morphology.—Short rods with rounded or somewhat pointed ends, about two to three times as long as broad—of about the same dimensions as the bacillus of typhoid fever.

Stains with the usual aniline colors and also by Gram's method; the longer rods from old cultures are irregularly stained.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Refractive bodies may be seen in some of the rods, which resemble spores and are stained by the method of Ernst, but they do not show the resistance of known spores to physical and chemical agents. Grows well in the usual culture media at the room temperature. Upon *gelatin plates* colonies are formed which resemble those of Friedländer's bacillus, but which gradually acquire a gray or grayish-white color. The prominent, convex, superficial colonies under a low power are finely granular and grayish-brown in color; the deep colonies are yellowish-brown in color, have slightly notched margins, and the surface is covered with minute projections. In stick cultures

a rather thin, circular layer forms about the point of puncture; this has the appearance of stearin; later it becomes grayish-white and the margins are marked by radiating lines. Upon the surface of *nutrient agar* a similar growth occurs which has a pale-brown or reddish-gray color. Upon *potato* in the incubating oven an abundant development occurs, forming a dry-looking layer of a grayish-brown color and having irregularly notched margins. Upon *blood serum* an abundant, grayish-white growth of cream-like consistence forms along the *impfstrich*; later this has a reddish gray color. This bacillus grows to the bottom of the line of puncture in stick cultures, and is no doubt a facultative anaërobic.

Pathogenesis.—Pathogenic for white mice and for guinea-pigs.

105. BACILLUS ENDOCARDITIDIS CAPSULATUS.

Obtained by Weichselbaum (1888) from thrombi and embolic infarctions in the spleen and kidneys of a man who died from endocarditis with formation of thrombi.

Morphology.—Resembles Friedländer's bacillus, and is frequently surrounded by a capsule, which may be stained; also forms long, curved filaments, in the protoplasm of which vacuoles may be observed in stained preparations.

Stains with the usual aniline colors, but not by Gram's method; by staining with fuchsin and carefully decolorizing with diluted alcohol the presence of a capsule may be demonstrated.

Biological Characters.—An *aërobic, non-liquefying* bacillus. Grows in the usual culture media at the room temperature.

In *gelatin stick cultures* development occurs along the line of puncture, and on the surface as a rather thin, white, dry layer which resembles stearin. In *agar plates* the superficial colonies are thin, about two millimetres in diameter and gray in color; under a low power the margins are transparent and colorless, and the centre resembles the deep colonies; these are very small and grayish-white in color; under a low power the surface is seen to be covered with tooth-like, projecting masses, the margin is dentate and has a pale-yellow color, while the centre is yellowish-brown.

Pathogenesis.—Rabbits are killed by the injection of a considerable quantity of a pure culture into the cavity of the abdomen or subcutaneously.

106. BACILLUS OF LESAGE.

Obtained by Lesage (1887) from the green-colored discharges of infants suffering from "green diarrhoea," and supposed to be the cause of this complaint (?). According to Baumgarten, this bacillus is probably identical with a well-known pigment-producing saprophyte—the *Bacillus fluorescens non-liquefaciens*.

Morphology.—Small bacilli with round ends, about 2.4μ long and 0.75 to 1μ broad; in old cultures may grow out into long filaments.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic, non-liquefying* (slight liquefaction in old cultures), *motile* bacillus. Forms spores. Grows slowly at the room temperature in the usual culture media, more rapidly at 25° to 35° C. Upon *gelatin plates* superficial colonies are formed which have irregularly dentate, leaf-like margins and a smooth surface; they produce a greenish color in the gelatin. In *gelatin stick cultures* a thin, smooth, transparent, greenish layer forms upon the surface, and in the course of four or five days the gelatin has acquired throughout a bright-green color. Upon *potato* a dark-green layer is formed. The cultures have the odor of old urine.

Pathogenesis.—The injection of a considerable quantity of a pure culture into the ear vein of a rabbit is said to have produced green diarrhoea, and the same result was obtained by mixing cultures with the food of these animals. These results have not yet been confirmed by other investigators.

107. BACILLUS OF DEMME.

Obtained by Demme (1888) from the fluid contents of the tumors and pustules of erythema nodosum, and also from the blood of the affected individual.

Morphology.—Bacilli with round ends, from 2.2 to 2.5 μ long and 0.5 to 0.7 μ broad; usually collected in smaller or larger groups.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic* (facultative anaërobic?) bacillus, which does not grow in nutrient gelatin at the room temperature. Grows in nutrient agar at 35° to 37° C. Forms spores. In *agar plates*, at 35° to 37° C., smooth, spherical, shining white colonies are formed in from forty-eight to sixty hours, which at the end of six or seven days may have the size of a small coin—five centimes; these are marked by lines radiating from the centre, which are slightly elevated above the surface of the colony and have a silvery lustre by obliquely reflected light; the margins of the colony are fringe-like, and after ten or twelve days conical offshoots are given off from this thready margin. In *agar stick cultures* growth occurs along the line of puncture in the form of a thorny column which has a paraffin-like lustre.

Pathogenesis.—According to Demme, when injected subcutaneously into guinea-pigs, or by rubbing pure cultures into the scarified skin, an eruption occurs which resembles that of erythema nodosum and is followed by a gangrenous condition of the skin. Rabbits, dogs, and goats proved to be refractory.

108. BACILLUS OEDEMATIS AËROBICUS.

Synonym.—A new bacillus of malignant oedema (Klein).

Obtained from garden earth by inoculation in guinea-pigs.

Morphology.—Bacilli from 0.8 to 2.4 μ in length and 0.7 μ thick; grow out into long filaments.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Does not form spores. Grows at the room temperature in the usual culture media. Upon *gelatin plates*, at the end of twenty-four hours, small, gray, punctiform colonies are developed; at the end of forty-eight hours the superficial colonies are seen as flat, grayish, transparent plaques, the margins of which are thin and irregularly notched; these attain a diameter of several millimetres in the course of a few days. The deep colonies do not exceed the diameter of a pin's head; they remain spherical, and by transmitted light have a brownish color. In *gelatin stick cultures* a white line of growth is developed along the track of the inoculating needle, and at the bottom of this isolated, punctiform colonies are seen; upon the surface a flat, thin, transparent, grayish layer with a dentate margin is developed. Upon the *surface of agar* a smeary, grayish-white stripe is developed along the *impfstrich*. Alkaline *bouillon*, at the end of twenty-four hours at 37° C., is densely clouded, and later contains numerous flocculi, but no pellicle upon the surface; at the end of twenty-four hours the reaction becomes strongly alkaline. Upon *potato* a viscid, yellowish stripe is developed along the line of inoculation. In deep cultures in nutrient gelatin gas bubbles are developed in from twenty-four to forty-eight hours; these are attached to the isolated colonies.

Pathogenic for guinea-pigs, rabbits, and white mice. The animals die within twenty-four hours—when very small quantities are injected subcutaneously into guinea-pigs they may live for two or three days and sometimes recover. The lethal dose of a bouillon culture is from one-fourth to one-half cubic centimetre, but one drop of the cedematous fluid from the subcutaneous connective tissue of an inoculated animal is infallibly fatal. In guinea-pigs an extensive inflammatory oedema is produced by subcutaneous inoculations; the spleen is but slightly enlarged. In rabbits but slight oedema

and a small spleen. In mice no oedema and a slightly enlarged spleen. The bacilli are found in the blood of the heart in small numbers, and are somewhat more numerous in the spleen, especially in mice.

109. BACILLUS OF LETZERICH.

Obtained by Letzerich (1887) from the urine of children suffering from "nephritis interstitialis primaria." Etiological relation not satisfactorily demonstrated.

Morphology.—Bacilli with round ends, straight or slightly curved, often forming filaments.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, liquefying* bacillus. Forms spores. Grows rapidly in nutrient gelatin at a comparatively low temperature—best at 14° C. Upon gelatin plates, at 14° C., complete liquefaction has occurred in from thirty-six to forty-eight hours, and a thin, white film covers the surface of the liquefied gelatin; the same in gelatin stick cultures.

Pathogenesis.—Rabbits injected in the cavity of the abdomen are said to die in about fourteen days. The autopsy shows an extensive abscess, enlargement and congestion of the kidneys, enlarged spleen, etc. The bacilli are found in great numbers in all of the organs.

110. BACILLUS OF SCHIMMELBUSCH.

Obtained by Schimmelbusch (1889) from the necrotic tissues at the boundary line of the still living tissues in cancrum oris, or noma. Etiological relation not proved.

Morphology.—Small bacilli with round ends; often united in pairs; may grow out into long filaments.

Stains best with an aqueous solution of gentian violet; does not stain by Gram's method.

Biological Characters.—An *aërobic, non-liquefying* bacillus. Grows in the usual culture media at the room temperature—better in the incubating oven at 30° to 37° C. Upon *gelatin plates* forms below the surface spherical, finely granular, grayish-white colonies, which come to the surface and form elevated masses with slightly dentate margins and an irregularly cleft surface. In *gelatin stick cultures* the growth along the line of inoculation is coarsely granular; upon the surface a broad, flat layer. Upon the surface of *agar*, in twenty-four hours at 37° C., a grayish-white layer along the line of inoculation, which is smooth and about three millimetres in breadth. Upon *potato*, at the end of two weeks, a broad, moist, grayish-white layer from two to three millimetres wide. Upon coagulated ascitic fluid, at the end of twenty-four hours, a thin layer along the impfstrich, from which lateral offshoots are given off.

Pathogenesis.—Cultures injected subcutaneously into rabbits produced local abscesses only; not pathogenic for mice or pigeons.

111. BACILLUS FÆTIDUS OZÆNÆ.

Obtained by Hajek (1888) from the nasal secretions of patients with ozæna. Etiological relation not proved.

Morphology.—Short bacilli, but little longer than broad; usually in pairs, or in chains of six to ten elements.

Stains with Löffler's solution of methylene blue or solutions of aniline colors in aniline water—not so well in aqueous solutions; does not stain by Gram's method.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the colonies, at the end of thirty-six hours, are scarcely visible, with well-defined but

somewhat irregular outlines; later liquefaction commences and crater-like depressions in the gelatin are formed, in which a gas bubble is seen; complete liquefaction occurs in the course of a few days. In *gelatin stick cultures* liquefaction occurs all along the line of inoculation, and is complete at the end of from eight to fourteen days. Upon *agar plates* the colonies are granular in the centre, and the margins, under a low power, are seen to be fringed. Upon the surface of agar a moist, slimy layer is formed along the impfstrich. Upon *potato*, at the end of twenty-four hours, a yellowish-brown layer is formed. Upon *blood serum* development is rapid in the form of a whitish layer, which extends over the whole surface. The cultures, and especially those kept in the incubating oven, give off a disagreeable putrefactive odor, which is most intense in the blood-serum cultures.

Pathogenesis.—Pathogenic for mice. When injected subcutaneously into rabbits it gives rise to intense local inflammation and progressive gangrene of the connective tissue.

112. BACILLUS OF LUMNITZER.

Obtained by Lumnitzer (1888) from the bronchial secretions of persons suffering from "putrid bronchitis." Etiological relation not demonstrated.

Morphology.—Bacilli with round ends, from 1.5 to 2 μ long, somewhat curved.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, motile* bacillus. Does not grow in nutrient gelatin at the room temperature. Grows slowly upon agar and more rapidly upon blood serum at 36° to 38° C. Forms spores. Upon *agar plates*, at 37° C., small, grayish-white colonies are formed in two or three days; upon the surface these form hemispherical masses which slowly increase in size. At the end of six or seven days the cultures give off a disagreeable odor, quite like that given off by the sputum of the cases of putrid bronchitis from which the bacillus was obtained. Upon the surface of *blood serum* the growth is rapid and forms grayish-white, shining colonies, of about one millimetre in diameter, which become confluent at the end of about four days, and cover the entire surface in eight or nine days.

Pathogenesis.—Causes a purulent inflammation when injected into the lungs of rabbits, which involves the bronchial tubes, the blood vessels, and the pulmonary alveoli; when injected subcutaneously produces inflammation and necrosis of the tissues.

113. BACILLUS OF TOMMASOLI.

Obtained by Tommasoli (1889) from the hairs of the head of a patient suffering from a form of sycosis supposed to be due to the presence of this parasite (?).

Morphology.—Short, straight bacilli, with round ends, from 1 to 1.8 μ long and from 0.25 to 0.3 μ broad; often united in chains containing four to six elements.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows slowly at the room temperature in the usual culture media. Upon *gelatin plates*, at the end of four days, the deep colonies are seen as small, white points, the superficial colonies as smooth discs of a grayish color. At the end of a month the deep colonies may be as large as a mustard seed; the superficial are thin, shining, and slimy, and have a diameter of one to two millimetres. In *gelatin stick cultures* a convex, shining, white mass is developed at the point of inoculation, and along the line of puncture in the course of five or six days a white line of growth is seen which consists of closely crowded, small colonies. Upon *agar* the development is very slow, and forms at first thin, slimy, grayish-white patches which are distributed along the impfstrich; later these become con-

fluent and form shining, wavy stripes. Upon *potato* the development is more rapid and forms elevated, sharply defined colonies, of granular appearance and of a chamois-yellowish-white color; later these become confluent; the potato acquires a dark-gray color and the culture gives off an intensely disagreeable odor.

Pathogenesis.—Pure cultures rubbed into the skin of man produce, at the end of twenty-four hours, intense itching, redness, and a vesicular eruption about the hairs; at the end of three days small pustules are formed, from which pure cultures may be recovered (Tommasoli). Subcutaneous injection into a rabbit produced no other result than the formation of a small abscess.

114. BACILLUS OF SCHOÛ.

Obtained by Schou (1885) in rabbits suffering from *vagus pneumonia* resulting from section of the vagi; found also in the buccal secretions of a healthy rabbit—one out of twenty-five examined.

Morphology.—Described as elliptical cocci, or diplococci, or as short, thick bacilli.

Stains with the aniline colors usually employed, but not by Gram's method.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Grows in the usual culture media at the room temperature. In *gelatin plates* forms spherical, opaque, granular colonies having a slightly rough surface. At the end of twenty-four hours, under the microscope, active movements are observed in these colonies, which are surrounded by a zone of diverging rays. In gelatin stick cultures liquefaction quickly occurs, and a copious white deposit, consisting of bacilli, is seen at the bottom of the tube.

Pathogenesis.—Pure cultures injected into the trachea, the pleural cavity, or the lungs are said to have produced fatal pneumonia in rabbits; a similar result was obtained from inhalation experiments.

115. BACILLUS NECROPHORUS.

Obtained by Löffler (1884) from rabbits which had been inoculated in the anterior chamber of the eye with small fragments of a broad condyloma.

Morphology.—Bacilli of various lengths, often forming long, slender, wavy filaments.

Biological Characters.—Does not grow in the ordinary culture media, but may be cultivated in neutral rabbit bouillon; a less favorable medium is blood serum from the horse. When small fragments of the organs of an infected animal are placed in rabbit bouillon they become enveloped, in the course of three or four days, in a cotton-like mass of filaments; later white flocculi are distributed through the medium, which consist of similar filaments loosely interlaced. The filaments may present swellings here and there, which are supposed to represent involution forms.

Pathogenesis.—Rabbits inoculated in the ear or in the anterior chamber of the eye with the flocculi from a bouillon culture, or with a small fragment of one of the organs of an infected animal, usually die at the end of eight days. At the autopsy a necrotic, cheesy process is found at the point of inoculation, and purulent foci, surrounded by inflamed or necrotic areas, in the lungs; also purulent collections in the myocardium; these were the principal pathological changes, but sometimes nodules were found in the abdominal viscera. The slender bacilli described were found in all of these localized centres of infection. Pathogenic also for white mice, which usually died in six days after being inoculated subcutaneously.

116. BACILLUS COPROGENES FŒTIDUS.

Synonym.—Darmbacillus of Schottelius.

Obtained by Schottelius (1885) from the intestinal contents of pigs which had died of Schweinerothlauf (rouget).

Morphology.—Resembles *Bacillus subtilis*, but is shorter, with rounded ends.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Forms spores in presence of oxygen in the course of three or four days at the room temperature; these are oval in form and are arranged in rows; when they germinate this occurs in a direction perpendicular to their long axis and to that of the filament in which they developed; as a result of this the newly formed rods lie parallel to each other. In *gelatin stick cultures* the growth upon the surface consists of a thin, transparent, grayish layer; along the line of puncture crowded, pale-yellow colonies are developed. The cultures give off an intense putrefactive odor. Upon *potato* a dry, grayish layer is formed, which may be about 0.5 millimetre in thickness.

Pathogenesis.—Not pathogenic for mice or for rabbits when injected in small amounts, but in considerable quantities causes fatal toxæmia in rabbits.

117. *BACILLUS OXYTOCUS PERNICIOSUS*.

Obtained by Wyssokowitsch from milk which had been standing for a long time.

Morphology.—Short bacilli with rounded ends, somewhat thicker and shorter than the lactic acid bacillus.

Biological Characters.—An *aërobic, non-liquefying* bacillus. In *gelatin plates* the deep colonies are small, spherical, finely granular, and of a yellowish or brownish-yellow color. The superficial colonies are hemispherical masses of a grayish-white color—by transmitted light, light-brown. They may have a diameter of one and one-half millimetres.

In *gelatin stick cultures* the growth is at first "nail-like"; later it extends over the entire surface of the gelatin. It causes coagulation of milk, with a sour reaction, within twenty-four hours. The cultures are without odor.

Pathogenesis.—Small doses are not pathogenic for mice or for rabbits, but considerable quantities injected into the circulation of rabbits cause their death in from three to twenty-two hours. Soon after the injection an abundant diarrhoea is developed. At the autopsy a hæmorrhagic inflammation of the intestinal mucous membrane is the principal pathological appearance observed.

118. *BACILLUS SAPROGENES* II.

Obtained by Rosenbach (1884) from the perspiration of foul-smelling feet.

Morphology.—Short bacilli with rounded ends.

Biological Characters.—*Aërobic and facultative anaërobic*. Characters of growth in gelatin, motility, etc., not given.

Streak cultures upon the surface of nutrient agar, at the end of twenty-four hours, cause the entire surface to be covered with minute, transparent colonies, which later become confluent and gradually somewhat opaque, forming a viscid, whitish gray layer. The odor of cultures resembles that of perspiring feet. Causes putrefaction of albuminous substances in the presence of oxygen, with evolution of stinking gases. In the absence of oxygen putrefactive changes also occurred, but less rapidly.

Pathogenesis.—When injected in considerable quantity into the knee joint or into the pleural cavity of rabbits, the animals succumb in from three to five days.

119. *BACILLUS OF AFANASSIEW*.

Obtained by Afanassiew (1887) from mucus and masses of pus coughed up by patients suffering from whooping cough. Etiological relation not demonstrated.

Morphology.—Bacilli from 0.6 to 2.2 μ long; solitary, in pairs, or in short chains.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Forms spores. Grows at the room temperature in the usual culture media. Upon *gelatin plates* the colonies are spherical or oval and of a light-brown color; under a low power they are seen to be finely granular, and later have a dark-brown color. Upon the surface of *gelatin stick cultures* a grayish-white layer is formed; but slight development occurs along the line of puncture. Upon the surface of *agar* a thick, gray layer forms along the line of inoculation. Upon *potato* yellowish, glistening, dew-like drops are first formed along the line of inoculation, and later a rather thick, brownish layer is formed which extends rapidly over the surface. Development is most rapid in the incubating oven.

Pathogenesis.—According to Afanassiew, pure cultures injected into the air passages or pulmonary parenchyma, in young dogs or in rabbits, produce bronchial catarrh, broncho-pneumonia, and attacks of spasmodic coughing resembling those of whooping cough. Death sometimes occurs. At the autopsy the bacillus is found in great numbers in the bronchial and nasal mucus.

120. PNEUMOBACILLUS LIQUEFACIENS BOVIS.

Obtained by Arloing from the lung of an ox which succumbed to an infectious form of pneumonia.

Morphology.—Slender, short bacilli, which rather resemble micrococci when cultivated in gelatin.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, non-motile* bacillus. Spore formation not observed; is killed by exposure for fifteen to twenty minutes to a temperature of 55° C. Grows in the usual culture media at the room temperature—better at 35° C. Forms white colonies in gelatin plates, and causes rapid liquefaction of the gelatin. Upon *potato* grows very rapidly as a white layer, which later has a brownish color.

Pathogenesis.—From one-half to one cubic centimetre of a pure culture injected beneath the skin of an ox, where the connective tissue is loose, causes the development of an acute abscess the size of a man's hand; after extending for two or three days this gradually becomes smaller and recovery occurs. When larger quantities are injected a fatal termination may result. Guinea-pigs and rabbits are less susceptible, and dogs are said to be immune.

121. BACILLUS PSEUDOTUBERCULOSIS.

Obtained by Pfeiffer (1889) from the organs of a horse suspected of having glanders and killed.

Morphology.—Rather thick bacilli with round ends; vary considerably in length—usually three to five times as long as broad.

Stains with fuchsin and Löffler's solution of methylene blue; does not stain by Gram's method.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of twenty-four hours, the superficial colonies are small, yellowish-brown plates, which increase rapidly in diameter; under a low power a central papilla is observed, around which the colony extends as a pale-yellow, peculiarly marbled, crystalline disc; the deep colonies are at first transparent, sharply defined spheres; on the third day, under a low power, they are seen to have a dark, finely granular central portion surrounded by a transparent zone; when not crowded upon the plate they may appear as yellowish-brown, finely granular, pear-shaped or lemon-shaped colonies. In *gelatin stick cultures* growth occurs along the line of puncture in the form of grayish-white, spherical colonies, more or less crowded above, and often isolated below, where by

transmitted light they are seen to have a brownish color ; upon the surface a grayish-white, concentric layer is formed about the point of inoculation in the course of five or six days, which later forms a disc with thickened margins. Upon the surface of *agar* the growth along the line of inoculation is abundant and viscid. Does not grow well upon *potato*. Upon *blood serum* forms transparent, drop-like colonies which have an opalescent appearance.

Pathogenesis.—Pathogenic for rabbits, guinea-pigs, hares, white mice, and house mice. Death occurs in from six to twenty days. At the autopsy the lymphatic glands are found to be enlarged and to have undergone caseation ; the liver and spleen are enlarged, the lungs oedematous and occasionally contain tuberculous-looking nodules. An abscess forms at the point of inoculation. Bacilli are found in the blood, the lymphatic glands, and the various organs.

122. BACILLUS GINGIVÆ PYOGENES.

Synonym.—*Bacterium gingivæ pyogenes* (Miller).

Obtained by Miller from an alveolar abscess and from deposit around the teeth "in a filthy mouth."

Morphology.—Short and thick bacilli with rounded ends, one to four times as long as broad; occur singly or in pairs.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* bacillus. Grows rapidly in the usual culture media. Upon gelatin plates it forms spherical colonies at the end of twenty-four hours, which have a yellowish color and well-defined margin; at the end of forty-eight hours liquefaction has progressed so far that the colonies have become confluent. In gelatin stick cultures liquefaction occurs rapidly in the form of a funnel, at the bottom of which a white deposit is formed. Upon the surface of *agar* a thick, moist growth occurs along the line of inoculation, which under the microscope has a slightly greenish-yellow tint and a fibrillated structure.

Pathogenesis.—Pathogenic for rabbits, guinea-pigs, and for white mice, when injected into the cavity of the abdomen in comparatively small amounts (0.25 cubic centimetre). At the autopsy peritonitis, sometimes purulent, is observed. Death occurs in from ten to twenty-four hours. The bacilli are found in the blood in small numbers. Subcutaneous injections in the animals mentioned produce a local abscess only.

123. BACILLUS DENTALIS VIRIDANS.

Found by Miller in the superficial layers of carious dentine.

Morphology.—Slightly curved bacilli with pointed ends; solitary or in pairs.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. In *gelatin plates* the colonies are spherical, and under a low power are colorless or have a slightly yellow tint; when not crowded they may present two or three concentric rings. In *gelatin stick cultures* growth occurs both upon the surface and along the line of puncture. Gelatin cultures acquire an opalescent-green color. Upon the surface of *agar* a thin growth with irregular margins occurs along the impf-strieh; this is bluish by transmitted light and greenish-gray by reflected light—colorless under the microscope.

Pathogenesis.—Injections into the cavity of the abdomen of white mice or of guinea-pigs usually cause fatal peritonitis in from one to six days; the bacilli are only found in the blood in small numbers, by the culture method. Subcutaneous injections in the animals mentioned produce severe local inflammation and suppuration.

124. BACILLUS PULPÆ PYOGENES.

Obtained by Miller from gangrenous tooth pulp.

Morphology.—Slightly curved bacilli with pointed ends; solitary or in pairs, or in chains of four to eight elements.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. In *gelatin plates* large, spherical, opaque, yellowish-brown colonies are formed. In *gelatin stick cultures* liquefaction occurs in the upper part of the tube and gradually extends downward, the liquefied gelatin being separated from the non-liquefied by a horizontal plane.

Pathogenesis.—Small quantities of a pure culture injected into the abdominal cavity of white mice proved fatal to these animals in from eighteen to thirty hours.

125. BACILLUS SEPTICUS KERATOMALACIÆ.

Obtained by Babes (1889) from the broken-down corneal tissues and from the various organs of a child which died of septicæmia following keratomalacia.

Stains with the usual aniline colors; deeply colored granules may often be seen at the extremities of the rods, or in the middle, in preparations stained with Löffler's solution.

Morphology.—Short, thick bacilli, thinning out at the ends; often united in pairs; may be surrounded by a capsule.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms white, slightly elevated, flat colonies with finely dentate margins. In *gelatin stick cultures* the growth is abundant both on the surface and along the line of puncture; gas bubbles are formed in the gelatin. Upon the surface of *agar* the growth along the line of inoculation is leaf-like, finely dentate, somewhat opalescent, and the culture has a slightly ammoniacal odor. Upon *blood serum* a semi-transparent, glistening film is formed, which has dentate margins.

Pathogenesis.—Pathogenic for rabbits and mice, less so for birds; not pathogenic for guinea-pigs. The animals die in from three to seven days. Inoculated into the cornea it causes a purulent keratitis.

126. BACILLUS SEPTICUS ACUMINATUS.

Obtained by Babes (1889) from the blood, the umbilical stump, and the various organs of a child which died five days after birth, apparently from septic infection.

Morphology.—Bacilli with lancet-shaped ends, somewhat resembling the bacillus of mouse septicæmia, but thicker. Often shows unstained places in the middle of the rods in stained preparations.

Stains readily with the usual aniline colors.

Biological Characters.—An *aërobic* bacillus; does not grow in gelatin at the room temperature. Spore formation not observed. Grows upon *blood serum* and upon *nutrient agar* at 37° C., in form of small, flat, circular, transparent, shining colonies, which become confluent and later form a yellowish layer. Blood serum is the most favorable medium.

Pathogenesis.—Pathogenic for rabbits and guinea-pigs, not for mice. The animals die in from two to six days, and the bacilli are found in their blood and in the various organs.

127. BACILLUS SEPTICUS ULCERIS GANGRÆNOSI.

Obtained by Babes (1889) from the blood and various organs of a boy who died from septicæmia following gangrene of the skin, etc.

Morphology.—Bacilli with round ends, oval or rod-shaped, about 0.5 to 0.6 μ thick.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* a sac-formed liquefaction occurs and a yellow deposit is seen at the bottom of the liquefied gelatin; gas bubbles are given off from the culture. Upon the surface of *agar* development occurs along the line of inoculation in the form of flat, grayish-yellow, transparent, varnish-like plaques. Upon *potato*, after several days, a brownish, shining, moist, transparent film is formed. Upon the surface of *blood serum* smooth, yellowish, transparent colonies are formed, under which the blood serum is softened, allowing these to sink below the surface.

Pathogenesis.—Pathogenic for mice and for guinea-pigs, which die in from one to two days. An abscess forms at the point of inoculation, which is covered with a dry, retracted crust.

128. BACILLUS OF TRICOMI.

Obtained by Tricomi (1886) from a case of senile gangrene.

Morphology.—Bacilli with round ends, about three μ long and one μ thick, solitary or in pairs; sometimes one end of a rod shows a club-shaped thickening.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic, liquefying, non-motile* bacillus. Forms spores. Grows in the usual culture media at the room temperature—better at 37° C.

Upon *gelatin plates*, at the end of twenty-four hours, the colonies are spherical, finely granular, and of a dirty-yellow color; after from thirty-six to forty-eight hours liquefaction of the surrounding gelatin occurs. In *gelatin stick cultures* closely crowded, small, white colonies are formed along the line of puncture; at the end of forty-eight hours liquefaction commences in funnel form, with formation of an air bubble above—like the cholera spirillum; later the entire gelatin is liquefied and becomes transparent, while a dirty-white collection of bacilli is seen at the bottom of the tube. Upon the surface of *agar* a white layer with irregular margins is formed, which later extends over the entire surface as a homogeneous, rather thin membranous film. Upon *potato*, at 37° C., dirty-white, milky colonies are formed, which later become confluent. Upon *blood serum* the growth is similar to that upon agar.

Pathogenesis.—The subcutaneous injection of one-half to one cubic centimetre of a gelatin culture is said by Tricomi to produce in rabbits and in guinea-pigs a gangrenous process resembling senile gangrene in man. The subcutaneous connective tissue is infiltrated with a foul-smelling serum, the muscles are soft and gray, and a portion of the skin has a mummified appearance. The gangrene extends over the abdomen, and death occurs in guinea-pigs in two to three days, in rabbits after four days, in house mice at the end of twenty-four hours; white mice are said to be immune.

129. BACILLUS ALBUS CADAVERIS.

Obtained by Strassmann and Stricker (1888) from the blood of two cadavers four days after death.

Morphology.—Bacilli about two and one-half μ long and 0.75 μ broad; also grow out into filaments of six μ or longer.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. In *gelatin plates* small, spherical, yellowish colonies are formed during the first twenty-four hours; later a radiating outgrowth occurs from the periphery, and liquefaction of the gelatin takes place. In *gelatin stick cultures* liquefaction begins within forty-eight hours, and forms a long funnel, at the opening of which is a cavity containing air; the liquefied gela-

tin is transparent, and a deposit of thick, granular masses accumulates at the bottom of the tube. Upon the surface of *agar* a thick, white layer is formed, which later is wrinkled and after a time gives off a putrefactive odor. Gelatin cultures give off an odor of sulphuretted hydrogen. Upon *potato* a soft, white or pale-yellow layer is formed, which in places is made up of small granules. The potato around the growth has a bluish-brown color.

Pathogenesis.—Subcutaneous injection of a small quantity (0.1 cubic centimetre) of a liquefied gelatin culture is fatal to mice in about six hours; the animals become comatose before death, and at the autopsy putrefactive changes are already observed; the bacillus can be recovered from the blood in cultures. Sterilized cultures also prove fatal to mice. Pathogenic also for guinea-pigs, which die in about twenty hours after receiving a subcutaneous inoculation.

130. BACILLUS VARICOSUS CONJUNCTIVÆ.

Obtained by Gombert (1889) from the healthy conjunctival sac of man.

Morphology.—Large bacilli with round ends, from two to eight μ long and about one μ broad; the shorter bacilli are often constricted in the middle.

Stains with the usual aniline colors.

Biological Characters.—An *aërobie* and *facultative anaërobie*, *liquefying*, *non-motile* bacillus. Grows very slowly in nutrient gelatin at 22° C.; rapidly in *agar* and upon *potato* at 37° C. In *gelatin stick cultures*, at the end of twenty-four hours, a circular layer having a grayish-white centre is developed upon the surface, and a scarcely visible grayish-white thread along the line of puncture. Liquefaction extends gradually from the surface without clouding or changing the gelatin, so that at the end of two weeks the gelatin is entirely liquefied without giving any other evidence of the presence of the microorganism. Upon *agar plates*, at 37° C., the deep colonies have a diameter of about four millimetres by the end of the fourth day; under a low power they are seen to be covered with minute, irregular, thorn-like projections, which subsequently increase in size; the centre of the colony is granular and opaque. The superficial colonies, under a low power, are seen to have an opaque central nucleus surrounded by a yellowish, finely granular, transparent peripheral zone; later the central portion is irregular and semi-opaque, surrounded by a broad marginal zone which consists of twisted and bent tapering offshoots having a dark contour. Upon the *surface of agar* a thin, white, dry, very adherent film is formed; a thick, white film forms upon the surface of the condensation water. Upon *potato* development is rapid at 37° C., forming at first a dry, white layer, which at the end of ten days covers the entire surface; it then has an irregular surface and fringed margins, is smooth, dry, and after a time has a reddish-brown color.

Pathogenesis.—When inoculated into the cornea of rabbits a grayish-white cloudiness is developed in twenty-four hours, around which the cornea is highly vascular; the animal recovers without the formation of an abscess. Injected into the conjunctiva it causes an intense hyperæmia.

131. BACILLUS MENINGITIDIS PURULENTÆ.

Obtained by Neumann and Schäffer (1887) from pus from beneath the pia mater in an individual who died of purulent meningitis.

Morphology.—Bacilli about two μ long and 0.6 to 0.7 μ broad; often grow out into long filaments, especially in gelatin cultures.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobie* and *facultative anaërobie*, *non-liquefying*, *motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature—better in the incubating oven. Upon *gelatin plates* the deep colonies, under a low power, are homogeneous, round or oval, pale brown, and with a smooth contour; the superficial colonies are

thin, moist, and transparent in appearance; later they have a grayish color, a coarsely granular surface, and are made up of flap-like layers. In *gelatin stick cultures* the superficial growth consists of broad, grayish layers, and a grayish-yellow growth is seen along the line of puncture, made up of crowded colonies. Upon *agar plates*, at the end of twenty-four hours at 37° C., thin colonies are developed, which have a granular surface, a smooth, more or less irregular outline, and a pale-brown color in the centre. Upon *potato* a scanty, moist, white layer develops along the line of inoculation. Upon *blood serum*, at 37° C., at the end of twenty-four hours a moist, shining layer about four millimetres broad is developed along the *impfstrich*; this is granular at the margins, and later more or less fissured.

Pathogenesis.—Subcutaneous injection produces in dogs, rabbits, guinea-pigs, and white mice a purulent inflammation in the vicinity of the point of inoculation.

132. BACILLUS SEPTICUS VESICÆ.

Obtained by Clado (1887) from the urine of a person suffering from cystitis.

Morphology.—Bacilli with round ends, 1.6 to 2 μ long and 0.5 μ thick; never united in pairs or chains.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Forms spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* small, spherical or oval colonies are developed throughout the gelatin, which rarely exceed the size of a pin's head; these are transparent, and yellowish-white in color; under a low power the centre is seen to be dark gray and is surrounded by a well-defined marginal zone of a pale-yellow color. In *gelatin stick cultures* the growth along the line of puncture is first seen as a delicate, whitish thread; at the end of six or seven days it is made up of lenticular colonies, one-third as large as a pin's head, arranged in two lines like piles of coin. Upon the surface the growth is scanty and consists of a thin layer around the point of inoculation, which has a jagged contour. Upon the surface of *agar* development is slow and forms a grayish-white stripe along the *impfstrich*. Upon *potato* a flat, dry, chestnut-brown layer is formed.

Pathogenesis.—Pathogenic for rabbits, guinea-pigs, and mice. Death appears to result from the toxic products formed, as well as from the multiplication of the bacilli in the inoculated animals.

133. BACILLUS OF GESSNER.

Synonym.—*Bacterium tholoideum* (Gessner).

Obtained by Gessner from the contents of the intestine of healthy persons.

Resembles in its morphology and in its growth in culture media *Bacillus lactis aërogenes* of Escherich.

Pathogenic for mice and for guinea-pigs.

134. BACILLUS CHROMO-AROMATICUS.

Obtained by Galtier (1888) from a pig which died from a general infectious malady characterized by broncho-pneumonia, pleuritis, enteritis, and swelling of the lymphatic glands.

Morphology.—Bacilli of medium size with rounded ends.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *motile* bacillus. Not observed to form spores. Grows in the usual culture media at the room temperature—better in the incubating oven. The cultures all produce a green or brown pigment and have an aromatic odor. In *gelatin stick cultures* a yellowish-white layer is formed upon the surface of the liquefied gelatin, which has a bright-green color; a yellowish-white

deposit accumulates at the bottom of the tube. Upon the surface of agar whitish colonies are formed, which coalesce to form a thin layer. Upon *potato* a tolerably thick, somewhat iridescent, brown layer is formed, which extends over the entire surface. In *bouillon*, at the end of twenty-four to forty-eight hours at 37° C., a greenish-yellow color is developed, first near the surface and later extending throughout the fluid, which acquires the color of a dilute solution of sulphate of copper; a whitish film forms upon the surface. In anaërobic cultures the color is a pale brown instead of green.

Pathogenesis.—Rabbits die at the end of two to three weeks after receiving an intravenous injection. At the autopsy they are found to have pneumonia with pleuritis and pericarditis.

135. BACILLUS CANALIS CAPSULATUS.

Obtained by Mori (1888) from sewer water.

Morphology.—Bacilli with round ends, elliptical or rod-shape in form, and from 0.9 to 1.6 μ thick; often surrounded with a broad capsule, which is always seen in preparations from the blood or tissues of an infected animal; sometimes in pairs with the acute ends of the rods in apposition, and surrounded by a single capsule.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* hemispherical, porcelain-white, sharply defined colonies, resembling those of Friedländer's bacillus, are developed at the end of twenty-four hours. In *gelatin stick cultures* development occurs along the line of puncture and upon the surface, forming a "nail-shaped" growth similar to that of Friedländer's bacillus (*Bacillus pneumoniae*) in the same medium. Upon *agar* a viscid and abundant growth is formed in the incubating oven at 37° C. Upon *potato* an abundant development in the form of a yellowish, moist, viscid layer, with irregular outlines. In *bouillon*, at the end of three or four days, a white film forms on the surface, especially in contact with the test tube.

Pathogenesis.—Mice die in two to three days after receiving a subcutaneous injection. Guinea-pigs and rabbits are immune.

136. BACILLUS CANALIS PARVUS.

Obtained by Mori (1888) from sewer water.

Morphology.—Bacilli with round ends, from 2 to 5 μ long and 0.8 to 1 μ broad.

Stains with the usual aniline colors, but not by Gram's method; the ends of the rods are more deeply stained than the central portion.

Biological Characters.—An *aërobic*, *non-liquefying*, *non-motile* bacillus. Not observed to form spores. Grows very slowly at the room temperature—more rapidly at 37° C. Upon *gelatin plates*, at the end of two to three weeks, extremely minute, homogeneous, pale-yellow colonies are developed. In *gelatin stick cultures* a thin, yellowish layer forms upon the surface at the end of three weeks. Upon the surface of *agar*, at 37° C., a dry, yellowish layer with jagged outlines is developed in two or three days. No growth occurs upon *potato*. Upon blood serum a thin, pale-green, dry layer is formed.

Pathogenesis.—Mice die in from sixteen to thirty hours after receiving a subcutaneous inoculation, guinea-pigs in about two days.

137. BACILLUS INDIGOGENUS.

Obtained by Alvarez (1887) from an infusion of the leaves of the indigo plant.

Morphology.—Bacilli with round ends, about 3 μ long and 1.5 μ thick,

often united in chains of six to eight elements. The cells are surrounded by a transparent capsule resembling that of Friedländer's bacillus.

Biological Characters.—An *aërobic, motile* bacillus. Upon *agar*, at 37° C., a yellowish-white layer is quickly developed and there is production of gas. According to Alvarez, this bacillus develops an indigo-blue color in a sterilized infusion of the leaves of the indigo plant.

Pathogenesis.—Guinea pigs die in from three to twelve hours from the intravenous injection of a pure culture.

138. BACILLUS OF KARTULIS.

Obtained by Koch (1883) and by Kartulis from the conjunctival secretions of persons suffering from a form of infectious catarrhal conjunctivitis which prevails in Egypt.

Morphology.—Resembles the bacillus of mouse septicæmia (*Bacillus murisepticus*) in its form and dimensions.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic* bacillus. Does not grow in nutrient gelatin at the room temperature. Upon the surface of *nutrient agar*, at 28° to 30° C., at the end of thirty to forty hours small, grayish-white points are developed along the impfstrich; later these become confluent and form an elevated, shining, dark-colored layer with irregular and often jagged margins.

Pathogenesis.—Out of six experimental inoculations, with pure cultures, made by Kartulis in the eyes of healthy individuals, four gave a negative result, one produced a catarrhal inflammation lasting for a week, in an eye which was blind from a previous attack of sclerochoroiditis, and one a conjunctivitis lasting for ten days in a perfectly healthy eye.

139. BACILLUS OF UTPADEL.

Obtained by Utpadel (1887) from the wards of a military hospital at Augsburg—in the “Zwischendeckenfüllung”; also by Gessner from the contents of the small intestine in man.

Morphology.—Bacilli with round ends, 1.25 to 1.5 μ long and 0.75 to 1 μ thick; often united in pairs or in chains of three elements.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Grows in the usual culture media at the room temperature. Spore formation not observed. Upon gelatin plates the superficial colonies are elevated and sometimes conical, and of a milk-white color. The deep colonies are round or oval; the centre is dark green and is surrounded by a brownish-green peripheral zone. Upon the surface of *agar* a yellowish-white layer is developed very slowly. The growth upon gelatin is rapid.

Pathogenesis.—When injected subcutaneously into cats, guinea-pigs, or mice it produces an extensive inflammatory oedema, resulting in the death of the animals.

140. BACILLUS ALVEI.

Synonym.—Bacillus of foul brood (of bees).

Obtained by Cheshire and Cheyne (1885) from the larvæ in hives infected with “foul brood.” The larvæ in the interior of cells in the comb die and become almost fluid as a result of parasitic invasion by this bacillus.

Morphology.—Bacilli with rounded ends, from 2.5 to 5 μ in length (average about 3.6 μ) and 0.8 μ in diameter. Grow out into filaments and form large oval spores which have a greater diameter than the rods in which they are developed—1.07 μ .

Stains readily with the aniline colors usually employed, also by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefy-*

ing, motile bacillus. Forms endogenous spores. Grows readily in the usual culture media at the room temperature.

In *gelatin plates* small, round or oval colonies are formed, which later become pear-shaped; a branching outgrowth occurs about the margins of the colonies, and especially from the small end of the pear-shaped mass. In streak cultures upon the surface of gelatin growth occurs first along the impfstrich, and from this an outgrowth occurs consisting of bacilli in a single row or in several parallel rows, and forming irregular or circular figures, from which other similar outgrowths occur; the branching outgrowths may anastomose. The gelatin is liquefied in the vicinity of these lines of growth, forming a network of channels. A similar growth is seen upon the surface of *gelatin stick cultures*, and along the line of puncture white, irregular masses are formed, from which rather coarse branches are given off which often have a club-shaped extremity. In older cultures the finer branches disappear, so that the secondary centres of growth are disconnected from the original colonies; complete liquefaction of the gelatin occurs in about two weeks; the liquefied gelatin has a yellowish color and peculiar odor. Upon the surface of nutrient agar, at 37° C., a white layer is formed. Upon *potato* the development is slow and results in the formation of a dry, yellowish layer. In *milk* coagulation first occurs, and the coagulum is subsequently dissolved; a slightly acid reaction is produced. This bacillus grows best in the incubating oven at 37°, and does not develop at temperatures below 16° C. The spores require for their destruction a temperature of 100° C. maintained for four minutes (determined by the writer, 1887).

Pathogenesis.—The introduction of pure cultures of this bacillus into hives occupied by healthy swarms causes them to become infected with foul brood; grown bees also become infected when given food containing the bacillus (Cheshire). Mice injected subcutaneously with a considerable quantity die within twenty-four hours, guinea-pigs in six days (Eisenberg). Small amounts injected beneath the skin of mice or rabbits produce no apparent result.

141. BACILLUS OF ACNE CONTAGIOSA OF HORSES.

Obtained by Dieckerhoff and Grawitz (1885) from pus and dried scales from the pustules of "acne contagiosa" of horses.

Morphology.—Short rods, straight or slightly bent, 0.2 μ in diameter.

Stains best with an aqueous solution of fuchsin, and also by Gram's method; does not stain well with Löffler's alkaline solution of methylene blue.

Biological Characters.—An *aërobie, non-liquefying* bacillus. In *gelatin stick cultures* a very scanty growth occurs along the line of puncture; upon the surface a white mass forms about the point of puncture. Upon *blood serum* and nutrient *agar* an abundant growth at the end of twenty-four hours at 37° C., consisting of white colonies along the impfstrich, which later have a yellowish-gray color. The growth is more abundant and rapid upon blood serum than upon other media.

Pathogenesis.—Pure cultures of the bacillus described are said by Dieckerhoff and Grawitz to produce typical acne pustules when rubbed into the skin of horses, calves, sheep, and dogs. When rubbed into the intact skin of guinea-pigs a phlegmonous erysipelatous inflammation was produced, and the animal died at the end of forty-eight hours with symptoms of toxæmia. Subcutaneous injections in guinea-pigs caused toxæmia and death at the end of twenty-four hours. At the autopsy a hæmorrhagic infiltration of the intestinal mucous membrane was observed; the bacilli were not found in the internal organs. In rabbits pure cultures rubbed into the intact skin caused a development of pustules and a severe inflammation of the subcutaneous connective tissue, from which the animal usually recovered. Subcutaneous injections in rabbits sometimes caused a fatal toxæmia. House mice, field mice, and white mice were not affected by the application of cultures, by

rubbing, to the uninjured skin, but succumbed to subcutaneous injections in twenty-four hours or between the fifth and tenth days. Those which died at a late date presented the pathological appearances which characterize pyæmia.

142. BACILLUS NO. I OF ROTH.

Obtained by Roth (1890) from old rags. Resembles *Bacillus coli communis* and Brieger's bacillus in its morphology and growth in various culture media, but, according to Roth, is distinguished from these bacilli by the fact that colonies upon gelatin plates are thicker and more opaque.

Pathogenesis.—Pathogenic for rabbits and for guinea-pigs when injected into the cavity of the abdomen; death usually occurs within twenty-four hours. The spleen is greatly enlarged, and the bacilli are found in cultures from the blood and various organs.

143. BACILLUS NO. II OF ROTH.

Obtained by Roth (1890) from old rags.

Morphology.—Bacilli with round ends, 0.6 to 1 μ broad and two to four times as long.

Stains with the usual aniline colors. When stained by Gram's method it is decolorized by alcohol.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* colonies resembling those of the colon bacillus are developed at the end of twenty-four hours; on the third day small, drop-like, shining, bluish-white colonies, around the periphery of which a commencing extension upon the surface of the gelatin is seen. Older colonies are seldom more than one-half centimetre in diameter, and are somewhat thicker than this; they are nearly transparent. Upon the surface of *gelatin stick cultures* a rather moist, yellowish-white layer with dentate margins is developed. Upon *potato* a colorless layer is developed, which later has a grayish color.

Pathogenic for rabbits and guinea-pigs when injected into the abdominal cavity.

144. BACILLUS OF OKADA.

Obtained by Okada (1891) from dust between the boards of a floor.

Morphology.—Short rods with round ends, about as long as *Bacillus murisepticus*, but somewhat thicker—about twice as long as thick; solitary or in pairs; in old cultures may grow out into filaments.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of two to three days, small, white, spherical colonies are developed. Under the microscope these are seen to be granular, pale-brown in color, and with slightly jagged margins; the superficial colonies after several days are considerably elevated above the level of the gelatin. In *gelatin stick cultures* development occurs as a white thread along the line of puncture, and upon the surface as a flat, milk-white layer which does not extend to the walls of the test tube. Upon *agar*, at 37° C., the growth is rapid and the surface is nearly covered at the end of eighteen hours with a milk-white layer; the condensation water is filled with a viscid mass of bacilli. Upon *blood serum* the growth is shining and almost transparent. In *bouillon* development is rapid, clouding the fluid throughout, and a cream-like layer forms upon the surface.

Pathogenesis.—Rabbits and guinea-pigs die in about twenty hours after receiving a subcutaneous injection of a half-syringeful of a bouillon culture, or from a small quantity (two öse) from a gelatin or agar culture. In

mice a minute quantity of a pure culture invariably proved fatal in about twenty hours. Four hours after the inoculation an abundant secretion from the lachrymal glands occurs, and soon after the eyes become completely closed.

According to Okada, this bacillus is differentiated from the bacillus of Brieger, and from Emmerich's bacillus which it greatly resembles, by the fact that it does not grow upon potato.

145. BACILLUS OF PURPURA HÆMORRHAGICA OF TIZZONI AND GIOVANNINI.

Obtained by Tizzoni and Giovannini (1889) from the blood of two children who died of purpura hæmorrhagica following impetigo.

Morphology.—Bacilli with round ends, from 0.75 to 1.3 μ long and 0.2 to 0.4 μ broad; often seen in pairs or in groups like streptococci.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the colonies at first resemble those of *Streptococcus pyogenes*. Upon the surface small, opaque points are seen at the end of forty-eight hours, which at the end of four to five days develop into spherical, yellowish-gray colonies with irregular margins, surrounded by a growth resembling tufts of curly hair. Upon *agar* the growth is similar, but more rapid and of a pale color, often with a central nucleus surrounded by a net-like marginal zone. Upon *blood serum* the growth is similar to that upon agar. Upon *potato*, at 37 C., a limited development occurs about the point of inoculation, which has a dark-yellow color. The cultures give off a very penetrating odor.

Pathogenesis.—Pathogenic for dogs, rabbits, and guinea-pigs when injected subcutaneously. Not pathogenic for white mice or pigeons. The symptoms resulting from a subcutaneous injection are said to be fever, albuminuria and, in some cases, anuria, hæmorrhagic spots upon the skin, convulsions; death occurs in from one to three days. At the autopsy there are found œdema about the point of inoculation, hæmorrhages in the skin and muscles, and sometimes in the internal organs and in serous cavities; the blood does not coagulate. The bacilli are found in the subcutaneous connective tissue, but not in the blood or in the various organs. Sections show coagulation necrosis of the liver cells and of the renal epithelium.

146. BACILLUS OF PURPURA HÆMORRHAGICA OF BABES.

Obtained by Babes (1890) from the spleen and lungs of an individual who died from purpura hæmorrhagica with symptoms of septicæmia. Resembles the bacillus previously described by Tizzoni and Giovannini, and still more that of Kolb; but, according to Babes, differs in some respects from both of these, although they all belong evidently to the same group.

Morphology.—Bacilli with rounded ends, oval or pear-shaped, about 0.3 μ thick, surrounded by a narrow capsule.

Stains with the aniline colors, but not deeply, and still less intensely by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature. In *gelatin stick cultures*, at the end of three days, a thin, transparent, irregular layer has developed upon the surface, and a whitish, punctate stripe along the line of inoculation. In *agar stick cultures* an abundant development occurs along the line of puncture, and at the end of three days the growth upon the surface consists of small, moist, transparent drops; later of larger, flat, shining, yellowish-white plaques which have ill-defined margins. Upon *blood serum* the development is somewhat more abundant in the form of small, white, moist colonies one to two millimetres broad. Upon *potato*, at the end of three days, moist, whitish drops with ill-defined margins.

Pathogenesis.—Inoculations in the conjunctivæ of rabbits produce ecchymoses of the conjunctiva. At the autopsy numerous hæmorrhagic extravasations are found in all the organs, especially in the lungs and liver; the spleen is enlarged; the bacilli can be recovered in pure cultures from the various organs. Old cultures proved to have lost their virulence. Pathogenic for mice, which die from general infection in the course of a few days; the spleen is enlarged, and hæmorrhages in the serous membranes are usually seen.

147. BACILLUS OF PURPURA HÆMORRHAGICA OF KOLB.

Obtained by Kolb (1891) from the various organs of three individuals who died in from two to four days from attacks characterized by suddenly developed fever, purpura, and albuminous urine.

Morphology.—Oval bacilli, usually in pairs, 0.8 to 1.5 μ long and 0.8 μ broad, surrounded by a narrow capsule, which is only seen distinctly in preparations from the organs.

Stains with the aniline colors, but not deeply, and still more feebly by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature. In *gelatin stick cultures*, at the end of four days, a very small, thin, hyaline growth is seen about the point of inoculation. The development is more abundant along the line of puncture. Upon the surface of agar a thin layer is formed with smooth margins. Upon *potato*, at the end of three to four days, a whitish, moist, shining stripe is seen along the *impfstrich* which is about three millimetres broad.

Pathogenesis.—Injections of 0.5 to 1 cubic centimetre of a bouillon culture into the abdominal cavity of rabbits cause symptoms of general infection in the course of a few days, and not infrequently hæmorrhagic extravasations are seen in the ear muscles. More than one cubic centimetre may cause death in from one to three days. At the autopsy hæmorrhagic extravasations are found in the subcutaneous tissues and in the serous and mucous membranes. The blood has little disposition to coagulate; the bacillus may be recovered in pure cultures from the various organs. In guinea-pigs local ecchymoses are sometimes produced, otherwise not pathogenic for this animal. Pathogenic for mice, which die from general infection, after being inoculated with a small quantity of a pure culture, in from two to three days; spleen enlarged; lymphatic glands often hæmorrhagic. Not fatal to dogs, but animals which were inoculated with one cubic centimetre of a bouillon culture and subsequently killed proved to have hæmorrhagic extravasations in the various organs.

148. BACILLUS HEMINECROBIOPHILUS.

Obtained by Arloing (1889) from a caseous lymphatic gland in a guinea-pig.

Morphology.—Bacilli which vary greatly in length and are sometimes so short as to resemble micrococci ('polymorphous'); usually from one to four μ long; in anaërobic cultures from eight to twenty μ .

Stains with the usual aniline colors.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *slightly motile* bacillus. Spore formation not observed. Grows rapidly in the usual culture media—best in the incubating oven at 35° C. The growth upon the surface of gelatin has a yellowish color. Upon *potato* a yellowish-white layer is developed.

Pathogenesis.—According to Arloing, this bacillus is not pathogenic when injected into healthy tissues in dogs, sheep, guinea-pigs, and rabbits, but when the tissues have previously been injured it produces a local oedema and necrotic changes, accompanied by gas formation. This is not peculiar to the microorganism described by Arloing, which appears to be one of the *Proteus* group.

XIV.

PATHOGENIC ANAEROBIC BACILLI.

STRICTLY anaërobic bacilli are not able to multiply in the blood of living animals ; but some of them may multiply in the subcutaneous connective tissue or in the muscles, when introduced by inoculation, and are pathogenic because of the local inflammatory or necrotic processes to which they give rise, or because they produce soluble toxic substances which are absorbed and cause death by their special action upon the nervous system or by general toxæmia.

149. BACILLUS TETANI.

Synonyms.—The bacillus of tetanus ; Tetanusbacillus, *Ger.*

Nicolaier (1884) produced tetanus in mice and rabbits by introducing garden earth beneath their skin, and showed that the disease might be transmitted to other animals by inoculations with pus or cultures in blood serum containing the tetanus bacillus, which, however, he did not succeed in obtaining in pure cultures. Carle and Rattone (1884) showed that tetanus is an infectious disease, which may be transmitted by inoculation from man to lower animals—a fact which has since been verified by the experiments of Rosenbach and others. Obtained in pure cultures by Kitasato (1889).

The writer produced tetanus in a rabbit in 1880 by injecting beneath its skin a little mud from the street gutters in New Orleans. The tetanus bacillus appears to be a widely distributed microorganism in the superficial layers of the soil in temperate and especially in tropical regions. In Nicolaier's experiments it was not found in soil from forests or in the deeper layers of garden earth.

Morphology.—Slender, straight bacilli, with rounded ends, which may grow out into long filaments. Spores are developed at one extremity of the bacilli, which are spherical in form and considerably greater in diameter than the rods themselves, giving the spore-bearing bacilli the shape of a pin.

Stains with the usual aniline colors and also by Gram's method. The method of Ziehl may be employed for double-staining bacilli and spores.

Biological Characters.—An *anaerobic, liquefying, motile* bacillus. *Forms spores.* Grows at the room temperature, in the absence of oxygen, in the usual culture media. Grows best at a temperature of 36° to 38° C.; in nutrient gelatin, at 20° to 25° C., development is first seen at the end of three or four days; does not grow at a temperature below 14° C. Spores are formed in cultures kept in the incubating oven at 36° C., at the end of thirty hours; in gelatin cultures at 20° to 25° C., at the end of a week (Kitasato). The bacilli exhibit voluntary movements which are not very active; those containing spores are not motile. It may be cultivated in an atmosphere of hydrogen, but does not grow in the presence of oxygen—strictly anaerobic—or in an atmosphere of carbon dioxide. The addition of one and one-half to two per cent of grape sugar to nutrient agar or gelatin causes the development to be more rapid

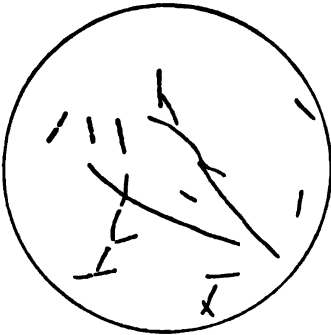


FIG. 159.

FIG. 159.—*Tetanus bacillus*, from a gelatin culture. $\times 1,000$. From a photomicrograph by Pfeiffer.



FIG. 160.

FIG. 160.—*Tetanus bacillus*, from an agar culture; spore-bearing rods. $\times 1,000$. From a photomicrograph by Pfeiffer.

and abundant. The culture medium should have a feebly alkaline reaction.

Colonies in *gelatin plates*, in an atmosphere of hydrogen, resemble somewhat colonies of *Bacillus subtilis*, the opaque central portion being surrounded by a circle of diverging rays; liquefaction is, however, much slower, and the resemblance is lost after a short time. Older colonies resemble the colonies of certain microscopic fungi, being made up of diverging rays. In long *gelatin stick cultures* development occurs along the line of puncture, at a considerable distance below the surface, in the form of a radiate outgrowth; the gelatin is slowly liquefied, and a small amount of gas is at the same time formed. In peptonized bouillon having a slightly alkaline reaction, under hydrogen gas, the development is abundant

and the cultures give off a characteristic odor—"brenzlichen Geruch" (Kitasato).

According to Kitasato, *blood serum* is not a very favorable medium for the growth of the tetanus bacillus, and—contrary to the statement of Kitt, Tizzoni, and others—it does not cause liquefaction of this medium.

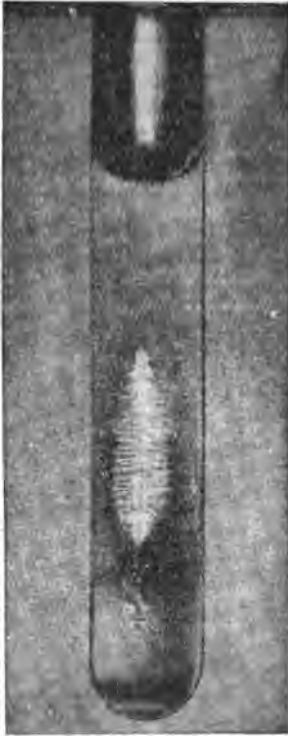


FIG. 161.—Culture of *Bacillus tetani* in nutrient gelatin. (Kitasato.)

The spores of the tetanus bacillus retain their vitality for months in a desiccated condition, and are not destroyed in two and one-half months when present in putrefying material (Turco). They withstand a temperature of 80° C. maintained for an hour, but are killed by five minutes' exposure to steam at 100° C. They are not destroyed in ten hours by a five-per-cent solution of carbolic acid, but did not grow after fifteen hours' exposure in the same solution. A five-per-cent solution of carbolic acid, to which 0.5 per cent of hydrochloric acid has been added, destroys them in two hours; in sublimate solution containing 1:1,000 of mercuric chloride they are destroyed at the end of three hours, or in thirty minutes when 0.5 per cent of hydrochloric acid is added to the solution. Kitasato succeeded in obtaining pure cultures from the pus formed in the vicinity of inoculation wounds, by destroying the associated bacilli after the tetanus bacilli had formed spores.

This was effected by heating cultures from this source for about an hour at a temperature of 80° C. The spores of the tetanus bacillus survived this exposure, and colonies were obtained from them in flat flasks especially devised for anaerobic cultures; from these colonies pure cultures in nutrient agar or gelatin—long stick cultures—or in peptonized bouillon were easily obtained.

Brieger (1886) first succeeded in obtaining from impure cultures of the tetanus bacillus a crystallizable toxic substance, called by him *tetanin*, which was found to kill small animals in very minute doses and with the characteristic symptoms of tetanus. More recently Kitasato and Weyl have obtained the same substance, by following Brieger's method, from a pure culture of this bacillus. From a

bouillon made from one and one-fourth kilogrammes of lean beef, with the addition of twenty-five grammes of peptone, they obtained 1.7118 grammes of hydrochlorate of tetanin. This proved fatal to white mice in six hours in the dose of 0.05 gramme, and a dose of 0.105 gramme caused characteristic tetanic convulsions and death within an hour. The bacteriologists last named also obtained from their cultures the *tetanotoxin* of Brieger. Two mice were inoculated subcutaneously with 0.003 gramme of this substance; one died at the end of five hours without the development of tetanic symptoms; the other survived. In addition to these substances, indol, phenol, and butyric acid were demonstrated to be present in cultures of the *tetanus bacillus*.

According to Kitasato, the *tetanus bacillus* does not become attenuated in its pathogenic potency by cultivation in artificial media, as is the case with many other pathogenic bacteria. The more recent researches of Brieger and Fränkel, and of Kitasato, show that the toxic ptomaine discovered by Brieger in 1886 is not the substance to which cultures of the *tetanus bacillus* owe their great and peculiar pathogenic power. The distinguished German chemist and his associate have succeeded in isolating from *tetanus* cultures a *toxalbumin* which is far more deadly than tetanin.

Pathogenesis.—The experiments of Kitasato (1889) show that pure cultures of the *tetanus bacillus* injected into mice, rabbits, or guinea-pigs produce typical tetanic symptoms and death. As the presence of this bacillus at the seat of injury, in cases of *tetanus* in man, has now been demonstrated by numerous observers, there is no longer any question that *tetanus* must be included among the traumatic infectious diseases, and that the bacillus of Nicolaier and of Kitasato is the specific infectious agent. Kitasato's recently published experiments (1890) show that cultures of the *tetanus bacillus* which have been sterilized by filtration through porcelain produce the same symptoms, and death, in the animals mentioned, as result from inoculation with cultures containing the bacillus. It is evident, therefore, that death results from the action of a toxic substance produced by the bacillus. This is further shown by the fact that the bacillus itself cannot be obtained in cultures from the blood or organs of an animal which has succumbed to an experimental inoculation with an unfiltered culture; but the blood of an animal killed by such an inoculation contains the *tetanus* poison, and when injected into a mouse causes its death with tetanic symptoms.

When a platinum needle is dipped into a pure culture of the *tetanus bacillus* and a mouse is inoculated with it subcutaneously, the animal invariably falls sick within twenty-four hours and dies of typical *tetanus* in two or three days. Rats, guinea-pigs, and rabbits

are killed in the same way by somewhat larger quantities—0.3 to 0.5 cubic centimetre (Kitasato). Pigeons are very slightly susceptible. The tetanic symptoms are first developed in the vicinity of the point of inoculation; if the animal is inoculated in the posterior portion of the body the hind legs first show tetanic contraction, if in the fore part of the body the muscles of the neck are first affected. At the autopsy there is a certain amount of hyperæmia at the point of inoculation, but no pus is formed; in inoculations with garden earth, or accidental inoculations in man, pus is commonly found in the vicinity of the inoculation wound. The various organs are normal in appearance. Kitasato says that he has not been able to demonstrate the presence of the bacillus or of spores in the spinal marrow, the nerves, muscles, spleen, liver, lungs, kidneys, or blood from the heart; nor has he been able to obtain cultures from the various organs. In mice which were inoculated at the root of the tail Kitasato was able to demonstrate the presence of the bacilli at the point of inoculation by the microscopical examination of an excised piece of the tissues for eight to ten hours after the inoculation; later than this they were not found. In pus from the inoculation wounds of men and animals accidentally infected the bacilli are present, but the formation of spores does not always occur. According to Kitasato, the sooner death has occurred after accidental inoculation the less likely are spores to be found in the rods, but from pus in which no spores are seen cultures of the bacillus may be obtained in which spores will develop in the usual manner.

Guinea-pigs are even more susceptible to the tetanus poison than mice, and rabbits less so. The amount of filtrate from a slightly alkaline bouillon culture required to kill a mouse is extremely minute—0.00001 cubic centimetre (Kitasato). The tetanic symptoms are developed within three days; if the animal is not affected within four days it escapes entirely. The tetanus poison is destroyed by a temperature of 65° C. maintained for five minutes, or 60° for twenty minutes, or 55° for an hour and a half; in the incubating oven at 37° C. it gradually loses its toxic potency; in diffuse daylight, also, its toxic power is gradually lost; in a cool, dark place it retains its original potency indefinitely; in direct sunlight it is completely destroyed in from fifteen to eighteen hours; it is not injured by being largely diluted with distilled water; it is destroyed in an hour by hydrochloric acid in the proportion of 0.55 per cent, terchloride of iodine destroys it in the proportion of 0.5 per cent, cresol in 1 per cent—one hour's exposure. In general it is destroyed by acids and by alkalis. Blood serum from cattle, horses, sheep, rabbits, rats, or guinea-pigs does not modify its toxic properties.

Recent researches by Tizzoni and Cattani show that tetanus spores preserved upon silk threads become attenuated after a time when preserved in a dark place in free contact with the air. Very virulent cultures liquefy gelatin, give off a very disagreeable odor, and have a decidedly alkaline reaction. Less virulent cultures quickly acquire an acid reaction. Cultures of which the virulence is very much attenuated grow more rapidly and abundantly than virulent cultures and produce more gas—in hydrogen at 37° C.; they do not liquefy gelatin and have no odor. In attenuated cultures degeneration forms are often seen, and the spores are frequently elongated or almost rod-shaped. Cultures preserved in various gases for thirteen to fourteen months invariably become attenuated.

Immunity.—Kitasato was not able to produce immunity in mice by inoculations with minute doses of the poison, or with a filtrate which had been exposed to various degrees of temperature by which its activity was diminished or destroyed. But immunity lasting for about two months was produced in rabbits by inoculating them with the filtrate from a culture of the tetanus bacillus and subsequently, in the same locality, with three cubic centimetres of a one-per-cent solution of terchloride of iodine; this last solution was injected subcutaneously in the same dose at intervals of twenty-four hours for five days. Of fifteen rabbits treated in this way six proved to be immune against large doses of a virulent culture of the tetanus bacillus. The same treatment was not successful in producing immunity in mice or guinea-pigs, but the important discovery was made that a small quantity of blood (0.2 cubic centimetre) from an immune rabbit, when injected into the abdominal cavity of a mouse, gave it immunity from the effects of inoculations with the tetanus bacillus. Moreover, mice which were first inoculated with a virulent culture of the bacillus, and, after tetanic symptoms had appeared, received in the cavity of the abdomen an injection of blood serum from an immune mouse, were preserved from death. The power of the blood of an immune animal to neutralize the tetanus poison was further shown by mixing the filtrate from a virulent culture with blood serum from an immune animal and allowing it to stand for twenty-four hours; a dose three hundred times greater than would have sufficed to kill a mouse proved to be without effect after such admixture with blood serum—as before stated, the blood serum of animals which are not immune has no effect upon the poison. The duration of immunity induced in this way was from forty to fifty days. Blood serum from an immune rabbit, preserved in a cool, dark room, retains its power of neutralizing the tetanus poison for about a week, after which time it gradually loses it. Having found that chickens have a natural immunity against tetanus, Kitasato made experiments

to ascertain whether their blood serum would also neutralize the tetanus poison; the result was negative.

That the tetanus poison is present in the blood of individuals who die from tetanus has been proved by Kitasato by injecting a small quantity (0.2 to 0.3 cubic centimetre) of blood from the heart of a fresh cadaver into mice; the animals develop typical tetanic symptoms and die in from twenty hours to three days.

Tizzoni and Cattani have recently (1891) reported results similar to those obtained by Kitasato. By repeated inoculations with gradually increasing doses of the tetanus poison they succeeded in making a dog and two pigeons immune, and found that blood serum from this immune dog, in very small amount, completely destroyed the toxic power of a filtrate from cultures of the tetanus bacillus—one to two drops of serum neutralized 0.5 cubic centimetre of filtrate after fifteen to twenty minutes' contact. They also ascertained that small amounts of blood serum from this immune dog injected into other dogs or white mice produced immunity in these animals; but they were not able to produce immunity in guinea-pigs or rabbits by the same method.

In a later communication (May, 1891) Tizzoni and Cattani give an account of their experiments made with a view to determining the nature of the substance in the blood serum of an immune animal which has the power of destroying the toxalbumin of tetanus—"tetanus antitoxin." They found, in the first place, that this antitoxin in blood serum is destroyed in half an hour by a temperature of 68° C.; further, that it does not pass through a dialyzing membrane; that it is destroyed by acids and alkalies. As a result of their researches they conclude that it is an albuminous substance having the nature of an enzyme.

Vaillard has succeeded in producing immunity in rabbits by repeated injections into the circulation of filtered cultures—in all twenty cubic centimetres—which had been exposed for one hour to a temperature of 60° C. At a temperature of 65° C. both the toxic and the immunizing action is destroyed.

150. BACILLUS OEDEMATIS MALIGNI.

Synonyms.—*Bacillus of malignant cedema*; *Vibrio septique* (Pasteur).

Discovered by Pasteur (1877); carefully studied by Koch (1881). This bacillus is widely distributed, being found in the superficial layers of the soil, in dust, in putrefying substances, in the blood of animals which have been suffocated (by invasion from the intestine), in foul water, etc.

It may usually be obtained by introducing beneath the skin of a

rabbit or a guinea-pig a small quantity of garden earth. The animal dies within a day or two, and this bacillus is found in the bloody serum effused in the subcutaneous connective tissue for a considerable distance about the point of inoculation.

Morphology.—Bacilli from 3 to 3.5 μ long and 1 to 1.1 μ broad;



FIG. 162.—*Bacillus oedematis maligni*, from subcutaneous connective tissue of inoculated guinea-pig. $\times 950$. (Baumgarten.)

frequently united in pairs, or chains of three elements; may grow out into long filaments 15 to 40 μ long—these are straight, or bent at an angle, or more or less curved. They resemble the bacillus of anthrax, but are not quite as broad, have rounded ends, and in stained preparations the long filaments are not segmented as is the case with the anthrax bacillus. By Löffler's method of staining they are seen to have flagella arranged around the periphery of the cells. Large, oval spores may be developed in the bacilli (not in the long filaments), which are of greater diameter than the rods, and produce a terminal or central swelling of the same, according to the location of the spore.

Stains readily by the aniline colors usually employed, but is decolorized when treated by Gram's method. In stained preparations the long filaments may present a somewhat granular appearance from unequal action of the staining agent.

Biological Characters.—A strictly anaerobic, liquefying, motile bacillus. Forms spores. Grows in the usual culture media



FIG. 163.—*Bacillus oedematis maligni*, from an agar culture, showing spores. $\times 1,000$ From a photomicrograph. (Fränkel and Pfeiffer.)

when oxygen is excluded—in an atmosphere of hydrogen. Grows at the room temperature—better in the incubating oven at 37° C. The spores are formed most abundantly in cultures kept in the incubating oven, but may also be formed at a temperature of 20° C. In the bodies of animals which succumb to an experimental inoculation

no spores are found immediately after death, but the bacilli multiply rapidly in the cadaver, and form spores when the temperature is favorable.

The malignant-œdema bacillus may be cultivated in ordinary *nutrient gelatin*, but its development is more abundant when one to two per cent of grape sugar has been added to the culture medium. In deep stick cultures in this medium development occurs at first only near the bottom of the line of puncture; the gelatin is liquefied and has a grayish-white, clouded appearance; an abundant development of gas occurs, and as this accumulates the growth and liquefaction of the gelatin extend upward.

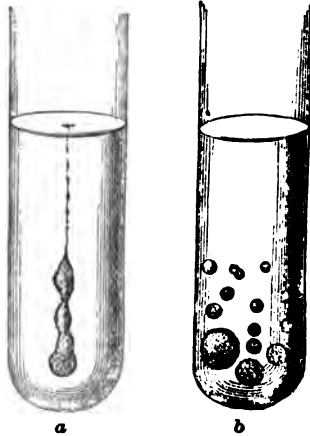


FIG. 164.—*Bacillus oedematis maligni*, cultures in nutrient gelatin; *a*, long stick culture; *b*, colonies at bottom of gelatin tube. (Flügge.)

when the bacilli are mixed in a test tube with gelatin which has been liquefied by heat, and which is then allowed to solidify. Spherical colonies are developed, in the course of two or three days, in the lower portion of the gelatin; these are filled with liquefied gelatin of a grayish-white color, and when examined with a low power are seen to be permeated with a network of filaments, while the periphery presents a radiate appearance. In *nutrient agar* growth also occurs at the bottom of a deep puncture; it has an irregular, jagged outline and a granular appearance; the considerable development at the deepest portion and gradual thinning out above give the growth a club shape; in the incubating oven there is an abundant development of gas, which often splits up the agar medium and forces the upper portion against the cotton stopper. An abundant development of gas also occurs in cultures in *blood serum*, and the medium is rapidly liquefied; at a temperature of 37° it is changed in a few days to a yellowish fluid, at the bottom of which some irregular, corroded fragments of the solidified serum may be seen. In *agar plates*, placed in a close receptacle from which oxygen is excluded, cloudy, dull-white colonies are formed which have irregular outlines and under the microscope are seen to be made up of branching and interlaced filaments radi-

ating from the centre. Cultures of the malignant-œdema bacillus give off a peculiar, disagreeable odor, which cannot, however, be designated as "putrefactive."

Pathogenesis.—Pathogenic for mice, guinea-pigs, rabbits, and, according to Kitt, for horses, dogs, goats, sheep, calves, pigs, chickens, and pigeons. According to Arloing and to Chauveau, cattle are immune. The disease is rarely developed except as a result of experimental inoculations, but horses occasionally have malignant œdema from accidental inoculation, and cases have been reported in man—"gangrène gazeuse." A small quantity of a pure culture injected beneath the skin of a susceptible animal gives rise to an extensive inflammatory œdema of the subcutaneous connective tissue and of the superficial muscles, which extends from the point of inoculation, especially towards the more dependent portions of the body. The bloody serum effused is without odor and contains little if any gas. But when malignant œdema results from the introduction of a little garden earth beneath the skin of a guinea-pig or other susceptible animal, the effused serum is frothy and has a putrefactive odor, no doubt from the presence of associated bacteria. Injections into the circulation do not give rise to malignant œdema, unless at the same time some bacilli are thrown into the connective tissue. While small animals usually die from an experimental inoculation with a moderately small quantity of a pure culture, larger ones (dogs, sheep) frequently recover. At the autopsy, if made at once, the bacilli are found in great numbers in the effused serum, but not in blood from the heart or in preparations made from the parenchyma of the various organs; later they may be found in all parts of the body as a result of post-mortem multiplication. This applies to rabbits and to guinea-pigs, but not to mice; in these little animals the bacilli may find their way into the blood during the last hours of life, and their presence may be demonstrated in smear preparations of blood from the heart or from the parenchyma of the spleen or liver. In mice the spleen is considerably enlarged, dark in color, and softened; in rabbits and guinea-pigs less so. With this exception the internal organs present no very notable pathological changes.

Animals which recover from malignant œdema are said to be subsequently immune (Arloing and Chauveau). Roux and Chamberlain have shown that immunity may be induced in guinea-pigs by injecting filtered cultures of the malignant-œdema bacillus (about one hundred cubic centimetres of a bouillon culture in three doses) into the abdominal cavity; or, better still, by the injection of filtered serum from animals which have recently succumbed to an experimental inoculation (one cubic centimetre repeated daily for even or eight days).

151. *BACILLUS CADAVERIS*.

Obtained by the writer (1899) from pieces of liver and kidney, from yellow-fever cadavers, which had been preserved for forty-eight hours in an antiseptic wrapping, at the summer temperature of Havana; also in two

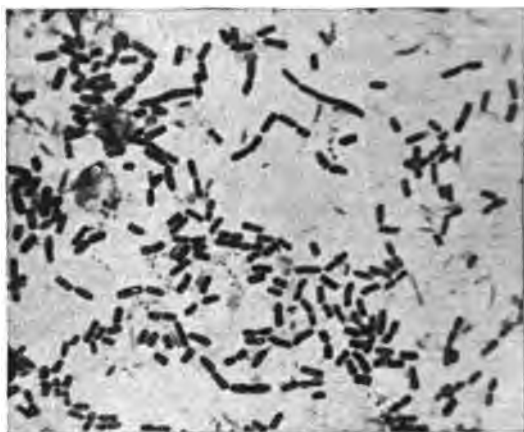


FIG. 165.—*Bacillus cadaveris*; smear preparation from liver of yellow-fever cadaver, kept twenty-four hours in an antiseptic wrapping. $\times 1,000$. From a photomicrograph. (Sternberg.)

cases from pieces of yellow-fever liver immediately after the autopsy; also from liver preserved in an antiseptic wrapping from comparative autopsies made in Baltimore.

Morphology.—Large bacilli with square or slightly rounded corners, from 1.5 to $4\ \mu$ in length and about $1.2\ \mu$ broad; frequently associated in pairs; may grow out into straight or slightly curved filaments of from 5 to $15\ \mu$ in length.

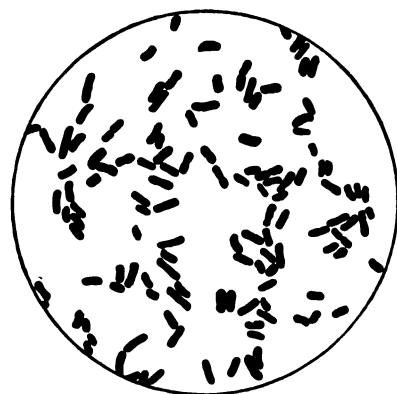


FIG. 166.—*Bacillus cadaveris*, from an anaerobic culture in glycerin-agar. $\times 1,000$. From a photomicrograph. (Sternberg.)

Biological Characters.—An anaerobic, non-motile bacillus; not cultivated in nutrient gelatin; not observed to form spores.

Bacillus cadaveris is a strict anaerobic and is difficult to cultivate. I have succeeded best with nutrient agar containing five per cent of glycerin, removing the oxygen thoroughly by passing a stream of hydrogen through the liquefied medium. The colonies in a glycerin-agar roll tube (containing hydrogen and hermetically sealed) are opaque, irregular in outline, granular, and of a white color by reflected light. The culture medium acquires an acid reaction as a result of the development of the bacillus.

Liver tissue containing this bacillus, after having been kept in an antiseptic wrapping for forty-eight hours, has a fresh appearance, a very acid reaction, and is without any putrefactive odor.

Pathogenesis.—Liver tissue containing this bacillus is very pathogenic for guinea-pigs when injected subcutaneously, and causes an extensive inflammatory oedema extending from the point of inoculation. Pure cultures of the bacillus are less pathogenic, and the few experiments which I made in Havana gave a somewhat contradictory result, recovery having occurred in one guinea-pig which received a subcutaneous injection of ten minims of liquid from an anaerobic culture in glycerin-agar, while another died at the end of twenty hours from a subcutaneous injection of three minims, with extensive inflammatory oedema in the vicinity of the point of inoculation.

152. BACILLUS OF SYMPTOMATIC ANTHRAX.

Synonyms.—Rauschbrandbacillus, *Ger.*; Bacille du charbon symptomatique, *Fr.*

First described by Bollinger and Feser (1878); carefully studied and its principal characters determined by Arloing, Cornevin, and Thomas (1880–83).

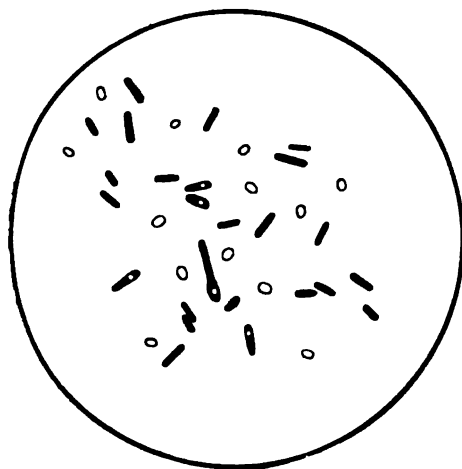


FIG. 167.

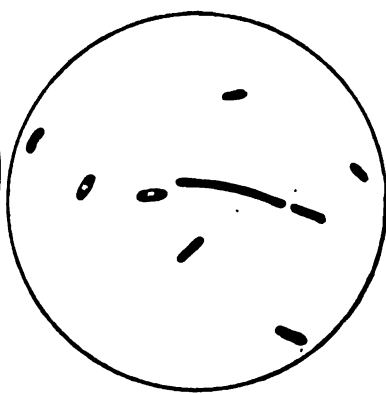


FIG. 168.

FIG. 167.—Bacillus of symptomatic anthrax, from an agar culture. $\times 1,000$. From a photomicrograph. (Frinkel and Pfeiffer.)

FIG. 168.—Bacillus of symptomatic anthrax, from muscles of inoculated guinea-pig. From a photomicrograph. (Roux.)

Found in the affected tissues of animals—principally cattle—suffering from “black leg,” “quarter evil,” or symptomatic anthrax (*Fr.*, “charbon symptomatique”; *Ger.*, “Rauschbrand”). The disease prevails during the summer months in various parts of Europe, and is characterized by the appearance of irregular, emphysematous swellings of the subcutaneous tissue and muscles, especially over the quarters, hence the name “quarter evil.” The muscles in the

affected areas have a dark color and contain a bloody serum in which the bacillus is found.

Morphology.—Bacilli with rounded ends, from three to five μ long and 0.5 to 0.6 μ broad; sometimes united in pairs, but do not grow out into filaments. The spores are oval, somewhat flattened on one side, thicker than the bacilli, and lie near the middle of the rods, but a little nearer to one extremity. The bacilli containing spores are somewhat spindle-formed (Kitasato). “Involution forms” are quite common in old cultures or in unfavorable media; in such cultures variously distorted and often greatly enlarged bacilli may be seen, some being greatly swollen in the middle — spindle-shaped. When properly stained, by Löffler’s method, a number of flagella are seen around the periphery of the cells.

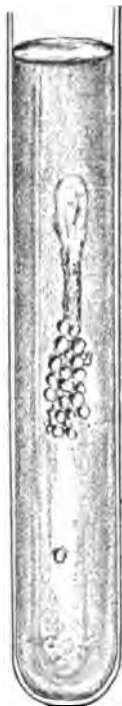


FIG. 169. — Bacillus of symptomatic anthrax; long stick culture in nutrient gelatin, ten days at 18°–20° C. (Kitasato.)

Stains with the aniline colors usually employed, but not by Gram’s method. Spore-bearing bacilli may be double-stained by first staining the spores by Ziehl’s method, and then the bacilli with a solution of methylene blue.

Biological Characters.—An anaerobic, liquefying, motile bacillus. *Forms spores.* Grows at the room temperature in the usual culture media, in the absence of oxygen, in an atmosphere of hydrogen, but not in carbon dioxide. This bacillus grows more rapidly and abundantly in nutrient agar or gelatin to which 1.5 to 2 per cent of grape sugar or five per cent of glycerin has been added. Colonies in gelatin, in an atmosphere of hydrogen, are at first spherical, with irregular outlines and a wart-like surface; later the gelatin is liquefied around them, and radiating filaments grow out into the gelatin, so that by transmitted light they present the appearance of an opaque central mass with an irregular surface surrounded by rays. In stick cultures in *nutrient gelatin*, at 20° to 25° C., at the end of two or three days

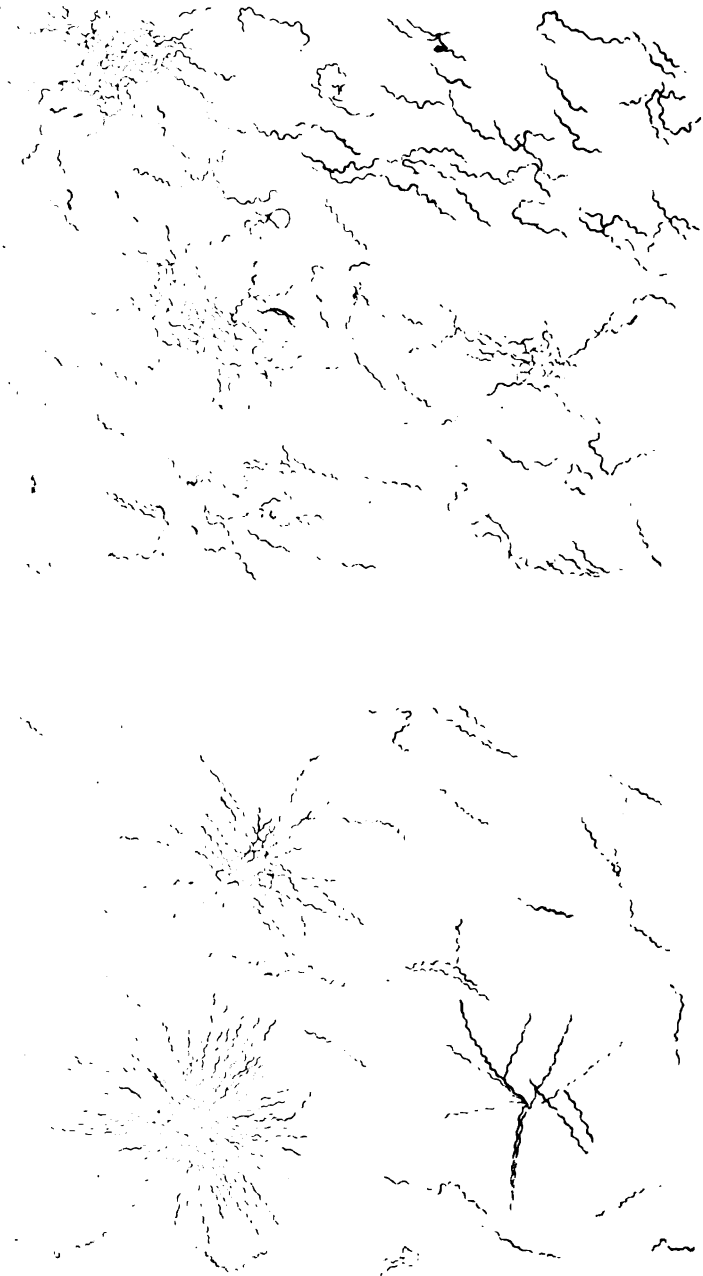
development occurs at the bottom of the line of puncture to within about two fingers’ breadth of the surface; the gelatin is slowly liquefied and considerable gas is formed. In old cultures the growth and liquefaction of the gelatin extend nearly to the surface. In *agar stick cultures*, in the incubating oven, development begins within a day or two and extends to within one finger’s breadth of the surface; considerable gas is evolved, and

the cultures have a peculiar, acid, penetrating odor. Development is most rapid at 36° to 38° C., but may occur at a temperature of 16° to 18° C.—not lower than 14°. Spores are quickly formed in cultures kept in the incubating oven—not so quickly at the room temperature. These withstand a temperature of 80° C. maintained for an hour, but are killed in five minutes by a temperature of 100° C. (in steam). In the bodies of infected animals spores are not formed until after the death of the animal, at the end of twenty-four to forty-eight hours (Kitasato).

The spores are destroyed by a five-per-cent solution of carbolic acid in ten hours, and the bacilli, in the absence of spores, in five minutes; a 1:1,000 solution of mercuric chloride destroys the spores in two hours (Kitasato). According to Kitasato, certain shining bodies of irregular form, which stain readily with the aniline colors, are to be seen in the rods as they are found in the bloody serum from an animal recently dead; but these are not spores, as some bacteriologists have supposed.

Pathogenesis.—Cattle, which are immune against malignant œdema, are most subject to infection by the bacillus of symptomatic anthrax, and the disease produced by this anaerobic bacillus prevails almost entirely among them; horses are not attacked spontaneously—i.e., by accidental infection—and when inoculated with a culture of this bacillus present only a limited local reaction. Swine, dogs, rabbits, fowls, and pigeons have but slight susceptibility, but the researches of Arloing, Cornevin, and Thomas, and of Roger show that by the addition of a twenty-per-cent solution of lactic acid to a culture its virulence is greatly increased, and animals which have but little susceptibility, like the rabbit or the mouse, succumb to such injections; similar results were obtained by Roger by the simultaneous injection of sterilized or non-sterilized cultures of *Bacillus prodigiosus* or of *Proteus vulgaris*. The guinea-pig is the most susceptible animal. When inoculated subcutaneously with a small quantity of a pure culture, or with spores attached to a silk thread, it dies in from twenty-four to thirty-six hours. At the autopsy a bloody serum is found in the subcutaneous tissues in the vicinity of the point of inoculation, and the muscles present a dark-red or black appearance similar to that in cattle affected with “black leg.” The internal organs present no notable pathological changes. Immediately after death the bacilli are found only in the effused serum and the affected tissues near the point of inoculation, but later they multiply in the cadaver and are found throughout the body. According to Kitasato, the cultures in solid media preserve their virulence for an indefinite period, but cultures in a bouillon made from the flesh of guinea-pigs soon lose their virulence. Cultures are readily attenuated by heat

according to the method of Toussaint and Chauveau ; a temperature of 42° to 43° C. is suitable. The pathogenic virulence of spores may also be attenuated by subjecting them to dry heat—a temperature of 80° to 100° C. maintained for several hours. For the production of immunity in cattle Arloing, Cornevin, and Thomas recommend the use of a dried powder of the muscles of animals which have succumbed to the disease, and which has been subjected to a suitable temperature to insure attenuation of the pathogenic virulence of the spores contained in it. Kitt, who has made extended experiments with this bacillus, recommends that the muscles be first dried at 32° to 35° C. and then powdered. Two vaccines are then prepared—a stronger vaccine by exposure of a portion of the powder to a temperature of 85° to 90° C. for six hours, and a weaker vaccine by exposure for six hours to a temperature of 100° to 104° C. (dry heat). Inoculations made with this attenuated virus—the weakest first and subsequently the least attenuated—give rise to a local reaction of moderate intensity, and the animal is subsequently immune from the effects of the most virulent material. Immunity may also be secured by intravenous inoculations ; or, in guinea-pigs, by inoculations with bouillon cultures which have been kept for a few days and as a result have lost their original virulence, or with cultures kept in an incubating oven at a temperature of 42° to 43° C.; or by inoculation with a very minute quantity of a pure culture ; or by an inoculation made into the extremity of the tail ; or by inoculations with filtered cultures (Roux and Chamberlain), or with cultures sterilized by heat (Kitasato). It has been claimed (Roux) that animals which have been made immune against symptomatic anthrax are also immune against malignant cedema. But in a carefully conducted series of experiments Kitasato was unable to confirm this ; he found that guinea-pigs which had an immunity against the most virulent cultures of the Rauschbrand bacillus succumbed invariably to malignant cedema when inoculated subcutaneously with the bacillus of malignant cedema.



**Spirillum Obermeieri in blood of two monkeys,
inoculated after removal of spleen.
(Soudakewitch).**

XV.

PATHOGENIC SPIRILLA.

153. SPIRILLUM OBERMEIERI.

Synonyms.—Spirochæte Obermeieri; Spirillum of relapsing fever; Die Recurrensspirochæte.

Discovered by Obermeier (1873) in the blood of persons suffering from relapsing fever.

This spirillum is present, in very great numbers, in the blood of relapsing-fever patients during the febrile paroxysms. It has not been found under any other circumstances, and its etiological relation to the disease with which it is associated is generally admitted.

Morphology.—Very slender, flexible, spiral or wavy filaments, with pointed ends; from sixteen to forty μ in length and considerably thinner than the cholera spirillum—about 0.1 μ . Koch has demonstrated the presence of flagella (Eisenberg).

Stains readily with the aniline colors, especially with fuchsin, Bismarck brown, and in Löffler's solution of methylene blue.

Biological Characters.—An *aërobic*, *motile* spirillum which has not been cultivated in artificial media. This spirillum appears to be a strict parasite, whose habitat is the blood of man. The disappearance of the parasite from the blood soon after the termination of a febrile paroxysm, and its reappearance during subsequent paroxysms, have led to the inference that it must form spores, but this has not been demonstrated. In fresh preparations from the blood the spirillum exhibits active progressive movements, accompanied by very rapid rotation in the long axis of the spiral filaments, or by undulatory movements. The movements are so vigorous that the comparatively large red blood corpuscles are seen, under the microscope, to be thrown about by the slender spiral filaments, which it is difficult to see in unstained preparations. When preserved in a one-half-per-cent salt solution they continue to exhibit active movements for a considerable time. Efforts to cultivate this spirillum in artificial media have thus far been unsuccessful, although Koch has observed an increase in the length of the spirilla and the formation of a tangled mass of filaments.

In experiments made by Heydenreich the spirillum was found to preserve its vitality (motility) for fourteen days at a temperature of

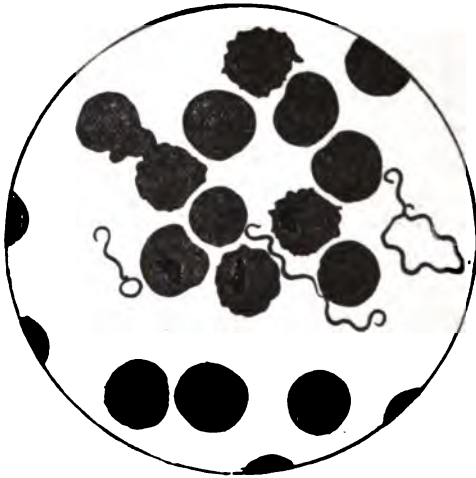


FIG. 170.—*Spirillum Obermeieri* in blood of man. $\times 1,000$. From a photomicrograph. (Fränkel and Pfeiffer.)

16° to 22° C., for twenty hours at 37° , and at 42.5° for two or three hours only.

Pathogenesis.—Causes in man the disease known as relapsing fever. Münch and Moczutkowsky have produced typical relapsing

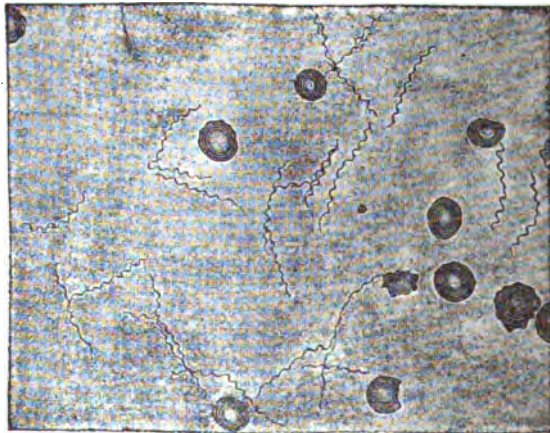


FIG. 171.—*Spirillum Obermeieri* in blood of an inoculated ape. $\times 700$. (Koch.)

fever in healthy persons by inoculating them with blood containing the spirillum of Obermeier. The spirilla are found in the blood during the febrile paroxysm, and for a day or two, at the outside, after

its termination ; sometimes they are present in great numbers, and at others can only be found by searching several microscopic fields; they are not present in the various secretions—urine, sweat, saliva, etc. In fatal cases the principal pathological changes are found in the spleen, which is greatly enlarged, and in the liver and marrow of the bones, which contain inflammatory and necrotic foci. Koch and Carter have succeeded in transmitting the disease to monkeys by subcutaneous inoculations with small amounts of defibrinated blood containing the spirillum. After an incubation period of several days typical febrile paroxysms were developed, during which the actively motile spirilla were found in the blood in large numbers. Blood from one animal, taken during the attack, induced a similar febrile paroxysm when inoculated into another of the same species—relapses, such as characterize the disease in man, were not observed. One attack did not preserve the animals experimented upon from a similar attack when they were again inoculated after an interval of a few days. Recently Soudakewitch (1891) has made successful inoculation experiments in monkeys, and has shown that in monkeys from which the spleen has previously been removed the spirilla continue to multiply very abundantly in the blood and the disease has a fatal termination, whereas in monkeys from which the spleen has not been removed the spirilla disappear from the blood within a few days after the access of the febrile paroxysm and the animal recovers.

154. SPIRILLUM ANSERUM.

Synonym.—*Spirochaeta anserina* (Sakharoff).

Obtained by Sakharoff (1890) from the blood of geese affected by a fatal form of septicæmia due to this spirillum. This disease prevails among geese in Caucasia, especially in swampy regions, appearing annually and destroying a large number of the domestic geese.

Morphology.—Resembles the spirillum of relapsing fever. The long and flexible spiral filaments, when the disease is at its height, are often seen in interlaced masses, around the margins of which radiate single filaments which by their movements cause the whole mass to change its place, as if it were a single organism. These masses are sometimes so large that a single one occupies the entire field of the microscope.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic*, *motile* spirillum. Not cultivated in artificial media. The movements are very active, resembling those of *Spirillum Obermeieri*, but cease in an hour or two in preparations made from the blood of geese containing it.

Pathogenesis.—A small quantity of blood from an infected goose inoculated into a healthy animal of the same species induces the disease after a period of incubation of four to five days. The infected goose ceases to eat, becomes apathetic, remaining in one place, and usually dies at the end of a week ; the temperature is increased, and in some cases there is diarrhoea. The spirilla are found in the blood at the outset of the malady, but after death they are not seen either in the blood or in the various organs. The heart and the liver are found to have undergone a fatty degeneration, and yellowish, cheesy granules the size of a millet seed are seen upon the surface of these organs. The spleen is soft and easily broken up by the fingers.

Inoculations into chickens and pigeons were without result; in one chicken the spirilla were found in the blood on the fourth day after inoculation, but the fowl recovered.

155. *SPIRILLUM CHOLERÆ ASIATICÆ.*

Synonyms.—*Spirillum* ("bacillus") of cholera; *Comma bacillus* of Koch; *Kommabacillus der Cholera Asiaticæ*; *Bacille-virgule cholérigène*.

Discovered by Koch (1884) in the excreta of cholera patients and in the contents of the intestine of recent cadavers.

The researches of Koch, made in Egypt and in India (1884), and subsequent researches by bacteriologists in various parts of the world, show that this spirillum—so-called "*comma bacillus*"—is constantly present in the contents of the intestine of cholera patients during the height of the disease, and that it is not found in the contents of the intestine of healthy persons or of those suffering from



FIG. 172.

FIG. 172.—*Spirillum cholerae Asiaticæ*. $\times 1,000$. From a photomicrograph. (Koch.)



FIG. 173.

FIG. 173.—*Spirillum cholerae Asiaticæ*, involutic forms. $\times 700$. (Van Ermengem.)

other diseases than cholera. The etiological relation of this spirillum to Asiatic cholera is now generally admitted by bacteriologists.

Morphology.—Slightly curved rods with rounded ends, from 0.5 to 2μ in length and about 0.3 to 0.4μ in breadth. The rods are usually but slightly curved, like a comma, but are occasionally in the form of a half-circle, or two united rods curved in opposite directions may form an S-shaped figure. Under certain circumstances the curved rods grow out into long, spiral filaments, which may consist of numerous spiral turns, and in hanging-drop cultures the S-shaped figures may also be seen to form the commencement of a spiral; in stained preparations the spiral character of the long filaments is often obliterated, or nearly so. When development is very rapid the short, curved rods or S-shaped spirals only are seen; but in hanging-drop cultures, or in media in which the develop.

ment is retarded by an unfavorable temperature, the presence of a little alcohol, etc., the long, spiral filaments are quite numerous, and bacteriologists generally agree that the so-called "comma bacillus" is really only a fragment of a true spirillum. By Löffler's method of staining the rods may be seen to have a single terminal flagellum. In old cultures the bacilli frequently lose their characteristic form and become variously swollen and distorted—involution forms. Hueppe has described the appearance of spherical bodies in the course of the spiral filaments, which he believes to be reproductive elements—so-called arthrospores.

Stains with the aniline colors usually employed, but not as quickly as many other bacteria; an aqueous solution of fuchsin is the



FIG. 174.

FIG. 174.—*Spirillum cholerae Asiaticae*; colonies upon gelatin plate, end of thirty hours. $\times 100$. Photograph by Fränkel and Pfeiffer.



FIG. 175.

FIG. 175.—*Spirillum cholerae Asiaticae*, from a gelatin culture. $\times 1,000$. From a photomicrograph. (Fränkel and Pfeiffer.)

most reliable staining agent; is decolorized by iodine solution—Gram's method. Sections may be stained with Löffler's solution.

Biological Characters.—An *aërobic* (facultative anaërobic), *liquefying*, *motile* spirillum. Grows in the usual culture media at the room temperature—more rapidly in the incubating oven. Does not grow at a temperature above 42° or below 14° C. Does not form endogenous spores (forms arthrospores, according to Hueppe?).

In *gelatin plate cultures*, at 22° C., at the end of twenty-four hours small, white colonies may be perceived in the depths of the gelatin; these grow towards the surface and cause liquefaction of the gelatin in the form of a funnel which gradually increases in

depth, and at the bottom of which is seen the colony in the form of a small, white mass; as a result of this the plates on the second or third day appear to be perforated with numerous small holes; later

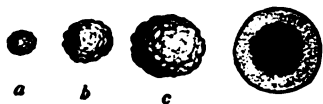


FIG. 176.—Colonies of the cholera spirillum; a, end of twenty hours; b, end of thirty hours; c, end of forty-eight hours; d, after liquefaction of the gelatin. (Flügge.)

the gelatin is entirely liquefied. Under a low power the young colonies, before liquefaction has commenced, present a rather characteristic appearance; they are of a white or pale-yellow color, and have a more or less irregular outline. the margins being rough and uneven: the texture is coarsely granular, and the surface looks as if it were covered with little fragments of broken glass, while

the colony has a shining appearance; when liquefaction commences an ill-defined halo is first seen to surround the granular colony, which by transmitted light has a peculiar roseate hue. In *stick cultures* in nutrient gelatin development occurs all along the line of inoculation.

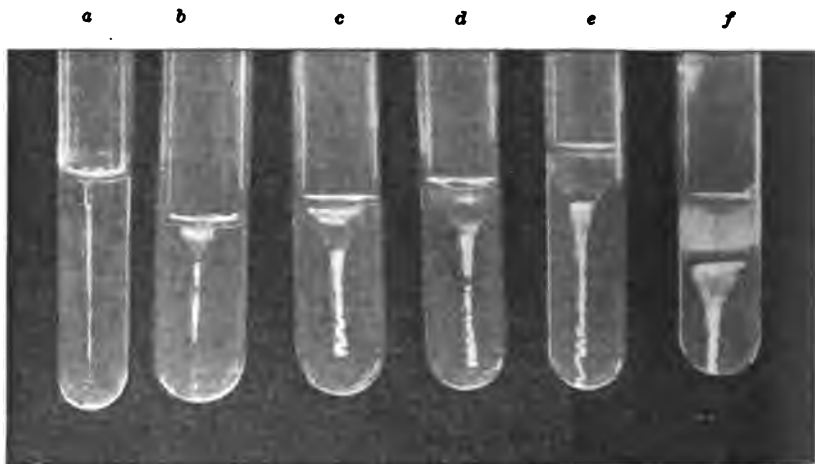


FIG. 177.—*Spirillum cholerae Asiaticae*; a, one day old; b, three days old; c, four days old; d, five days old; e, seven days old; f, ten days old. From photographs by Koch.

but liquefaction of the gelatin first occurs only near the surface; on the second day, at 22° C., a short funnel is formed which has a comparatively narrow mouth, and the upper portion of which contains air, while just below this is a whitish, viscid mass; later the funnel increases in depth and diameter, and at the end of from four to six days may reach the edge of the test tube; in from eight to fourteen days the upper two-thirds of the gelatin is completely liquefied. Owing to the slight liquefaction which occurs along the line of growth during the first three or four days, the central mass which

had formed along the line of inoculation settles down as a curled or irregularly bent, yellowish-white thread in the lower part of a slender tube filled with liquefied gelatin, the upper part of which widens out and is continuous with the funnel above. Upon the surface of *nutrient agar* a moist, shining, white layer is formed along the line of inoculation—impfstrich. *Blood serum* is slowly liquefied by this spirillum. Upon the surface of cooked *potato*, in the incubating oven, a rather thin and semi-transparent brown or grayish-brown layer is developed. In *bouillon* the development is rapid and abundant, especially in the incubating oven ; the fluid is only slightly

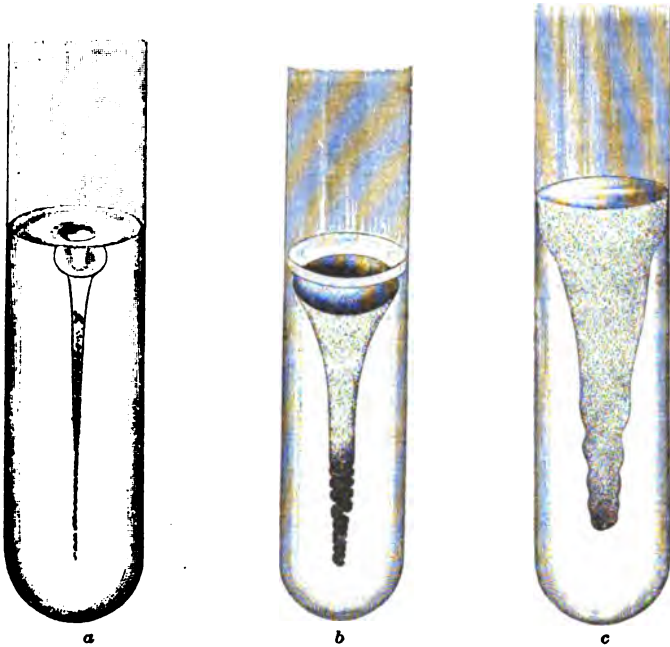


FIG. 178.—Cultures in nutrient gelatin, at the room temperature (16° to 18° C.), at the commencement of the fourth day; a, *Spirillum cholerae Asiaticae*; b, *Spirillum tyrogenum*; c, *Spirillum* of Finkler and Prior. (Baumgarten.)

clouded, but the spirilla accumulate at the surface, forming a wrinkled membranous layer. *Sterilized milk* is also a favorable culture medium. In general this spirillum grows in any liquid containing a small quantity of organic pabulum and having a slightly alkaline reaction. An acid reaction of the culture medium prevents its development, as a rule, but it has the power of gradually accommodating itself to the presence of vegetable acids, and grows upon potatoes—in the incubator only—which have a slightly acid reaction. Abundant development occurs in bouillon which has been diluted with eight or ten parts of water, and the experiments of Wolffhügel

and Riedel show that it also multiplies to some extent in sterilized river or well water, and that it preserves its vitality in such water for several months. But in milk or water which contains other bacteria it dies out in a few days. Gruber and Schottelius have shown, however, that in bouillon which is greatly diluted the cholera spirillum may take the precedence of the common saprophytic bacteria, and that they form upon the surface of such a medium the characteristic wrinkled film. Koch found in his early investigations that rapid multiplication may occur upon the surface of moist linen, and also demonstrated the presence of this spirillum in the foul water of a "tank" in India which was used by the natives for drinking purposes. In the experiments of Bolton (1886) the cholera spirillum was found to multiply abundantly in distilled water to which bouillon was added in the proportion of fifteen to twenty-five parts in one thousand.

The thermal death-point of the cholera spirillum in recent cultures in flesh-peptone-gelatin, as determined by the writer (1887), is 52° C., the time of exposure being four minutes; a few colonies only developed after exposure to a temperature of 50° for ten minutes. In Kitasato's experiments (1889) ten or even fifteen minutes' exposure to a temperature of 55° C. was not always successful in destroying the vitality of the spirillum, although in certain cultures exposure to 50° for fifteen minutes was successful. He was not, however, able to find any difference between old and recent cultures as regards resistance to heat or to desiccation. In a moist condition this spirillum retains its vitality for months—as much as nine months in agar and about two months in liquefied gelatin. It is quickly destroyed by desiccation, as first determined by Koch, who found that it did not grow after two or three hours when dried in a thin film on a glass cover. In Kitasato's experiments (1889) the duration of vitality was found to vary from a few hours to thirteen days, the difference depending largely upon the thickness of the film. When dried upon silk threads they may retain their vitality for a considerably longer time (Kitasato). Very numerous experiments have been made to determine the amount of various disinfecting agents required to destroy the vitality of this microorganism. We give below the results recently reported by Boer (1890), whose experiments were made in Koch's laboratory. Experiments upon a culture in bouillon kept for twenty-four hours in the incubating oven, time of exposure two hours: hydrochloric acid, 1:1,350; sulphuric acid, 1:1,300; caustic soda, 1:150; ammonia, 1:350; mercuric cyanide, 1:60,000; gold and sodium chloride, 1:1,000; silver nitrate, 1:4,000; arsenite of soda, 1:400; malachite green, 1:5,000; methyl violet, 1:1,000; carbolic acid, 1:400; creolin, 1:3,000; lysol, 1:500. In

Bolton's experiments (1887) mercuric chloride was effective in two hours in the proportion of 1:10,000; sulphate of copper, 1:500.

The low thermal death-point and comparatively slight resisting power for desiccation and chemical agents indicate that this spirillum does not form spores, and most bacteriologists agree that this is the case. Hueppe, however, has described a mode of spore formation which is different from that which occurs among the bacilli, viz., the formation of so-called arthrospores; these are said to be developed in the course of the spiral threads, not as endogenous refractive spores, but as spherical bodies which have a somewhat greater diameter than the filament and are somewhat more refractive. This mode of spore formation has not been observed by Kitasato and other bacteriologists who have given attention to the question, and cannot be considered as established. In competition with the ordinary putrefactive bacteria the cholera spirillum soon disappears, and, as determined by Neffelman and by Kitasato, they only survive for a few days when mixed with normal fæces.

A test for the presence of the cholera spirillum has been found by Bujwid and by Dunham in the reddish-violet color produced in bouillon cultures containing peptone, or in cultures in nutrient gelatin, when a small quantity of sulphuric acid is added to the culture. According to Fränkel, this test serves to distinguish it from the ordinary bacteria of the intestine and from the Finkler-Prior spirillum, but not from Metschnikoff's spirillum ("vibrio"). The reaction is shown by bouillon cultures which have been in the incubating oven for ten or twelve hours, and by gelatin cultures in which liquefaction has occurred. The sulphuric acid used should be quite pure; the color quickly appears and is reddish-violet or purplish-red. According to Salkowski, the red color is due to the well-known indol reaction, which in cultures of the cholera spirillum is exceptionally intense and rapid in its development. A test which is said to distinguish cultures of the cholera spirillum from the spirillum of Deneke and that of Finkler-Prior, has been proposed by Cahen. This consists in adding a solution of litmus to the bouillon and in making the culture at 37° C. The cholera cultures show on the following day a decoloration which does not occur at this temperature with the other spirilla named.

For determining as promptly as possible whether certain suspected excreta contain cholera spirilla, a little of the material may be used to inoculate greatly diluted bouillon, gelatin plates being made at the same time. At the end of ten or twelve hours the cholera spirillum, if present, will already have formed a characteristic wrinkled film upon the surface; a little of this should be used to start a new culture in diluted bouillon, and a series of gelatin plates made from

it, after which the color test may be applied. The result of this, in connection with the morphology of the microorganisms forming the film and the character of growth in the gelatin plates, will establish the diagnosis if the cholera spirillum is present in considerable numbers. If but few are present in the original material it may be necessary to make two or more series of plates and bouillon cultures before a pure culture can be obtained and a positive diagnosis made.

Brieger has succeeded in isolating several toxic ptomaines from cultures of the cholera bacillus, some of which had previously been obtained from other sources—cadaverin, putrescin, creatinin, methyl-guanidin. In addition to these he obtained two toxic substances not previously known. One of these is a diamin, resembling trimethylenediamin; it gave rise to cramps and muscular tremor in inoculated animals. The other poison reduced the frequency of the heart's action and the temperature of the body in the animals subjected to experiment. In more recent researches made by Brieger and Fränkel (1890) a toxalbumin was obtained from cholera cultures which, when injected subcutaneously into guinea-pigs, caused their death in two or three days, but had no effect upon rabbits.

Pfeiffer has recently (1892) published his extended researches relating to the cholera poison. He finds that recent aerobic cultures of the cholera spirillum contain a specific toxic substance which is fatal to guinea-pigs in extremely small doses. This substance stands in close relation with the bacterial cells and is perhaps an integral part of the same. The spirilla may be killed by chloroform, thymol, or by desiccation without apparent injury to the toxic potency of this substance. It is destroyed, however, by absolute alcohol, by concentrated solutions of neutral salts, and by the boiling temperature, and secondary toxic products are formed which have a similar physiological action but are from ten to twenty times less potent.

Similar toxic substances were obtained by Pfeiffer from cultures of Finkler-Prior's spirillum and from *Spirillum Metschnikovi*. The spirillum is not found in the blood or in the various organs of individuals who have succumbed to an attack of cholera, but it is constantly found in the alvine discharges during life and in the contents of the intestine examined immediately after death; frequently in almost a pure culture in the colorless "rice-water" discharges. It is evident, therefore, that if we accept it as the etiological agent in this disease, the morbid phenomena must be ascribed to the absorption of toxic substances formed during its multiplication in the intestine. In cases which terminated fatally after a very brief sickness Koch found but slight changes in the mucous membrane of the intestine, which was slightly swollen and reddened; but in more protracted cases the follicles and Peyer's patches were reddened around their margins, and

an invasion of the mucous membrane by the "comma bacilli" was observed in properly stained sections; they penetrated especially the follicles of Lieberkühn, and in some cases were seen between the epithelium and basement membrane. As a rule, the spirillum is not present in vomited matters, but Koch found it in small numbers in two cases and Nicati and Rietsch in three. In about one hundred cases in which Koch examined the excreta, or the contents of the intestine of recent cadavers, during his stay in Egypt, in India, and in Toulon, his "comma bacillus" was constantly found, and other observers have fully confirmed him in this particular—Nicati and Rietsch in thirty-one cases examined at Marseilles; Pfeiffer, twelve cases in Paris; Schottelius in cases examined in Turin; Ceci in

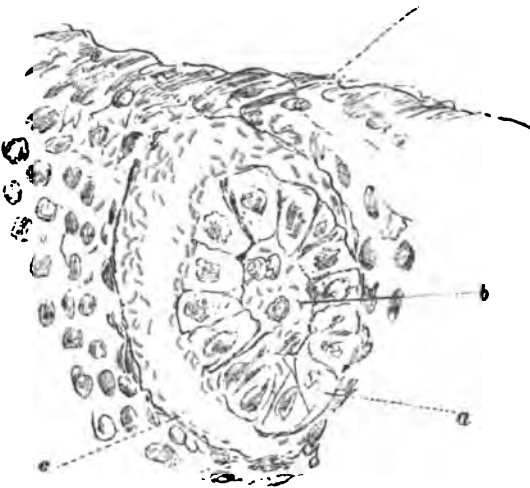


FIG. 179.—Section through mucous membrane of intestine from cholera cadaver; a tubular gland (a) is cut obliquely; in the interior of this (b), and between the epithelial and basement membrane, are numerous spirilla. $\times 600$. (Flügge.)

Genoa, etc. On the other hand, very numerous control experiments made by Koch and others show that it is not present in the alvine discharges of healthy persons or in the contents of the intestine of those who die from other diseases. In the writer's extended bacteriological studies of the excreta, and contents of the intestine of cadavers, in yellow fever, he has not once encountered any microorganism resembling the cholera spirillum.

As none of the lower animals are liable to contract cholera during the prevalence of an epidemic, or as a result of the ingestion of food contaminated with choleraic excreta, we have no reason to expect that pure cultures of the spirillum introduced by subcutaneous inoculation or by the mouth will give rise in them to a typical attack of

cholera. Moreover, it has been shown by experiment that this spirillum is very sensitive to the action of acids, and is quickly destroyed by the acid secretions of the stomach, of man or the lower animals, when the functions of this organ are normally performed. By a special method of procedure, however, Nicati and Rietsch, and Koch, have succeeded in producing in guinea-pigs choleraic symptoms and death. The first-named investigators injected cultures of the spirillum into the duodenum, after first ligating the biliary duct; the animals experimented upon died, and the intestinal contents contained the spirillum in large numbers. The fact that this procedure involves a serious operation which alone might be fatal, detracts from the value of the results obtained. Koch's experiments on guinea-pigs are more satisfactory, and, having been fully controlled by comparative experiments, show that the "comma bacillus" is pathogenic for these animals when introduced in a living condition into the intestine. This was accomplished by first neutralizing the contents of the stomach with a solution of carbonate of soda—five cubic centimetres of a five-per-cent solution, injected into the stomach through a pharyngeal catheter. For the purpose of restraining intestinal peristalsis the animal also receives, in the cavity of the abdomen, a tolerably large dose of laudanum—one gramme tincture of opium to two hundred grammes of body weight. The animals are completely narcotized by this dose for about half an hour, but recover from it without showing any ill effects. Soon after the administration of the opium a bouillon culture of the cholera spirillum is injected into the stomach through a pharyngeal catheter. As a result of this procedure the animal shows an indisposition to eat and other signs of sickness, its posterior extremities become weak and apparently paralyzed, and, as a rule, death occurs within forty-eight hours. At the autopsy the small intestine is found to be congested and is filled with a watery fluid containing the spirillum in great numbers. Comparatively large quantities of a pure culture injected into the abdominal cavity of rabbits or of mice often produce a fatal result within two or three hours; and Nicati and Rietsch have obtained experimental evidence of the pathogenic power of filtered cultures not less than eight days old. The most satisfactory evidence that this spirillum is able to produce cholera in man is afforded by an accidental infection which occurred in Berlin (1884), in the case of a young man who was one of the attendants at the Imperial Board of Health when cholera cultures were being made for the instruction of students. Through some neglect the spirillum appears to have been introduced into his intestine, for he suffered a typical attack of cholera, attended by thirst, frequent watery discharges, cramps in the extremities, and partial suppression of urine. Fortunately he

recovered ; but the genuine nature of the attack was shown by the symptoms and by the abundant presence of the "comma bacillus" in the colorless, watery discharges from his bowels. Nicati and Rietsch observed a certain degree of attenuation in the pathogenic power of the spirillum after it had been cultivated for a considerable time at 20° to 25° C. ; and the observation has since been made that cultures which have been kept up from Koch's original stock have no longer the primitive pathogenic potency.

Cunningham, as a result of recent researches made in Calcutta (1891), arrives at the conclusion that Koch's "comma bacillus" cannot be accepted as the specific etiological agent in this disease. This conclusion is based upon the results of his own bacteriological studies, which may be summed up as follows : *First*, in many undoubted cases of cholera he has failed to find comma bacilli. *Second*, in one case he found three different species. *Third*, in one case the reaction with acids could not be obtained. From sixteen cases in which Cunningham made cultures he obtained ten different varieties of comma bacilli, the characters of which he gives in his published report. It may be that in India, which appears to be the permanent habitat of the cholera spirillum, many varieties of this microorganism exist ; but extended researches made in the laboratories of Europe show that Cunningham is mistaken in supposing that spirilla resembling Koch's "comma bacillus" are commonly present in the intestine of healthy persons. The view advocated is that during the attack these spirilla are found in increased numbers because conditions are more favorable for their development, but that they have no etiological import. The writer would remark that, in very extended researches made in the United States and in Cuba, he has never found any microorganism resembling Koch's cholera spirillum in the fæces of patients with yellow fever or of healthy individuals, or in the intestinal contents of yellow-fever cadavers.

156. SPIRILLUM OF FINKLER AND PRIOR.

Synonym.—*Vibrio proteus*.

Obtained by Finkler and Prior (1884) from the fæces of patients with cholera nostras, after allowing the dejecta to stand for some days. Subsequent researches have not sustained the view that this spirillum is the specific cause of cholera morbus.

Morphology.—Resembles the spirillum of Asiatic cholera, but the curved segments ("bacilli") are somewhat longer and thicker and not so uniform in diameter, the central portion being usually thicker than the somewhat pointed ends; forms spiral filaments, which are not as numerous, and are usually shorter than those formed by the cholera spirillum. In unfavorable media involution forms are common—large oval, spherical, or spindle-shaped cells, etc. Has a single flagellum at one end of the curved segments, which is from one to one and one-half times as long as these.

Stains with the usual aniline colors—best with an aqueous solution of fuchsin.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *motile* spirillum. Spore formation not demonstrated. Grows in the usual culture media at the room temperature. Upon gelatin plates small, white, punctiform colonies are developed at the end of twenty-four hours, which under the microscope are seen to be finely granular and yellowish or yellowish-brown in color; liquefaction of the gelatin around these colonies progresses rapidly, and at the end of forty-eight hours is usually complete in plates where they are numerous. Isolated colonies on the second day form saucer-shaped depressions in the gelatin the size of lentils, having a sharply defined border. In *gelatin stick cultures* liquefaction progresses much more rapidly than in similar cultures of the cholera spirillum, and a stocking-shaped pouch of liquefied gelatin is already seen on the second day, which rapidly increases in dimensions, so that by the end of a week the gelatin is usually completely liquefied; upon the surface of the liquefied medium a whitish film is seen. Upon *agar* a moist, slimy layer, covering the entire surface, is quickly developed. The growth in *blood serum* is rapid and



FIG. 180.

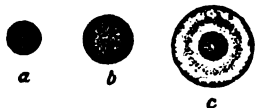


FIG. 181.

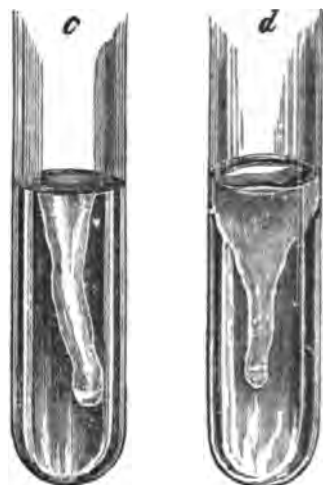


FIG. 182.

FIG. 180.—Spirillum of Finkler and Prior, from a gelatin culture. $\times 1,000$. From a photomicrograph. (Fränkel and Pfeiffer.)

FIG. 181.—Spirillum of Finkler and Prior; colonies upon gelatin plate; a, end of sixteen hours; b, end of twenty-four hours; c, end of thirty-six hours. $\times 80$. (Flügge)

FIG. 182.—Spirillum of Finkler and Prior; culture in nutrient gelatin; c, two days old; d, four days old. (Flügge.)

causes liquefaction of the medium. Upon *potato* this spirillum grows at the room temperature and produces a slimy, grayish-yellow, glistening layer, which soon extends over the entire surface. The cholera spirillum does not grow upon potato at the room temperature. The cultures of the Finkler-Prior spirillum give off a tolerably strong putrefactive odor, and, according to Buchner, in media containing sugar an acid reaction is produced as a result of their development. They have a greater resistance to desiccation than the cholera spirillum.

Pathogenesis.—Pathogenic for guinea-pigs when injected into the stomach by Koch's method, after previous injection of a solution of carbonate of soda, but a smaller proportion of the animals die from such injections (Koch). At the autopsy the intestine is pale, and its watery contents,

which contain the spirilla in great numbers, have a penetrating, putrefactive odor.

157. SPIRILLUM TYROGENUM.

Synonyms.—Spirillum of Deneke; Käsespirillen.

Obtained by Deneke (1885) from old cheese.

Morphology.—Curved rods and long, spiral filaments resembling the spirilla of Asiatic cholera. The diameter of the curved segments is somewhat less than that of the cholera spirillum, and the turns in the spiral filaments are lower and closer together. The diameter of the "commas" is uniform throughout, so that this spirillum more closely resembles the cholera spirillum than does that of Finkler and Prior.

Stains with the usual aniline colors—best with an aqueous solution of fuchsin.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *motile* spirillum. Spore formation not demonstrated. Grows in the usual culture media at the room temperature—more rapidly than the cholera spirillum and less so than that of Finkler and Prior. Upon *gelatin plates* small, punctiform colonies are developed, which on the second day are about the size of a pin's head and have a yellowish color; under the microscope they are seen to be coarsely granular, of a yellowish-green color in the centre and paler towards the margins. The outlines of the colonies are sharply defined at first, but later, when liquefaction has commenced, the sharp contour is no longer seen. At first liquefaction of the gelatin causes funnel-shaped cavities resembling those formed by the cholera spirillum, but liquefaction is more rapid. In *gelatin stick cultures* liquefaction occurs all along the line of puncture, and the spirilla sink to the bottom of the liquefied gelatin in the form of a coiled mass, while a thin, yellowish layer forms upon the surface; complete liquefaction usually occurs in about two weeks. Upon the surface of *agar* a thin, yellowish layer forms along the imp-

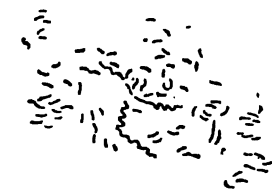


FIG. 153.—*Spirillum tyrogenum*. $\times 700$. (Flügge.)

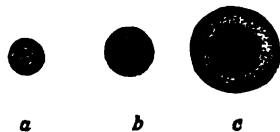


FIG. 154.—*Spirillum tyrogenum*; colonies in gelatin plate; a, end of sixteen hours; b, end of twenty-four hours; c, end of thirty-six hours. $\times 80$. (Flügge.)

a thin, yellow layer is usually developed (not always—Eisenberg); this contains, as a rule, beautifully formed, long, spiral filaments.

Pathogenesis.—Pathogenic for guinea-pigs when introduced into the stomach by Koch's method; three out of fifteen animals treated in this way succumbed.

158. SPIRILLUM METSCHNIKOWI.

Synonym.—*Vibrio Metschnikovi* (Gaméléia).

Obtained by Gaméléia (1888) from the intestinal contents of chickens dying of an infectious disease which prevails in certain parts of Russia during the summer months, and which in some respects resembles fowl cholera. The experiments of Gaméléia show that the spirillum under consideration is the cause of the disease referred to, which he calls gastro-enteritis cholERICA.

Morphology.—Curved rods with rounded ends, and spiral filaments; the curved segments are usually somewhat shorter, thicker, and more decidedly curved than the "comma bacillus" of Koch. The size differs very considerably in the blood of inoculated pigeons, the diameter being sometimes twice as great as that of the cholera spirillum, and at others about the same. A single, long, undulating flagellum may be seen at one extremity of the spiral filaments or curved rods in properly stained preparations.

Stains with the usual aniline colors, but not by Gram's method.

Biological Characters.—An *aërobie* (facultative anaërobie 1), *liquefying*, *motile* spirillum. According to Gamaléia, endogenous spores are formed by this spirillum; but Pfeiffer does not confirm this observation, and it must



FIG 185.—Spirillum Metschnikovi; culture in nutrient gelatin, end of forty-eight hours. From a photograph. (Fränkel and Pfeiffer.)

be considered extremely doubtful in view of the slight resistance to heat—killed in five minutes by a temperature of 50° C. Grows in the usual culture media at the room temperature. Upon *gelatin plates* small, white, punctiform colonies are developed at the end of twelve to sixteen hours; these rapidly increase in size and cause liquefaction of the gelatin, which is, however, much more rapid with some than with others. At the end of three days large, saucer-like areas of liquefaction may be seen, resembling that produced by the Finkler-Prior spirillum and the contents of which are turbid, while other colonies have produced small, funnel-shaped cavities filled with transparent, liquefied gelatin and resembling colonies of the cholera spirillum of the same age. Under the microscope the larger liquefied areas are seen to contain yellowish-brown granular masses which are in active movement, and the margins are surrounded by a border of radiating filaments. In *gelatin stick cultures* the growth resembles that of the cholera spirillum, but the development is more rapid. Upon *agar*, at 37° C., a yellowish layer resembling that formed by the cholera spirillum is quickly developed. Upon *potato* no growth occurs at the room temperature, but at 37° C. a yellowish-brown or chocolate-colored layer is formed, which closely resembles that produced by the cholera spirillum under the same circumstances. In *bouillon*, at 37° C., development is extremely rapid, and the liquid becomes clouded and opaque, having a grayish-white color, while a thin, wrinkled film forms upon the surface. When muriatic or sulphuric acid is added to a culture in peptonized bouillon a red color is produced similar to that produced in cultures of the cholera spirillum, and even more pronounced. In *milk*, at 35° C., rapid development occurs, and the milk is coagulated at the end of a week; the precipitated casein accumulates at the bottom of the tube in irregular masses and is not redissolved. The milk acquires a strongly acid reaction and the spirilla quickly perish.

Pathogenesis.—Pathogenic for chickens, pigeons, and guinea-pigs; rabbits and mice are refractory except for very large doses. Chickens suffering from the infectious disease caused by this spirillum remain quiet and somnolent, with ruffled feathers; they have diarrhoea; the temperature is not elevated above the normal, as is the case in chicken cholera. At the autopsy the most constant appearance is hyperæmia of the entire alimentary canal. A grayish-yellow liquid, more or less mixed with blood, is found in considerable quantity in the small intestine; the spleen is not enlarged and the organs generally are normal in appearance. In adult chickens the spirillum is not found in the blood, but in young ones its presence may be verified by the culture method and by inoculation into pigeons, which die in from twelve to twenty hours after being inoculated with two to four cubic centimetres. The pathological appearances in pigeons correspond with those found in chickens, but usually the spirillum is found in great numbers in blood taken from the heart. A few drops of a pure culture inoculated subcutaneously in pigeons or injected into the muscles cause their death in eight to twelve hours. Gameléia claims that the virulence of cultures is greatly increased by successive inoculations in pigeons, but Pfeiffer has shown that very minute doses are fatal to pigeons and that no decided increase of virulence occurs as a result of successive inoculations. According to Gameléia, chickens may be infected by giving them food contaminated

with the cultures of the spirillum, but pigeons resist infection in this way. Guinea-pigs usually die in from twenty to twenty-four hours after receiving a subcutaneous inoculation; at the autopsy an extensive subcutaneous œdema is found in the vicinity of the point of inoculation, and a superficial necrosis may be observed; the blood and the organs generally contain the "vibrio" in great numbers, showing that the animals die from general infection—acute septicæmia. When infection occurs in these animals by way of the stomach the intestine will be found highly inflamed and its liquid contents will contain numerous spirilla.

Gameléia has shown that pigeons and guinea-pigs may be made immune by inoculating them with sterilized cultures of the spirillum—sterilized by heat at 100° C. Old cultures contain more of the toxic substance than those of recent date. Thus two to three cubic centimetres of a culture twenty days old will kill a guinea-pig when injected subcutaneously, while five cubic centimetres of a culture five days old usually fail to do so. According to Pfeiffer, old cultures have a decidedly alkaline reaction, and their toxic power is neutralized by the addition of sulphuric acid.

Gameléia has claimed that by passing the cholera spirillum of Koch through a series of pigeons, by successive inoculation, its pathogenic power is greatly increased, and that when sterilized cultures of this virulent variety of the "comma bacillus" are injected into pigeons they become immune against the pathogenic action of the "vibrio Metschnikoff," and the reverse. Pfeiffer (1889), in an extended and carefully conducted research, was not able to obtain any evidence in support of this claim.

XVI.

BACTERIA IN INFECTIOUS DISEASES NOT PROVED TO BE DUE TO SPECIFIC MICROÖRGANISMS.

In the present chapter we shall give a brief account of the researches which have been made relating to the presence of bacteria, in a number of diseases, in which these researches have thus far failed to settle in a definite manner the etiology of the diseases named. For convenience of reference we shall arrange these diseases in alphabetical order.

ALOPECIA.

Robinson (1888) claims to have found, in sections from the diseased skin in a case of alopecia areata, micrococci having a diameter of about $0.8\ \mu$, usually united in pairs and associated in zoöglæa masses. They were located for the most part in the lymph spaces of the central portion of the chorium. They stained with the usual aniline colors and also by Gram's method. No culture or inoculation experiments were made.

Kasaui (1889) obtained from the margins of the affected patches in alopecia areata a bacillus about two to three times as long as broad, and which formed spores. It was attached to hairs withdrawn from the diseased patches, and was easily cultivated in various media.

Vaillard and Vincent (1890), in a form of alopecia resembling favus, obtained by cultivation, from hairs pulled out from the diseased patches, a micrococcus; this was also found in the hair follicles in stained sections. The diameter of this micrococcus was about one μ ; it was easily stained with the aniline colors and by Gram's method; grew in nutrient gelatin, causing liquefaction; did not grow well upon potato; was pathogenic for mice. When applied to the surface of the body of guinea-pigs or rabbits, by rubbing, alopecia resulted similar to that in the cases from which the micrococcus was first obtained.

BERI-BERI.

Lacerda (1887) claims to have demonstrated the presence of cocci, sometimes united in chains, in the blood and tissues of persons affected with beri-beri, and also to have produced in rabbits, by inoculation with his cultures, certain symptoms resembling those which characterize this disease.

Pekelharing and Winkler (1887) have also obtained by cultivation, from the blood of patients with beri-beri, various forms of bacteria, but principally cocci; these are described as being usually associated in pairs or in irregular groups, as forming a milk-white mass upon agar, and as liquefying gelatin. According to the authors named, injection into rabbits of cultures of this coccus gave rise to multiple nerve degeneration, such as is seen in cases of beri-beri in man.

Eykmann (1888) failed to obtain cultures from the blood of patients with beri-beri, but demonstrated the presence of slender bacilli similar to those

which Pekelharing and Winkler encountered in some of their cases. These failed to grow in the usual culture media.

In his latest communication upon the subject Pekelharing says that in twelve cases out of fifteen he obtained cultures of micrococci, and bacilli in three out of fifteen. From his inoculation experiments he concludes that the micrococci found are the cause of the morbid phenomena which characterize the disease.

When in Rio de Janeiro (1887) the writer collected blood from the finger from four typical cases of beri-beri, selected by Dr. Lacerda, and introduced it into the usual culture media. The result of this experiment was negative, agreeing in this regard with the results obtained by Eykmann.

BRONCHITIS.

Lumnitzer (1888) obtained from the sputum of a patient with putrid bronchitis a bacillus which proved to be pathogenic for mice and for rabbits, and the cultures of which gave off a characteristic odor, similar to that of the putrid bronchial secretion in his patient.

Picchini (1889), in three cases of "croupous bronchitis," made culture experiments and isolated three different micrococci; one developed upon nutritive gelatin as a grayish-white mass and did not liquefy; one as a reddish-gray mass, also non-liquefying; the third form was always associated with these two.

CARCINOMA.

Various microorganisms have occasionally been found in carcinomatous growths, and especially in those which have undergone ulceration; but that any one of these bears an etiological relation to such malignant tumors has not been demonstrated.

CEREBRO-SPINAL MENINGITIS.

Various microorganisms have been found by bacteriologists in the exudate of cerebro-spinal meningitis, and there seems to be but little doubt that the meningeal inflammation is due to their presence, as the bacteria usually found are pathogenic for certain of the lower animals, and when introduced into a serous cavity they give rise to a fibrinous or purulent inflammatory process. The researches of Weichselbaum, Netter, and others show that *Micrococcus pneumoniae crouposa* ("diplococcus pneumoniae") is the microorganism most frequently found, and next to this the *Diplococcus intercellularis meningitidis* of Weichselbaum. *Streptococcus pyogenes* has also been found in a certain proportion of the cases—four out of twenty-five cases of purulent meningitis studied by Netter.

Bonome, in a series of cases studied by him, obtained a micrococcus closely resembling *Micrococcus pneumoniae crouposa*, but which he believes not to be identical with it (see *Micrococcus* of Bonome, No. 40).

For further details see the descriptive accounts of the microorganisms above referred to.

CHANCROID.

Ducrey, in an extended research (1890), was not able to cultivate any specific microorganism from the pus of soft chancres, or of buboes resulting

from these ulcers. Various common microorganisms were obtained in cultures from the chancroidal ulcers, but a negative result was obtained in his cultures from the pus of buboes. In pustules developed upon the arm from the inoculation of chancroidal virus he found constantly a bacillus which did not grow in artificial cultures. This was about $1.48\ \mu$ long and $0.5\ \mu$ thick, with round ends. It was readily stained with a solution of fuchsin, but not by Gram's method.

CHOLERA INFANTUM.

The researches of Booker and of Jeffries do not support the idea that cholera infantum is due to the presence of a specific microorganism in the intestine, but rather that the symptoms are due to the absorption of toxic products formed in the alimentary canal, or in the child's food before it is ingested, as a result of the multiplication and ferment action of various microorganisms, and especially of certain putrefactive bacteria. The common putrefactive bacillus, *Proteus vulgaris*, and other species nearly related to this, were found by Booker in a considerable proportion of his cases, and he is disposed to believe that these putrefactive bacteria play an important part in the development of the morbid phenomena which characterize the disease. Jeffries, after reviewing the various theories which have been advanced in explanation of the etiology of cholera infantum, says: "Bacteria I believe to be at the bottom of the disease—that is, rule bacteria out of all foods and the alimentary canal, and summer diarrhoea would cease to be." Upon another page of his memoir he says: "Passing a step further, the symptoms, pathology, and etiology of summer diarrhoea stand in close relationship with the group of symptoms first clearly brought to light by Panum as putrid infection. The animals poisoned by the injection of putrid fluids, sterile or not, sicken and die with two variable groups of symptoms: one referable to the nervous system, the other to the intestines, diarrhoea being a prominent symptom, and the autopsy revealing inflammatory changes in the intestine."

CHOLERA NOSTRAS.

What has been said above with reference to cholera infantum applies as well to cholera nostras. This has not been shown to be due to the presence in the alimentary canal of a particular microorganism; but it can scarcely be doubted that the morbid phenomena are induced by the development of toxic substances as a result of the ferment action of various species of bacteria.

Finkler and Prior (1884) obtained from the fæces of patients with cholera nostras a spirillum which they supposed to be the specific cause of this disease, but subsequent researches have not confirmed their conclusion. Thus, in seven cases studied by bacterio-

logical methods in Koch's laboratory during the years 1885, 1886, and 1887, the spirillum of Finkler and Prior was not found in a single instance (Frank).

CONJUNCTIVITIS.

The various forms of conjunctivitis have been ascribed to the specific action of bacteria. That this is true as regards gonorrhœal ophthalmia is now generally admitted, and there is some reason to believe that the bacillus discovered by Koch and studied by Kartulis (see *Bacillus* of Kartulis) is the cause of one form of "Egyptian catarrhal conjunctivitis." The non-infectious forms of conjunctivitis can scarcely be supposed to be due to the action of specific microörganisms; but it is probable that an inflammation resulting from any cause, such as a chemical or mechanical irritant, may be aggravated and become chronic as a result of the presence of various microörganisms, and especially of the pyogenic micrococci.

With reference to the so-called bacillus of xerosis, the researches of Schreiber, made in Neisser's laboratory, show that it is not peculiar to xerosis, but that it is often found quite as abundantly in other eye affections and also in the healthy conjunctival sac.

CORYZA.

Hajek found in the secretions of acute nasal catarrh a large diplococcus, called by him "*Diplococcus coryzæ*," and probably identical with the diplococcus previously obtained by Klebs from the same source. This was most abundant during the early stage of the disease, when the secretion from the nasal mucous membrane was thin and abundant; later various other microörganisms were encountered in greater numbers, and among them Friedländer's bacillus. There is no satisfactory evidence that the diplococcus of Hajek or any other known bacteria are directly concerned in the etiology of this affection. To what extent chronic nasal catarrh is due to the action of microörganisms is also uncertain, but it appears probable that they play an important part in maintaining such inflammations; and in *ozæna* the offensive odor of the nasal secretions is no doubt due to the presence of certain bacteria, whatever may be the relation of these to the morbid process which gives rise to the chronic discharge. (See *Bacillus foetidus ozænæ* of Hajek.)

CYSTITIS.

Various species of bacteria have been found in the urine in cases of cystitis, and it appears probable that some of these are directly or indirectly concerned in keeping up the vesical inflammation in chronic cases of this disease. Whether any one of the species found is capable of producing cystitis when introduced into the healthy bladder is uncertain. On the other hand, we have rather numerous observations which show that there may be bacteriuria without cystitis. Thus Schottelius and Reinhold report a case of heart disease in which certain bacilli were found in the urine in considerable numbers, and in a pure culture, during the entire time that the

patient was under observation, and in which there was no cystitis or other symptoms that could be ascribed to the presence of this bacillus.

In the extended researches of Rovsing—thirty cases of cystitis—the following results were obtained: In one case diagnosed as cystitis no bacteria were found; in three cases culture experiments gave a negative result, but the tubercle bacillus was found in the urine by microscopical examination—in these cases the urine was strongly acid; in twenty-six cases the urine was ammoniacal, and in all of these bacteria were found—usually but a single species. All of these grew in the usual culture media except the tubercle bacillus, which in two cases was associated with some other species, and all produced alkaline fermentation in sterile urine when added to it in pure cultures. The following species were found: Tubercle bacillus, *Staphylococcus pyogenes aureus*, *Staphylococcus pyogenes albus*, *Staphylococcus pyogenes citreus*, *Streptococcus pyogenes ureæ* (n. sp.), *Diplococcus pyogenes ureæ* (n. sp.), *Coccobacillus pyogenes ureæ* (n. sp.), *Micrococcus pyogenes ureæ flavus* (n. sp.), *Diplococcus ureæ trifolius* (n. sp.), *Streptococcus ureæ rugosus* (n. sp.), *Diplococcus ureæ* (n. sp.), *Coccobacteria ureæ* (n. sp.).

Pure cultures of all of these species introduced into the bladder of rabbits failed to induce cystitis, even when injected in considerable quantities. But when retention of urine was effected artificially for six to twelve hours, allowing time for ammoniacal fermentation to occur, cystitis was developed. When the pyogenic species were introduced under these circumstances a suppurative inflammation of the mucous membrane occurred; the non-pyogenic species caused a catarrhal cystitis. Rovsing records the important fact, as bearing upon the etiology of cystitis, that in twenty of the cases examined the bladder had been invaded by the finger or by instruments prior to the development of cystitis.

Lundström (1890) isolated from alkaline urine obtained from patients with cystitis two species of staphylococci—*Staphylococcus ureæ candidus* and *Staphylococcus ureæ liquefaciens*; from albuminous, acid urine he obtained *Streptococcus pyogenes*. Krogus obtained from the urine of individuals suffering from cystitis a bacillus which he calls *Urobacillus liquefaciens septicus*. Schnitzler (1890) found the same bacillus, or one very similar to it, in thirteen out of twenty cases of purulent cystitis examined by him. In eight of these cases it was obtained from the urine in pure cultures, and in five it was associated with other bacteria. In twelve of these twenty cases the cystitis resulted directly from catheterization; in the others it occurred in individuals suffering from stricture or from calculus.

When cultures of this bacillus were injected into a vein in rabbits, the animals died in from three to eight days, and in every

instance an intense nephritis was observed at the autopsy—twice with the formation of small abscesses. The bacillus was found in the blood and the organs generally. Injections into the bladder of rabbits almost always gave rise to a severe purulent cystitis—large rabbits were selected and great care taken not to injure the mucous membrane of the bladder. Schnitzler was not able to induce cystitis in rabbits by injecting in the same way considerable quantities of a culture of *Staphylococcus pyogenes aureus*.

Guyon (1888) did not succeed in inducing cystitis by the injection of pure cultures of various microörganisms into the bladder, unless he at the same time produced an artificial retention of urine. His experimental results are therefore in accord with those of Rovsing, who found that without mechanical injury, or artificial retention until ammoniacal fermentation had occurred, no results followed his injections into the bladder.

DENGUE.

McLaughlin (1886) has claimed to find micrococci in the blood of patients suffering from dengue. No satisfactory evidence of their etiological relation has been presented, and his observations have not yet been confirmed by other investigators.

ECZEMA EPIZOÖTICA.

Synonym.—Foot and mouth disease.

This is an infectious disease of horned cattle, characterized by a vesicular eruption in the mouth and about the feet. It affects also sheep and pigs and may be communicated to man.

Up to the present time no satisfactory demonstration has been made of the specific infectious agent; but Schottelius has recently (1892) described a microörganism which he thinks may bear an etiological relation to the disease. His inoculation experiments do not, however, sustain this view, inasmuch as the characteristic vesicles were never developed in inoculated calves, and experiments upon other animals gave a negative result. In young cattle small doses (one cubic centimetre) of a bouillon culture gave rise to a slight fever and loss of appetite, while larger doses produced an intense fever, salivation, and great debility. But recovery occurred at the end of five or six days without any aphthous eruption. Schottelius obtained from the clear contents of the vesicles in the mouth various bacteria which he believes to have been accidentally present. After making a considerable number of culture experiments his attention was attracted by a spherical microörganism, united in chains, which grew very slowly in the ordinary culture media. This he describes as follows:

The individual cells vary greatly in diameter, and are considerably larger than known micrococci; they are associated in longer or shorter chains, and are endowed with active movements. According to Schottelius, they belong to the "*streptocytæ*" rather than to the streptococci. They do not stain readily with methylene blue, but may be stained with gentian violet and by Gram's method. Development does not occur at temperatures below 37° to 39° C. The most suitable culture medium was found to be bouillon or glycerin agar to which formate of soda had been added (amount?). Growth occurred in an atmosphere of CO₂ as well as in atmospheric air. Upon plates of nutrient agar—containing glycerin and formate of soda—at 37° C.,

very delicate, almost transparent colonies developed; they were of a pearl-gray color, with an irregular, rosette-like margin; in the course of several weeks they attained a diameter of one to one and one-half millimetres. Upon *potato* a scanty, grayish-white, dry layer is developed. Under the most favorable conditions the development was very slow—not more rapid than that of the tubercle bacillus.

EMPHYEMA.

A. Fränkel (1888), as a result of his bacteriological studies in twelve cases of empyema, divides the cases into four groups. In one group of three cases *Streptococcus pyogenes* was the only microorganism obtained in his cultures or seen in stained preparations of pus from the pleural cavity. In a second group of three cases, occurring in the course of a pneumonia, the only microorganism present was "*diplococcus pneumoniae*" (*Micrococcus pneumoniae crouposæ*). The third group included four cases of tubercular empyema; in one of these tubercle bacilli only were found in pus from the pleural cavity, in one case streptococci were found, and in two no microorganisms were found. In the fourth group of two cases the empyema resulted from the opening of an abscess into the pleural cavity, and streptococci were found in the pus.

Netter, in a series of forty-six cases examined by him, found *Micrococcus pneumoniae crouposæ* in fourteen. Koplik (1890) found the same microorganism in seven cases examined by him, and *Streptococcus pyogenes* in two cases.

ENDOCARDITIS.

The experimental evidence relating to endocarditis is similar to that in cystitis. The injection of the microorganisms found attached to the diseased structures into the circulation of lower animals does not produce endocarditis unless the valves have been previously injured by mechanical violence or by chemical irritants. If some doubt remains among pathologists as to the etiological relation of the microorganisms found, the serious secondary results of the mycotic invasion are well established. In a series of twenty-nine cases studied by Weichselbaum (1885–1888) the following results were obtained: In eight the result of culture experiments and microscopical examination was negative; in seven "*diplococcus pneumoniae*" (*Micrococcus pneumoniae crouposæ*) was found; in six *Streptococcus pyogenes*; in two *Staphylococcus pyogenes aureus*; in two *Bacillus endocarditidis griseus* (Weichselbaum); in one *Micrococcus endocarditidis rugatus* (Weichselbaum); in one *Bacillus endocarditidis capsulatus* (Weichselbaum); in two cases a bacillus which he did not succeed in cultivating. For further details see the descriptions of microorganisms referred to.

ERYTHEMA.

Cordua (1885) obtained from a series of cases of an erysipelatoid skin affection of the fingers and hands, which he identified as corresponding with erythema exudativum multiforme of Hebra, a micrococcus resembling *Staphylococcus pyogenes albus* in its biological characters, but which he describes as being three to four times as large. Inoculations in animals were without result, but two inoculations upon his own hand produced a dark-red tumefaction in the vicinity of the point of inoculation resembling that in the individuals from whom he obtained his cultures.

In two cases of "polymorphous erythema" Haushalter (1887) isolated a streptococcus which did not produce an erysipelatous inflammation when inoculated into the ear of rabbits, and which he supposed to be a different species from the now better known *Streptococcus pyogenes* (?). In five cases of erythema nodosum in children Demme obtained a bacillus which his inoculation experiments proved to be pathogenic, and which was perhaps concerned in the etiology of the skin affection from which his cultures were obtained (see *Bacillus* of Demme, No. 107).

GRANULOMA FUNGOIDES (MYCOSIS FUNGOIDES).

Rindfleisch (1885) and Auspitz (1885) report the presence of streptococci in the capillary vessels of the papillary body and of the subcutaneous tissue in the affected localities in cases of this disease. That the streptococcus differs from *Streptococcus pyogenes*, as Auspitz supposes, has not been definitely established.

HYDROPHOBIA.

Notwithstanding the extended researches made, especially in Pasteur's laboratory, the etiology of hydrophobia still remains unsettled. It has been demonstrated by experiment that the virus of the disease is located in the brain, spinal marrow, and nerves of animals which have succumbed to the disease, as well as in the salivary secretions of rabid animals, and that the disease may be transmitted by intravenous inoculation, or by introducing a small quantity of virus beneath the dura mater, with greater certainty than by subcutaneous inoculations. But the exact nature of this virus has not been determined. The fact that a considerable interval elapses after inoculation before the first symptoms are developed indicates that there is a multiplication of the virus in the body of the infected animal; and this is further shown by the fact that after death the entire brain and spinal marrow of the animal have a virulence equal to that of the material with which it was inoculated in the first instance. The writer's experiments (1887) show that this virulence is neutralized by a temperature of 60° C. maintained for ten minutes—a temperature which is fatal to all known pathogenic bacteria in the absence of spores. But recent experiments show that certain toxic products of bacterial growth are destroyed by the same temperature. We are, therefore, not justified in assuming that the morbid phenomena are directly due to the presence of a living microörganism; and, indeed, it seems probable, from what we already know, that the symptoms developed and the death of the animal are due to the action of a potent chemical poison of the class known as toxalbumins. But, if this is true, we have still to account for the production of the toxic albuminoid substance, and, in the present state of knowledge, have no other way to explain its increase in the body of the infected animal than the supposition that a specific, living germ is present in the virulent material, the introduction of which into the body of a susceptible animal gives rise to morbid phenomena characterizing an attack of rabies.

Pasteur and his associates have thus far failed to demonstrate the presence of microörganisms in the virulent tissues of animals which have succumbed to an attack of rabies. Babes has obtained micrococci in cultures

from the brain and spinal cord of rabid animals, and states in his article on hydrophobia in "*Les Bactéries*" (second edition, page 791) that pure cultures of the second and third generations induced rabies in susceptible animals; but his own later researches do not appear to have established the etiological relation of this micrococcus.

Gibier (1884) has reported the presence of spherical refractive granules, resembling micrococci, in the brain of rabid animals, which he demonstrated by rubbing up a little of the cerebral substance with distilled water. As these supposed micrococci did not stain with the usual aniline colors and were not cultivated, it appears very doubtful whether the refractive granules seen were really microorganisms.

Fol (1885) claims to have demonstrated the presence of minute cocci, 0.2 μ in diameter, in sections of spinal cord from rabid animals, by Weigert's method of staining. The cords were hardened in a solution of bichromate of potash and sulphate of copper, colored with a solution of hematoxylin, and decolorized in a solution of ferrocyanide of potash and borax.

The writer (1887) has made similar preparations, carefully following the method as described by Fol, but was not able to demonstrate the presence of microorganisms in the numerous sections made. Nor have the observations of Fol been confirmed by the researches of other bacteriologists who have given their attention to the subject since the publication of his paper.

With reference to the results of Pasteur's protective inoculations, we may say that it is now pretty generally admitted that the published statistics demonstrate the prophylactic value of the method as practised at the Pasteur Institute in Paris.

In a paper by Perdrix published in the *Annales de l'Institut Pasteur*, vol. iv., 1890, the following statistics are given for the years 1886-89:

Year.	Number treated.	Deaths.	Mortality.
1886.....	2,671	25	0.94%
1887.....	1,770	13	0.73
1888.....	1,622	9	0.55
1889.....	1,830	6	0.33
Total	7,893	53	0.67%

This table includes only those deaths which occurred at least fifteen days after the treatment was terminated. When all deaths are included the figures are as follows:

Year.	Number treated.	Deaths.	Mortality.
1886.....	2,683	36	1.34%
1887.....	1,778	21	1.18
1888.....	1,625	13	0.74
1889.....	1,834	10	0.54
Total.....	7,919	79	0.92%

In the statistics of the Pasteur Institute the cases have from the commencement been classified as follows:

A. Persons bitten by animals proved to be rabid by experimental inoculations in other animals.

B. Persons bitten by animals examined by veterinary surgeons and pronounced by them to be rabid.

C. Persons bitten by animals suspected of being rabid.

The following table gives the results in accordance with this classification:

Year.	A.			B.			C.		
	Number treated.	Died.	Mortality.	Number treated.	Died.	Mortality.	Number treated.	Died.	Mortality.
1886...	223	5	2.15%	1,931	24	1.24%	518	7	1.35%
1887....	357	2	0.56	1,161	15	1.29	260	4	1.54
1888....	403	7	1.74	974	4	0.41	248	1	0.40
1889...	848	4	1.15	1,188	3	0.25	298	3	1.00
Total.	1,341	18	1.34%	5,254	46	0.88%	1,324	15	1.13%

At the meeting of the Tenth International Medical Congress in Berlin (1890), Babes reported that in the Pasteur Institute at Bucharest about three hundred persons are inoculated yearly, with a mortality of 0.4 per cent, in cases bitten by dogs, most of which were demonstrated to have been rabid by inoculation experiments made at the Institute.

The recent researches of Tizzoni and Schwartz (1892) show that the blood of rabbits which have an artificial immunity against rabies contains an antitoxine which has the power of neutralizing the virus of rabies, either in a test tube or in the body of an inoculated animal. Their experiments indicate the possibility of curing rabies in man by subcutaneous inoculations of this antitoxine extracted from the blood serum of immune rabbits.

ICTERUS.

Karlinsky (1890), in a series of five cases of "infectious icterus" attended with fever, found in the blood, during the height of the attack, curved bacilli from two to six μ long and one-third to one μ broad, which were readily stained by the usual aniline colors, but not by Gram's method. These he did not succeed in cultivating in any of the culture media usually employed.

Ducamp (1890) has also given an account of a "slight epidemic of infectious icterus," which he supposes to have been due to microörganisms.

LEPROSY.

No satisfactory experimental demonstration that the *Bacillus lepræ* is the cause of the disease with which it is associated has yet been made; but there is very little doubt among bacteriologists and pathologists that such is the case. For the facts relating to its presence in leprous tissues, its morphology, etc., the reader is referred to the descriptive account of *Bacillus lepræ* (No. 53, page 394).

MALARIA.

Klebs and Tommasi-Crudeli, as a result of researches made by them in the vicinity of Rome (1879), announced the discovery of a bacillus which they supposed to be the cause of malarial fevers—their *Bacillus malarie*. The writer repeated their experiments the following year (1880) in the vicinity of New Orleans, and reported as follows:

"Among the organisms found upon the surface of swamp mud near New Orleans, and in the gutters within the city limits, are some which closely resemble, and perhaps are identical with, the *Bacillus malarie* of

Klebs and Tommasi-Crudeli; but there is no satisfactory evidence that these or any of the other bacterial organisms found in such situations, when injected beneath the skin of a rabbit, give rise to a malarial fever corresponding with the ordinary paludal fevers to which man is subject.

"The evidence upon which Klebs and Tommasi-Crudeli have based their claim of the discovery of a *Bacillus malarie* cannot be accepted as sufficient; (a) because in their experiments and in my own the temperature curve in the rabbits experimented upon has in no case exhibited a marked and distinctive paroxysmal character; (b) because healthy rabbits sometimes exhibit diurnal variations of temperature (resulting apparently from changes in the external temperature) as marked as those shown in their charts; (c) because changes in the spleen such as they describe are not evidence of death from malarial fever, inasmuch as similar changes occur in the spleens of rabbits dead from septicæmia produced by the subcutaneous injection of human saliva; (d) because the presence of dark-colored pigment in the spleen of a rabbit cannot be taken as evidence of death from malarial fever, inasmuch as this is frequently found in the spleens of septicæmic rabbits."

Later researches have also failed to confirm the supposed discovery of Klebs and Tommasi-Crudeli, and it is now generally admitted that there is no satisfactory evidence in favor of the view that microorganisms of this class are concerned in the etiology of the malarial fevers. On the other hand, we have now very extended observations which indicate that the blood parasite discovered by Laveran (1881) in the blood of patients suffering from various forms of malarial fever bears an etiological relation to fevers of this class. This hæmatozoön belongs to quite a different class of microorganisms. It was first described by Laveran as the *Oscillaria malarie*, but is more frequently spoken of at present as the *Plasmodium malarie*.

MEASLES.

The etiology of measles and of the specific eruptive febrile diseases generally still remains unsettled. The occasional presence of micrococci in the blood of patients with measles, which has been noted, is without doubt due to a secondary or mixed infection by one of the common pyogenic micrococci. In pneumonia occurring during the course of an attack of measles the *Micrococcus pneumoniae croupose* is usually found in the pulmonary exudate.

No great importance can be attached to the observations made, with reference to the presence of microorganisms in this disease, prior to the introduction of Koch's plate method and the use of solid culture media for the differentiation of bacteria similar in their morphology. Recently (1892) Canon and Pielicke, of Berlin, have announced the discovery of a minute bacillus in the blood of patients (fourteen) with measles, which may turn out to be the specific infectious agent in this disease. But this cannot be considered demonstrated by the researches thus far made. (See *Bacillus* of Canoñ and Pielicke, No. 486.)

MENINGITIS.

See Cerebro-spinal meningitis, page 515.

NEPHRITIS.

The various microorganisms which have occasionally been found in the urine of cases of nephritis are probably not directly related to the renal disease. Numerous observations are on record which show that pathogenic microorganisms present in the blood or tissues may find their way into the urine during the course of the disease due to their presence. In these cases it is probable that the passage of bacteria into the urine depends upon structural changes in the kidneys; but that these changes are directly due to the bacteria is by no means established. As an example we may mention that

the bacillus of typhoid fever is occasionally found in the urine during an attack of this disease.

Letzerich (1887) has described a form of nephritis which he ascribes to a bacillus found by him in the urine and in sections of the kidneys of rabbits inoculated with a culture of this bacillus.

Lustgarten and Manneberg (1887) in three cases of acute Bright's disease found streptococci in the urine, which they suppose to have had an etiological relation to the renal disease. The following year Manneberg reported eleven additional cases, in eight of which he found the same streptococcus, which he believes to be different from *Streptococcus pyogenes*, but this cannot be considered as established. Nor has he shown that the streptococcus obtained by him from the urine was present in the kidneys of his patients, or that pure cultures of this streptococcus produce acute nephritis when inoculated into lower animals.

OSTEOMYELITIS.

The evidence with reference to the presence of *Staphylococcus pyogenes aureus* in acute osteomyelitis and its probable etiological relation to the cases in which it is found, is given in the article descriptive of this microörganism; but the researches of Kraske (1886) and of Lamelongue and Achard (1890) show that the "golden staphylococcus" is not always found in osteomyelitis. The last-named investigators, in a series of thirteen cases, found *Staphylococcus pyogenes aureus* in four only, and in one of these *Staphylococcus pyogenes albus* was also present; in three cases *Staphylococcus pyogenes albus* was obtained in pure cultures; in two cases it was associated with *Streptococcus pyogenes*; and in two cases a streptococcus was found which resembled *Streptococcus pyogenes* and yet differed from it in some particulars.

OTITIS MEDIA.

In otitis media various microörganisms have been found in pus obtained by paracentesis of the tympanum, as well as in the chronic discharge after perforation; and there can be but little doubt that these microörganisms are responsible, directly or indirectly, for the inflammatory process and pus formation. The following species are most frequently found in the purulent discharge in recent cases of otitis media: *Micrococcus pneumoniae crouposæ* ("diplococcus pneumoniae"), *Streptococcus pyogenes*, *Staphylococcus pyogenes albus*, *Staphylococcus pyogenes aureus*, Friedländer's bacillus. The following have also been found occasionally: *Staphylococcus tenuis*, *Bacillus tenuis*, *Micrococcus tetragenus*, *Bacillus pyocyaneus*.

According to Zaufal, *Micrococcus pneumoniae crouposæ* is most frequently found in cases which result from exposure to cold, while the ordinary pus cocci are more frequently found in otitis which is secondary to specific febrile diseases.

OZÆNA.

The researches of Thost, Klamann, Hajek, and others show that Friedländer's bacillus is present in the nasal secretions in a considerable proportion of the cases of ozæna, but its etiological relation to the morbid condition which gives rise to the offensive discharge has not been established.

Thost found this bacillus in twelve out of seventeen cases studied by him, and frequently almost in a pure culture; but he also found it in rhinitis from syphilitic ulceration, from polypus, and in simple coryza.

Hajek found Friedländer's bacillus in seven out of ten cases studied by him, but it was associated with various other species of bacteria, and especially with the pyogenic micrococci and with *Bacillus fluorescens liquefaciens*. He also obtained almost constantly his *Bacillus foetidus ozænæ* (No. 111), which appears to have been the cause of the foetid odor of the nasal discharge.

Marano (1890) in ten cases of ozæna found a capsule bacillus in the nasal secretions which closely resembles Friedländer's bacillus, but which he believes not to be identical with it.

PAROTITIS.

No demonstration of a specific microorganism in mumps has been made, but in non-specific, suppurative parotitis one or other of the pyogenic micrococci appears to be the cause of the inflammation and pus formation. In parotitis occurring as a complication of pneumonia *Micrococcus pneumoniæ crouposæ* has been found as the only microorganism in pus from the inflamed gland (Testi, Duplay).

PEMPHIGUS.

Demme (1886) has cultivated a diplococcus from a case of acute pemphigus which possibly is related to this disease (see *Micrococcus* of Demme, No. 27, page 319). The same coccus was found by Dähnhardt in a similar case.

PERITONITIS.

That peritonitis usually results from the presence of microorganisms in the cavity of the abdomen seems to be pretty well established by experimental evidence and by bacteriological researches in cases of this disease. Mechanical irritants, like finely powdered glass (writer's experiments), introduced into the cavity of the abdomen of rabbits, do not cause peritonitis unless microorganisms are introduced at the same time; the minute fragments of glass become encysted and the animal remains in good health. But Pernice has shown that peritonitis may be induced in rabbits and in

guinea-pigs by injecting into the cavity of the abdomen various chemical substances, such as concentrated mineral acids, acetic acid, phenol, nitrate of silver, etc. It is also demonstrated by numerous experiments that pure cultures of various bacteria injected into the cavity of the abdomen of the animals mentioned may produce a fibrinous or a purulent peritonitis. Among these is the *Bacillus coli communis*, which is constantly present in the intestine of healthy persons; and in peritonitis following perforation of the bowel it is probable that this bacillus is responsible, in part at least, for the intense peritoneal inflammation which so quickly occurs. In puerperal peritonitis the pus cocci, and especially *Streptococcus pyogenes*, appear to be the usual cause of the inflammatory process.

Weichselbaum has observed two cases of primary peritonitis and pleuritis apparently induced by *Micrococcus pneumoniae crouposæ*, as this microörganism was found in the exudate into the peritoneal cavity. The same author, in a case of peritonitis resulting from rupture of the spleen in the course of typhoid fever, obtained a pure culture of the typhoid bacillus from the peritoneal cavity.

The recently published (1891) results of A. Fränkel's researches are as follows: In thirty-one cases examined pure cultures were obtained in twenty, viz.: *Bacillus coli communis*, nine times; streptococci, seven times; *Bacillus lactis aërogenes*, twice; *Micrococcus pneumoniae crouposæ*, once; *Staphylococcus pyogenes aureus*, once. In three cases *Bacillus coli communis* was present in association with other bacilli, and in four cases the bacteriological examination gave a negative result.

PLEURITIS.

See Empyema, page 520.

PLEURO-PNEUMONIA OF CATTLE.

This is an infectious disease, the etiology of which is still undetermined, notwithstanding the researches of numerous bacteriologists. Various bacteria have been isolated from the exudate into the pulmonary alveoli, but there is no satisfactory proof that any one of these is the specific cause of the disease.

PURPURA HÆMORRHAGICA.

See account of bacilli found in purpura hæmorrhagica by Babes (No. 146), Kolb (No. 147), and Tizzoni and Giovannini (No. 145).

RHINOSCLEROMA.

See *Bacillus* of rhinoscleroma (No. 58).

SCARLET FEVER.

The specific infectious agent in scarlet fever has not been demonstrated. In the diphtheritic exudate frequently seen in the angina of scarlet fever a

streptococcus is commonly found which appears to be identical with *Streptococcus pyogenes*; and in the secondary affections which occur in the course of this disease or during convalescence, when suppuration occurs, one or the other of the common pyogenic micrococci is usually found and is doubtless the cause of the local inflammatory process. (See *Otitis media*, page 525.)

SYPHILIS.

The etiology of syphilis has not been determined by the researches of bacteriologists. For an account of the microorganisms which have been encountered in syphilitic lesions the reader is referred to the article on the *Bacillus of Lustgarten* (No. 55).

"TEXAS FEVER" OF CATTLE.

Billings (1888) has announced the discovery of a bacillus in the blood of cattle suffering from Texas fever, which he supposes to be the cause of this disease, but the investigations of other bacteriologists have failed to confirm the alleged discovery. It appears probable that a mistake in diagnosis was made, and that the disease studied by Billings was an infectious form of septicæmia in cattle similar to the *Rinderseuche* of German authors. The microorganism which he has described as coming from the blood of the infected animals resembles in its morphology *Bacillus septicæmiæ hæmorrhagicæ* (No. 61), and, if not identical with this widely distributed species, appears to be very nearly related to it.

TYPHUS FEVER.

The etiology of typhus fever has not been determined in a definite manner. Hlava (1888) has described a "*streptobacillus*" which he supposes to be concerned in the etiology of this extremely contagious disease; but it has been shown by other investigators that this bacillus is not constantly present, and there is no satisfactory evidence that it is the specific infectious agent. Thoinot and Calmette encountered the streptobacillus of Hlava and various other microorganisms in a certain proportion of the cases examined by them; but no one of these was constantly present. "An interesting organism," which they did not succeed in cultivating, was found in the blood of all cases examined by the investigators last mentioned.

VARICELLA.

Various microorganisms have been found in the contents of the vesicles and pustules of varicella, but there is no evidence that any one of these bears an etiological relation to this specific eruptive fever.

VARIOLA AND VACCINIA.

The etiology of small-pox still remains undetermined. The common pus cocci and various other microorganisms are found in the characteristic pustular eruption, and various microorganisms have been isolated from vaccine vesicles; but no one of these has been shown to possess the specific pathogenic power of unfiltered lymph from the same source. The experiments of Carstens and Coert show that the specific virulence of vaccine lymph is destroyed by ten minutes' exposure to a temperature of 54° C. And the writer's experiments show that various disinfecting agents tested—chlorine, sulphur dioxide, nitrous acid—destroy the infective virulence of lymph dried upon ivory points in about the same proportion as is required for the destruction of some of the best-known pathogenic bacteria. But this does not prove that virulence depends upon the presence of a living microorganism, however probable this appears, for certain toxalbumins are likewise

destroyed by a correspondingly low temperature and by the action of various chemical disinfectants.

YELLOW FEVER.

In the writer's "Report upon the Prevention of Yellow Fever by Inoculation," submitted in March, 1888, the following conclusions are given as the result of his investigations at the date of this report:

Conclusions.

Facts relating to the endemic and epidemic prevalence of yellow fever, considered in connection with the present state of knowledge concerning the etiology of other infectious diseases, justify the belief that yellow fever is due to a living microörganism capable of development under favorable local and meteorological conditions external to the human body, and of establishing new centres of infection when transported to distant localities.

Inasmuch as a single attack of yellow fever, however mild, protects, as a rule, from future attacks, there is reason to hope that similar protection would result if a method could be discovered of inducing a mild attack of the disease by inoculation or otherwise.

The hypothetical yellow-fever germ, multiplying external to the human body in unsanitary places in tropical regions where the disease is endemic, or during the summer months in the area of its occasional epidemic prevalence, establishes infected localities, and susceptible persons contract yellow fever by exposure in these infected areas. We infer, therefore, *a priori*, that the yellow-fever germ invades the system by the respiratory tract, by the alimentary canal, or from the general surface of the body, and it should be found in the blood and tissues, or in the alimentary canal, or upon the surface.

Another possibility presents itself, viz., that the germ multiplying in unsanitary localities external to the body produces a volatile poison which contaminates the air, and that an attack is induced by the toxic effects of this potent chemical poison. The more or less prolonged period of incubation—two to five days—in numerous cases in which the attack has been developed after removal from the infected locality, is opposed to this latter hypothesis.

In the light of what is known of the etiology of other infectious diseases, the hypothesis that the germ really finds entrance to the body of the person attacked and multiplies within it is that which presents itself as most probable, and it hardly seems worth while to consider any other, unless this is proved by a complete investigation not to be true.

Naturally the attention of investigators has first been given to a search for the "germ" in the blood of those attacked and in the blood and tissues of the victims of the malady.

The researches made up to the present time have failed to demonstrate the constant presence of any microörganism in the blood and tissues of those attacked.

My own researches, recorded in the foregoing report, show that no such microörganism as Dr. Domingos Freire, of Brazil, has described in his published works, or as he presented to me as his yellow-fever germ at the time of my visit to Brazil, is found, as he asserts, in the blood and tissues of typical cases of yellow fever. There is no satisfactory evidence that the method of inoculation practised by Dr. Domingos Freire has any prophylactic value.

The claims of Dr. Carmona y Valle, of Mexico, to have discovered the specific cause of yellow fever have likewise no scientific basis, and he has failed to demonstrate the protective value of his proposed method of prophylaxis.

It is highly important, in the interest of science and of the public health, that further investigations be made by more exact methods which have

been perfected since Drs. Freire and Carmona made their researches, and with which they were evidently not familiar.

The failure thus far to find a specific microorganism in the blood or tissues makes it desirable that a thorough research should be made with reference to the microorganisms present in the alimentary canal, for it is possible that in yellow fever, as in cholera, the disease is induced by a microorganism which multiplies in this situation. Additional researches are also required before we can say definitely that there is no germ demonstrable in the blood and tissues. Having exhausted our resources by the method of direct examination, and by cultures from blood drawn during life, it is highly desirable that various culture media should be inoculated with material taken, with proper precautions, from the various organs at the earliest possible moment after death.

The results of subsequent investigations made by the writer in Cuba during the summers of 1888 and 1889 are given in the following summary statement from the Transactions of the Tenth International Medical Congress (Berlin, 1890):

Bacteriological Researches in Yellow Fever.

The report relates to investigations made in Havana, Cuba, during the summers of 1888 and 1889, in Decatur, Alabama, during the autumn of 1888, and in the pathological and biological laboratories of the Johns Hopkins University during the winters of 1888 and 1889.

Forty-two autopsies were made in typical cases of yellow fever and seventeen autopsies in other diseases for comparative researches.

Aërobic and anaërobic cultures were made from the blood, the liver, the kidney, the urine, the stomach, and the intestine.

The experimental data recorded in this report show that:

The specific infectious agent in yellow fever has not been demonstrated.

The most approved bacteriological methods fail to demonstrate the constant presence of any particular microorganism in the blood and tissues of yellow-fever cadavers.

The microorganisms which are sometimes obtained in cultures from the blood and tissues are present in comparatively small numbers; and the one most frequently found (*Bacterium coli commune*) is present in the intestine of healthy individuals, and consequently its occasional presence cannot have any etiological import.

A few scattered bacilli are present in the liver, and probably in other organs, at the moment of death. This is shown by preserving portions of liver, obtained at a recent autopsy, in an antiseptic wrapping.

At the end of twenty-four to forty-eight hours the interior of a piece of liver so preserved contains a large number of bacilli of various species, the most abundant being those heretofore mentioned as occasionally found in fresh liver tissue, viz., *Bacterium coli commune* and *Bacillus cadaveris*.

Blood, urine, and crushed liver tissue obtained from a recent autopsy are not pathogenic in moderate amounts for rabbits or guinea-pigs.

Liver tissue preserved in an antiseptic wrapping at a temperature of 28° to 30° C., for forty-eight hours, is very pathogenic for guinea-pigs when injected subcutaneously.

This pathogenic power appears to be due to the microorganisms present and to the toxic products developed as a result of their growth. It is not peculiar to yellow fever, inasmuch as material preserved in the same way at comparative autopsies, in which death resulted from accident or other diseases, has given a similar result.

Having failed to demonstrate the presence of a specific "germ" in the blood and tissues, it seems probable that it is to be found in the alimentary canal, as is the case in cholera. But the extended researches made, and recorded in the present report, show that the contents of the intestines of yel-

low-fever cases contain a great variety of bacilli, and not a nearly pure culture of a single species, as is the case in recent and typical cases of cholera.

Comparatively few liquefying bacilli are found in the *feces* discharged during life, or in the intestinal contents collected soon after death from yellow-fever cadavers. On the other hand, non-liquefying bacilli are very abundant.

The one most constantly and abundantly present is the *Bacterium coli commune* of Escherich.

This is associated with various other bacilli, some of which are strict anaërobics and some facultative anaërobics.

Among the facultative anaërobics is one—my *Bacillus X*—which has been isolated by the culture method in a considerable number of cases and may have been present in all. This bacillus has not been encountered in the comparative experiments made. It is very pathogenic for rabbits when injected into the cavity of the abdomen.

It is possible that this bacillus is concerned in the etiology of yellow fever, but no satisfactory evidence that this is the case has been obtained by experiments on the lower animals, and it has not been found in such numbers as to warrant the inference that it is the veritable infectious agent.

All other microörganisms obtained in pure cultures from yellow-fever cadavers appear to be excluded, either by having been identified with known species, or by having been found in comparative researches made outside of the area of yellow-fever prevalence, or by the fact that they have only been found in small numbers and in a limited number of cases.¹

Finally we remark that many facts relating to the origin and extension of yellow-fever epidemics give support to the inference that the specific infectious agent is present in the dejecta of those suffering from the disease, and that accumulations of faecal matter, and of other organic material of animal origin, furnish a suitable nidus for the development of the "germ" when climatic conditions are favorable for its growth.

It may be that such a nidus is essential, and that the culture media usually employed by bacteriologists do not afford a suitable soil for this particular microbe.

It is also possible that its development depends upon the presence of other microörganisms found in faecal matter, which give rise to chemical products required for the development of this one.

Some of the microörganisms present in the dejecta of yellow-fever patients, as shown by stained smear preparations, have not developed in the cultures made, either aerobic or anaërobic. One extremely slender filiform bacillus, which can only be seen with high powers and which is quite abundant in some of my preparations, has never been obtained in the cultures made, and no doubt there are others in the same category.

That the yellow-fever germ is a strict anaërobic, or that it will only grow in a special nidus, may be inferred from certain facts relating to the extension of epidemics.

There is no evidence that yellow fever is propagated by contamination of the supply of drinking water, as frequently, and probably usually, occurs in the case of typhoid fever and cholera. Moreover, epidemics extend in a more deliberate manner and are restricted within a more definite area than is the case with cholera and typhoid fever. It is usually at least ten days or two weeks after the arrival of an infected vessel or of a person sick with the disease before cases of local origin occur; and these cases occur in the immediate vicinity of the imported case or infected vessel. When the disease has effected a lodgment the area of infection extends slowly and usually has well-defined boundaries. In towns and cities having a common water sup-

¹The possibility, of course, remains that the specific infectious agent in yellow fever may belong to an entirely different class of microörganisms from the bacteria, or that it may be ultra-microscopic or not capable of demonstration in the tissues by the staining methods usually employed by bacteriologists.

ply one portion remains perfectly healthy, while another, and usually the most filthy portion, may be decimated by the scourge.

The experimental evidence recorded, and the facts just stated, seem to justify the recommendation that the dejecta of yellow-fever patients should be regarded as infectious material, and that such material should never be thrown into privy vaults or upon the soil until it has been completely disinfected.

This rule thoroughly enforced, together with an efficient quarantine service and proper attention to the sanitary police of our exposed seaport cities, would, I believe, effectually prevent this pestilential disease from again obtaining a foothold within the limits of the United States.

XVII.

CLASSIFICATION OF PATHOGENIC BACTERIA.

A. BACTERIA BELIEVED TO BE THE CAUSE OF INFECTIOUS DISEASES IN MAN.

Traumatic infections: *Staphylococcus pyogenes aureus* (No. 1); *Staphylococcus pyogenes albus* (No. 2); *Streptococcus pyogenes* (No. 5).

Gonorrhœal infections: *Micrococcus gonorrhœæ* (No. 6).

Croupous pneumonia: *Micrococcus pneumoniæ crouposæ* (No. 8).

Anthrax ("wool-sorters' disease"): *Bacillus anthracis* (No. 45).

Typhoid fever: *Bacillus typhi abdominalis* (No. 46).

Diphtheria: *Bacillus diphtheriæ* (No. 47).

Epidemic influenza: *Bacillus of influenza* (No. 52).

Tuberculosis: *Bacillus tuberculosis* (No. 53).

Leprosy: *Bacillus lepræ* (No. 54).

Glanders: *Bacillus mallei* (No. 55).

Tetanus: *Bacillus tetani* (No. 149).

Relapsing fever: *Spirillum Obermeieri* (No. 153).

Cholera: *Spirillum cholerae Asiaticæ* (No. 155).

B. BACTERIA ASSOCIATED WITH DISEASES OF MAN WHICH HAVE BEEN SUPPOSED, ON MORE OR LESS SATISFACTORY EVIDENCE, TO BE THE CAUSE OF THESE DISEASES.

Pneumonia: *Bacillus of Friedländer* (No. 7).

Meningitis: *Diplococcus intercellularis meningitidis* (No. 9); *Micrococcus pneumoniæ crouposæ* (No. 8); *Bacillus meningitidis purulentæ* (No. 131).

Biskra button—"Clou de Biskra": *Micrococcus of Heydenreich* (No. 26).

Pemphigus acutus: *Micrococcus of Demme* (No. 27).

Bright's disease: *Streptococcus of Manneberg* (No. 28); *Bacillus of Letzerich* (No. 109).

Endocarditis: *Staphylococcus pyogenes aureus* (No. 1); *Micrococcus pneumoniae crouposa* (No. 8); *Micrococcus endocarditidis rugatus* (No. 29); *Bacillus endocarditidis griseus* (No. 104); *Bacillus endocarditidis capsulatus* (No. 105).

Syphilis: *Bacillus* of Lustgarten (No. 56); *Bacillus* of Eve and Lingard; *Micrococcus* of Disse and Taguchi.

Rhinoscleroma: *Bacillus* of rhinoscleroma (?) (No. 58).

Erythema nodosum: *Bacillus* of Demme (No. 107).

Green diarrhoea of infants: *Bacillus* of Lesage (No. 106).

Noma: *Bacillus nomæ* (?) (No. 110).

Ozæna: *Bacillus foetidus ozænae* (No. 111).

Bronchitis: *Bacillus* of Lumnitzer (No. 112).

Sycosis: *Bacillus* of Tommasoli (No. 113).

Whooping cough: *Bacillus* of Afanassiew (No. 119).

Influenza: *Micrococcus* of Kirchner (No. 38); *Micrococcus* No. II. of Fischel (No. 39).

Measles: *Bacillus* of Canon.

Senile gangrene: *Bacillus* of Tricomi (No. 128).

Cystitis: *Bacillus septicus vesicæ* (No. 132).

Egyptian ophthalmia: *Bacillus* of Kartulis (No. 138).

Trachoma: *Micrococcus* of trachoma (?) (No. 17).

Purpura hæmorrhagica: *Bacillus* of Babes (No. 146); *Bacillus* of Kolb (No. 147); *Bacillus* of Tizzoni and Giovannini (No. 145).

Cholera infantum: *Proteus vulgaris* (No. 97).

Cholera nostras: *Spirillum* of Finkler and Prior (No. 156).

Peritonitis: *Bacillus coli communis* (No. 89); *Staphylococcus pyogenes aureus* (No. 1); *Streptococcus pyogenes* (No. 5).

Pleuritis: *Micrococcus pneumoniae crouposa* (No. 8).

C. BACTERIA PROVED TO BE THE CAUSE OF INFECTIOUS DISEASES IN THE LOWER ANIMALS.

Anthrax: *Bacillus anthracis* (No. 45).

Tuberculosis: *Bacillus tuberculosis* (No. 53).

Glanders: *Bacillus mallei* (No. 55).

Septicæmia in cattle—"Rinderseuche": *Bacillus septicæmiæ hæmorrhagicæ* (No. 61).

Swine plague—"Schweineseuche," Löffler and Schütz: *Bacillus septicæmiæ hæmorrhagicæ* (No. 61).

Symptomatic anthrax—"black leg" in cattle; *Bacillus* of symptomatic anthrax (No. 152).

Farcy in cattle: *Bacillus* of Nocard (No. 60).

Septicæmia in deer—"Wildseuche": *Bacillus septicæmiæ hæmorrhagicæ* (No. 61).

Hog cholera : Bacillus of hog cholera (No. 63).

Swine plague, Marseilles : Bacillus of swine plague (No. 65).

Septicæmia in ferrets : Bacillus of swine plague, Marseilles (No. 65).

"*Myko-desmoids*" of the horse : Micrococcus botryogenus (No. 19).

Septicæmia in geese : Spirillum anserum (No. 154).

Bovine mastitis : Micrococcus of bovine mastitis (No. 21); Streptococcus of mastitis in cows (No. 31).

Mastitis in sheep : Micrococcus of gangrenous mastitis in sheep (No. 30).

Pneumonia in horses : Diplococcus of pneumonia in horses (No. 32).

Contagious coryza in horses—"Druse der Pferde" : Streptococcus coryzæ contagiosæ equorum (No. 33).

Hog erysipelas—"rouget," "rothlauf" : Bacillus erysipelatos suis (No. 67).

Septicæmia in parrots—"gray parrot disease" : Streptococcus perniciosus psittacorum (No. 43).

Diphtheria in pigeons : Bacillus diphtheriæ columbarum (No. 49).

Fowl cholera : Bacillus septicæmiæ hæmorrhagicæ (No. 61); Bacillus gallinarum (No. 77).

Cholera in ducks : Bacillus of Cornil and Toupet (No. 62).

Grouse disease : Bacillus of grouse disease (No. 76).

Septicæmia in fowls : Spirillum Metschnikovi (No. 158).

Septicæmia in frogs, etc. : Bacillus hydrophilus fuscus (No. 81).

Infectious diseases of bees : Bacillus alvei (No. 140); Bacillus of Canestrini (No. 295).

Infectious diseases of silkworms : Streptococcus bombycis (No. 24); Nosema bombycis (No. 25).

D. BACTERIA FROM VARIOUS SOURCES FOUND BY EXPERIMENT TO BE MORE OR LESS PATHOGENIC FOR LOWER ANIMALS.

From the pus of abscesses and open wounds : ..

Staphylococcus pyogenes aureus (No. 1).

Staphylococcus pyogenes albus (No. 2).

Streptococcus pyogenes (No. 5).

Staphylococcus pyosepticus (No. 42).

Bacillus of Belfanti and Pascarola (No. 64).

Bacillus pyogenes fœtidus (No. 72).

Bacillus pyocyanus (No. 95).

Micrococcus of Heydenreich (No. 26).

Proteus of Karlinsky (No. 98).

From infected animals :

Micrococcus of bovine pneumonia (No. 22).

Hæmatococcus bovis (No. 34).

Pseudo-diplococcus pneumoniae (No. 36).

Bacillus of Ribbert (No. 51).

Bacillus enteritidis (No. 75).

Bacillus capsulatus (No. 80).

Bacillus of Schou (No. 114).

Pneumobacillus liquefaciens bovis (No. 120).

From the buccal, nasal, or bronchial secretions of man :

Bacillus of Friedländer (No. 7).

Micrococcus pneumoniae crouposæ (No. 8).

Staphylococcus salivarius pyogenes (No. 10).

Micrococcus salivarius septicus (No. 15).

Micrococcus tetragenus (No. 18).

Micrococcus of Manfredi (No. 20).

Micrococcus gingivæ pyogenes (No. 35).

Micrococcus No. II. of Fischel (No. 39).

Bacillus of rhinoscleroma (No. 58).

Bacillus crassus sputigenus (No. 71).

Bacillus smaragdinus foetidus (No. 78).

Bacillus tenuis sputigenus (No. 82).

Bacillus of Lumnitzer (No. 112).

Bacillus of Afanassiew (No. 119).

Bacillus gingivæ pyogenes (No. 122).

Bacillus dentalis viridans (No. 123).

Bacillus pulpæ pyogenes (No. 124).

From fæces :

Bacillus coprogenus parvus (No. 68).

Bacillus cavicida (69).

Bacillus coli communis (No. 89).

Bacillus lactis aërogenes (No. 90).

Bacillus leporis lethalis (No. 94).

Bacillus of Lesage (No. 106).

Bacillus coprogenes foetidus (No. 116).

Bacillus of Gessner (No. 133).

Spirillum of Finkler and Prior (No. 156).

From urine :

Streptococcus of Manneberg (No. 28).

Bacillus of Letzerich (No. 109).

Bacillus septicus vesicæ (No. 132).

From cadavers and from putrefying material :

Diplococcus intercellularis meningitidis (No. 9).

Streptococcus septicus liquefaciens (No. 37).

Bacillus of Koubasoff (No. 59).

Bacillus cavicida Havaniensis (No. 70).

Proteus hominis capsulatus (No. 73).

Bacillus erysipelatos suis (mouse septicæmia) (No. 67).

Bacillus septicæmiæ hæmorrhagicæ (rabbit septicæmia) (No. 61).

Proteus capsulatus septicus (No. 74).

Bacillus pneumosepticus (No. 79).

Bacillus acidiformans (No. 92).

Bacillus cuniculicida Havaniensis (93).

Bacillus pseudo-tuberculosis (No. 121).

Bacillus septicus keratomalaciæ (No. 125).

Bacillus septicus acuminatus (No. 126).

Bacillus septicus ulceris gangrænosi (No. 127).

Bacillus albus cadaveris (No. 129).

Bacillus meningitidis purulentæ (No. 131).

Bacillus chromo-aromaticus (No. 134).

Bacillus cadaveris (No. 151).

Proteus vulgaris (No. 97).

Proteus mirabilis (No. 99).

Proteus Zenkeri (No. 100).

Proteus septicus (No. 101).

Proteus lethalis (No. 102).

From the soil :

Streptococcus septicus (No. 23).

Bacillus septicus agrigenus (No. 66).

Bacillus œdematis aërobicus (No. 108).

Bacillus œdematis maligni (No. 150).

Bacillus tetani (No. 149).

Bacillus muscoides (No. 401).

From various sources :

Bacillus oxytocus perniciosus (No. 117)—from milk.

Bacillus saprogenes II. (No. 118)—from sweating feet.

Bacillus canalis capsulatus (No. 135)—from sewers.

Bacillus canalis parvus (No. 136)—from sewers.

Bacillus indigogenus (No. 137)—from leaves of the indigo plant.

Bacillus of Utpadel (No. 139)—from dust.

Bacillus of Okada (No. 144)—from dust.

Spirillum tyrogenum (No. 157)—from cheese.

Micrococcus endocarditidis rugatus (No. 29)—cardiac valves.

Bacillus endocarditidis griseus (No. 104)—cardiac valves.

Bacillus endocarditidis capsulatus (No. 105)—cardiac valves.

PART FOURTH.

SAPROPHYTES.

- I. BACTERIA IN THE AIR. II. BACTERIA IN WATER. III. BACTERIA IN THE SOIL. IV. BACTERIA ON THE SURFACE OF THE BODY AND OF EXPOSED MUCOUS MEMBRANES. V. BACTERIA OF THE STOMACH AND INTESTINE. VI. BACTERIA OF CADAVERS AND OF PUTREFYING MATERIAL FROM VARIOUS SOURCES. VII. BACTERIA IN ARTICLES OF FOOD. VIII. NON-PATHOGENIC MICROCOCCI. IX. NON-PATHOGENIC BACILLI. X. NON-PATHOGENIC SPIRILLA. XI. LEPTOTRICHÆ AND CLADOTRICHÆ. XII. ADDITIONAL SPECIES OF BACTERIA NOT CLASSIFIED. XIII. BACTERIOLOGICAL DIAGNOSIS.

I.

BACTERIA IN THE AIR.

THE saprophytic bacteria are found wherever the organic material which serves as their pabulum is exposed to the air under conditions favorable to their growth. The essential conditions are presence of moisture and a suitable temperature. The organic material may be in solution in water or in the form of moist masses of animal or vegetable origin, and the temperature may vary within considerable limits— 0° to 70° C. But the species which takes the precedence will depend largely upon special conditions. Thus certain species multiply abundantly in water which contains comparatively little organic pabulum, and others require a culture medium rich in albuminous material or in carbohydrates; some grow at a comparatively low or high temperature, while others thrive only at a temperature of 20° to 40° C. or have a still more limited range; some require an abundant supply of oxygen, and others will not grow in the presence of this gas. Our statement that saprophytic bacteria are found wherever the organic material which serves as their pabulum is exposed to the air—under suitable conditions—relates to the fact that it is through the air that these bacteria are distributed and brought in contact with exposed material. It is a matter of common laboratory experience that sterilized organic liquids quickly undergo putrefactive decomposition when freely exposed to the air, and may be preserved indefinitely when protected from the germs suspended in the air by means of a cotton air filter. But the organic pabulum required for the nourishment of these bacteria is not found in the air in any considerable amount, and if they ever multiply in the atmosphere it must be under very exceptional conditions. Their presence is due to the fact that they are wafted from surfaces where they exist in a desiccated condition, and, owing to their levity, are carried by the wind to distant localities. But, under the law of gravitation, when not exposed to the action of currents of air they constantly fall again upon exposed surfaces, which, if moist, retain them, or from which, if dry, they are again wafted by the next current of air. Under these circumstances it is easy to understand why, as deter-

mined by investigation, more bacteria are found near the surface of the earth than at some distance above the surface, more over the land than over the ocean, more in cities with their dust-covered streets than in the country with its grass-covered fields.

Careful experiments have shown that bacteria do not find their way into the atmosphere from the surface of liquids, unless portions of the liquid containing them are projected into the air by some mechanical means, such as the bursting of bubbles of gas. Cultures of pathogenic bacteria freely exposed to the air in laboratories do not endanger the health of those who work over them; but if such a culture is spilled upon the floor and allowed to remain without disin-

fection, when it is desiccated the bacteria contained in it will form part of the dust of the room and might be dangerous to its occupants. Bacteria do not escape into the air from the surface of the fluid contents of sewers and cesspools, but changes of level may cause a deposit upon surfaces, which is rich in bacteria, and when dried this material is easily carried into the atmosphere by currents of air.

Tyndall's experiments (1869) show that in a closed receptacle in which the air is perfectly still all suspended particles are after a time deposited on the floor of the closed air chamber. And common experience demonstrates the fact that the dust of the atmosphere is carried by the wind from exposed surfaces and again deposited when the air is at rest. This dust as deposited, for example, in our dwellings contains innumerable bacteria in a desiccated condition, and the smallest quantity of it introduced into a sterile organic liquid will cause it to undergo putrefactive decomposition, and by bacteriological methods it will be found to contain various species of bacteria. Such dust also contains the spores of various mould fungi which are present in the atmosphere, usually in greater numbers than the bacteria. The mould fungi are air plants

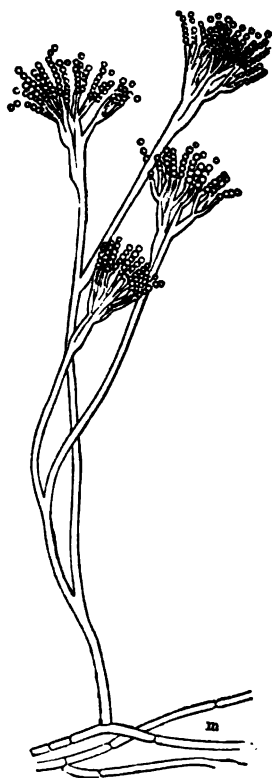


FIG. 186. — *Penicillium glaucum*; *m*, mycelium, from which is given off a branching pedicel bearing spores. $\times 150$.

which vegetate upon the surface of moist organic material and form innumerable spores, which are easily wafted into the air, both on account of their low specific gravity and minute size, and because they

are borne upon projecting pedicles by which they are removed from the moist material upon which and in which the mycelium develops (Fig. 186), and, being dry, are easily carried away by currents of air.

Bacteriologists have given much attention to the study of the microorganisms suspended in the atmosphere, with especial reference to hygienic questions. The methods and results of these investigations will be considered in the present section.

Pasteur (1860) demonstrated the presence of living bacteria in the atmosphere by aspirating a considerable quantity of air through a filter of gun-cotton or of asbestos contained in a glass tube. By dissolving the gun-cotton in alcohol and ether he was able to demonstrate the presence of various microorganisms by a microscopical examination of the sediment, and by placing the asbestos filters in sterilized culture media he proved that living germs had been filtered out of the air passed through them.

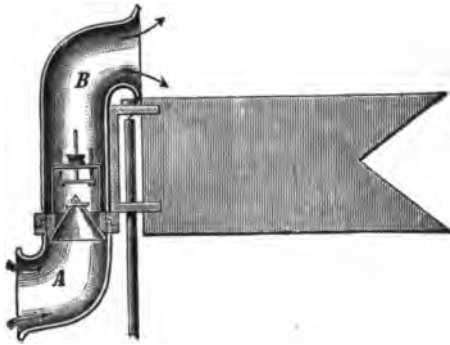


FIG. 187.

A method employed by several of the earlier investigators consisted in the collection of atmospheric moisture precipitated as dew upon a surface cooled by a freezing mixture. This was found to contain living bacteria of various forms. The examination of rain water, which in falling washes the suspended particles from the atmosphere, gave similar results.

The first systematic attempts to study the microorganisms of the air were made by Maddox (1870) and by Cunningham (1873), who used an aëroscope which was a modification of one previously described by Pouchet. In the earlier researches of Miquel a similar aëroscope was used. This is shown in Fig. 187. The opening to the cylindrical tube A is kept facing the wind by means of a wind vane, and when the wind is blowing a current passes through a small aperture in a funnel-shaped partition which is properly placed in the cylindrical tube. A glass slide, upon the lower surface of which a

mixture of glycerin and glucose has been placed, is adjusted near the opening of the funnel, at a distance of about three millimetres, so that the air escaping through the small orifice is projected against it. By this arrangement a considerable number of the microorganisms present in the air, as well as suspended particles of all kinds, are arrested upon the surface of the slide and can be examined under the microscope or studied by bacteriological methods. But an aëroscope of this kind gives no precise information as to the number of living germs contained in a definite quantity of air. The microscopical examination also fails to differentiate the bacteria from particles of various kinds which resemble them in shape, and the microorganisms seen are for the most part spores of various fungi mingled with pollen grains, vegetable fibres, plant hairs, starch granules, and amorphous granular material.

Another method, which has been employed by Cohn, Pasteur, Miquel, and others, consists in the aspiration of a definite quantity of air through a culture liquid, which is then placed in an incubating oven for the development of microorganisms washed out of the air which has been passed through it. This method shows that bacteria of different species are present, but gives no information as to their relative number, and requires further researches by the plate method to determine the characters of the several species in pure cultures.

A far simpler method consists in the exposure of a solid culture medium, which has been carefully sterilized and allowed to cool on a glass plate or in a Petri's dish, for a short time in the air to be examined. Bacteria and mould fungi deposited from the air adhere to the surface of the moist culture medium, and form colonies when the plate, enclosed in a covered glass dish, is placed in the incubating oven. The number of these colonies which develop after exposure in the air for a given time enables us to estimate in a rough way the number of microorganisms present in the air of the locality where the exposure was made; and the variety of species is determined by examining the separate colonies, each of which is, as a rule, developed from a single germ. By exposing a number of plates at different times this method enables us to determine what species are most abundant in a given locality and the comparative number in different localities, as determined by counting the colonies after exposure for a definite time—*e.g.*, ten minutes. Of course we will only obtain evidence of the presence of such aërobic bacteria as will grow in our culture medium. The anaërobic bacteria may be studied by placing plates exposed in a similar way in an atmosphere of hydrogen. Bacteria which grow slowly and only under special conditions, like the tubercle bacillus, would be likely to escape observation, as the mould fungi and common saprophytes would take complete pos-

session of the surface of the culture medium before the others had formed visible colonies. Students will do well to employ this simple and satisfactory method for the purpose of making themselves familiar with the more common atmospheric organisms, and they will find the shallow glass dishes with a cover, known as Petri's dishes, very convenient for the purpose. These dishes should be sterilized in the hot-air oven and sufficient sterile nutrient gelatin or agar poured into them to cover the bottom. After the culture medium has become solid by cooling, the exposure may be made by simply removing the cover and replacing it at the end of the time fixed upon.



FIG. 188.

To determine in a more exact way the number of microorganisms contained in a given quantity of air will require other methods. But we may say, *en passant*, that such a determination is usually not of great scientific importance. The number is subject to constant fluctuations in the same locality, depending upon the force and direction of the wind. If we have on one side of our laboratory a dusty street and on the other a green field, more bacteria will naturally be found when the wind blows from the direction of the street than when it comes from the opposite direction; or, if the air is filled with dust from recently sweeping the room, we may expect to find very

many more than when the room has been undisturbed for some time. The painstaking researches which have already been made have established in a general way the most important facts relating to the distribution of atmospheric bacteria, but have failed to show any definite relation between the number of atmospheric bacteria and the prevalence of epidemic diseases. In the apparatus of Hesse, Fig. 188, a glass tube, having a diameter of four to five centimetres and a length of half a metre to a metre, is employed. In use this is supported upon a tripod, as shown in the figure, and air is drawn through it by a water aspirator consisting of two flasks, also shown. The upper flask being filled with water, this flows into the lower flask by siphon action, and upon reversing the position of the flasks number one is again filled. By repeating this operation as many times as desired a quantity of air corresponding with the amount of water passed from the upper to the lower flask is slowly aspirated through the horizontal glass tube. The microorganisms present are deposited upon nutrient gelatin previously allowed to cool upon the lower portion of the large glass tube. The air enters through a small opening in a piece of sheet rubber which is tied over the extremity of the horizontal tube, and before the aspiration is commenced this opening is covered by another piece of sheet rubber tied over the first. Experience shows that when the air is slowly aspirated most of the germs contained in it are deposited near the end of the tube through which it enters. The colonies which develop upon the nutrient gelatin show the number and character of living microorganisms contained in the measured quantity of air aspirated through the apparatus. The method with a soluble filter of pulverized sugar, to be described hereafter, is preferable when exact results are desired; and for the purpose of determining the relative abundance and the variety of microorganisms present in the atmosphere of a given locality the exposure of nutrient gelatin in Petri's dishes is far simpler, and, as a rule, will furnish all the information that is of real value.

In his extended researches made at the laboratory of Montsouri, in Paris, Miquel has used various forms of apparatus and has obtained interesting results; but his method of *ensemencements fractionnés* requires a great expenditure of time and patience, and the more recent method with soluble filters is to be preferred.

In his latest modification of the method referred to Miquel used a flask like that shown in Fig. 189. From twenty to forty cubic centimetres of distilled water are introduced into this flask. The cap A contains a cotton air filter and is fitted to the neck of the flask by a ground joint. This is removed during the experiment. The tube C is connected with an aspirator. It contains two cotton or asbestos

filters, *c* and *b*. The cap being removed and the aspirator attached, the air is drawn through the water, by which suspended germs are arrested; or if not they are caught by the inner cotton plug *b*. The sealed point of the tube *B* is now broken off, and the contents of the flask equally divided in thirty to forty tubes containing bouillon, which are placed in the incubating oven. Twenty-five cubic centimetres of bouillon are also introduced into the flask, and the cotton plug *b* is pushed into it so that any bacteria arrested by it may develop. If one-fourth or one-fifth of the bouillon tubes show a development of bacteria it is inferred that each culture originated from a single germ, and the number present in the amount of air drawn through the flask is estimated from the number of tubes in which development occurs.

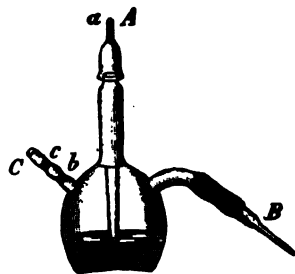


FIG. 189.

The method adopted by Straus and Würtz is more convenient and more reliable in its results. This consists in passing the air by means of an aspirator through liquefied nutrient gelatin or agar. The apparatus shown in Fig. 190 is used for this purpose. Two cotton plugs are placed in the tube *B*, to which the aspirator is attached, and after the determined quantity of air has been passed through the liquefied medium the inner plug is pushed down with a sterilized

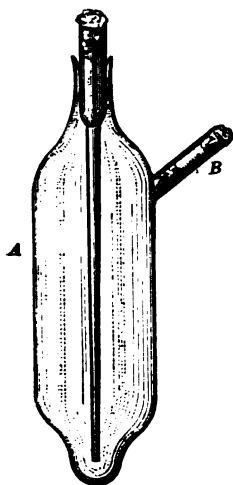


FIG. 190.

platinum needle so as to wash out in the culture medium any germs arrested by it. Finally the gelatin or agar is solidified upon the walls of the tube *A* by rotating it upon a block of ice or under a stream of cold water. It is now put aside for the development of colonies, which are counted to determine the number of germs present in the quantity of air passed through the liquefied culture medium. The main difficulty with this apparatus is found in the fact that the nutrient gelatin foams when air is bubbled through it; for this reason an agar medium is to be preferred. In using this it will be necessary to place the liquefied agar in a bath maintained at 40° C. Foaming of the gelatin is prevented by adding a drop of olive oil before sterilization in the steam sterilizer. But this interferes with the transparency of the medium.

In the earlier experiments upon atmospheric organisms Pasteur used a filter of asbestos, which was subsequently washed out in a

culture liquid. A filter of this kind washed out in liquefied gelatin or nutrient agar would give more satisfactory results, as the culture medium could be poured upon plates or spread upon the walls of a test tube and the colonies counted in the usual way. Petri prefers to use a filter of sand, which he finds by experiment arrests the microorganisms suspended in the atmosphere, and which is subsequently distributed through the culture medium. The sand used is such as



FIG. 191.

has been passed through a wire sieve having openings of 0.5 millimetre in diameter. This is sterilized by heat, and is supported in a cylindrical glass tube by small wire-net baskets. The complete arrangement is shown in Fig. 191. Two sand filters, *c*, and *c*₂, are used, the lower one of which serves as a control to prove that all microorganisms present in the air have been arrested by the upper one. The upper filter is protected, until the aspirator attached to the tube *h* is put in operation, by a sterile cotton plug, not shown in the figure which represents the filter in use. Petri uses a hand air pump as an aspirator, and passes one hundred litres of air through the sand in from ten to twenty minutes. The sand from the two filters is then distributed in shallow glass dishes and liquefied gelatin is poured over it; this is allowed to solidify and is put aside for the development of colonies. The principal objection to this method is the presence of the opaque particles of sand in the culture medium. This objection has been overcome by the use of soluble filters, a method first employed by Pasteur and since perfected by Sedgwick and by Miquel. The most useful material for the purpose appears to be cane sugar, which can be sterilized in the hot-air oven at 150° C. without undergoing any change in its physical characters. Loaf sugar is pulverized in a mortar and passed through two sieves in order to remove the coarser grains and the

very fine powder, leaving for use a powder having grains of about one-half millimetre in diameter. This powdered sugar is placed in a glass tube provided with a cap having a ground joint and a cotton plug to serve as an air filter (*A*, Fig. 192), or in a tube such as is shown at *B*, having the end drawn out and hermetically sealed. Two cotton plugs are placed at the lower portion of the tube, at *a* and at *b*.

Glass tubing having a diameter of about five millimetres is used in making these tubes, and from one to two grammes of powdered sugar is a suitable quantity to use as a filter. The whole apparatus is sterilized for an hour at 150°C . in a hot-air oven after the pulverized sugar has been introduced. Before using it will be necessary to pack the sugar against the supporting plug *a* by gently striking the lower end of the tube, held in a vertical position, upon some horizontal surface; and during aspiration the tube must remain in a vertical position, or nearly so, in order that the sugar may properly fill its entire calibre. The aspirator is attached to the lower end of the tube by a piece of rubber tubing. When the tube B is used the sealed extremity is broken off at the moment that the aspirator is set in action, and it is again sealed in a flame after the desired amount of air has been passed through the filter. The next step consists in dissolving the sugar in distilled water or in liquefied gelatin. To insure the removal of all the sugar the cotton plug *a* may be pushed out with a sterilized glass rod, after removing *b* with forceps. From fifty to five hundred cubic centimetres of distilled water, contained in an Erlenmeyer flask and carefully sterilized, may be used, the amount required depending upon circumstances relating to the conditions of the experiment. By adding five or ten cubic centimetres of this water, containing the sugar and microorganisms arrested by it, to nutrient gelatin or agar liquefied by heat, and then making Es-march roll tubes, the number of germs in the entire quantity is easily estimated by counting the colonies which develop in the roll tubes.

Sedgwick and Tucker, in a communication made to the Boston Society of Arts, January 12th, 1888, were the first to propose the use of a soluble filter of granulated sugar for collecting atmospheric germs. Their complete apparatus consists of an exhausted receiver, from which a given quantity of air is withdrawn by means of an air pump. A vacuum gauge is attached to the receiver, which is coupled

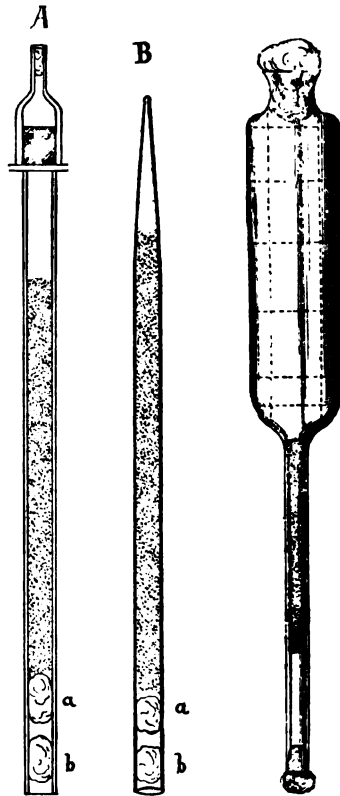


FIG. 192.

FIG. 193.

with the glass tube containing the granulated-sugar filter by a piece of rubber tubing. Instead of transferring the soluble filter to gelatin in test tubes, they use a large glass cylinder having a slender stem, in which the sugar is placed (Fig. 193). After the aspiration liquefied gelatin is introduced into the large glass cylinder, which is held in a horizontal position; the sterilized cotton plug is then replaced in the mouth of the cylinder, the sugar is pushed into the liquefied gelatin and dissolved, and by rotating the cylinder upon a block of ice the gelatin is spread upon its walls as in an Esmarch roll tube. For convenience in counting the colonies lines are drawn upon the surface of the cylinder, dividing it into squares of uniform dimensions.

GENERAL RESULTS OF RESEARCHES MADE.

As already stated, the presence of bacteria in the atmosphere depends upon their being wafted by currents of air from surfaces where they are present in a desiccated condition. That they are not carried away from moist surfaces is shown by the fact that expired air from the human lungs does not contain microorganisms, although the inspired air may have contained considerable numbers, and there are always a vast number present in the salivary secretions. The moist mucous membrane of the respiratory passages constitutes a germ trap which is much more efficient than the glass slide smeared with glycerin used in some of the aërosopes heretofore described, for it is a far more extended surface. As a matter of fact, most of the suspended particles in inspired air are deposited before the current of air passes through the larynx.

Air which passes over large bodies of water is also purified of its germs and other suspended particles. The researches of Fischer show that at a considerable distance from the land no germs are found in the atmosphere over the ocean, and that it is only upon approaching land that their presence is manifested by the development of colonies upon properly exposed gelatin plates.

Uffelmann found, in his researches, that in the open fields the number of living germs in a cubic metre of air averaged two hundred and fifty, on the sea coast the average was one hundred, in the courtyard of the University of Rostock four hundred and fifty. The number was materially reduced after a rainfall and increased when a dry land wind prevailed.

Frankland found that fewer germs were present in the air in winter than in summer, and that when the earth was covered with snow the number was greatly reduced, as also during a light fall of snow; the air of towns was found to be more rich in germs than the

air of the country ; the lower strata of the atmosphere contained more than the air of elevated localities.

Von Freudenreich also found that the air of the country contained fewer germs than that of the city. Thus in the city of Berne a cubic metre of air often contained as many as two thousand four hundred germs, while the maximum in country air was three hundred. His results corresponded with those of Miquel in showing that the number of atmospheric organisms is greater in the morning and the evening, between the hours of 6 and 8, than during the rest of the day. Neumann, whose researches were made in the Moabite Hospital, found the greatest number of bacteria in the air in the morning after the patients able to sit up had left their beds and the wards had been swept. The number of germs was then from eighty to one hundred and forty in ten litres of air, while in the evening the number fell to four to ten germs in ten litres.

Miquel has given the following summary of results obtained in his extended experiments, made in Paris during the years 1881, 1882, and 1883 :

	Number of Germs in a Cubic Metre of Air.	
	Air of Laboratory, Montsouris.	Air of Park, Mont souris.
Average for 1880.....	215	71
" " 1881.....	348	62
" " 1882.....	550	51

Rue de Rivoli, average for one year, 750 ; summit of Pantheon, 28 ; Hôtel-Dieu, 1880, average for four months, male ward 6,300, female ward 5,120 ; La Piété Hospital, average of fifteen months, 11,100.

It must be remembered that the figures given relate both to bacteria and to the spores of mould fungi, and that the latter are commonly the most numerous when the experiment is made in the open air. Petri has shown that when gelatin plates are exposed in the air the relative number of spores of mould fungi deposited upon them is less than is obtained in aspiration experiments.

The number of colonies which develop on exposed plates does not represent the full number of bacteria deposited, for these colonies very frequently have their origin in a dust particle to which several bacteria are attached, or in a little mass of organic material containing a considerable number.

It is generally conceded that sea air and country air are more wholesome than the air of cities, and especially of crowded apartments, in which the number of bacteria has been shown to be very much greater. But it would be a mistake to ascribe the sanitary value of sea, country, and mountain air to the relatively small num-

ber of bacteria present in such air. There are other important factors to be considered, and we have no satisfactory evidence that the number of saprophytic bacteria present in the air has an important bearing upon the health of those who respire it. We do know that the confined air of crowded apartments, and especially of factories in which a large quantity of dust is suspended in the air, predisposes those breathing such air to pulmonary diseases and lowers the general standard of health. But it has not been proved that this is due to the presence of bacteria. Infectious diseases may, under certain circumstances, be communicated by way of the respiratory passages as a result of breathing air containing in suspension pathogenic bacteria; but there is reason to believe that this occurs less frequently than is generally supposed.

Krüger has shown that the dust of a hospital ward in which patients with pulmonary consumption expectorated occasionally upon the floor contained tubercle bacilli. This was proved by wiping up the dust on a sterilized sponge, washing this out in bouillon, and injecting this into the cavity of the abdomen of guinea-pigs. Two animals out of sixteen injected became tuberculous. In pulmonic anthrax, which occasionally occurs in persons engaged in sorting wool—"wool-sorters' disease"—infection occurs as a result of the respiration of air containing the spores of the anthrax bacillus.

Among the non-pathogenic saprophytes found in the air certain aërobic micrococci appear to be the most abundant, and, as a rule, bacilli are not found in great numbers or variety. In some localities various species of sarcinæ are especially abundant. The following is a partial list of the species which have been shown by the researches of various bacteriologists to be occasionally present in the air. But, as heretofore remarked, their presence is to be regarded as accidental, and so far as we know there is no bacterial flora properly belonging to the atmosphere:

Micrococcus ureæ (Pasteur), *Diplococcus roseus* (Bumm), *Diplococcus citreus conglomeratus* (Bumm), *Micrococcus radiatus* (Flügge), *Micrococcus flavus desidens* (Flügge), *Micrococcus flavus liquefaciens* (Flügge), *Micrococcus tetragenus versatilis* (Sternberg), *Micrococcus pyogenes aureus* (Rosenbach), *Micrococcus pyogenes citreus* (Passet), *Micrococcus cinnabæus* (Flügge), *Micrococcus flavus tardigradus* (Flügge), *Micrococcus versicolor* (Flügge), *Micrococcus viticulosus* (Katz), *Micrococcus candidans* (Flügge), *Pediococcus cerevisiæ* (Balcke), *Sarcina lutea* (Schröter), *Sarcina rosea* (Schröter), *Sarcina aurantiaca*, *Sarcina alba*, *Sarcina candida* (Reinke), *Bacillus tumescens* (Zopf), *Bacillus subtilis* (Ehrenberg), *Bacillus multipediculosus* (Flügge), *Bacillus mesentericus fuscus* (Flügge), *Bacillus mesentericus ruber* (Globig), *Bacillus inflatus* (A. Koch), *Bacillus mesentericus vulgaris*, *Bacillus prodigiosus*, *Bacillus aërophilus* (Liborius), *Bacillus pestifer* (Frankland), *Spirillum aureum* (Weibel), *Spirillum flavescens* (Weibel), *Spirillum flavum* (Weibel), *Bacillus Havaniensis* (Sternberg).

In the recent researches of Welz, made in the vicinity of Freiburg, twenty-three different micrococci and twenty-two bacilli were obtained from the air.

II.

BACTERIA IN WATER.

THE water of the ocean, of lakes, ponds, and running streams necessarily contains bacteria, as they are constantly being carried into it by currents of air passing over the neighboring land surfaces, and by rain water which washes suspended microorganisms from the atmosphere; and, as such water contains more or less organic material in solution, many of the saprophytic bacteria multiply in it abundantly. It is only in the water of springs and wells which comes from the deeper strata of the soil that they are absent. The number and variety of species present in water from any given source will depend upon conditions relating to the amount of organic pabulum, the temperature, the depth of the water, the fact of its being in motion or at rest, its pollution from various sources, etc. The comparatively pure water of lakes and running streams contains a considerable number of bacteria which find their normal habitat in such waters and which multiply abundantly in them, notwithstanding the small quantity of organic matter and salts which they contain. The water of stagnant, shallow pools, and of sluggish streams into which sewage is discharged, contains a far greater number and a greater variety of species.

The study of these bacteria in water has received much attention on account of the sanitary questions involved, relating to the use of water from various sources for drinking purposes. In the present section we shall first give an account of the methods of bacteriological water analysis, and then a condensed statement of results obtained in the very numerous investigations which have been made.

A very important point to be kept in view is the fact that a great increase in the number of bacteria present, in samples of water collected for investigation, is likely to occur if these samples are kept for some time. A water which, for example, contains only two hundred to three hundred bacteria per cubic centimetre when the examination is made at once, may contain several thousand at the end of twenty-four hours, and at the end of the second or third day twenty thousand or more may be present in the same quantity.

Later, on account of the exhaustion of organic pabulum, the number is again reduced as the bacteria present gradually lose their vitality. Under these circumstances it is evident that an estimate of the number of bacteria present in water from a given source can have no value, unless a sample is tested by bacteriological methods within a short time after it has been collected. Not more than an hour or two should be allowed to elapse, especially in warm weather. By placing the water upon ice the time may be extended somewhat, but Wolffhügel has shown that the number of germs is gradually diminished when water is preserved in this way, and it will be safest to make an immediate examination when this is practicable.

The *collection* may be made in a sterilized Erlenmeyer flask provided with a cotton air filter, or in a bottle having a ground-glass stopper which has been wrapped in tissue paper and sterilized for an hour or more at 150° C. in the hot-air oven. Or the small flasks with a long neck may be used, as first recommended by Pasteur. These are prepared as follows : The bulb is first gently heated, and the extremity of the tube dipped into distilled water, which mounts into

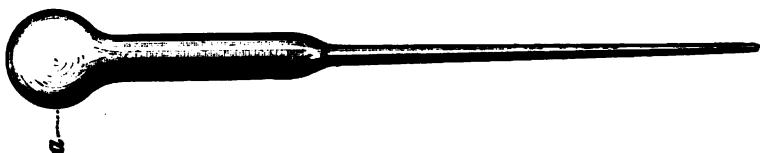


FIG. 194.

the bulb as it cools ; the water is then made to boil, and when all but a drop or two has escaped and the bulb is filled with steam the extremity of the tube is hermetically sealed. When the steam has condensed by the cooling of the bulb a partial vacuum is formed, and the tube is ready for use at any time. It is filled with water by breaking off the sealed extremity under the surface of the water of which a sample is desired. This is done with sterilized forceps, and care must be taken that the exterior of the tube is properly sterilized before the collection is made. The end is immediately sealed in the flame of a lamp. A difficulty with these vacuum tubes is that they are so completely filled with water that this cannot be readily drawn from them again in small quantities. The writer therefore prefers to make the collection in a tube shaped as shown in Fig. 194, in which a partial vacuum is formed just before the collection by heating the air in the bulb. The water mounts into the tube as the air in the bulb cools, and is readily forced out again for making cultures by applying gentle heat to the bulb. As a lamp is needed to seal the end of the tube in either case, there is no special advantage in having a vacuum formed in advance, and, as stated, the vacuum tubes are so

nearly filled with water that it is not so simple a matter to obtain the contents for our culture experiments without undue exposure to atmospheric germs. In practice small glass bottles with ground-glass stoppers will be found most convenient, and, when properly sterilized, are unobjectionable. They should be filled at a little distance below the surface, as there is often a deposit of dust upon the surface of standing water, and sometimes a delicate film made up of aerobic bacteria. When water is to be obtained from a pump or a hydrant it should be allowed to flow for some time before the collection is made. To collect water at various depths the apparatus shown in Fig. 195 is recommended by Lepsius. An iron frame supports an inverted flask, A, filled with sterilized mercury and containing about three hundred cubic centimetres. The flask B is intended to receive the mercury when, at the desired depth, it is allowed to flow through the capillary tube *b*. This is sealed at the extremity and bent as shown in the figure. By pulling upon the cord *c* this tube is broken, and as the mercury flows from the flask this is filled with water through the tube *a*. The extremity of the broken tube *b* is closed by the mercury in the flask B when A is full of water, and the apparatus can be brought to the surface with only such water as was collected at the depth from which a sample was desired.

The *bacteriological analysis* is made by adding a definite quantity of the water under investigation to liquefied gelatin or agar-gelatin, and making a plate or Esmarch roll tube, which is put aside for the development of colonies. Miquel and others have preferred to use liquid cultures and the method of fractional cultivation described in the previous section. The use of a solid culture medium has, however, such obvious advantages that we do not consider it necessary to do more than refer to the other method as one which, when applied with skill and patience, may give sufficiently accurate results.



FIG. 195.

The amount of water which should be added to the usual quantity of liquefied flesh-peptone-gelatin in a test tube, in order that the colonies which develop may be well separated from each other and easily counted, can only be determined by experiment. If the water is from an impure source a single drop may be too much, and it will be necessary to dilute it with distilled water recently sterilized. But for ordinary potable water it will usually be best, in a first experiment, to make two trials, one with one cubic centimetre and one with one-half cubic centimetre added to the liquefied nutrient gelatin. The water in the collecting bottle should be shaken, to distribute the bacteria which may have settled to the bottom, before drawing off by means of a sterilized pipette the amount used for the experiment, and the germs present in it are to be distributed through the liquefied gelatin by gently moving the tube to and fro.

Koch's method of preparing a gelatin plate is illustrated in Fig. 196. A glass dish, containing ice water and covered with a large

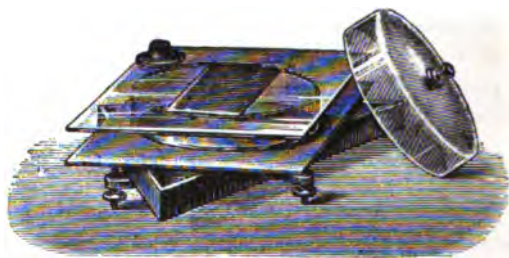


FIG. 196.

plate of glass, is supported upon a levelling tripod. By means of a spirit level this is adjusted to a horizontal position, so that when the liquefied gelatin is poured upon the smaller sterilized glass plate, seen in the centre of the large plate of glass, it will not flow, but may be evenly distributed over the surface by means of a sterilized glass rod. The glass cover resting against the side of the apparatus is placed over the gelatin plate while it is cooling, to protect it from atmospheric germs, and when the gelatin is hard the plate is transferred to a shallow glass dish, which is kept at a temperature of about 20° C. for several days for the development of colonies. It is difficult to count colonies when more than five thousand develop upon a plate of the usual size, and for this reason it will be best to repeat the experiment with a smaller quantity of water from the same source, if this is at hand, rather than to attempt to count an overcrowded plate. Before pouring the gelatin upon the plate the lip of the test tube containing it should be sterilized by passing it through a flame. The liquefied gelatin should be carefully distributed to cover a rect-

angular surface and leaving a margin of about one centimetre around the edge of the plate. The Koch's dish in which the gelatin plate is placed for the development of colonies should be carefully sterilized by heat or by washing it out with a sublimate solution. A circular piece of filtering paper, saturated with sublimate solution or distilled water, is placed at the bottom of the lower dish to keep the air in a moist condition and prevent drying of the gelatin. Usually two or three plates made at the same time are placed one above the other on glass supports made for this purpose. If many liquefying organisms are present it will be necessary to count the colonies before these run together—usually on the second day; but in the absence of liquefying colonies it is best to wait until the third, or even the fifth day, as the number of visible colonies and the ease of counting them will be greater than at an earlier date. The development of a few scattered liquefying colonies which threaten to spoil the plate may be arrested by taking up the liquefied gelatin from each with a bit of filtering paper, and then, by means of a camel's-hair brush, applying a solution of potassium permanganate to the margin of the colony. The growth of colonies of mould fungi, which have developed from spores from the atmosphere falling upon the plate while it is exposed, can be checked by the application of collodion containing bichloride of mercury.

Counting of the colonies is a simple matter when they are few in number; when they are numerous it is customary to place the plate over a dark background, and to place above it a glass plate divided into square centimetres by lines ruled with a diamond. By means of a lens of low power the colonies in a certain number of squares are counted and the average taken. This multiplied by the number of square centimetres in the gelatin-covered surface gives approximately the entire number of colonies which have developed from the amount of water used in the experiment.

Instead of using Koch's original plate method, as above described, the shallow, covered glass dishes recommended by Petri may be employed. These are from one to one and one-half centimetres high and from ten to fifteen centimetres in diameter. The liquefied gelatin is poured into the lower dish and the cover at once placed over it. The gelatin does not dry out very soon, but, if necessary, several of these Petri's dishes may be placed in a larger jar, which serves as a moist chamber.

The roll tubes of Esmarch may also be used, and have the advantage that accidental colonies from air-borne germs are excluded. The counting of colonies is not quite as easy, but by the use of a mounted lens especially designed for the purpose it is attended with no great difficulty. The surface of the tube is divided into squares

by colored lines, and the number of colonies in several squares is counted in order to obtain an average and estimate the entire number.

Water which contains numerous liquefying bacteria had better be examined by the use of nutrient agar instead of gelatin; and in very warm weather it will be necessary to use an agar medium, as ten-per-cent gelatin is likely to melt if the temperature goes above 22° C. A difficulty in the use of agar for plates consists in the liability of the film to slip from the glass. This may be remedied to some extent by adding a few drops of a concentrated solution of gum acacia to the liquefied agar medium. Petri's dishes are well adapted for the use of the agar medium, as the objection referred to does not apply to them. The gelatin-agar medium, containing 5 per cent of gelatin and 0.75 per cent of agar, may also be used with advantage in the bacteriological analysis of water. Much stress was at one time laid upon the enumeration of liquefying colonies, upon the supposition that the liquefying bacteria were especially harmful as compared with the non-liquefying, and that a water containing many liquefying colonies was to be looked upon with suspicion. We now know, however, that there are many common and harmless saprophytes which cause the liquefaction of gelatin, and that some of the most dangerous pathogenic bacteria do not liquefy gelatin. This distinction has therefore no special value, and the question for bacteriologists to-day is not how large is the comparative number of liquefying colonies, but what species are represented by the colonies present, liquefying and non-liquefying, and what are the special pathogenic properties of each. The answer to these questions, in the case of any particular water supply, calls for special knowledge and great patience and care in the isolation in pure cultures, and careful study of the various species present.

It is now generally recognized that a mere enumeration of the number of colonies which develop from a water under investigation is not a sufficient indication upon which to found an opinion as to its potability. An excessive number of bacteria is an indication that the water contains a large amount of the organic material which serves as pabulum for these microorganisms. But the chemists are able to determine the amount of organic matter present in water with greater precision; and, as we have seen, the number of bacteria may increase many-fold in water which is kept standing in the laboratory for two or three days in a well-corked bottle. As a matter of fact, the enumeration of bacteria in water, although it has given us results of scientific interest, has not materially added to the methods previously applied for estimating the sanitary value of water obtained from various sources for drinking purposes. But the bacte-

riological examination may prove to be of great value if it succeeds in demonstrating the presence of certain pathogenic bacteria and in thus preventing the use of a dangerous water. We do not mean to say, however, that an enumeration of the bacteria present in drinking water has no practical value. An excessive number indicates an excessive amount of organic pabulum, which may have come from a dangerous source; and the dangerous pathogenic bacteria are not only more likely to be present in such water, but they can more readily multiply in it, while in a pure water they would fail to increase in number, and, as has been shown by experiment, would die out within a short time.

The number of bacteria present in *rain water*, or in *snow* which has recently fallen, varies greatly at different times. Naturally the number is greater when the surface of the earth is dry and the atmosphere loaded with dust by currents of wind passing over it, and less when the surface is moist and the atmosphere has been purified by recent rains.

In snow from the surface of a glacier in Norway, Schmelck found two bacteria and two spores of mould fungi per cubic centimetre of water from the melted snow. Ganowski, in experiments made with freshly fallen snow collected in the vicinity of Kiew, obtained the following results: February 2d, 1888: temperature of the air, $7.2^{\circ}\text{C}.$; snowfall, 0.1 millimetre; number of bacteria in 1 cubic centimetre of water from melted snow, 34 in one sample and 38 in another. February 20th, 1888: temperature, $11.1^{\circ}\text{C}.$; snowfall, 1.1 millimetres; number of bacteria in one sample, 203, in another 384.

Miquel obtained from rain water collected at Montsouris during a rainy season 4.3 germs per cubic centimetre; in rain water collected in the centre of the city of Paris, 19 per cubic centimetre.

Hail has also been shown to contain bacteria in considerable numbers. Bujwid found in hailstones which fell at Warsaw 21,000 bacteria in 1 cubic centimetre; but this is exceptional, and is supposed to be due to the fact that surface water had been carried into the air by the storm and frozen. Fontin examined hail which fell in St. Petersburg, and obtained an average of 729 bacteria per cubic centimetre of water from the melted hail.

River water has been carefully examined by numerous bacteriologists in various localities and at different seasons of the year. We give below some of the results reported:

Water of the Seine at Choisy, before reaching Paris, 300; at Bercy, 1,200; at Saint-Denis, after receiving the sewer water from the city, 200,000 germs per cubic centimetre (Miquel).

Water of the Spree beyond Köpenick, 82,000; two hundred steps below the mouth of the Wuhle, 118,000; in Berlin above the mouth

of the Panke, 940,000; below the mouth of the Panke, 1,800,000 (Koch).

Water of the Main above the city of Würzburg, in the month of February, 520; below the city, 15,500 (Rosenberg).

Water of the Potomac, at Washington, in 1886: January, 3,774; February, 2,536; March, 1,210; April, 1,521; May, 1,064; June, 348; July, 255; August, 254; September, 178; October, 75; November, 116; December, 967 (Theobald Smith).

The Thames, in the autumn of 1885, in the vicinity of London Bridge two hours after high water, contained 45,000 germs per cubic centimetre; the water of the Lea at Lea Bridge, 4,200,000 (Bisch-off).

The Neva inside the city of St. Petersburg, in September, 1883, contained 1,500 in one sample and 1,040 in another; in November (20th), 6,500 (Poehl).

The water of the Oder, collected within the limits of the city of Stettin, was found by Link to contain from 5,240 to 15,000 bacteria per cubic centimetre; that of the Limmat, at Zurich, 346 in one specimen and 508 in another (Cramer).

Lake water, as a rule, contains fewer bacteria than river water.

Wolffhügel, in researches extending from July, 1884, to July, 1885, obtained from the water of the Tegeler Lake an average of 396 bacteria per cubic centimetre. Cramer obtained an average of 168 per cubic centimetre during the months of October, December, and January, 1884, from the water of Lake Zurich; in June of the same year the average of 42 examinations gave 71 per cubic centimetre. In Lake Geneva, Fol and Dunant obtained from water collected some distance from the shore an average of 38 bacteria per cubic centimetre.

Ice which is usually collected from lakes and rivers contains a greater or less number of bacteria, according to the depth and purity of the water. The ice used in Berlin, collected from the surface of lakes and rivers in the vicinity of the city, contains from a few hundred to 25,000 bacteria to the cubic centimetre (Fränkel). In the experiments of Heyroth samples of ice from the same source gave less than 100 per cubic centimetre in three, from 100 to 500 in eight, from 500 to 1,000 in six, from 1,000 to 5,000 in seven, and 14,400 in one.

Prudden obtained from Hudson River ice, put up six miles below the city of Albany, an average of 398 bacteria per cubic centimetre from transparent ice, and in the superficial "snow ice" 9,187. Ice collected lower down the river contained an average of 189 in the transparent and 3,693 in the snow ice.

Ice from the Dora at Turin was found by Bordoni-Uffreduzzi to contain from 120 to 3,546 bacteria per cubic centimetre.

Hydrant water, as supplied to cities, has received the attention of numerous investigators. The water supply of Berlin was examined by Plagge and Proskauer at intervals of a week from June, 1885, to April, 1886. Their tabulated results show considerable variations. We give the figures for a single day, June 30th, 1885: Stralauer works, water of the Spree, unfiltered 4,400, filtered 53; Tegeler works, water of the lake, unfiltered 880, filtered 44; high reservoir at Charlottenberg, 71; 75 W. Wilhelmstrasse, 121; Friedrichstrasse, 41-42 S. W., 160; Schmidstrasse, 165 E., 51; Friedrichstrasse, 126 N., 151; Weinmeisterstrasse, 15 C., 63.

Wells which are supplied by water from deep strata contain few bacteria, unless contaminated by surface water in which they are usually very abundant. Roth examined the water of sixteen surface wells in Belgard, which has a very porous subsoil, and found from 4,500 to 5,000 bacteria in three, from 7,800 to 15,000 in six, from 18,000 to 35,000 in six, and 130,000 per cubic centimetre in one.

Forty-seven wells in Stettin, the water of which was examined by Link, gave the following results: Less than 100 in six, 100 to 500 in twenty-one, and in the remainder (sixteen) from 1,000 to 18,000.

Sixty-four wells in Mainz examined by Egger, and 53 in Gotha by Becker, gave more favorable results; the number of wells in the former city, in which less than 100 colonies developed from 1 cubic centimetre, was 34, and in the latter the same (34). Bolton examined the water of 13 wells in Göttingen, and found but 1 in which the number of colonies from 1 cubic centimetre was less than 100; in 12 the number varied from 180 to 4,940.

The water of *deep wells* and *springs* may be entirely free from bacteria, or nearly so. Egger found in the water of an artesian well at Mainz 4 bacteria per cubic centimetre, and the same number was found by Hueppe in the deep well at the Wiesbaden slaughter-house. The artesian well at the gasworks of Kiel was found by Brenning to contain from 6 to 30 bacteria per cubic centimetre. In a spring at Batiolettes, Fol and Dunant found 57 bacteria per cubic centimetre. Fürbringer obtained from springs at Jena 156 from one, 51 from another, 32 from another, and 109 from another. The water supplied to Danzig from the Prangenaaur Spring was found in several experiments to be free from bacteria (Freimuth).

In a summary of results obtained in various German cities Tie-mann and Gärtner find that sixty-nine per cent of the wells from which samples of water were examined contained less than 500 bacteria per cubic centimetre.

The *water of sewers* is naturally rich in bacteria. Miquel found that at Clichy the sewer water contained 6,000,000 bacteria per cubic centimetre. Bischoff found in water from London sewers 7,500,000,

and numerous observations show that the number of bacteria in river water is greatly increased in the vicinity of and below the mouths of city sewers.

We conclude from the experimental data recorded that water containing less than 100 bacteria to the cubic centimetre is presumably from a deep source and uncontaminated by surface drainage, and that it will usually be safe to recommend such water for drinking purposes, unless it contains injurious mineral substances. Water that contains more than 500 bacteria to the cubic centimetre, although it may in many cases be harmless, is to be looked upon with some suspicion, and water containing 1,000 or more bacteria is presumably contaminated by sewage or surface drainage and should be rejected or filtered before it is used for drinking purposes. But, as heretofore stated, the danger does not depend directly upon the number of bacteria present, but upon contamination with pathogenic species which are liable to be present in surface water and sewage. In swallowing a glassful of pure spring water a number of bacteria from the buccal cavity are washed away and carried into the stomach, which, if enumerated, would doubtless far exceed in numbers those found in the most impure river water.

The number of bacteria does not depend alone upon the amount of organic pabulum contained in a water, and cannot be depended upon in forming an estimate of this; for, as has been shown by Bolton, certain water bacteria multiply abundantly in water containing comparatively little organic matter, while other species fail to grow unless the quantity is greater. In a water containing considerable nutrient material the water bacteria may be restrained in their development by other species present until the amount of pabulum is reduced so that these no longer thrive, when the common water bacteria will take the precedence, and an enumeration may show a greater number of colonies than at first. But, in general, water rich in organic material contains a greater number of bacteria and a greater variety of species than that which is comparatively pure.

That certain bacteria may multiply in water which has been carefully sterilized has been shown by Bolton and others. Two common water bacteria—*Micrococcus aquatilis* and *Bacillus erythrosporus*—multiplied abundantly in doubly distilled water, and when this water was again sterilized and re-inoculated with one of these species the same abundant increase occurred. This was repeated six times with the same result (Bolton). Computing the number of these water bacteria in ten cubic centimetres of distilled water at twenty millions, and estimating their specific gravity at one, and the diameter of the individual cells at one μ , the total weight of the entire

number, according to Bolton, would be less than one-hundredth of a milligramme, and at least three-fourths of this must consist of water. The organic material represented by this number of bacteria would therefore be so minute that it might be supplied by dust particles accidentally falling into the distilled water.

Rosenberg has shown that while many of the species which he obtained in pure cultures from the water of the river Main multiplied in sterilized distilled water, other species quickly died out in such water. The growth of certain bacteria depends not only upon the quantity of nutritive material present, but upon its quality, the conditions in this regard being widely different for different species.

In view of the facts heretofore stated bacteriologists are now giving more attention to a careful study of the *kinds of bacteria present* in their examinations of water. Rosenberg, in his examinations of the water of the Main in the vicinity of Würzburg (1886), found that before the river reached the city the water contained more micrococci than bacilli, but that after receiving the sewage of the city the number of bacilli was greatly in excess.

Adametz (1888) has described eighty-seven species obtained by him from water in the vicinity of Vienna; Maschek found fifty-five different species in the drinking water used at Leitmeritz; and Tils (1890) has described fifty-nine species obtained by him from the city water supply at Freiburg.

Among the *pathogenic bacteria* which are liable to find their way into water used for drinking purposes, the most important, from a sanitary point of view, are the bacillus of typhoid fever and the spirillum of Asiatic cholera. Both of these microorganisms are present in great numbers in the excreta of persons suffering from the specific forms of disease to which they give rise, and are consequently liable to contaminate wells and streams which receive surface water, when such excreta are thrown upon the surface or into sewers, etc. Epidemics of these diseases have frequently been traced to the use of such contaminated water, and in a few instances the presence of these specific disease germs in water has been demonstrated by bacteriological methods. Laboratory experiments indicate, however, that an increase of these pathogenic bacteria in drinking water is not likely to occur, except under special conditions, and that they die out after a time, being at a disadvantage in the struggle for existence constantly going on among the numerous species which have their normal habitat in water.

Bolton, Frankland, and others have shown that the anthrax bacillus, not containing spores, dies out in hydrant water within five or six days. In the experiments of Kraus the anthrax bacillus added to well water, not sterilized, at a temperature of 10.5° C., was still

present in a living condition on the second day, but no colonies developed after the third day; the typhoid bacillus died out between the fifth and seventh days; the cholera spirillum was no longer found on the second day. In the meantime the common water bacteria had increased in numbers enormously. Similar results have been reported by Hochstetter and others. Hueppe, in ten experiments in which the typhoid bacillus was added to well water of a bad quality, found that in two no development of this bacillus occurred after the fifth day, while a few colonies developed in the other experiments as late as the tenth day. In these experiments the temperature was comparatively low (10.5° C.). At a higher temperature the experiments of Wolffhügel and Riedel show that an increase may take place. At the room temperature (about 20° C.) the typhoid bacillus added to distilled water, to well water, and to Berlin hydrant water was still present, in some instances, at the end of thirty-two days. And it was found that in some cases a decrease in the number occurred, then a notable increase, and finally a second diminution.

Koch found the cholera spirillum in a water tank at Calcutta during a period of fourteen days, and in his experiments showed that it preserved its vitality in well water for thirty days, in Berlin sewer water for six to seven days, and in the same mixed with faeces for twenty-seven hours only. In the experiments of Nicati and Rietsch the cholera spirillum preserved its vitality in distilled water for twenty days, in sewer water (of Marseilles) thirty-eight days, in water of the harbor for eighty-one days. The numerous experiments recorded by the observers named, and by Bolton, Hueppe, Hochstetter, Maschek, Kraus, and others, show that while the cholera spirillum may sometimes quickly die out in distilled water, in other experiments it preserves its vitality for several weeks (Maschek), and that it lives still longer in water of bad quality, such as is found in sewers, harbors, etc. Bolton found that for its multiplication a water should contain at least 40 parts in 100,000 of organic material, while the typhoid bacillus grew when the proportion was considerably less than this—6.7 parts in 100,000.

Russell has recently (1891) studied the bacterial flora of the Gulf of Naples, and of the mud at the bottom of this gulf, collected at various depths up to eleven hundred metres. His investigations show that sea water does not contain as many bacteria as an equal volume of fresh water; that bacteria are found in about equal numbers in water from the surface and in that from various depths; that the mud at the bottom constantly contains large numbers of bacteria; that some of the species isolated grow best in a culture medium containing sea water.

At a depth of 50 metres the water contained 121 bacteria per cubic

centimetre, and the mud from the bottom 245,000 ; at 100 metres the water contained 10 and the mud 200,000 per cubic centimetre ; at 500 metres the water contained 22 and the mud 12,500 per cubic centimetre ; at 1,100 metres the mud contained 24,000.

The following new species were obtained by Russell from the source mentioned : *Bacillus thalassophilus*, *Cladothrix intricata*, *Bacillus granulosus*, *Bacillus limosus*, *Spirillum marinum*, *Bacillus litoralis*, *Bacillus halophilus*.

The bacterial flora of fresh and sea water is very extensive, as will be seen by the following list of species which have been described by various bacteriologists who have given their attention to its study :

NON-PATHOGENIC MICROCOCCI.

Micrococcus aurantiacus (Cohn), *Micrococcus luteus* (Cohn), *Micrococcus violaceus* (Cohn), *Micrococcus flavus liquefaciens* (Flügge), *Micrococcus flavus desidens* (Flügge), *Micrococcus radiatus* (Flügge), *Micrococcus cinnabareus* (Flügge), *Micrococcus flavus tardigradus* (Flügge), *Micrococcus versicolor* (Flügge), *Micrococcus agilis* (Ali-Cohen), *Micrococcus fuscus* (Maschek), *Diplococcus luteus* (Adametz), *Pediococcus albus* (Lindner), *Micrococcus cerasinus siccus* (List), *Micrococcus citreus* (List), *Micrococcus aquatilis* (Bolton), *Micrococcus fervidus* (Adametz), *Micrococcus plumosus* (Bräutigam), *Micrococcus viticulosus* (Katz), *Micrococcus cremoides* (Zimmermann), *Micrococcus carneus* (Zimmermann), *Micrococcus concentricus* (Zimmermann), *Micrococcus rosetaceus* (Zimmermann), *Micrococcus ureæ* (Pasteur), *Weisser Streptococcus* (Maschek), *Wurmformiger Streptococcus* (Maschek), *Micrococcus aërogenes* (Miller), *Sarcina alba*, *Sarcina candida* (Reinke), *Sarcina lutea*.

PATHOGENIC MICROCOCCI.

Staphylococcus pyogenes aureus (Rosenbach), *Micrococcus* of Heydenreich—" *Micrococcus Biskra*."

NON-PATHOGENIC BACILLI.

Bacillus arborescens (Frankland), *Bacillus viscosus* (Frankland), *Bacillus aquatilis* (Frankland), *Bacillus liquidus* (Frankland), *Bacillus nubilus* (Frankland), *Bacillus vermicularis* (Frankland), *Bacillus aurantiacus* (Frankland), *Bacillus coerules* (Smith), *Bacillus glaucus* (Maschek), *Bacillus albus putidus* (Maschek), *Bacillus fluorescens liquefaciens*, *Bacillus fluorescens nivalis* (Schmolck), *Bacillus lividus* (Plagge and Proskauer), *Bacillus rubidus* (Eisenberg), *Bacillus sulfureum* (Holschewnikoff), *Bacillus violaceus*, *Bacillus gasoformans* (Eisenberg), *Bacillus liquefaciens* (Eisenberg), *Bacillus phosphorescens indicus* (Fischer), *Bacillus phosphorescens indigenus* (Fischer), *Bacillus phosphorescens gelidus* (Katz), *Bacillus smaragdino-phosphorescens* (Katz), *Bacillus argenteo-phosphorescens* Nos. I., II., and III. (Katz), *Bacillus cyaneo-phosphorescens* (Katz), *Bacillus argenteo-phosphorescens liquefaciens* (Katz), *Bacillus ramosus*, *Bacillus subtilis* (Ehrenberg), *Proteus sulfureus* (Lindenborn), *Bacillus aureus* (Adametz), *Bacillus brunneus* (Adametz), *Bacillus flavocoriaceus* (Adametz), *Bacillus fluorescens non-liquefaciens*, *Bacillus latericeus* (Adametz), *Bacillus stolonatus* (Adametz), *Bacillus berolinensis indicus* (Cläßen), *Bacillus erythrosporus* (Eidam), *Bacillus luteus* (List), *Bacillus aquatilis sulcatus* Nos. 1, 2, 3, 4, and 5 (Weichselbaum), *Bacillus albus* (Eisenberg), *Bacillus multipediculosus* (Flügge), *Bacillus Ziirnianum* (List), *Bacillus fulvus* (Zimmermann), *Bacillus helvolus* (Zimmermann), *Bacillus ochraceus* (Zimmer-

mann), *Bacillus plicatus*, *Bacillus devorans* (Zimmermann), *Bacillus gracilis* (Zimmermann), *Bacillus guttatus* (Zimmermann), *Bacillus implexus* (Zimmermann), *Bacillus punctatus* (Zimmermann), *Bacillus radiatus aquatilis* (Zimmermann), *Bacillus vermiculosus* (Zimmermann), *Bacillus constrictus* (Zimmermann), *Bacillus fluorescens aureus* (Zimmermann), *Bacillus fluorescens longus* (Zimmermann), *Bacillus fluorescens tenuis* (Zimmermann), *Bacillus fuscus* (Zimmermann), *Bacillus rubefaciens* (Zimmermann), *Bacillus subflavus* (Zimmermann), *Bacillus janthinus* (Zopf), *Bacillus mycoides* (Flügge), *Bacillus tremelloides* (Tils), *Bacillus cuticularis* (Tils), *Bacillus filiformis* (Tils), *Bacillus ubiquitus* (Jordan), *Bacillus circulans* (Jordan), *Bacillus superficialis* (Jordan), *Bacillus reticularis* (Jordan), *Bacillus rubescens* (Jordan), *Bacillus hyalinus* (Jordan), *Bacillus cloacæ* (Jordan), *Bacillus delicatulus* (Jordan), *Bacillus violaceus laurentius* (Jordan).

PATHOGENIC BACILLI.

Bacillus typhi abdominalis (Eberth, Gaffky), *Bacillus erysipelatos suis* (" *Bacillus murisepticus*," Koch), *Bacillus septicæmiæ hæmorrhagicæ* (" *Bacillus cuniculicida*," Koch), *Proteus vulgaris* (Hauser), *Proteus mirabilis* (Hauser), *Bacillus canalis capsulatus* (Mori), *Bacillus canalis parvus* (Mori), *Spirillum cholerae Asiaticæ* (" *Comma bacillus*," Koch), *Bacillus coli communis* (Escherich), *Bacillus hydrophilus fuscus* (Sanarelli), *Bacillus venenosus* (Vaughan), *Bacillus venenosus brevis* (Vaughan), *Bacillus venenosus invisibilis* (Vaughan), *Bacillus venenosus liquefaciens* (Vaughan).

III.

BACTERIA IN THE SOIL.

SURFACE soil, and especially that which is rich in organic matter, contains very numerous bacteria of many different species. Some of these are of special interest on account of their pathogenic power. Thus the bacillus of malignant cedema and the bacillus of tetanus have been shown to be widely distributed species, which have been obtained by investigators in various parts of the world by inoculating susceptible animals—guinea-pigs or mice—with a little rich surface soil. Other species are interesting because of their action in nitrification and in the destructive decomposition of organic material by which it is fitted for assimilation by the higher plants. Many of the bacteria present in the soil are strictly anaërobic, and in attempts to estimate the number and kind of microorganisms present in a given sample this fact must be kept in view.

The simplest *method of studying* the bacteria in the soil consists in introducing a small quantity into liquefied gelatin in test tubes, and, after carefully crushing it with a sterilized glass rod and thoroughly mixing it with the gelatin, making roll tubes in the usual way. Some of these should be put up for anaërobic cultures—*i.e.*, the tube should be filled with an atmosphere of hydrogen according to Fränkel's method. If the object in view is to estimate the number of bacteria in a given sample of soil the difficulty is encountered that, however finely crushed, the little masses of earth are likely to contain numerous bacteria, and we cannot safely assume that each colony originates from a single germ. Thoroughly washing a small quantity of soil, by agitation, in a considerable quantity of distilled water, and then adding a definite quantity of the water to nutrient gelatin and making roll tubes or plates, as in water analysis, suggests itself as a simple method ; but Fränkel has shown that it is far from being reliable when the object is to estimate the number of bacteria. He obtained more uniform and accurate results by introducing the earth at once into liquefied gelatin and crushing it as thoroughly as possible with a strong platinum wire, after which as thorough a mixture as possible was effected by tilting the tube up

and down. But for the purpose of obtaining pure cultures from single colonies of the various species present, we should prefer to wash the earth in distilled water and to allow the sediment to settle before taking a portion of the water to add to the nutrient medium.

In some experiments made in 1881 Koch ascertained that in soil which had not been disturbed but few bacteria were to be found at the depth of a metre; and this fact has since been established by the extended researches of Fränkel, who devised a special boring instrument for obtaining samples of earth from different depths. Miquel, in 1879, estimated the number of bacteria in one gramme of earth collected in the park of Montsouris, Paris, at a depth of twenty centimetres, at 700,000; and in a cultivated field which had been treated with manure, at 900,000. The following results were obtained by Adametz: One gramme of earth from a sandy soil contained at the surface 380,000, at a depth of twenty to twenty-five centimetres 400,000; the same quantity of clayey soil contained at the surface 500,000, at a depth of twenty to twenty-five centimetres 460,000.

In experiments made by Beumer (1886) and by Maggiora (1887) considerably greater numbers were found, but the last-named observer, in some instances at least, kept the earth for some time after collecting it, which may have materially influenced the result. Beumer obtained from a specimen of sandy humus taken from a depth of three metres 45,000,000 to the gramme; at four metres, 10,000,000; at five metres, 8,000,000; at six metres, 5,000,000. These specimens were obtained from the vicinity of hospitals at Greifswald. In a churchyard, at a depth of four metres, the number in one experiment was 1,152,000, and in another 1,278,000.

Fränkel has given special attention to the examination of undisturbed soil not in the immediate vicinity of dwellings. In samples from a fruit orchard near Potsdam he found that the superficial layers contained from 50,000 to 350,000 germs per cubic centimetre. The greatest number was not immediately upon the surface, but at from one-quarter to one-half metre below the surface. The number was found to be greater in summer than in winter, the maximum being in July and August. At a depth of three-quarters of a metre to a metre and a half there was a very great and abrupt diminution in the number of germs. From 200,000 at one-half metre the number fell to 2,000 at a depth of a metre, from 250,000 at three-quarters of a metre to 200 at one metre, etc., and at a depth of one and one-half metres, in some instances, no more living germs were obtained. In other experiments a few colonies developed from earth obtained at a depth of three or four metres, but these were slow in making their appearance, and often several days, or even weeks, elapsed before they became visible in Esmarch roll tubes. In experiments with sur-

face soil, on the contrary, a multitude of colonies developed within twenty-four to forty-eight hours, and, as many liquefying bacteria were present, it was necessary to make the enumeration on the first or second day, at which time, no doubt, many of the bacteria present had not yet formed visible colonies. The results obtained have, therefore, only a relative value.

The most important fact developed by Fränkel's researches is that in virgin soil there is a dividing line at a depth of from three-quarters to one and one-half metres, below which very few bacteria are found, and that, consequently, the "ground-water region" is free from micro-organisms, or nearly so, notwithstanding the immense numbers present in the superficial layers.

The extended researches of Maggiora, made in the vicinity of Turin, led him to the following conclusions :

1. The number of germs in desert and forest soils is much smaller, other conditions being equal, than in cultivated lands, and in these it is less than in inhabited localities.

2. In desert soils the number of germs bears a relation (a) to the geological epoch to which the lands belong, and, within certain limits, to the height above the level of the sea—the older the soil and the greater the altitude, other things being equal, the fewer the germs ; (b) to the compactness and aëration of the soil—the more compact and impermeable to air the smaller the number of germs capable of developing in gelatin ; (c) to the nature of the soil—sandy soils contain fewer germs than soils rich in clay and in humus.

3. In cultivated lands the number of germs augments with the activity of cultivation and the strength of the fertilizers used.

4. In inhabited localities the number of germs in the superficial layers is very great. In the deep layers it usually diminishes rapidly, as is the case in all other soils.

As to the kinds of bacteria present, and their biological characters and functions in preparing organic material for assimilation by the plants whose roots penetrate the soil, we have yet much to learn. Fränkel remarks that the species most frequently encountered in the deeper strata of the soil were three bacilli which also abound in the superficial layers—viz., the "hay bacillus," the "wurzel bacillus," and the "hirnbacillus." In all eleven bacilli were isolated and cultivated. Micrococci were only found four times, and spirilla not at all. Mould fungi were more abundant, and especially one previously obtained from the air by Hesse and called by him "brauner Schimmelpilz." Anaërobic bacilli, contrary to expectation, were not obtained in Fränkel's researches, and no pathogenic species were found in the deeper layers of the soil. We have already referred to the fact that the bacillus of malignant œdema and the bacillus of tetanus, two pathogenic, anaërobic species, are common in rich surface soil in various parts of the world.

The results obtained in the researches referred to, in which nutrient gelatin was used as a culture medium, are no doubt very incomplete, not only on account of the liquefaction of the gelatin by common liquefying bacilli before other species present have formed visible colonies, but also because this is not a favorable culture medium for some of the species present in the soil. Thus Frankland has succeeded in isolating a nitrifying ferment which he calls "*Bacillo-coccus*," which grows abundantly in bouillon, but fails to grow in nutrient gelatin. Winogradski has also obtained in pure cultures a nitrifying ferment from the soil in the vicinity of Zurich, which he has called "*Nitromonas*."

Comparatively few micrococci are found in the soil, while in the air they are usually found to be more abundant than bacilli. This is perhaps due to the fact that the bacilli are more promptly destroyed by desiccation and the action of sunlight.

Several bacteriologists have made investigations relating to the duration of vitality of pathogenic bacteria in the soil. Fränkel found that in Berlin the bacillus of anthrax, in Esmarch roll tubes, when buried in the soil at a depth of two metres, only occasionally gave evidence of growth, and at three metres no development occurred. The comparatively low temperature at this depth was no doubt an important factor in influencing the result. The cholera spirillum in the months of August, September, and October grew at a depth of three metres, but in the remaining months of the year failed to grow at two, while growth occurred at one and one-half metres. The bacillus of typhoid fever grew at three metres during the greater portion of the year.

Giaxa has made extended and interesting experiments with the cholera spirillum, cultures of which he added to different kinds of soil (garden earth, clay, sand) and placed at different depths below the surface—one-quarter, one-half, and one metre. Some of the earth was sterilized and some was not. In the unsterilized earth he found the cholera spirillum in considerable numbers at the end of twenty-four hours at the greatest depth tested (one metre), but at the end of forty-eight hours it had disappeared in five experiments out of seven—the lowest temperature at this depth was 20° C. In the sterilized soil the result was different; the cholera spirillum was present in enormous numbers at the end of four days at a depth of a metre, and was still found in smaller numbers at the end of twelve days, but had disappeared at the end of twenty-one days. These results indicate that the presence of common saprophytes in the soil is prejudicial to the development of the cholera spirillum, and that under ordinary circumstances it succumbs in the struggle for existence with these more hardy microorganisms.

The recently published researches of Proskauer (1891) confirm those of Fränkel and others as to the rapid diminution in the number of bacteria in the deeper layers of the soil. They also agree with those of Gärtner in showing that in the soil of churchyards the number of bacteria diminishes greatly in the soil beneath the layer containing coffins. In general the influence of dead bodies upon the bacteria in the soil in the vicinity of coffins was very slight; in the subsoil of the graveyard there were not many more bacteria than in similar soil outside of this. Reimers had previously shown that samples of earth from two graves, in one of which the body had been buried for thirty-five years and in the other for one and one-half years, gave similar results when examined by bacteriological methods.

Manfredi has recently (1892) published the results of his extended investigations relating to the dust in the streets of Naples. The number of bacteria varied greatly in different parts of the city. In streets where the traffic was least and hygienic conditions the best the average number was 10,000,000 per gramme. In dirty and busy thoroughfares the average was 1,000,000,000, and in certain localities the number was even five times as great as this. Injections into guinea-pigs gave a positive result in seventy-three per cent of the animals experimented upon. Among the known pathogenic bacteria obtained in this way were the pus cocci (in eight), the *Bacillus tuberculosis* (in three), the bacillus of malignant oedema, and the tetanus bacillus.

In the recently published memoir of Fülles (1891) the following species are described as having been found by him in the soil at Freiburg, Germany:

MICROCOCCLI.

(a) *Non-liquefying*.—*Micrococcus aurantiacus* (Cohn), *Micrococcus candidus* (Cohn), *Micrococcus luteus* (Cohn), *Micrococcus candicans* (Flügge), *Micrococcus versicolor* (Flügge), *Micrococcus cinnabareus* (Flügge), *Micrococcus cereus albus* (Passet), *Micrococcus fervitosus* (Adametz), *Rother coccus* (Maschek).

(b) *Liquefying*.—*Micrococcus flavus liquefaciens* (Flügge), *Micrococcus flavus desidens* (Flügge), *Diplococcus luteus* (Adametz), *Sarcina lutea*.

NON-PATHOGENIC BACILLI.

(a) *Non-liquefying*.—*Bacillus fluorescens putidus* (Flügge), *Bacillus muscoides* (Liborius), *Bacillus scissus* (Frankland), *Bacillus candicans*, *Bacillus diffusus* (Frankland), *Bacillus filiformis* (Tils), *Bacillus luteus* (Flügge), *Fluorescent water bacillus* (Eisenberg), *Bacillus viridis pallescens* (Frick), *Bluish-green fluorescent bacillus* (Adametz), *Bacillus stolonatus* (Adametz), *Bacillus Zürnianum* (List), *Bacillus aërogenes* (Miller), *Bacillus* No. 1 and *Bacillus* No. 2 (Fülles).

(b) *Liquefying*.—*Bacillus ramosus liquefaciens* (Flügge), *Bacillus liquidus* (Frankland), *Bacillus ramosus*—"wurzel bacillus," *Bacillus subtilis*

(Ehrenberg), *Bacillus mesentericus fuscus* (Flügge), *Bacillus mesentericus vulgatus* (Flügge), *Bacillus fluorescens liquefaciens* (Flügge), Lemon-yellow bacillus (Maschek), Green-yellow bacillus (Eisenberg), Gas-forming bacillus (Eisenberg), Gray bacillus (Maschek), *Bacillus prodigiosus* (Ehrenberg), *Proteus mirabilis* (Hauser), *Proteus vulgaris* (Hauser), *Bacillus mesentericus vulgatus*, *Bacillus cuticularis* (Tils), "Weisser bacillus" (Eisenberg).

(c) *Pathogenic*.—*Bacillus oedematis maligni* (Koch).

In addition to the above the following species have been described by other authors: *Bacillus liquefaciens magnus* (Lüderitz), *Bacillus radiatus* (Lüderitz), *Bacillus solidus* (Lüderitz), *Bacillus mycoides roseus* (Scholl), *Bacillus viscosus* (Frankland), *Bacillus candicans* (Frankland), *Bacillus poliformis* (Liborius), *Clostridium foetidum* (Liborius).

Pathogenic species.—*Staphylococcus pyogenes aureus* (Rosenbach), *Bacillus tetani* (Nicolai), *Streptococcus septicus* (Nicolai), *Pseudo-oedema bacillus* (Liborius), *Bacillus septicus agrigenus* (Nicolai), *Bacillus* of Utpatel.

IV.

BACTERIA OF THE SURFACE OF THE BODY AND OF EXPOSED MUCOUS MEMBRANES.

GREAT numbers of bacteria of various species multiply upon the *surface of the human body*, where they find the necessary pabulum in the excretions from the skin and the exfoliated epithelium. Evidently the number will be largely influenced by the clothing worn, the atmospheric conditions as to heat and moisture, personal habits, etc. The writer has frequently inoculated culture media with a drop of sterilized fluid which had been placed upon the surface of the body of patients in hospitals and of healthy persons. By friction with a platinum needle at the point where the drop of fluid is applied the surface is washed and a little epithelium detached. Cultures may always be obtained by inoculating nutrient media from a drop of fluid applied in this way. Micrococci of various species, including the pus cocci, are very commonly encountered; sarcinæ and various bacilli are also frequently met with. Even the hands, which by reason of their exposure and frequent ablutions are freer from exfoliated epithelium than portions of the body covered with clothing, have constantly attached to their surface a considerable number of bacteria. This is shown by the experiments of Kümmel and Forster, of Fürbringer and others, with reference to the disinfection of the hands. Forster found that after the most careful cleaning of the hands with soap, water, and a brush, contact of the fingers with nutrient gelatin always resulted in the development of a greater or less number of colonies.

Bordoni-Uffreduzzi, in his researches relating to the bacteria of the skin, obtained in pure cultures five different species of micrococci and two bacilli. Pure cultures of his *Bacterium graveolens*, which was usually found between the toes, gave off a disagreeable odor like that observed from this locality in certain individuals. In his researches made in Havana the writer frequently encountered in cultures from the surface, associated with various micrococci, his *Micrococcus tetragenus versatilis*.

Fürbringer found quite frequently in the spaces beneath the fin-

ger nails *Staphylococcus pyogenes aureus* associated with various other microorganisms. A similar result had previously been reported by Bockhart.

In his examinations of water from various sources Miquel found that "wash-water" from the floating laundries on the Seine contained more bacteria than water from any other source, even than the water of the Paris sewers. His enumeration gave twenty-six million germs per cubic centimetre.

Hohein has enumerated the colonies developing from underclothing worn for various lengths of time and made of different kinds of material. A piece of the goods to be tested was sewed fast to the underclothing, so as to come in immediate contact with the body; at the end of a given time a fragment one-quarter of a centimetre square was cut up as fine as possible and distributed in nutrient gelatin. Plates were made and the colonies counted at the end of five or six days.

In an experiment in which sterilized woven goods were worn next to the skin of the upper arm the following results were obtained: Linen goods, at the end of one day 28, two days 4,180 colonies; cotton goods, end of one day 105, end of two days 1,870; woollen goods, end of one day 606, end of two days 6,799. When the material had been in contact with the skin for four days the colonies which developed were so numerous that they could not be counted.

Maggiore isolated twenty-two species of bacteria from his cultures inoculated with epidermis from the foot. None of these proved to be pathogenic for mice, rabbits, or guinea-pigs. Several gave off a strong odor of trimethylamin, similar to that of sweating feet.

The following species have been found upon the surface of the body:

Non-pathogenic.—*Diplococcus albicans tardus* (Unna and Tommasoli), *Diplococcus citreus liquefaciens* (Unna and Tommasoli), *Diplococcus flavus liquefaciens tardus* (Unna and Tommasoli), *Staphylococcus viridis flavescens* (Guttmann), *Bacillus graveolens* (Bordoni-Uffreduzzi), *Bacillus epidermidis* (Bordoni), *Ascobacillus citreus* (Unna and Tommasoli), *Bacillus fluorescens liquefaciens minutissimus* (Unna and Tommasoli), *Bacillus aureus* (Unna and Tommasoli), *Bacillus ovatus minutissimus* (Unna and Tommasoli), *Bacillus albicans pateriformis* (Unna and Tommasoli), *Bacillus spiniferus* (Unna and Tommasoli), *Bacillus* of Scheurlen, *Micrococcus tetragenus versatilis* (Sternberg), *Bacillus Havaniensis liquefaciens* (Sternberg).

Pathogenic.—*Staphylococcus pyogenes albus*, *Staphylococcus pyogenes aureus*, *Streptococcus pyogenes*, *Diplococcus* of Demme, *Bacillus* of Demme, *Bacillus* of Schimmelbusch, *Bacillus* of Tommasoli, *Bacillus saprogenes* II. (Rosenbach), *Bacillus parvus ovatus* (Löffler).

SURFACE OF MUCOUS MEMBRANES.

Cultures made from the *conjunctivæ* of healthy persons usually show the presence of various micrococci, and sometimes of bacilli.

In diseased conditions these are more numerous than in health, but the pus cocci are not infrequently found in healthy eyes.

As bacteria are constantly present in the air, they are necessarily deposited upon the moist mucous membrane of the *nose* during inspiration. Indeed, it would appear as if an important function of this extended mucous membrane is to purify the air from suspended particles, and it has been shown by experiment that expired air is practically free from bacteria. The greater number of those contained in inspired air are deposited upon the mucous membrane of the anterior and posterior nares. In culture experiments made by Von Besser, Wright, and others the nasal mucus was found to contain a great variety of bacteria; among others the pus cocci were frequently found by both of the observers mentioned. In eighty-one cases Von Besser found the "*diplococcus pneumoniae*" fourteen times, *Staphylococcus pyogenes aureus* fourteen times, *Streptococcus pyogenes* seven times, and Friedländer's bacillus twice. Twenty-eight of the cases examined were convalescents in hospital; among these the pathogenic species mentioned were found less frequently than in other individuals. The following non-pathogenic species were isolated: *Micrococcus liquefaciens albus* in twenty-two cases, *Micrococcus albus* in nine cases, *Micrococcus cumulatus tenuis* in fourteen cases, *Micrococcus flavus liquefaciens* in three cases, *Bacillus striatus albus* in ten cases, etc.

Paulsen (1890) made thirty-one cultures in nutrient gelatin from sixteen persons and thirty-three in nutrient agar from twenty-two persons, with the following result: Eleven remained sterile, nineteen showed not more than ten colonies, sixteen less than one hundred, twelve more than one hundred, and in six the number was so great that they could not be counted. Micrococci were more numerous than bacilli; of these a "sulphur-yellow coccus" in tetrads was found in eight individuals. Various species of liquefying cocci, resembling the pus cocci, were isolated, but the conclusion was reached that none of these were identical with the staphylococci of pus, which Von Besser and Wright both found in a considerable proportion of the culture experiments made by them.

Very extended researches have been made with reference to the bacteria present in the human *mouth*, which show that numerous species are constantly present in the buccal secretions and upon the surface of the moist mucous membrane. Some of these are occasional and accidental, while others appear to have their normal habitat in the mouth, where the conditions as to temperature, moisture, and presence of organic pabulum are extremely favorable for their development. A minute drop of saliva spread upon a glass slide, dried, and stained with one of the aniline colors, will always be

found to contain an immense number of bacteria of various forms. Some of these are attached to epithelial cells and some scattered about singly or in groups. Among those seen in a single specimen we will usually find cocci in tetrads, in chains, and in irregular groups, bacilli of various dimensions, and occasionally spirilla. According to Prof. Miller, of Berlin, the following species "almost invariably occur in every mouth: *Leptothrix innominata*, *Bacillus buccalis maximus*, *Leptothrix buccalis maxima*, *Iodococcus vaginatus*, *Spirillum sputigenum*, *Spirochaete dentium*. All of these fail to grow in ordinary culture media. Miller has made extended attempts to obtain cultures by varying the medium used and attempting to imitate as nearly as possible the natural medium in which they are found; but his attempts have been unsuccessful, or nearly so—"only line cultures afforded a limited growth, but the colonies never developed more than fifteen to twenty cells, and a transference to a second plate proved futile, no further growth taking place."

Up to the year 1885 Miller had isolated twenty-two different species of bacteria from the human mouth. Ten of these were cocci, five short bacilli, six long bacilli, and one a spirillum. Later the same author cultivated eight additional species. Vignal has isolated and described seventeen species obtained by him in pure cultures from the healthy human mouth; most of these are bacilli, and Miller, who found micrococci to be more numerous, supposes the difference in results to be due to the fact that many of the cocci do not grow in nutrient gelatin, which was the medium employed by Vignal. In the researches of the last-named author the following species were obtained most frequently, in the order given: 1. *Bacterium termo*. 2. *Bacillus e* (*Bacillus ulna*?). 3. Potato bacillus. 4. *Coccus a*. 5. *Bacillus b*. 6. *Bacillus d*. 7. *Bacillus c* (*Bacillus alvei*?). 8. *Bacillus subtilis*. 9. *Staphylococcus pyogenes albus*. 10. *Staphylococcus pyogenes aureus*.

Among the species above enumerated we find two of the most common pus cocci, *Staphylococcus albus* and *aureus*, but no mention is made of another important pathogenic micrococcus which is frequently found in the healthy human mouth, viz., the micrococcus of sputum septicæmia, first named by the writer *Micrococcus Pasteuri*. This does not grow at ordinary temperatures, and consequently would not be obtained in gelatin plate cultures. Very different results have been reported by different observers as to the frequency with which these pathogenic cocci are found in the buccal cavity. Black found in the saliva of ten healthy individuals the *Staphylococcus pyogenes aureus* seven times, *Staphylococcus pyogenes albus* four times, and *Streptococcus pyogenes* three times. On the other hand, Netter found *Staphylococcus aureus* only seven times in

one hundred and twenty-seven individuals examined. Miller also has rarely found the pus cocci in the mouths of healthy persons. *Streptococcus pyogenes* was not found by Vignal in his extended researches. The experiments of the writer, of Vulpian, Fränkel, Netter, Claxton, and others show that the micrococcus which in 1885 I named *Micrococcus Pasteuri*, and which is identical with the "diplococcus pneumoniae" of German authors, is frequently present in the healthy human mouth—now called *Micrococcus pneumoniae crouposa*. Netter examined the saliva of one hundred and sixty-five healthy individuals and obtained it in fifteen per cent of the number examined.

Another pathogenic micrococcus which is frequently present in the mouths of healthy persons is the *Micrococcus tetragenus* of Koch. The following pathogenic bacteria have also been isolated and described: *Bacillus crassus sputigenus* (Kreibohm), *Bacillus salivarius septicus* (Biondi). The *Streptococcus septo-pyæmicus* of Biondi is described as having characters identical with those of the *Streptococcus pyogenes* of Rosenbach. Two other pathogenic species described by Biondi were each found in a single case only. Miller has described the following pathogenic species isolated and studied by him: *Micrococcus gingivæ pyogenes*, *Bacterium gingivæ pyogenes*, *Bacillus dentalis viridans*, *Bacillus pulpæ pyogenes*.

Vignal has tested a considerable number of microorganisms, obtained by him in his cultures from the healthy human mouth, with reference to their peptonizing action upon various kinds of food, with the idea that some of them may have an important physiological function of this kind. Out of nineteen species he found ten which, after a longer or shorter time, dissolved fibrin, nine which dissolved gluten, ten which dissolved casein, and five which dissolved albumin; nine changed lactose into lactic acid, seven inverted cane sugar, seven caused the fermentation of glucose, and seven coagulated milk.

Recently (1891) Sanarelli has shown that normal saliva has the power of destroying the vitality of a limited number of certain pathogenic bacteria, including the following species: *Staphylococcus pyogenes aureus*, *Streptococcus pyogenes*, *Micrococcus tetragenus*, *Bacillus typhi abdominalis*, *Spirillum cholerae Asiaticæ*. When to ten cubic centimetres of saliva, sterilized by filtration through porcelain, the above-mentioned pathogenic bacteria were added in small numbers by means of a platinum needle carried over from a pure culture, no development occurred, and at the end of twenty-four hours the bacteria introduced were incapable of growth in a suitable medium. But when this amount of filtered saliva was inoculated with a large platinum loop—an öse—a certain number of the bacteria

survived, and at the end of three or four days an abundant development occurred. At first, however, the number of living cells was considerably diminished. In saliva to which one öse of a culture of *Staphylococcus aureus* was added thirteen thousand eight hundred and forty colonies developed in a plate made immediately after inoculation, while a plate made at the end of twenty-four hours contained but one hundred and thirty-two colonies, and one at the end of forty-eight hours had but eight colonies. Subsequently multiplication occurred, and a plate made on the ninth day after inoculation contained so many colonies that they could not be counted.

The diphtheria bacillus was not destroyed in filtered saliva, but did not multiply in it. On the other hand, it proved to be a very favorable medium for the development of *Micrococcus pneumoniae crouposæ*.

Mucus from the surface of the *meatus urinarius* of man and woman, or from the *vagina*, will always be found to contain various bacteria; but the bladder, the uterus, and Fallopian tubes in healthy individuals are free from microorganisms. Winter has isolated twenty-seven different species from vaginal and cervical mucus, and reports that he found *Staphylococcus pyogenes albus* in one-half of the cases examined. A streptococcus was also encountered which resembled *Streptococcus pyogenes*, although not positively identified with it. Samschin, on the other hand, failed to obtain the pus cocci in vaginal mucus from healthy women.

Dönderlein, Von Ott, and others have carefully examined the *lochial discharge* with reference to the presence of bacteria. The first-named author found that in healthy women the lochial discharge obtained from the uterus was free from germs, but when collected from the vagina various microorganisms were obtained. In one case in which some fever existed *Staphylococcus pyogenes aureus* was found in the vagina, while the discharge from the uterus was free from germs. In five cases of puerperal fever *Streptococcus pyogenes* was obtained in the lochial discharge from the uterus. The results of Von Ott correspond with those of Dönderlein. Czerniewski, in the lochia of fifty-seven healthy women, found the *Streptococcus pyogenes* but once, while in the lochial discharge of fatal cases of puerperal fever it was always present.

Steffeck (1892) has examined the vaginal secretion of twenty-nine pregnant females who had not been subjected to digital examination, and found *Staphylococcus pyogenes albus* in nine, *Staphylococcus pyogenes aureus* in three, and *Streptococcus pyogenes* in one. These results indicate that puerperal septicæmia from self-infection may occur in exceptional cases. In seventeen of the twenty-nine cases examined none of these pyogenic micrococci were found.

The following species have been obtained from the nasal and buccal secretions :

FROM THE NOSE.

Non-pathogenic.—*Micrococcus nasalis* (Hajek), *Diplococcus coryzæ* (Hajek), *Micrococcus albus liquefaciens* (Von Besser), *Micrococcus cumulatus tenuis* (Von Besser), *Micrococcus tetragenus subflavus* (Von Besser), *Diplococcus fluorescens foetidus* (Klamann), *Micrococcus foetidus* (Klamann), *Vibrio nasalis* (Weibel), *Bacillus striatus flavus* (Von Besser), *Bacillus striatus albus* (Von Besser).

Pathogenic.—*Staphylococcus pyogenes aureus*, *Staphylococcus pyogenes albus*, *Streptococcus pyogenes*, *Bacillus* of Friedländer, *Bacillus* of rhinoscleroma (?), *Bacillus foetidus ozænæ* (Hajek), *Bacillus mallei* (Löffler), *Bacillus smaragdinus foetidus* (Reimann).

FROM THE MOUTH.

Non-pathogenic.—*Micrococcus roseus* (Eisenberg), *Micrococcus* A, B, C, D, E of Podbielskij, *Sarcina pulmonum* (Hauser), *Sarcina lutea*, *Micrococcus candicans* (Flügge), *Bacillus* of Miller, *Bacillus virescens* (Frick), *Vibrio rugula*, *Vibrio lingualis* (Weibel), *Pseudo-diphtheria bacillus* (Von Hoffmann), *Bacillus mesentericus vulgatus*, *Bacillus subtilis*, *Bacillus a, b, c, d, e, f, g, h, i, and j* of Vignal, *Bacillus subtilis similis*, *Bacillus radiceformis* (Eisenberg), *Bacillus luteus*, *Bacillus fluorescens non-liquefaciens*, *Bacillus ruber*, *Bacillus viridiflavus*, *Proteus Zenkeri*, *Bacillus G, H, I, J, K, L, M, N, and Vibrio O and P* of Podbielskij, *Vibrio viridans* (Miller), *Micrococcus nexifer* (Miller), *Iodococcus magnus* (Miller), *Ascococcus buccalis* (Miller), *Bacillus fuscans* (Miller).

Pathogenic.—*Staphylococcus pyogenes albus*, *Staphylococcus pyogenes aureus*, *Staphylococcus salivarius septicus* (Biondi), *Streptococcus pyogenes*, *Micrococcus salivarius septicus* (Biondi), *Micrococcus tetragenus* (Gaffky), *Micrococcus gingivæ pyogenes* (Miller), *Streptococcus septo-pyæmicus* (Biondi), *Streptococcus articulorum* (Löffler), *Micrococcus* of Manfredi, *Micrococcus pneumoniae crouposæ*—" *Micrococcus Pasteuri*" (Sternberg); *Bacillus diphtheriæ* (Löffler), *Bacillus tuberculosis* (Koch), *Bacillus* of Friedländer, *Bacillus bronchitidis putridæ* (Lumnitzer), *Bacillus septicæmiæ hæmorrhagicæ*, *Bacillus gingivæ pyogenes* (Miller), *Bacillus pulpæ pyogenes* (Miller), *Bacillus dentalis viridans* (Miller), *Bacillus crassus sputigenus* (Kreibohm), *Bacillus saprogenes* No. 1 (Rosenbach), *Bacillus pneumoniae agilis** (Schou), *Bacillus pneumoniae* of Klein, *Bacillus pneumosepticus* (Babes).

V.

BACTERIA OF THE STOMACH AND INTESTINE.

As the secretions of the mouth contain numerous bacteria, these must constantly find their way to the stomach, but conditions are not favorable for their development when the stomach is in a healthy state and its secretions normal. Under certain circumstances, however, there may be an abundant development in the stomach of species which give rise to various fermentations, and no doubt dyspeptic symptoms are frequently due to this cause. In the present section we are, however, only concerned with the bacteria of the healthy stomach. Most of these, we think, are to be considered as only temporarily and accidentally present in this viscus as the result of the swallowing of the buccal secretions and of food and drink containing them.

The experiments of Straus and Würtz and of others show that normal gastric juice possesses decided germicidal power, which is due to the free hydrochloric acid contained in it. Hamburger (1890) found that gastric juice containing free acid is almost always free from living microorganisms, and that it quickly kills the cholera spirillum and the typhoid bacillus, but has no effect upon anthrax spores. Straus and Würtz found that the cholera spirillum is killed by two hours' exposure in gastric juice obtained from dogs, the typhoid bacillus in two to three hours, anthrax bacilli in fifteen to twenty minutes, and the tubercle bacillus in from eighteen to thirty-six hours. The experiments of Kurlow and Wagner, made with gastric juice obtained from the stomach of healthy men by means of a stomach sound, gave the following results: Anthrax bacilli without spores failed to grow after exposure to the action of human gastric juice for half an hour, but spores were not destroyed in twenty-four hours; the typhoid bacillus was killed in one hour; the cholera spirillum, the bacillus of glanders, and *Bacillus pyocyaneus* were all destroyed at the end of half an hour; the pus cocci showed greater resisting power. Certain bacteria have a greater resisting power for acids than any of those above mentioned, and some of them may consequently pass through the healthy stomach to the intestine

in a living condition, but there is good reason to believe that the spirillum of cholera or the bacillus of anthrax would not. On the other hand, the tubercle bacillus and the spores of other bacilli can, no doubt, pass through the stomach to the intestine without losing their vitality.

Of nineteen species isolated by Vignal in his cultures from the healthy human mouth, the greater number resisted the action of the gastric juice for more than an hour, and six species which did not form spores were found to retain their vitality in gastric juice for more than twenty-four hours.

In making a bacteriological analysis of the contents of the healthy stomach the more resistant microorganisms and those which form spores will naturally be found in greater or less numbers, inasmuch as some of them are likely to be present in food and water ingested.

Van Puteren (1888) obtained a variety of microorganisms in very considerable numbers from the stomachs of infants fed upon unsterilized cow's milk, but in healthy nursing infants the number was much smaller, especially when the mouth was washed out with distilled water immediately before and after nursing. In 18 per cent of the cases no microorganisms were found under these circumstances, and in 41 per cent the number fell below one thousand per cubic centimetre. Among the nursing infants examined (eighty-five) the following species were most numerous: *Monilia candida*, *Bacillus lactis aërogenes*, a non-liquefying coccus, *Staphylococcus pyogenes aureus*, *Bacillus subtilis*. In infants fed upon cow's milk (eleven) *Bacillus lactis aërogenes* was present in 45.4 per cent of the cases, and *Staphylococcus pyogenes aureus* in 27.2 per cent, non-liquefying cocci in 54.4 per cent, liquefying cocci in 72.7 per cent, *Bacillus subtilis* in 36.3 per cent, and *Bacillus butyricus* (Hueppe) in all of the cases; next to these *Bacillus flavescens liquefaciens* was the most abundant. The author named reaches the conclusion that no species is constant and that the presence of those found depends upon accidental circumstances.

Abelous (1889) found in his own stomach, washed out while fasting, a considerable number of species of bacteria, viz.: *Sarcina ventriculi*, *Bacillus pyocyaneus*, *Bacillus lactis aërogenes*, *Bacillus subtilis*, *Bacillus mycoides*, *Bacillus amylobacter*, *Vibrio rugula*, and eight other undescribed bacilli and one coccus. All of these microorganisms were able to resist the action of hydrochloric acid in the proportion of 1.7 grammes in 1,000 grammes of water. Several were found to be facultative anaerobics.

The action of the bacteria isolated by him was tested by Abelous upon various alimentary substances. The time required to effect changes, such as the digestion of fibrin, the changing of starch

into glucose, etc., was found to be so long that there was no reason to suppose that any one of the microorganisms tested was concerned in ordinary stomach digestion.

In the *intestine* conditions are favorable for the development of many species of saprophytic bacteria, and the smallest quantity of excrementitious material from the bowels, spread upon a glass slide and stained with one of the aniline colors, will be found to contain a multitude of microorganisms of this class, of various forms. Among these are certain species which have their normal habitat in the intestine, and which may always be obtained in cultures from this source, while others, having been present in food or water ingested, and having escaped destruction in the acid juices of the stomach, are accidentally and temporarily present. These latter may or may not increase in the organic pabulum which abounds in the intestine, according as the conditions are favorable or otherwise. The strictly aerobic bacteria could not multiply because of the absence of oxygen, and the species encountered are for the most part anaerobics or facultative anaerobics. The *Bacillus coli communis* of Escherich, which is the most constant and abundant species found in the intestine of man and of certain of the lower animals, is a facultative anaerobic, which grows readily in the ordinary culture media, either in the presence of oxygen or in an atmosphere of hydrogen. But certain other bacteria of the intestine are strictly anaerobic and do not grow readily in the media commonly employed by bacteriologists.

Escherich has shown that in new-born infants the meconium is free from bacteria. At the end of twelve to eighteen hours after birth bacteria appear in the alvine discharges, and the number is already considerable at the expiration of the first twenty-four hours of independent existence. The species first found are cocci and yeast cells which no doubt come from the atmosphere, having been deposited upon the moist mucous membrane of the mouth and swallowed with the buccal secretions. When the meconium is replaced by "milk faeces" these contain in large numbers the *Bacillus coli communis*, heretofore spoken of as the most common species found in the intestine of adults. Another species associated with this, but not so abundant, is the *Bacillus lactis aerogenes* of Escherich. Other bacilli and cocci are found occasionally in smaller numbers. These bacilli do not liquefy gelatin, and, as a rule, the microorganisms found in the alvine discharges of healthy persons are non-liquefying bacteria. Escherich's researches led him to the conclusion that the *Bacillus lactis aerogenes* is constantly present in the small intestine of milk-fed children as the most prominent species, and that its multiplication there is favored by the presence of milk

sugar, and that *Bacillus coli communis* finds the most favorable conditions for its growth in the large intestine.

Brieger, in 1884, isolated from fæces and carefully studied two bacilli, one of which has since been called by his name. This is a non-liquefying bacillus which is very pathogenic for guinea-pigs, and which in its morphology and characters of growth closely resembles the *Bacillus coli communis* of Escherich. Indeed, a number of non-liquefying bacilli, differing but slightly in their morphological and biological characters, have been obtained by various investigators from the alimentary canal of man and the lower animals, and it is still a question whether they are to be regarded as distinct species or as varieties of the "colon bacillus" of Escherich. The bacillus obtained by Emmerich from cholera cadavers in Naples belongs to this group, and, if not identical with the colon bacillus, resembles it so closely that its differentiation is extremely difficult. Brieger's bacillus forms propionic acid in solutions containing grape sugar. A second bacillus obtained by him from the same source resembles the "pneumococcus" of Friedländer; this causes the fermentation of saccharine solutions, with production of ethyl alcohol.

Bienstock (1883) isolated four species of bacilli from normal fæces, two of which are comparatively large and resemble *Bacillus subtilis* in their morphology and in the formation of spores. A third species is described as an extremely slender pathogenic bacillus, resembling the bacillus of mouse septicæmia. The fourth species is an actively motile bacillus which forms end spores, causing the rods to have the form of a drumstick. This is said to cause the decomposition of albumin, with production of ammonia and carbon dioxide. Later researches do not sustain Bienstock's conclusion that the bacilli described by him are the principal forms found in normal fæces.

Among the species encountered by Escherich, in addition to those mentioned above (*Bacillus coli communis* and *Bacillus lactis aërogenes*), are the following: *Proteus vulgaris*, found three times in meconium, and constantly in the fæces of dogs fed upon flesh; *Streptococcus coli gracilis*, found in meconium, but not during the period of nursing, is constantly present in the intestine when a flesh diet is employed.

The intestine of carnivorous and omnivorous animals contains a greater number of bacteria than that of the herbivora, and in the large intestine they are far more numerous than in the small intestine (De Giæxa). Sucksdorf has enumerated the colonies developing from one milligramme of fæces from individuals on mixed diet. He obtained an average of 380,000 from a series of observations in which the maximum was 2,300,000 and the minimum 25,000.

The following species have been isolated from fæces and the contents of the intestine of cadavers :

Non-pathogenic.—*Streptococcus coli gracilis* (Escherich), *Micrococcus aërogenes* (Miller), *Micrococcus tetragenus versatilis* (Sternberg), *Micrococcus ovalis* (Escherich), "Yellow liquefying staphylococcus" (Escherich), "Porzellancoccus" (Escherich), *Bacillus subtilis*, *Bacillus aërogenes* (Miller), *Bacterium aërogenes* (Miller), *Bacillus lactis erythrogenes* (Hueppe), *Clostridium foetidum* (Liborius), *Bacillus muscoides* (Liborius), *Bacillus putrificus coli* (Bienstock), *Bacillus subtilis similis* I. and II. (Bienstock), *Bacillus Zopfii*, *Bacillus liquefaciens communis* (Sternberg), *Bacillus intestinalis liquefaciens* (Sternberg), *Bacillus intestinalis motilis* (Sternberg), *Bacillus fluorescens liquefaciens* (Flügge), "Colorless fluorescent liquefying bacillus" (Escherich), "Yellow liquefying bacillus" (Escherich), *Bacillus mesentericus vulgaris*, Bacilli of Booker, A to T, first series; α to s , second series; Bacilli of Jeffries A to Z, and α , β .

Pathogenic.—*Staphylococcus pyogenes aureus*, *Bacillus typhi abdominalis*, *Bacillus septicæmiæ hæmorrhagicæ*, *Bacillus* of Belfanti and Pascaroia, *Bacillus enteritidis* (Gärtner), *Bacillus* of Lesage, *Bacillus pseudomurisepticus* (Bienstock), *Bacillus coli communis* (Escherich), *Bacillus lactis aërogenes* (Escherich), *Bacillus cavidia* (Brieger), *Bacillus* of Emmerich, *Bacillus coprogenes foetidus* (Schottelius), *Bacillus* of Utpadel, *Bacillus leporis lethalis* (Sternberg), *Bacillus acidiformans* (Sternberg), *Bacillus cuniculicida Havaniensis* (Sternberg), *Bacillus cadaveris* (Sternberg), *Bacillus cavidia Havaniensis* (Sternberg), *Proteus vulgaris* (Hauser), *Bacillus tuberculosis*, *Spirillum cholerae Asiaticæ*, *Spirillum* of Finkler and Prior.

VI.

BACTERIA OF CADAVERS AND OF PUTREFYING MATERIAL FROM VARIOUS SOURCES.

THE putrefactive changes which occur so promptly in cadavers, when temperature conditions are favorable, result chiefly from post-mortem invasion of the tissues by bacteria contained in the alimentary canal. But it is probable that under certain circumstances microorganisms from the intestine may find their way into the circulation during the last hours of life, and that the very prompt putrefactive changes in certain infectious diseases in which the intestine is more or less involved are due to this fact. The writer has made numerous experiments in which a portion of liver or kidney removed from the cadaver at an autopsy made soon after death—one to six hours—has been enveloped in an antiseptic wrapping and kept for forty-eight hours at a temperature of 25° to 30° C. In every instance there has been an abundant development of bacteria, although as a rule none were obtained from the same material immediately after the removal of the organ from the body. This shows that a few scattered bacteria were present. The same result was obtained in cases of sudden death from accident, as from portions of liver or kidney removed from the bodies of persons dying of yellow fever, tuberculosis, and other diseases.

Numerous researches show that the blood of healthy men and animals is free from bacteria, and that saprophytic bacteria injected into a vein soon disappear from the circulation; and recent experiments show that blood serum has decided germicidal power. But in spite of this fact the experiments of Wyssokowitsch show that certain bacteria injected into the circulation may be deposited in the liver, the spleen, and the marrow of the bones, and there retain their vitality for a considerable time. The spores of *Bacillus subtilis* were found by the observer named to preserve their vitality in the liver or spleen of animals into which they had been injected, for a period of two or three months. In the writer's experiments the microorganisms which first developed in fragments of liver preserved in an antiseptic wrapping were certain large anaërobic bacilli, and especially

my *Bacillus cadaveris*, together with the *Bacillus coli communis* of Escherich, my *Bacillus hepaticus fortuitus*, and other non-liquefying bacilli of the "colon group."

These bacteria did not give rise to a putrefactive odor, and the fragment of liver when cut into had a fresh appearance and a very acid reaction. Later, putrefactive changes occurred and *Proteus*

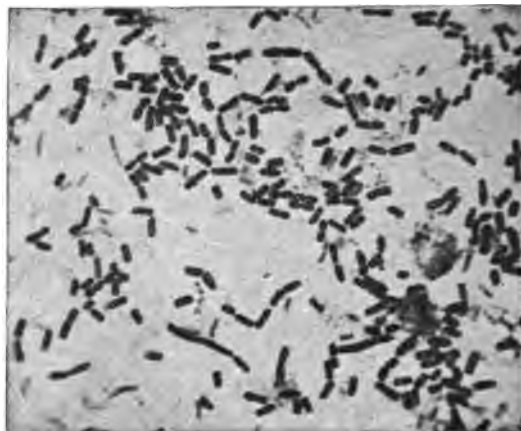


FIG. 197.—Smear preparation from liver of yellow-fever cadaver, kept forty-eight hours in an antiseptic wrapping. $\times 1,000$. From a photomicrograph. (Sternberg.)

vulgaris and other putrefactive bacteria obtained the precedence. Evidently all of these species must have been present in the liver at the time it was removed from the cadaver, although in such small numbers that they were rarely seen in smear preparations or obtained in cultures from the fresh liver tissue. The appearance of a smear preparation from the interior of a fragment preserved for forty-eight hours in an antiseptic wrapping is shown in Fig. 197.

The horribly offensive gases which are given off from dead animals in a state of putrefaction appear to be due to certain large anaërobic bacilli which are found in such material, and which have not yet been thoroughly studied owing to the difficulty of cultivating them in artificial media; among them is a large bacillus with round ends which forms an oval spore at one extremity of the rather long rod. This the writer has described under the name of *Bacillus cadaveris grandis*, Fig. 198.

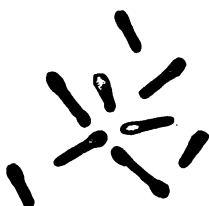


FIG. 198.

In the interior of a putrefying mass of this kind only those bacteria are found which are able to grow in the absence of oxygen, but aerobic saprophytes may multiply upon the surface of

such a mass, or in organic liquids to which the air has free access. Among the most common putrefactive bacteria are the *Proteus vulgaris*, *Proteus mirabilis*, and *Proteus Zenkeri* of Hauser. Formerly the minute motile bacteria found in putrefying animal infusions, etc., were commonly spoken of as belonging to the species "*Bacterium termo*," but recent researches show that several different species were included under this name by those whose researches were made before the introduction of Koch's method for isolating and differentiating microorganisms of this class by the use of solid culture media. The different species of *Proteus* are all facultative anaerobics. They are more or less pathogenic, and according to Hauser produce a chemical poison which, when injected into small animals, causes death with all of the symptoms of putrid intoxication. The bacillus of mouse septicæmia, which was first obtained by Koch from a putrefying meat infusion, is also pathogenic, as are the writer's *Bacillus cadaveris* and various other anaerobic bacteria found in putrefying material.

Some account of the various products of putrefaction and the microorganisms concerned in their production will be found in Section IV., Part Second, of the present volume.

VII.

BACTERIA IN ARTICLES OF FOOD.

Milk always contains bacteria, unless drawn with special precautions into a sterilized flask. In the healthy udder of the cow it is sterile, but in tuberculous cows, when the milk glands are involved, tubercle bacilli may find their way into the milk in considerable numbers. As ordinarily obtained and preserved, milk is greatly exposed to bacterial contamination from various sources; desquamated cuticle from the external surface of the udder and from the hands of the milker, and floating particles from the air of the stable, fall into it at the very moment it is drawn, and it is subsequently contaminated by bacteria from the air, and from water used in washing the receptacles in which it is placed or added to it by the thrifty milkman. As it furnishes an excellent nutrient medium for many of the bacteria which are thus introduced into it, under favorable conditions of temperature it quickly undergoes changes due to the multiplication in it of one or more of these microorganisms. The acid fermentation and coagulation of the casein which so constantly occurs is completely prevented by sterilizing fresh milk in flasks provided with a close-fitting cork or cotton air filter. Numerous researches have been made with reference to the microorganisms found in milk and the various fermentations to which they give rise. Naturally a great variety of species will be found in an extended research, but all are accidentally present, and only those demand special attention which produce the various fermentations of this fluid commonly encountered, or which have special pathogenic properties.

Several different bacteria produce an acid fermentation and consequent coagulation of milk, but the usual agent in producing this fermentation is the *Bacillus acidi lactici*, which is identical with the "ferment lactique" of Pasteur. When a pure culture of this bacillus is introduced into sterilized milk kept at a temperature of 25° to 30° C., coagulation occurs in from fifteen to twenty-four hours. A uniform, gelatinous mass is produced which does not subsequently become dissolved (Adametz). Various other bacteria produce a similar change, including a number of common water bacteria, several spe-

cies of *sarcina*, *Staphylococcus pyogenes aureus*, and other pus cocci. Usually coagulation is due to the combined action of several bacteria, among which *Bacillus acidi lactici* is apt to be the most prominent.

Other bacteria produce coagulation without the lactic acid fermentation. This appears to be due to the formation of a soluble ferment which acts like rennet, causing the coagulation of milk which has a neutral or slightly alkaline reaction. The coagulated casein in this case is subsequently redissolved. The bacteria which produce this change for the most part form spores, while the lactic acid ferments do not. If, therefore, milk is heated nearly to the boiling point the acid-forming bacteria will be destroyed and the spores of the other species surviving will give rise to coagulation without the production of lactic acid. Among the more common microorganisms of this group are the *Bacillus butyricus* (Hueppe), *Bacillus mesentericus vulgatus*, Löffler's "white milk-bacillus," and the bacilli described by Duclaux under the generic name of *Tyrothrix*.

Other fermentations are produced by certain chromogenic bacteria, and these, as a rule, are not as harmless from a sanitary point of view as those above referred to. Blue milk is produced by the presence of *Bacillus cyanogenus*, yellow milk by *Bacillus synxanthus* (Schröter) and by a species obtained by List from the fæces of a sheep and another found by Adametz in cheese. The well-known *Bacillus prodigiosus* produces its characteristic red pigment when present in milk, and a bluish-red color is caused by *Bacterium lactis erythrogenes* (Hueppe).

Viscous fermentation in milk is produced by several different bacteria, among others by a micrococcus studied by Schmidt-Mühlheim, and a short bacillus isolated by Adametz—*Bacillus lactis viscosus*. Milk which has undergone this change is unwholesome as food; it is recognized by the long filaments which are produced when it is touched with any object and this is slowly withdrawn.

The Caucasian milk ferment, *Bacillus Caucasicus*, produces a special fermentation, which has been referred to in Section IV., Part Second (page 132).

Various pathogenic bacteria have occasionally been found in milk in addition to the tubercle bacillus already referred to. Thus Adametz found *Staphylococcus pyogenes aureus* in two samples which had been submitted to him for examination, one of which had given rise to vomiting and diarrhoea. Wyssokowitsch cultivated from milk which had been standing some time a pathogenic bacillus, named by him *Bacillus oxytocus pernicius*.

The special microorganism which produces the poisonous ptomaine called by Vaughan tyrotoxin has not yet been isolated; nor do we know the exact cause of scarlet fever, although there is evi-

dence that this disease has been spread by the use of contaminated milk, as have also diphtheria and typhoid fever, which diseases are due to bacilli now well known. As the cholera spirillum grows readily in milk, this disease could no doubt also be transmitted in the same way.

Recently (1892) Sedgwick and Batchelder have examined a large number of specimens of milk obtained in Boston and vicinity, for the purpose of determining the number of bacteria present. They found, as an average of several trials, that milk obtained in a clean stable, from a well-kept cow, by milking in the usual way into a sterilized bottle, contained 530 bacteria per cubic centimetre. "When, however, the milkman used the ordinary milk pail of flaring form, seated himself with more or less disturbance of the bedding, and vigorously shook the udder over the pail during the usual process of milking," the numbers were very much higher—on an average 30,500 per cubic centimetre immediately after milking. The average of fifteen samples taken from the tables of persons living in the suburbs of Boston was 69,143 per cubic centimetre. The average of fifty-seven samples of Boston milk, obtained directly from the milk wagons and plated at once, was 2,355,500 per cubic centimetre. The average of sixteen samples from groceries in the city of Boston was 4,577,000 per cubic centimetre.

Prof. Renk found in the milk supply of Halle from 6,000,000 to 30,000,000 bacteria per cubic centimetre—a number considerably exceeding that usually found in the sewage of American cities (Sedgwick).

In fresh *butter* of good quality but few microorganisms are found, but in "cheesy butter" having a disagreeable odor Kreuger has found a great number of bacteria. Among these the most numerous were an oval coccus, *Micrococcus acidi lactici* (Kreuger), a slender bacillus closely resembling, and possibly identical with, the *Bacillus fluorescens*, and the *Bacillus acidi lactici* of Hueppe.

Duclaux (1887) has isolated from different kinds of *cheese* no less than eleven different species of bacteria, which he believes are concerned in the "ripening process." Seven of these are aerobic and four anaerobic species. Adametz (1889) has also isolated and studied a number of species to which he attributes the ripening of cheese.

Meats, even when salted and smoked, may contain living pathogenic bacteria which were present prior to the death of the animal, and, when not properly preserved, are of course liable to be invaded by putrefactive bacteria.

The researches of Foster (1889) show that the typhoid bacillus, the pus cocci, the tubercle bacillus, and the bacillus of swine plague resist the action of a saturated solution of salt for weeks and even for

months; and the same observer found that the ordinary processes of salting and smoking did not destroy the tubercle bacillus in the flesh of a cow which had succumbed to tuberculosis. Beu has made cultures from a large number of specimens of fresh, salted, and smoked meats and fish, with the general result that the fresh and salted meats were found to contain a limited number of bacteria of various species, and that smoking for several days did not insure the destruction of these microorganisms. In specimens of sausage six days' smoking did not destroy a liquefying bacillus which was present, but at the end of six weeks' exposure to smoke this bacillus no longer grew, while a non-liquefying bacillus present in the same specimen had not been destroyed. Fourteen days' smoking sufficed to destroy all the microorganisms in a specimen of bacon, but this was not sufficient for the interior portions of a ham. Among the bacteria obtained by Beu from smoked meats he mentions the following: *Staphylococcus cereus albus*, *Proteus vulgaris*, *Staphylococcus pyogenes aureus*, *Bacillus liquefaciens viridis*, etc. The number of colonies which developed from a fragment, the size of a mustard seed to that of a flaxseed, taken from the interior of the meats examined, was usually small; and the presence of a few scattered bacteria of these common species has no significance from a sanitary point of view, except as showing that pathogenic bacteria may survive in infected meats after they have been exposed to the usual processes of salting and smoking.

Petri, in experiments upon the bacillus of swine plague (*Schweine-rotlauf*), arrived at the following results:

The flesh of swine which died of this disease preserved its infectious properties after having been preserved in brine for several months, and the same flesh salted or pickled for a month and then smoked for fourteen days contained the *rotlauf* bacillus in a living and unattenuated condition. At the end of three months virulent *rotlauf* bacilli were still obtained from a smoked ham, but they were no longer found at the end of six months.

Schrank (1888) has made cultures from both the albumin and the yolk of *fresh eggs*, and finds that they are free from bacteria. He thinks that, as a rule, putrefactive bacteria obtain access to the interior through injured places in the shell, although exceptionally the egg may be infected with them in the oviduct of the fowl. The usual bacteria concerned in the putrefactive changes in eggs are, according to the author mentioned, a variety of *Proteus vulgaris* and *Bacillus fluorescens putidus*.

Peters (1889) has studied the flora of the "*sauerteig*" used in Germany as yeast for leavening bread. In addition to the numerous cells of three species of *Saccharomyces*, he finds that bacilli are present in great numbers, as shown by direct microscopical examination and

culture experiments. He describes five species, designated *Bacillus* A, B, C, D, and E, which are commonly present, and to which the acid fermentation of the dough is ascribed.

In Graham bread which had undergone changes making it unfit to eat, Kratschmer and Niemilowicz have found the *Bacillus mesentericus vulgatus*, which appears to have been the cause of the fermentation, which was produced in bread having a slightly alkaline reaction by inoculating it with a pure culture of this bacillus. The infected bread has a brownish color, a peculiar odor, and becomes sticky and viscid.

Uffelmann (1890) has also studied the bacteria in spoiled rye bread, and obtained, in addition to common mould fungi, *Bacillus mesentericus vulgaris* and *Bacillus liodermus*.

Bernheim (1888) has examined various grains used as food, with reference to the presence of bacteria, and claims to have demonstrated their presence by staining thin sections, and also by cultures, in corn, wheat, rye, barley, and peas. He supposes that they find their way from the earth through the roots and stems of plants. This appears to be very doubtful, in view of the researches of other observers, and further researches are necessary before we can accept the fact as demonstrated that they are usually present in healthy kernels of the grains mentioned.

VIII.

NON-PATHOGENIC MICROCOCCI.

MANY of the saprophytic micrococci and bacilli have already been described in the sections devoted to pathogenic bacteria (Part Third). We propose at present to give an account of the morphological and biological characters which distinguish those microorganisms which have not been shown to possess pathogenic power. But it must be remembered that in many instances the bacteria described in this and the following sections have not been tested at all, or only very imperfectly tested, with reference to this point; and no doubt some of them, if tested upon the various animals usually employed in experiments of this kind, would prove to be more or less pathogenic. On the other hand, many of the saprophytes heretofore described as pathogenic only produce marked morbid phenomena in susceptible animals when they are injected beneath the skin, into a serous cavity, or into the circulation in considerable quantities. The experiments of Buchner show that very many of the common saprophytes usually classed as non-pathogenic give rise to a local abscess when sterilized cultures are injected subcutaneously into rabbits or guinea-pigs. In short, there is no well-defined dividing line between the pathogenic and non-pathogenic bacteria, and some of those now described as non-pathogenic, as the result of more extended experiments, will no doubt eventually be transferred to the list of pathogenic bacteria.

159. MICROCOCCUS FLAVUS LIQUEFACIENS (Flügge).

Found in the air and in water.

Morphology.—Tolerably large micrococci, in pairs or in irregular groups.

Biological Characters.—An *aërobic, liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms small, yellow colonies, which under a low power are seen to be spherical or oval, with a finely granular surface and a yellowish-brown color; lines radiate from the centre through a zone of transparent liquefied gelatin to the sharply defined border, and later the colonies, which have a diameter of four to six millimetres, resemble a wagon wheel. In *gelatin stick cultures* smooth, spherical, yellow colonies form upon the surface; these become confluent and form a yellow layer, which by the slow liquefaction of the gelatin becomes depressed; at the end of five days the gelatin is liquefied to a depth of about two millimetres and a yellowish, flocculent deposit is seen at the bottom of the yellowish-white fluid. Upon *potato* a deep-yellow layer with irregular margins is quickly developed.

160. MICROCOCCUS FLAVUS DESIDENS (Flügge).

Found in air and water.

Morphology.—Small micrococci, usually in pairs, but sometimes seen in groups of three or in short chains.

Biological Characters.—An *aërobic, liquefying, chromogenic* micrococcus. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies appear as white or yellow points; under a low power they are seen as oval, yellowish-brown, finely granular discs. The superficial colonies are circular, with irregular margins, and are not elevated above the level of the gelatin; by the fourth day they may attain a diameter of five to ten millimetres; they have a brownish-yellow color; the gelatin is gradually liquefied, and the colony which sinks below the surface is surrounded by a ring of liquefied gelatin from one to four millimetres broad. In *gelatin stick cultures* a slimy, yellowish-brown layer of limited extent is formed upon the surface, and a confluent, porcelain-white mass along the line of puncture; at the end of eight days liquefaction has occurred under the superficial layer to a depth of three to four millimetres, forming a cylinder filled with a thick fluid, to the bottom of which the surface growth gradually sinks. Upon *potato* a slimy, yellowish-brown layer with irregular outlines is slowly developed.

161. MICROCOCCUS AGILIS (Ali-Cohen).

Found in water.

Morphology.—Micrococci, one μ in diameter, usually in pairs, occasionally in tetrads or in chains; have extremely slender flagella, which are four to five μ in length.

Biological Characters.—An *aërobic, liquefying* (very slowly), *motile, chromogenic* micrococcus. Grows at the room temperature in the usual culture media—not in the incubator at 37° C. This micrococcus is distinguished by its *active movements* and by the presence of a long flagellum, which may be demonstrated by Löffler's method of staining. In *gelatin stick cultures* growth occurs along the line of inoculation, and liquefaction commences at the end of three to four weeks; sometimes only a dry, funnel-shaped cavity is formed. Upon *agar* and upon *potato* a pink layer is slowly developed.

162. MICROCOCCUS FUSCUS (Maschek).

Found in water.

Morphology.—Micrococci, which are often elliptical, or even in the form of short rods (bacilli?).

Biological Characters.—An *aërobic, liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms spherical colonies which under a low power present the appearance of being finely cleft and vary in color from pale-brown to black; liquefaction quickly occurs. In *gelatin stick cultures* but scanty growth occurs along the line of puncture; upon the surface a sepia-brown layer is formed and the gelatin is quickly liquefied. Gelatin cultures have a strong putrefactive odor. Upon *potato* a slimy, brown layer is formed, which becomes almost black.

163. DIPLOCOCCUS CITREUS CONGLOMERATUS (Bumm).

Obtained from gonorrhœal pus and from the air—in dust.

Morphology.—Diplococci, consisting of two hemispherical elements separated by a narrow cleft, and closely resembling the *Micrococcus gonorrhœæ*; about 1.5 μ in diameter; frequently in tetrads, usually united in conglomerate masses.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefy-*

ing, chromogenic micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms lemon-yellow colonies, which throw out tongue-like projections upon the surface of the gelatin and have wave-like margins; the surface is at first moist and shining, later cleft and scaly. In *gelatin stick cultures* development occurs along the line of puncture and on the surface; liquefaction, beginning near the surface, progresses slowly; a yellowish layer floats upon the surface, and later settles to the bottom of the tube.

164. DIPLOCOCCUS CITREUS LIQUEFACIENS (Unna).

Found on the skin of persons suffering from eczema seborrhoeicum.

Morphology.—Small, oval cocci, in pairs or in tetrads, often in irregular groups or in short chains; the diameter of a single element in a pair is from 0.4 to 0.1 μ .

Biological Characters.—An *aërobic, liquefying, chromogenic micrococcus*. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of four days, the superficial colonies are grayish-white, flat, circular discs the size of a mustard seed; at the end of eight days they are grayish-yellow, opaque, and about one to two millimetres in diameter; at the end of two weeks they are lemon-yellow, concave, and begin to sink into a shallow funnel of liquefied gelatin; under a low power they are seen to be finely granular. The deep colonies appear at first as white points; under the microscope they are seen to be spherical or oval, brownish-yellow, and have sharply defined outlines. In *gelatin stick cultures*, at the end of six days, a thin, shining, yellowish layer has formed on the surface; at the end of two weeks the gelatin is softened and a thick, yellowish-white, flocculent deposit is seen, while upon the surface of the liquefied medium is an irregular, plate-shaped, deep lemon-yellow layer; at the end of three weeks the liquefaction has extended to a depth of about six millimetres; the liquefied gelatin is opaque and of a yellow color. Upon the surface of *agar* a yellowish-brown layer with irregular margins is quickly developed; upon *potato*, at the end of two weeks, a grayish-yellow layer.

165. DIPLOCOCCUS FLAVUS LIQUEFACIENS TARDUS (Unna).

Found upon the skin of individuals suffering from eczema seborrhoeicum.

Morphology.—Biscuit-formed diplococci, resembling the "gonococcus"; each element in a pair is from 0.5 to 0.8 μ in diameter.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, chromogenic micrococcus*. Grows in the usual culture media at the room temperature—very slowly upon gelatin and potato, more rapidly on agar. Upon *gelatin plates*, at the end of eight days, the superficial colonies are very small, circular, shining, pale grayish-yellow discs; at the end of three weeks they are as large as a hempseed and of a chrome-yellow color; later they become greenish-yellow and float in a circular zone of transparent, liquefied gelatin. The deep colonies are at first punctiform; later they are small, opaque spheres of an olive brownish-yellow color. In *gelatin stick cultures* a thin, yellowish-white, slimy layer is slowly developed upon the surface; at the end of three weeks this is from three to four millimetres in diameter and irregular in outline; as it becomes older the color is dark-yellow or greenish-yellow; a thin, yellowish growth develops along the line of puncture; at the end of four weeks the surface is depressed without being really liquefied; in eight weeks about half of the gelatin in the tube is liquefied and transparent. The surface growth first floats upon the liquefied medium; later it settles to the bottom as a thick, flocculent, yellow deposit, and the gelatin acquires a yellow color. Upon the surface of *agar* a thick, slimy, yellowish white layer with wavy margins is developed; later this has a greenish-yellow color. Upon *potato* a sulphur-yellow layer is formed.

166. DIPLOCOCCUS FLUORESCENS FÆTIDUS (Klamann).

Obtained from the posterior nares.

Morphology.—Diplococci, about $1.4\ \mu$ in diameter (the pair), often arranged in chains containing from six to ten elements.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *chromogenic* micrococcus. Grows in the usual culture media at the room temperature—better at 37°C . Upon *gelatin plates* the superficial colonies are at first gray or brownish circular masses, which soon sink below the surface of the liquefying gelatin and later form a crater-like depression, in the centre of which is seen a brownish-gray sediment, while the surrounding gelatin has a grass-green or violet color. In *gelatin stick cultures* a circular, shallow, pale-gray, saucer-shaped cavity forms at the surface, and a purse-like pouch along the line of puncture; a shining, iridescent film floats upon the surface of the liquefied gelatin, and a greenish-gray sediment accumulates at the bottom; finally the gelatin is completely liquefied and has a green color above, while a violet-colored film floats upon the surface. Upon *agar* a granular, brownish-gray layer is quickly developed. Upon *potato* a finely granular layer, which after a time acquires a dark, bluish-green color, while the potato around it is colored blue. The color is changed to red by acids.

167. DIPLOCOCCUS LUTEUS (Adametz).

Found in water.

Morphology.—Micrococci, usually in pairs, of 1.2 to $1.3\ \mu$ in diameter; sometimes observed in chains of eight to ten elements or in irregular groups.

Biological Characters.—An *aërobic*, *liquefying*, *motile*, *chromogenic* micrococcus. Grows in the usual culture media at the room temperature. This micrococcus is described by Adametz as *actively motile*. Upon *gelatin plates*, at the end of three days, circular, pale-yellow, viscous colonies are developed, which have a diameter of about one millimetre. Under a low power they are seen to be granular and brownish-yellow in the centre, while the margins are pale-yellow; at the end of six days the colonies are of an intense yellow color and about three millimetres in diameter. In *gelatin stick cultures* growth occurs rapidly upon the surface only, at first as a circular, lemon-yellow layer marked with concentric circles; at the end of about ten days the gelatin at the surface acquires an intense brownish-red color, which extends downward in a cloud-like manner, gradually diminishing in intensity; liquefaction commences at the end of several weeks. Upon the surface of *agar* a viscid yellow layer is formed along the impf-strich, and the medium acquires a brownish-red color. Upon *potato* a dirty-yellow layer, which subsequently has a brownish color, is developed; this gives off the characteristic odor of penicillum cultures. In *milk coagulation* of the casein is produced in about five days.

168. DIPLOCOCCUS ROSEUS (Bumm).

Found in the air.

Morphology.—Diplococci resembling the "gonococcus," the elements of a pair being hemispherical and separated by a tolerably broad cleft; the diameter, measured from pole to pole, is from 1 to $1.5\ \mu$.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *chromogenic* micrococcus. Grows in nutrient gelatin at the room temperature. Upon *gelatin plates* slightly elevated, pink colonies are developed, which under the microscope are seen to be finely granular and irregular in outline. In *gelatin stick cultures* an abundant development occurs upon the surface and along the line of puncture; this has a pink color; the gelatin is slowly liquefied after a considerable time.

169. MICROCOCCUS CREMOIDES (Zimmermann).

Found in water.

Morphology.—Micrococci, about $0.8\ \mu$ in diameter, arranged in grape-like masses.

Biological Characters.—An *aërobic, liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are small and yellowish-white in color; under a low power they are seen to be spherical, granular, and yellow or brownish-gray. Superficial colonies have an irregular, "gnawed" margin, and cause a saucer-like liquefaction of the gelatin, at the bottom of which a yellowish-white mass, arranged in concentric rings, may be seen; around the margin delicate outgrowths into the unliquefied gelatin may be seen. In *gelatin stick cultures* liquefaction occurs along the line of puncture in three or four days; an air bubble is usually seen near the surface, and below this an accumulation of a yellowish-white color; the liquefied gelatin below this is transparent for some distance, and the bottom of the narrow channel is again filled with a yellowish-white mass of micrococci; at the end of a week the liquefied channel measures about eleven millimetres at the surface and a yellowish-white film floats upon the liquefied gelatin. Upon the surface of *agar* a yellowish-white layer with irregular margins and a lustre like that of amber is developed. Upon *potato* a tolerably abundant, cream-colored layer extends over the surface.

170. MICROCOCCUS ROSEUS (Eisenberg).

Found in sputum of a patient with influenza.

Morphology.—Micrococci of 0.8 to $1\ \mu$ in diameter, solitary or in irregular groups.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature, and at 37°C . without production of color. Upon *gelatin plates*, at the end of three to four days, minute pink colonies are formed; later liquefaction commences about the colonies and progresses slowly. In *gelatin stick cultures* development occurs slowly both upon the surface and along the line of puncture; the growth is at first colorless; after three to four days a small, round, pink layer is formed, which is depressed in the centre; at the end of a week the color resembles that of a red azalea blossom, and liquefaction commences; at the end of three weeks the gelatin is about half-liquefied and a pink sediment is seen. Upon the *surface of agar*, at the room temperature, a soft, dark-pink layer is formed along the *impfstrich* in two days; at 37°C . a similar development occurs in twenty-four hours, but without color. Upon *potato*, at the end of three to four days, a cherry-red streak is seen along the *impfstrich*; this gradually becomes darker and covers the entire surface; the growth then resembles that of *Bacillus prodigiosus*.

171. MICROCOCCUS AURANTIACUS (Cohn).

Found in water.

Morphology.—Spherical or slightly oval cocci, 1.3 to $1.5\ \mu$ in diameter, solitary, in pairs, or in irregular groups.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* spherical or elliptical colonies of an orange-yellow color and smooth, shining surface. In *gelatin stick cultures* a small, button-like, yellow growth develops upon the surface, and after a considerable time minute yellow colonies are seen along the line of puncture. Upon *agar* an orange-yellow layer is formed, and upon *potato* a slimy, yellow growth.

172. MICROCOCCUS CERASINUS SICCUS (List).

Found in water.

Morphology.—Micrococci, from 0.25 to 0.32 μ in diameter, solitary or in pairs.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows best at 37° C. Does not grow well in nutrient gelatin. Upon the *surface of agar* a dry, cherry-red layer is quickly developed. Upon *potato* the surface is quickly covered with a cherry-red layer. The pigment is not soluble in water, alcohol, or ether, and is not changed by acids or alkalis.

173. MICROCOCCUS VERSICOLOR (Flügge).

Found in water.

Morphology.—Micrococci, in pairs or in irregular groups.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are at first white points; later yellow, opaque, finely granular spheres. The superficial colonies are irregular in outline, slimy, and have a pearly lustre; they may attain a diameter of two to ten millimetres. In *gelatin stick cultures* small, spherical, yellow colonies develop along the line of puncture, and a layer with irregular, "gnawed" margins and a pearly lustre upon the surface. Upon *agar* a slimy, opaque layer of a yellowish-brown color. Upon *potato* a slimy layer is quickly developed.

174. MICROCOCCUS OF DANTEC.

Obtained by Dantec (1891) from salted codfish which had undergone changes characterized by a red color and an offensive odor.

Morphology.—Micrococci, from three to five μ in diameter, often marked by a line of commencing binary division.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Forms a red pigment. In *gelatin plates* small, disc-shaped colonies of a red color are slowly developed; these rarely measure more than a millimetre in diameter. In *gelatin stick cultures* development is slow; along the line of puncture the growth has a yellowish color; on the surface it is of a pale-red, and later of deeper-red color. Upon *agar* the development is more rapid than upon gelatin. It grows upon dried codfish without production of pigment, except when it is associated with other microorganisms—especially a liquefying coccus which is often found with it.

Not pathogenic.

175. MICROCOCCUS CARNEUS (Zimmermann).

Found in water.

Morphology.—Micrococci, having a diameter of about 0.8 μ , united in irregular, grape-like masses.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows best at the room temperature; very scanty development at 30° to 33° C. Upon *gelatin plates* the deep colonies are small, spherical, and grayish-white in color. Superficial colonies are but slightly elevated, circular in outline, and of a grayish-red to pale-red color; under the microscope they are seen as circular discs with a more opaque, reddish-gray centre surrounded by a somewhat more transparent zone, and this by a second, still paler zone; in older cultures the distinct zones are no longer to be distinguished, but the reddish-brown color fades out from the centre towards the margin of the colonies. In *gelatin stick cultures*, at the end of five days, a thin, circular, pale-pink layer, with irregular outlines and about

3.5 millimetres in diameter, is developed upon the surface, and a finely granular, white growth along the line of puncture. Upon the *surface of gelatin* a flesh-red layer, which later acquires a violet hue, is formed along the *impfstrich*. Upon *agar* the growth is similar but more abundant, and the margins are coarsely toothed. Upon *potato* an abundant red layer is developed.

176. MICROCOCCUS CINNABAREUS (Flügge).

Found in air and in water.

Morphology.—Large, spherical cocci, frequently associated in pairs or in tetrads.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are first seen as minute points at the end of four days; under a low power they are seen to be oval or lenticular, with a well-defined contour and of a dark reddish-brown color. The superficial colonies, at the end of four days, are from 0.5 to 1 millimetre in diameter and brick-red; at the end of eight days they project from the gelatin in button-shape and are cinnabar-red. In *gelatin stick cultures* isolated white colonies are seen along the line of puncture at the end of four to five days, and upon the surface a button-like mass of moderate dimensions is developed, which is first pink and later cinnabar-red. Upon *potato* a cinnabar-red layer is slowly developed.

177. MICROCOCCUS CEREUS ALBUS (Passet).

Obtained by Passet (1885) in the pus of acute abscesses (two cases out of thirty-three examined), and by Tils (1890) from the Freiburg water supply.

Morphology.—Large cocci, 1.16μ in diameter, solitary or associated in irregular groups.

Biological Characters.—An *aërobic, non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms superficial colonies, which attain a diameter of one to two millimetres and resemble drops of stearin or white wax. In *gelatin stick cultures* grows upon the surface as a grayish-white layer with irregular, thickened margins, resembling a drop of stearin; scanty growth along the line of puncture. Upon *potato* a dirty-white layer of moderate thickness is developed.

178. MICROCOCCUS CEREUS FLAVUS (Passet).

Obtained by Passet (1885), in a single case out of thirty-three examined, from the pus of an acute abscess.

Morphology.—Micrococci of irregular dimensions, associated in irregular groups and occasionally in chains.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* lemon-yellow colonies are developed, which attain a diameter of one to two millimetres. In *gelatin stick cultures* the growth around the point of inoculation resembles a drop of stearin or wax with elevated margins and has a yellow color; a scanty yellow streak is developed along the line of puncture. Upon *potato* a citron-yellow layer is formed.

179. MICROCOCCUS CITREUS.

Synonym.—Crème-farbigiger micrococcus (List).

Found in water.

Morphology.—Large, spherical cocci, from 1.5 to 2.2μ in diameter, solitary, in pairs, or in chains of eight or more elements.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micro-

coccus. Grows in the usual culture media at the room temperature—better at 37° C. Upon *gelatin plates* forms upon the surface dirty pale-yellow or cream-colored colonies, which after several days have a diameter of 0.5 to 0.8 centimetre and are about 0.5 millimetre thick; these have usually irregular outlines and a moist, shining appearance. In *gelatin stick cultures* very scanty growth occurs along the line of puncture. Upon *agar* a pale-yellow layer is formed. Upon *potato*, at 37° C., an abundant growth occurs, forming a yellow layer.

180. MICROCOCCUS FERVIDOSUS (Adametz).

Found in water.

Morphology.—Small, round cocci, 0.6 μ in diameter, in pairs or in irregular groups.

Biological Characters.—An *aërobic, non-liquefying* micrococcus. Grows in the usual media at the room temperature. Upon *gelatin plates*, at the end of four to five days, the deep colonies appear as white points, which under a low power have a pale-yellow color and resemble dewdrops; upon the surface transparent, yellow colonies with irregular, jagged edges are developed; later these are granular in the centre and have a brownish color, while the marginal zone is yellowish and slightly wrinkled. In *gelatin stick cultures* a thin, circular layer with finely toothed margins forms upon the surface, and a granular growth along the line of puncture. In glycerin-gelatin numerous gas bubbles of various sizes are developed in the medium. Upon *agar* a circular, milk-white, slimy layer is formed, which later has a pearly lustre. Upon *potato* a dirty-white layer with irregular margins.

181. MICROCOCCUS FLAVUS TARDIGRADUS (Flügge).

Found in the air and in water.

Morphology.—Large, spherical cocci, usually associated in irregular groups; sometimes have peculiar dark poles.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are spherical or oval, dark chrome-yellow, and from 0.4 to 0.6 millimetre in diameter; under a low power they appear to have a dark olive-green color. The superficial colonies are from 0.5 to 1 millimetre in diameter, have a smooth, varnished-looking surface, and are slightly elevated above the surface of the gelatin at the middle; under a low power the centre is grayish-yellow and the margin paler. In *gelatin stick cultures*, at the end of eight days, a row of small, spherical, isolated yellow colonies is developed along the line of puncture.

182. MICROCOCCUS LUTEUS (Cohn).

Found in water.

Morphology.—Oval cocci, from 1 to 1.2 μ in diameter, associated in zoëglæa masses; the intercellular substance is easily soluble in water.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. The yellow pigment produced is not soluble in water, ether, or alcohol, and is not changed by acids or alkalies. Grows in the usual culture media at the room temperature. Upon *gelatin plates* sulphur-yellow, superficial colonies are developed, which have irregular outlines and may attain a diameter of 4 millimetres and a thickness of 0.5 millimetre; under a low power they are seen to be granular. In *gelatin stick cultures* a yellow layer forms about the point of puncture and a granular growth along the line of inoculation. Upon *agar* a yellow, slimy layer. Upon *potato* an intensely yellow layer with irregular margins, which after a time has a wrinkled surface.

183. MICROCOCCUS VIOLACEUS (Cohn).

Found in water.

Morphology.—Elliptical cocci, frequently united in chains.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms superficial colonies of hemispherical form and violet color. In *gelatin stick cultures* scanty growth along the line of puncture, and upon the surface a hemispherical mass of violet-blue color. Upon *agar* a violet-blue layer. Upon *potato* a violet-colored streak is formed along the impfstrich.

184. STAPHYLOCOCCUS VIRIDIS FLAVESCENS (Guttmann).

Found in the vesicles of varicella.

Morphology.—Micrococci of irregular dimensions, solitary, in pairs, or in irregular groups; does not differ in morphology from *Staphylococcus pyogenes aureus*.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of two days, small, greenish-yellow colonies become visible; under a low power these are seen to be spherical and slightly granular upon the surface—less so at a later date. In *gelatin stick cultures* growth occurs both upon the surface and along the line of puncture, of a grayish-yellow color. Upon *agar*, at the end of twenty-four hours at 37° C., a greenish-yellow growth occurs along the line of puncture. An abundant development occurs upon *potato* in twenty-four hours at 37° C.

185. MICROCOCCUS OCHROLEUCUS (Prove).

Found in urine of man.

Morphology.—Micrococci, from 0.5 to 0.8 μ in diameter, solitary, in pairs, or in short chains.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* micrococcus. The pigment is soluble in alcohol, insoluble in water, and is decolorized by acids. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of twenty-four hours, small, colorless colonies are developed, surrounded by a somewhat elevated and wavy border; later branching offshoots are given off from the margin and the centre acquires a sulphur-yellow color. In *gelatin stick cultures* a thin, colorless, superficial layer is quickly developed; this in three or four days acquires a sulphur-yellow color. The growth upon *potato* is scanty and is scarcely visible before the fifth day. Old gelatin cultures give off a peculiar odor.

186. MICROCOCCUS ACIDI LACTICI LIQUEFACIENS (Kreuger).

Found in "cheesy butter."

Morphology.—Oval cocci, from 1 to 1.5 μ in diameter, frequently associated in pairs or in tetrads.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying* micrococcus. Grows best at the room temperature. Upon *gelatin plates* small, white colonies are developed at the end of three days, which under the microscope are seen to have deeply cleft margins; the gelatin about the colonies is gradually liquefied. In *gelatin stick cultures* a white, granular growth occurs along the line of puncture, and a funnel-shaped liquefaction of the gelatin occurs by the third day; liquefaction progresses rapidly, and a dirty-white, slightly wrinkled layer forms upon the surface, while the gelatin below is clouded. In *milk* coagulation occurs in three days at 20° to 25° C., and lactic acid is formed; a clear layer of serum is seen above the homogeneous coagulated mass of casein, which is not subsequently peptonized.

187. MICROCOCCUS AËROGENES (Miller).

Found in the alimentary canal.

Morphology.—Large oval cocci.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms spherical colonies of dark color and smooth contour. In *gelatin stick cultures* development occurs along the line of puncture, with a brownish-yellow color, and a flat, button-like, grayish-white, soft mass is formed upon the surface; slight liquefaction occurs after some days. Upon the *surface of agar* a yellowish-white, pap-like layer is formed. The growth upon *potato* is similar to that upon agar. Possesses a great resistance against the action of acids.

188. MICROCOCCUS ALBUS LIQUEFACIENS (Von Besser).

Very common in the nasal mucus of healthy persons.

Morphology.—Micrococci, about twice as large as *Staphylococcus pyogenes albus*, spherical or elliptical; in irregular groups or in chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* produces a stocking-shaped pouch of liquefied gelatin, and after a time complete liquefaction—sometimes only partial and very tardy liquefaction. Upon *agar plates* white, shining colonies with an elevation at the centre and of the margin, with a depression between, 0.5 centimetre or more in diameter; under a low power the centre appears brownish and is surrounded by a dark zone, this by a more transparent, grayish-brown zone, and finally by an opaque marginal ring. Upon *potato* a shining, white layer is developed.

189. MICROCOCCUS FËTIDUS (Klamann).

Found in the posterior nares of man.

Morphology.—Micrococci of irregular dimensions, solitary, in pairs, in short chains, or in irregular groups; the diplococci measure $1.4\ \mu$ in diameter.

Biological Characters.—An *aërobic*, *liquefying* micrococcus. Grows at the room temperature in the usual culture media—not so well in the incubating oven. Upon *gelatin plates* oval or spherical white colonies are slowly developed. In *gelatin stick cultures* a milk-white, shining, elevated mass, with a knobby surface, is developed about the point of puncture; later this presents a central prominence surrounded by concentric circles and acquires a brownish color; liquefaction occurs slowly and the cultures develop a disagreeable odor like that of ozæna. Upon the *surface of agar* a whitish, irregular layer is slowly developed and extends over the entire surface. Upon *potato* a slimy, irregular growth, of a pale reddish-gray color and a knobby surface, which gives off an intense and disagreeable odor like that of ozæna.

190. MICROCOCCUS RADIATUS (Flügge).

Found in the air and in water.

Morphology.—Micrococci, from 0.8 to $1\ \mu$ in diameter, solitary, in short chains, or in irregular groups.

Biological Characters.—An *aërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of two days, white colonies with a yellowish-green shimmer, about one millimetre in diameter, are developed; under a low power these appear yellowish-brown and have starfish-like outgrowths; at the end of four days a delicate, regularly-arranged, radiating aureole surrounds the colonies, out-

side of which a second and finally a third similar "Strahlenkranz" is often developed; liquefaction progresses slowly about the colonies. In *gelatin stick cultures* feathery outgrowths occur at intervals along the line of puncture, radiating horizontally into the gelatin; liquefaction commences near the surface as a pointed funnel and gradually extends downward. Upon *potato* a yellowish-brown layer is quickly developed.

191. DIPLOCOCCUS ALBICANS AMPLUS.

Synonym.—Gray-white micrococcus (Bumm).

Found in mucus from the healthy vagina.

Morphology.—Diplococci resembling the "gonococcus" in form, but considerably larger, from 2 to 2.8 μ in diameter; the diplococci are usually solitary, but sometimes are in groups of three or four.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* micrococcus. Grows at the room temperature in the usual culture media. Upon *gelatin plates* slightly elevated, grayish-white colonies are formed. In *gelatin stick cultures* growth occurs upon the surface and along the line of puncture as a grayish-white stripe; after a time liquefaction commences under the surface growth.

192. MICROCOCCUS CANDICANS (Flügge).

Very common in the air and in water.

Morphology.—Spherical cocci, from 1 to 1.2 μ in diameter, associated in irregular groups.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of two days, the deep colonies are spherical and white or yellowish in color, of from 0.4 to 0.5 millimetre in diameter; under the microscope they are seen to be finely granular, dark-brown spheres. Upon the surface milk-white, shining colonies with irregular outlines, which under the microscope are seen to be finely granular and to have jagged margins. In *gelatin stick cultures* a confluent white growth forms along the line of puncture, and a button-like mass upon the surface. Upon *potato* a slimy, white layer is quickly developed.

193. MICROCOCCUS CANDIDUS (Cohn).

Found in water.

Morphology.—Small, perfectly spherical cocci, from 0.5 to 0.7 μ in diameter, united in zoöglæa masses—the intercellular zoöglæa substance is soluble in water.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms snow-white colonies with irregular outlines, which under the microscope are seen to be slightly granular. In *gelatin stick cultures* a flat, milk-white layer is formed about the point of puncture; very scanty development along the line of inoculation. Upon *agar* the same as on gelatin.

194. MICROCOCCUS ACIDI LACTICI (Marpmann).

Found in cow's milk.

Morphology.—Large cocci, solitary or in pairs.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms at the end of twenty-four hours punctiform, yellowish-white, lustreless colonies. In *gelatin stick cultures* a thin, yellowish layer forms upon the surface, which is thickest in the middle, thin and transparent at the

margin, and without lustre. In *milk*, at the end of twelve hours, a red color is developed, which disappears at the end of twenty-four hours, when coagulation has occurred as a result of the formation of lactic acid.

195. MICROCOCCUS LACTIS VISCOSUS.

Synonym.—Micrococcus of bitter milk (Conn).

Found in cream which had a bitter taste.

Morphology.—Micrococci of moderate dimensions, frequently united in pairs; in agar cultures forms short chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature—more rapidly at 35° C. Upon *gelatin plates* forms small, spherical colonies, which, as liquefaction commences, spread out upon the surface as a thin, granular mass. In *gelatin stick cultures* liquefaction commences at the surface, forming a shallow cavity, and rapidly progresses until the gelatin is entirely liquefied; the liquefied gelatin is extremely viscid. Upon *agar* a shining, homogeneous white layer is developed. Upon *potato* white, shining masses, which are more or less separated from each other. In *bouillon* an abundant development occurs and a thin film is formed upon the surface; the bouillon becomes very viscous. In *milk* growth is rapid and the milk acquires a bitter taste; at 35° C. coagulation occurs in twenty-four hours and the milk has an acid reaction; the coagulum is soft and soon commences to dissolve from the peptonizing action of the ferment, but solution is not complete. Cultures in gelatin and bouillon are especially viscid, and may be drawn out into threads which are scarcely visible and are as much as three metres long. The acid formed by the growth of this micrococcus in milk is butyric.

196. SPHÆROCOCCUS ACIDI LACTICI (Marpmann).

Found in fresh cow's milk.

Morphology.—Very small, oval cocci, in pairs or in short chains.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms porcelain-white colonies, the size of a pin's head, upon the surface of the gelatin. In *gelatin stick cultures* growth is scanty along the line of puncture; upon the surface a layer is developed which has sloping, toothed margins, and at the end of six weeks acquires a pale-yellow color. *Milk* acquires a reddish color, and is coagulated at the end of twenty-four hours, with formation of lactic acid.

197. MICROCOCCUS AQUATILIS (Bolton).

Very common in water.

Morphology.—Small cocci, associated in irregular groups.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms circular, porcelain-white, slightly elevated colonies; under a low power the deep colonies are seen to be mulberry-like in form, with roughly toothed contour and a pale-yellowish color; the superficial colonies are circular and are surrounded by a narrow, homogeneous marginal zone, while the interior is peculiarly marked, resembling a schematic drawing of a section of a liver acinus. In *gelatin stick cultures* growth occurs both on the surface and along the line of puncture, of a white color. Upon the surface of *agar* a white layer is formed.

198. MICROCOCCUS CONCENTRICUS (Zimmermann).

Found in water.

Morphology.—Micrococci, 0.9 μ in diameter, associated in irregular masses.

Biological Characters.—An *aërobic, non-liquefying* micrococcus. Grows in the usual culture media—best at the room temperature. Upon *gelatin plates* the deep colonies appear as small, bluish-gray points; the superficial colonies are at first small, bluish-gray discs, which later are irregular in outline; at the end of five days they are about three millimetres broad and consist of a central grayish-white disc surrounded by a bluish-gray ring with irregular outlines. Under a low power the deep colonies are seen to be spherical and granular, of a pale-brown or yellowish-green color, and in the interior usually several concentric rings are observed; the superficial colonies show in the interior a darker disc with irregular margins and marked by fine radiating fissures; around this is an irregular marginal zone of a pale-brown color and granular in appearance, and this is enclosed in a white, shining border. In *gelatin stick cultures* a thin, bluish-gray layer forms upon the surface, which consists of a number of concentric rings of growth arranged around the point of puncture as a centre. Upon *agar* a broad, smooth, shining layer with toothed margins and of bluish-gray white color. On *potato* a thin, slimy, yellowish-gray layer.

199. MICROCOCCUS CUMULATUS TENUIS (Von Besser).

Very common in nasal mucus of man.

Morphology.—Large oval cocci, associated in masses.

Biological Characters.—An *aërobic and facultative anaërobic, non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* grows along the line of puncture as a delicate white stripe composed of small colonies; upon the surface as a flat, transparent layer with slightly elevated margins. Upon *agar plates* as thick, transparent drops about 0.2 millimetre in diameter; under a low power these are seen to have a large brown nucleus surrounded by a grayish-brown zone having wrinkled margins; later the colonies attain a diameter of 0.5 centimetre, and appear as flat, transparent discs with a large central nucleus. Scarcely any growth upon *potato*. In *bouillon* a considerable deposit accumulates at the bottom of the tube, and the liquid is nearly transparent above.

200. MICROCOCCUS PLUMOSUS (Bräutigam).

Found in water.

Morphology.—Micrococci, $0.8\ \mu$ in diameter, associated in zoöglæa.

Biological Characters.—An *aërobic, non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* yellowish-white colonies are developed, which send out tongue-like processes and the margins of which are abruptly thickened. In *gelatin stick cultures*, from along the line of puncture, at certain points, long, delicate, white offshoots are given off into the gelatin, which resemble needle-like crystals; similar offshoots from the layer upon the surface of the gelatin are also seen; these consist of cocci in masses, arranged like chains of pearls. Upon *potato* an irregular, yellowish-white layer with tongue-like offshoots.

201. MICROCOCCUS ROSETTACEUS (Zimmermann).

Found in water.

Morphology.—Spherical or elliptical cocci, from 0.7 to $1\ \mu$ in diameter, associated in grape-like masses.

Biological Characters.—An *aërobic, non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are small, grayish-white, and usually spherical in form; under a low power they are sometimes seen to be lenticular or mussel-shaped. The superficial colonies are rather broad, shining, yellowish-gray drops, with more or less irregular outlines. In *gelatin stick cultures* a thin,

gray, rosette-like layer of irregularly circular contour develops upon the surface; very scanty growth along the line of puncture. Upon *agar* a smooth, shining, gray layer with finely toothed margins. Upon *potato* a yellowish-gray layer is quickly formed.

202. MICROCOCCUS UREÆ (Pasteur).

Found in the air and in ammoniacal urine.

Morphology.—Micrococci, from 0.8 to 1 μ in diameter, solitary, in pairs, in tetrads, or in short chains; also in zoöglæa masses.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature—better at 30° to 35° C. Upon *gelatin plates* forms, at the end of twenty-four hours, small, white, pearly, shining colonies of smooth surface and sharply defined outline; at the end of ten days these are large, flat colonies resembling a drop of stearin. In *gelatin stick cultures* development occurs along the line of puncture in form of a thin, tenacious thread. Old cultures have a paste-like odor.

According to Von Jaksch, a very favorable medium for the growth of this micrococcus is made by adding to one litre of water, magnesia sulphate one-sixteenth gramme, potassium hypophosphite one-eighth gramme, potassium sodium tartrate five grammes, urea five grammes. In urine and solutions containing urea carbonate of ammonia is formed during the development of this micrococcus—ammoniacal fermentation.

203. MICROCOCCUS UREÆ LIQUEFACIENS (Flügge).

Found in ammoniacal urine.

Morphology.—Spherical cocci, from 1.25 to 2 μ in diameter, solitary, in chains of three to ten elements, or in irregular groups.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms, at the end of twenty-four hours, small, white, punctiform colonies, which under a low power appear as well-defined, dark-gray spheres; after they reach the surface of the gelatin the colonies become considerably larger, have a yellowish-brown color and a central nucleus consisting of the original deep colony; the surface of the superficial colonies is granular, the outline becomes gradually wavy; liquefaction of the gelatin around the colonies occurs gradually. In *gelatin stick cultures* a confluent, white growth develops along the line of puncture, and liquefaction quickly occurs and extends to the walls of the tube; finally one-half or more of the gelatin is liquefied and has a whitish, clouded appearance, while a thick, yellowish-white deposit is seen at the bottom.

204. MICROCOCCUS VITICULOSUS (Katz).

Found in the air and in water.

Morphology.—Oval micrococci, from 1 to 1.2 μ in diameter; forms thick zoöglæa masses.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows rapidly in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are seen to consist of hair-like branches given off from a centre, and which for some distance form a delicate network; under a low power these branches are seen to be made up of zoöglæa masses of various dimensions united in chaplets. The superficial colonies extend rapidly as a thin, jelly-like, clouded white layer, from which fine threads are given off into the deeper layers of the gelatin. In *gelatin stick cultures* growth occurs along the line of puncture as a delicate network of threads; upon the surface a feathery growth occurs along the line of inoculation. Upon *potato* a dry, dirty-white layer is quickly developed.

205. DIPLOCOCCUS ALBICANS TARDISSIMUS (Eisenberg).

Synonym.—Milk-white micrococcus (Bumm).

Found in secretions from the vagina and cervix, especially in the vaginal secretions of puerperal women.

Morphology.—Diplococci resembling the "gonococcus," and consisting of two biscuit-shaped halves separated by a cleft which is not as broad as that seen in *Micrococcus gonorrhoeae*; the diameter, from pole to pole, averages about $1.25\ \mu$. In unstained preparations the cleft is not seen and the diplococci appear as spherical, highly refractive bodies.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows very slowly at the room temperature in the usual culture media. Upon *gelatin plates* forms extremely small, punctiform colonies, which under a low power are seen to be spherical, opaque, and brownish-green in color; at the end of two weeks they may attain a diameter of two millimetres. In *gelatin stick cultures* small, isolated, grayish-white colonies are developed, after some days, along the line of puncture, and a thin, whitish, stearin-like layer with irregularly dentate margins is slowly developed upon the surface. Upon the *surface of agar* a thin, moist, grayish-white layer with dentate margins is slowly developed.

206. DIPLOCOCCUS ALBICANS TARDUS (Unna).

Found upon the surface of the body in individuals having eczema seborrhoeicum.

Morphology.—Diplococci consisting of two oval elements, with the long diameters in parallel planes, from 0.7 to $0.8\ \mu$ long and $0.6\ \mu$ broad; often associated in short chains or in irregular groups.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are usually oval, dark-yellow, and at the end of eight days are as large as a mustard seed. The superficial colonies are circular in outline, with well-defined margins, elevated, grayish-yellow, and at the end of eight days one to two millimetres in diameter; under a low power they are seen to be granular, grayish-yellow, with shining margins; at the end of five weeks they are gray and present two or three zones of different dimensions, with an elevated circular centre and finally with thin, slimy, dentate margins; under the microscope finely granular projections are seen upon the surface. In *gelatin stick cultures*, at the end of three weeks, a thin, waxy-looking, yellowish-white layer with finely dentate margins develops upon the surface and a scanty growth has occurred along the line of puncture. At the end of five weeks this superficial layer may have a diameter of one centimetre. Upon the *surface of agar*, at the end of five weeks, a yellowish-gray streak with irregular, dentate margins and a dull lustre is formed along the line of inoculation.

207. STAPHYLOCOCCUS ALBUS LIQUEFACIENS.

Synonym.—White liquefying staphylococcus (Escherich).

Found occasionally in the alvine discharges of healthy infants.

Morphology.—Micrococci of from 0.8 to $1.2\ \mu$ in diameter, occasionally oval in form and three μ in long diameter; associated in irregular groups—considerably larger than *Staphylococcus pyogenes albus*.

Biological Characters.—An *aërobic*, *liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms spherical, white colonies, which after some time cause a gradual liquefaction of the surrounding gelatin. In *gelatin stick cultures* a scanty development is seen along the line of puncture at the end of three to four days, and gradual liquefaction of the gelatin occurs in funnel form; the liquefied gelatin is viscid, of syrupy consistence, and slightly clouded; the

surface is covered by a white layer of micrococci; development usually ceases before complete liquefaction has occurred. Upon *agar* and upon *blood serum* a white layer is developed, which presents nothing characteristic; blood serum is not liquefied. Upon *potato* a very scanty, thin, colorless layer, which later appears as a collection of white, button-like masses.

208. MICROCOCCUS OVALIS (Escherich).

Found frequently in meconium and fæces of milk-fed infants.

Morphology.—Micrococci of from 0.2 to $0.3\ \mu$ in diameter, frequently seen as oval cells 0.6 to $0.7\ \mu$ long and $0.3\ \mu$ broad, in the middle of which a commencing line of division may sometimes be seen; sometimes in short chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms very small colonies which are in no way characteristic. In *gelatin stick cultures* small, white colonies are developed along the line of puncture, and no development occurs upon the surface, or a scanty, colorless ring of growth surrounds the point of puncture. Upon *potato* a tolerably abundant growth occurs, consisting of a small, white layer. In *milk* it causes an acid reaction, and coagulation after several days.

209. DIPLOCOCCUS CORYZÆ (Hajek).

Found in nasal mucus in acute nasal catarrh.

Morphology.—Large diplococci, flattened along the line of contact; resemble short bacilli with round ends.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms white, glass-like, slightly elevated colonies. In *gelatin stick cultures* the growth resembles that of Friedländer's bacillus at first, but the superficial growth is flatter after some days. Upon the *surface of agar* forms a diffuse layer.

210. MICROCOCCUS FINLAYENSIS (Sternberg).

Obtained by Finlay in cultures from the liver and spleen of a yellow-fever cadaver.

Morphology.—Micrococci, from 0.5 to $0.7\ \mu$ in diameter, solitary, in pairs, or occasionally in groups of four; also in irregular masses. Like other staphylococci, the cocci are seen, in properly stained preparations, to be made up of two hemispherical portions.

Biological Characters.—An *aërobic*, *liquefying*, *chromogenic* micrococcus. Grows slowly at the room temperature in the usual culture media. In *gelatin stick cultures* growth occurs along the line of puncture and liquefaction near the surface; the cup-shaped cavity which is slowly formed is lined with a very viscid, opaque, pale-yellow layer of cocci. Upon the *surface of agar* a viscid layer having a pale-yellow color is formed. Not pathogenic for rabbits or guinea-pigs.

211. MICROCOCCUS OF FREIRE.

Presented to the writer, at the time of his visit to Brazil (1887), by Dr. Domingos Freire, as his yellow-fever germ—so-called *Cryptococcus xanthogenicus*.

Morphology.—Micrococci, from 0.5 to $0.8\ \mu$ in diameter, solitary, in pairs, or in irregular agglomerations; like other staphylococci, groups of four and chains of three or four elements are occasionally formed.

Stains with the aniline colors usually employed and by Gram's method.

Biological Characters.—An *aërobic, liquefying* staphylococcus. Grows in the usual culture media at the room temperature. Is killed by exposure to a temperature of 60° C. for ten minutes. Vitality not destroyed by long exposure to a freezing temperature. Development occurs at a comparatively low temperature—10° to 15° C. Preserves its vitality for several months in cultures. In *gelatin stick cultures* development occurs along the line of puncture and liquefaction in cup shape near the surface; the cocci accumulate at the bottom of the cup as a milk-white deposit; later the gelatin may be completely liquefied at the upper part of the tube for half an inch or more, and the non-liquefied gelatin forms a horizontal floor upon which a milk-white deposit is seen. In *agar stick cultures* an irregular, white, opaque growth is seen along the line of puncture, and a soft, milk-white layer, with irregular outlines, forms upon the surface. Upon *potato* a milk-white layer, from three to five millimetres wide, is developed along the line of inoculation at the end of forty-eight hours at 37° C. Does not coagulate *milk*.

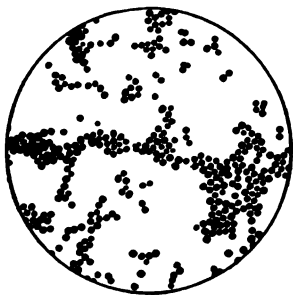


FIG. 199.



FIG. 200.

FIG. 199.—Micrococcus of Freire, from a gelatin culture. $\times 1,000$. From a photomicrograph. (Sternberg.)

FIG. 200.—Culture of Freire's micrococcus in nutrient gelatin, end of eight days at 22° C. From a photograph. (Sternberg.)

In the writer's experiments this micrococcus has not proved to be pathogenic for guinea-pigs. According to Freire, it is not pathogenic for these animals in the winter months, but in summer, at Rio de Janeiro, it is fatal to guinea-pigs and to small birds.

212. STREPTOCOCCUS COLI GRACILIS (Escherich).

Found in the fæces of healthy children on flesh diet—not during the period of nursing (Escherich).

Morphology.—Micrococci, from 0.2 to 0.4 μ in diameter, usually in S-shaped chains containing from six to twenty elements. In agar cultures the chains are shorter, and upon potato they are rarely seen; the elements in a chain are often elongated and show indications of commencing transverse division.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying* streptococcus. Grows rapidly in the usual culture media at the room temperature. Upon *gelatin plates* the colonies are at first small, spherical, and well defined, later bulging and lying at the bottom of a funnel of lique-

fied gelatin; by reflected light the colonies are of a whitish lustre. In *gelatin stick cultures* development occurs along the line of puncture, and liquefaction commences on the second day along the entire line, forming a tube of rather uniform dimensions; the liquefied gelatin is slightly clouded, and a very slight deposit of a white color is seen at the bottom of the tube; by the eighth to tenth day liquefaction is complete; in older cultures a finely granular, whitish deposit is seen, while the gelatin above is quite transparent and has an acid reaction. Upon *agar* a very scanty superficial growth occurs. *Blood serum* is not liquefied, but small, shining scales are developed upon the surface. Upon young *potato* a thin layer of white, button like colonies develops upon the surface; no growth occurs on old potato. *Milk* is coagulated and acquires an acid reaction after a considerable time.



FIG. 201.—*Str. coli gracilis*, from a liquefied gelatin culture. $\times 970$. (Escherich.)

213. STREPTOCOCCUS ACIDI LACTICI (Grotenfeld).

Found in coagulated milk in Finland.

Morphology.—Spherical or oval cocci, from 0.5 to $1\ \mu$ long and 0.3 to $0.6\ \mu$ thick, associated in long chains.

Biological Characters.—An *anaerobic* (not strict), *non-liquefying* streptococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms spherical, white colonies—best when the plate is covered with a layer of gelatin to exclude the air. In *gelatin stick cultures* an abundant development along the line of puncture only. In *milk* causes coagulation of the casein and an acid reaction.

214. STREPTOCOCCUS GIGANTEUS URETHRÆ (Lustgarten).

Found in the normal human urethra.

Morphology.—Spherical cocci, 0.8 to $1\ \mu$ in diameter, associated in chains which often contain many hundred elements, and are often united in thick, tangled masses.

Biological Characters.—An *aerobic* micrococcus. Grows slowly in nutrient agar at 37°C .; does not grow at the room temperature. Upon *agar plates* elongated, drop-like colonies are slowly formed, which are not elevated above the surface; these are easily overlooked in reflected light, and in transmitted light are iridescent; the colonies are often shaped like a clover leaf; they never become confluent, and attain their greatest development by the eighth day; transplantation to other agar tubes is rarely successful. In *streak cultures* upon agar development occurs for the most part in the condensation water as a flocculent deposit.

215. STREPTOCOCCUS ALBUS.

Synonym.—Weisser Streptococcus (Maschek).

Found in Freiburg water by Tils.

Morphology.—Streptococci, which show independent movements when undergoing binary division (Tils).

Biological Characters.—An *aerobic*, *liquefying* streptococcus. Grows in the usual culture media at the room temperature. Upon *gelatin plates* superficial colonies are developed, which are flat, circular in outline, and bounded by a white margin; saucer-shaped liquefaction quickly occurs, and under a low power a pale-brown cloud is seen in the centre. Upon *agar* the disc-shaped colonies have a darker interior. In *gelatin stick cultures* the development is chiefly upon the surface; liquefaction progresses rapidly, and a white deposit is formed. Upon *potato* a slimy, white layer is quickly developed.

216. STREPTOCOCCUS VERMIFORMIS.

Synonym.—Wurmförmiger Streptococcus (Maschek).

Found in Freiburg water by Tils.

Morphology.—Streptococci, which show slow, vermiform, progressive movements; the juxtaposition of several cocci in a chain gives the appearance of bacilli, which are again united in chains (Tils).

Biological Characters.—An *aërobic, liquefying streptococcus*. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms yellowish-white colonies, which sink into the gelatin as liquefaction occurs; about the more transparent interior a darker liquefied ring is seen, which is bounded by a white border; under a low power the interior is seen to be finely granular and pale-brown in color; the margin has a radiate appearance. In *gelatin stick cultures* growth occurs upon the surface and along the line of puncture, causing rapid liquefaction of the gelatin; a dirty-yellow deposit accumulates at the bottom of the tube. Upon *potato* a dirty-yellow layer is slowly developed; later this becomes somewhat darker.

217. STREPTOCOCCUS BREVIS (Von Lingelsheim).

Obtained from normal human saliva, and differentiated from *Streptococcus pyogenes* by Von Lingelsheim (1891).

Morphology.—Micrococci, solitary, in pairs, or in short chains—seldom as many as eight to ten elements.

Biological Characters.—An *aërobic and facultative anaërobic, non-liquefying streptococcus*. Grows in the usual culture media at the room temperature—more rapidly than *Streptococcus pyogenes*. *Bouillon* is uniformly clouded by the growth of this streptococcus, while the streptococci which form long chains leave the bouillon transparent and form conglomerate masses. In blood serum of the ox the growth of this streptococcus exactly resembles that of *Streptococcus pyogenes*, long chains being formed which are associated in conglomerate masses. Upon *gelatin plates*, at the end of twenty-four hours, forms punctiform colonies, which under the microscope are usually seen to be spherical in form with an even contour. Upon the *surface of agar* a thin, homogeneous, yellowish-gray layer is developed along the impistrich—not composed of separate colonies; along the line of puncture in nutrient agar the colonies are largest below and are frequently flattened or irregular in form. In *gelatin stick cultures* the development at first resembles that of "*Streptococcus longus*" (*Streptococcus pyogenes*), but after three days a funnel-like cavity forms near the surface of the gelatin, which finally extends downward to a distance of four to five millimetres, while below this, usually separated by a space in which there is no evidence of growth, colonies the size of a pin's head are developed along the line of puncture in rows. Upon *potato*, at 37° C., an abundant development occurs within forty-eight hours, in the form of a confluent, white layer, easily stripped from the surface.

STREPTOCOCCUS CADAVERIS (Sternberg).

Possibly identical with *Streptococcus brevis*, described above. Found by the writer in the liver of a yellow-fever cadaver.

Morphology.—Micrococci, in pairs, or in short chains which are usually made up of cocci in pairs, or oval elements which show, more or less distinctly, commencing binary division; diameter about 0.5 μ .

Biological Characters.—An *aërobic and facultative anaërobic, non-liquefying streptococcus*. Grows in the usual culture media at the room temperature. Grows readily in a decidedly acid medium. In *gelatin stick cultures* opaque, spherical colonies are formed along the line of puncture—larger and more opaque than in similar cultures of *Streptococcus pyogenes*.

made at the same time; later the isolated colonies at the bottom of the line of puncture are coarsely granular and irregular in outline; no growth upon the surface of gelatin. Upon the *surface of agar*, in Havana, a white mass

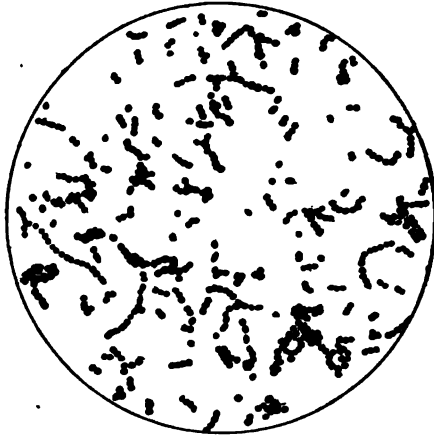


FIG. 202.—*Streptococcus cadaveris*, from a culture in *agua coco*. $\times 1,000$. From a photomicrograph. (Sternberg.)

formed about the point of puncture; later, in Baltimore, a thin, translucent layer developed upon the surface of agar cultures. Very thin, white growth upon *potato* at the end of twelve days. In *bouillon* and *agua coco* it forms



FIG. 208.—*Streptococcus Havanensis*, from acid vomit of yellow-fever patient, kept twenty four hours in a collecting tube. $\times 1,000$. From a photomicrograph. (Sternberg.)

little flocculi made up of chains. In *old agar cultures* it is very small and not in chains. In *veal broth*, long chains, in which the cocci are larger than usual; the elements vary considerably in size in the same chain.

218. *STREPTOCOCCUS HAVANIENSIS* (Sternberg).

Found in acid vomit of a yellow-fever patient in military hospital, Havana.

Morphology.—Micrococci, from 0.6 to 0.9 μ in diameter, associated in long chains which are made up of cocci in pairs or of oval elements showing commencing transverse division.

Biological Characters not determined.

219. STREPTOCOCCUS LIQUEFACIENS (Sternberg).

Obtained from liver of yellow-fever cadavers, also from contents of intestine.

Morphology.—Spherical or short oval micrococci, from 0.4 to 0.6 μ in diameter, solitary, in pairs, or in short chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* streptococcus. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* liquefaction occurs rapidly along the entire

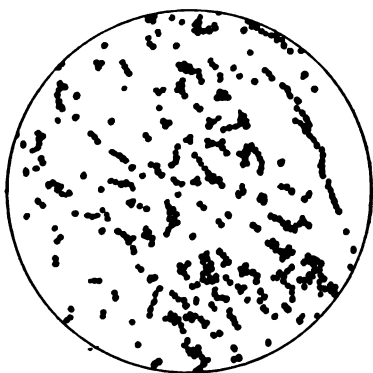


FIG. 204.

FIG. 204.—*Streptococcus liquefaciens*, from anaërobic culture in nutrient gelatin. $\times 1,000$. From a photomicrograph. (Sternberg.)



FIG. 205.

FIG. 205.—*Streptococcus liquefaciens*; culture in nutrient gelatin, end of seven days at 22° C. (Sternberg.)

line of puncture, and by the end of a week the gelatin is completely liquefied; it is but slightly opalescent, and a scanty white deposit forms at the bottom of the tube. In *agar stick cultures* a scanty growth occurs at the point of puncture, and closely crowded colonies are developed along the line of inoculation. Upon *potato* a thin and limited, dry, white layer is formed along the line of inoculation in four to five days. Not pathogenic for rabbits or guinea-pigs.

220. MICROCOCCUS TETRAGENUS VERSATILIS (Sternberg).

Synonym.—*Micrococcus tetragenus febris flavæ* (Finlay).

Obtained by Dr. Finlay, of Havana, from his "mosquito cultures"—from the excrement of mosquitoes which he had allowed to draw blood from yellow-fever patients; by the writer from cultures from the surface of the body of patients in hospital in Havana, and from the air.

Morphology.—Micrococci, in tetrads, or in irregular groups containing three or more united elements; varies greatly in the size and grouping of the elements, which are sometimes less than 0.5 μ in diameter and may measure 1.5 μ . This micrococcus would probably be classed as a *sarcina* by some bac-

teralogists, but division in three directions, forming packets, has not been observed.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic, liquefying, chromogenic* micrococcus. Grows in the usual culture media at the room temperature. Colonies in gelatin roll tubes are spherical, opaque, at first of a pale-yellow and later of a lemon-yellow color; liquefaction around the colony usually does not commence for several days and progresses slowly. In *gelatin stick cultures* very scanty development occurs along the line of puncture, and the gelatin is liquefied in cup shape by the surface growth; at the bottom of the liquefied gelatin a viscid, pale-yellow mass accumulates. Upon the *surface of agar* a thick, viscid, yellow layer is formed along the impfstrich, which

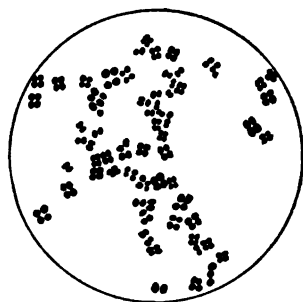


FIG. 206.

FIG. 206.—*Micrococcus tetragenus versatilis*, from a single colony in nutrient gelatin. $\times 1,000$. From a photomicrograph. (Sternberg.)



FIG. 207.

FIG. 207.—*Micrococcus tetragenus versatilis*; culture in nutrient gelatin, end of two weeks at 22° C. (Sternberg.)

gradually extends over the entire surface; the color varies from cream-yellow to lemon-yellow, and the surface is moist and shining. The growth upon *potato* is similar to that upon agar. Not pathogenic for rabbits or guinea-pigs.

221. PEDIOCOCCUS ALBUS (Lindner).

Found in well water.

Morphology.—Micrococci, solitary, in pairs, or in tetrads; frequently in pseudo-sarcina—accidental—groups.

Biological Characters.—An *aërobic, liquefying* micrococcus. Grows in the usual culture media at the room temperature. Is destroyed by a temperature of 60° C. in eight minutes. Upon *gelatin plates* forms spherical colonies, around which liquefaction rapidly occurs, allowing the colonies to fall to the bottom of the liquefied gelatin, where they form irregular flocculi. In *gelatin stick cultures*, at the end of twenty-four hours, a deep channel of liquefaction has already formed, at the bottom of which a white, loose sediment collects; by the fourth day a pale-orange color is developed. Upon the *surface of agar* a broad, dry layer forms along the impfstrich; old cultures have an orange color. Upon *potato* a dirty-white layer is developed. Produces an acid reaction in culture media.

222. *PEDIOCOCCUS ACIDI LACTICI* (Lindner).

Found by Lindner in mash from malt and in hay infusion.

Morphology.—Micrococci, from 0.6 to 1 μ in diameter, solitary, in pairs, or in tetrads—division in two directions.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows very slowly in the usual culture media at the room temperature. Upon *gelatin plates* small colonies are developed, which are first colorless and later yellowish-brown. In *agar stick cultures* a moist, thin, colorless layer is slowly developed upon the surface. Upon *potato* a scanty, scarcely visible growth. Lactic acid is produced by the growth of this tetragenus in saccharine solutions.

223. *PEDIOCOCCUS CEREVISIÆ* (Balcke).

Found in beer and in the air of breweries.

Morphology.—Micrococci, solitary, in pairs, or in tetrads.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* micrococcus. Grows slowly in the usual culture media at the room temperature. Is killed by a temperature of 60° C. maintained for eight minutes. Upon *gelatin plates* forms small, colorless colonies, which later are yellowish-brown. In *gelatin stick cultures* a white, leaf-like layer is formed on the surface and a grayish-white growth along the line of puncture. Upon *agar* a moist, grayish-white, iridescent layer with smooth margins. Upon *potato* a scanty, scarcely visible growth.

224. *MICROCOCCUS TETRAGENUS MOBILIS VENTRICULI* (Mendoza).

Found in the contents of the stomach.

Morphology.—Micrococci, in tetrads, enclosed in a transparent capsule.

Biological Characters.—An *aërobic*, *non-liquefying*, *motile* (?) tetragenus. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* forms superficial colonies which are circular in outline, with well-defined margins, and dirty-white in color; under a low power they are seen to be finely granular, both on the surface and in the interior. In *gelatin stick cultures* grows upon the surface as a dirty-white layer; old cultures have a "sugar color" and give off an odor like that of skatol. The growth upon *agar* is similar to that upon gelatin.

225. *MICROCOCCUS TETRAGENUS SUBFLAVUS* (Von Besser).

Found in nasal mucus.

Morphology.—Spherical or oval cocci of medium size, usually associated in tetrads.

Biological Characters.—An *aërobic* and *facultative anaërobic* tetragenus. Does not grow in nutrient gelatin. Grows at the room temperature in nutrient agar. Upon *agar plates* forms flat, dirty-white colonies, 0.5 cubic centimetre in diameter, with shining surface and somewhat wrinkled margins; under a low power these are seen to have a small brown nucleus surrounded by a grayish-brown, irregularly striped zone, around which is a wrinkled margin. The deep colonies are dark-green and punctiform, of very irregular form. Upon the *surface of agar* a broad, flat, grayish-white layer is formed, which in three or four days commences to turn brown at the margins, and at the end of four weeks has the color of cultures of *Staphylococcus pyogenes aureus*. Upon *potato* a pale-brown layer is developed along the line of inoculation.

226. *SARCINA AURANTIACA*.

Found in the air and in water.

Morphology.—Micrococci, in pairs, with hemispherical elements, in tetrads.

or in packets containing eight or more elements, as a result of division in three directions. The cells are smaller than those of *Sarcina lutea*.

Biological Characters.—An *aërobic, liquefying, chromogenic sarcina*. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms small, spherical, granular colonies of an orange-yellow color. In *gelatin stick cultures* liquefaction occurs slowly along the line of inoculation, and in old cultures a yellow deposit is seen at the bottom of the transparent, liquefied gelatin. Upon the *surface of agar* an abundant, shining, golden-yellow layer is formed. Upon *potato* development is scanty and slow, being limited to the line of inoculation.

227. *SARCINA LUTEA* (Schröter).

Found in the air.

Morphology.—Micrococci, $1\ \mu$ or more in diameter, associated in pairs, in groups of four, or in packets containing eight or more elements.

Biological Characters.—An *aërobic, liquefying, chromogenic sarcina*. Grows in the usual culture media at the room temperature. Upon *gelatin plates* small, spherical, yellow colonies are slowly developed. In *gelatin stick cultures* development occurs only on the surface in the form of a coarsely granular, yellow layer of moderate dimensions; liquefaction of the gelatin occurs in old cultures and progresses slowly. Upon the *surface of agar* a rather thick, canary-yellow layer is quickly developed. Upon *potato* a sulphur-yellow streak is slowly developed along the line of inoculation.



FIG. 208.—*Sarcina lutea*, from an agar culture. $\times 1,000$. From a photomicrograph. (Fränkel and Pfeiffer.)

228. *SARCINA FLAVA* (De Bary).

Found by Lindner in beer.

Morphology.—Small micrococci in packets of sixteen to thirty-two elements, which may be associated in irregular masses or united in larger packets of cubical form.

Biological Characters.—An *aërobic, liquefying, chromogenic sarcina*. Grows in the usual culture media at the room temperature. Upon *gelatin plates* spherical, yellow colonies are developed, and liquefaction of the gelatin around them commences in about four days. Upon the *surface of agar* forms a yellow layer. Upon *potato* the growth is scanty and limited to the impfstich, yellow in color.

229. *SARCINA ROSEA* (Schröter).

Found in the air.

Morphology.—Large cocci in cubical packets with rounded corners, and about $8\ \mu$ thick.

Biological Characters.—An *aërobic, liquefying, chromogenic sarcina*. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* liquefaction occurs rapidly, and old cultures acquire a red color, which by the addition of sulphuric acid is changed to bluish-green. Upon *agar* growth is slow, and small, cartilaginous-looking masses are developed. Upon *potato* an abundant, intensely red layer is quickly developed. In *bouillon* development is rapid and a red sediment is deposited.

230. *SARCINA ALBA* (Eisenberg).

Found in the air and in water.

Morphology.—Small cocci, associated in pairs or in tetrads.

Biological Characters.—An *aërobic, liquefying* (very slightly) sarcina. Grows very slowly at the room temperature in the usual culture media. Upon *gelatin plates* forms small, round, white colonies. In *gelatin stick cultures* forms a white, button-like mass at point of puncture; scanty growth along line of inoculation. Upon *potato* grows very slowly in the form of a yellowish-white layer limited to the line of inoculation.

231. *SARCINA CANDIDA* (Reinke).

Found in the air about breweries.

Morphology.—Micrococci of 1.5 to 1.7 μ in diameter, solitary, in pairs, or in tetrads; under certain circumstances, as for example in hay infusion, multiplication occurs in three directions, forming sarcina-like packets.

Biological Characters.—An *aërobic, liquefying* sarcina. Grows slowly at the room temperature in the usual culture media. Upon *gelatin plates* forms shining white, spherical colonies, which later have a yellowish color and are surrounded by liquefied gelatin. In *gelatin stick cultures* causes rapid liquefaction along the line of puncture. Upon the *surface of agar* forms a white, moist, shining layer with smooth margins.

232. *SARCINA PULMONUM* (Hauser).

Found in the sputum of patients with phthisis.

Morphology.—Micrococci, from 1 to 1.5 μ in diameter; usually associated in tetrads, but may form cubical groups of eight cells.

Biological Characters.—An *aërobic, non-liquefying* sarcina. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* in three days white, punctiform colonies are seen; later these still remain small; under a low power they are seen to be coarsely granular. In *gelatin stick cultures* the development is very scanty along the line of puncture; upon the surface a round, sharply defined layer of a pearl-gray color is developed; later this becomes tolerably thick, is grayish-brown in color, glistening, and has more or less folded and irregular margins. The growth upon *potato* is very scanty and is confined to the line of inoculation.

This sarcina is said by Hauser to form endogenous spores which may be demonstrated by Neisser's method of staining, and which have great resistance to heat. When cultivated *in urine* it causes ammoniacal decomposition of the urea.

233. *SARCINA VENTRICULI* (Goodsir).

Found in the contents of the stomach of man and animals.

Morphology.—Spherical or slightly oval cells, about 2.5 μ in diameter; united in cubical groups with rounded corners, containing eight elements, and then associated in larger "packets."

Biological Characters.—An *aërobic, non-liquefying* sarcina. Grows rapidly at the room temperature in suitable media. Upon *gelatin plates*, at the end of thirty-six to forty-eight hours, hemispherical, yellowish colonies are developed upon the surface; these contain cocci in pairs and tetrads, but not in cubical packets. In *hay infusion* development occurs upon the surface in the form of small, brownish scales, while a brownish, flocculent deposit is seen at the bottom of the tube; the sarcina form is well developed. Upon *potato* a dry, colorless strip develops along the *impfstrich* in twenty-four hours; later this acquires a chrome-yellow color. Upon the surface of *blood serum* flat, round, pale-yellow colonies are formed along the line of inoculation.

234. MICROCOCCUS AMYLOVORUS (Burrill).

Synonym.—Micrococcus of pear blight.

Found in the diseased branches of pear trees affected with "blight"—"fire blight." Discovered by Burrill (1878) and carefully studied by Arthur (1886).

Morphology.—Oval cells, from 1 to 1.25 μ long and 0.5 to 0.75 μ broad; usually solitary, often in pairs, and occasionally in a linear series of four. In fluid culture media form zoöglæa, which are spherical or oblong and may be from thirty to forty μ long and twenty to thirty μ broad; these zoöglæa masses may be solitary, or united in pairs or short chains; they are found especially upon the surface of the culture medium, and after a time present a wrinkled appearance, giving some resemblance to the external surface of the brain.

Biological Characters.—An *aërobic, non-liquefying, motile* micrococcus (bacillus?). Grows in various media at the room temperature. According to Arthur, this micrococcus exhibits active "swarming movements" when undergoing rapid multiplication in a favorable medium: "The bacteria move rapidly back and forth, in and out, among each other, but never in a straight line to any distance." An *infusion of potato* constitutes a favorable culture medium. In this medium, at a temperature of 25° to 30° C., turbidity commences in twenty-four hours and bubbles of carbon dioxide are given off; in forty-eight hours the liquid is entirely turbid and a thin, whitish pellicle has formed on the surface, at the same time a sediment commences to form at the bottom of the tube. Upon the surface of *nutrient gelatin* development is very scanty and scarcely perceptible. In the interior small, punctiform colonies are developed; these are spherical or oval, and may attain a diameter of 0.5 millimetre. Upon slices of unripe pear, in two or three days, fine milky drops like beads of dew are developed. Upon *potato* a thin, moist, somewhat yellowish layer is developed.

The experiments of Burrill and of Arthur show that the disease known as pear blight is produced by applying cultures of this "micrococcus" to healthy branches of the pear tree.

235. ASCOCOCCUS BILLROTHII.

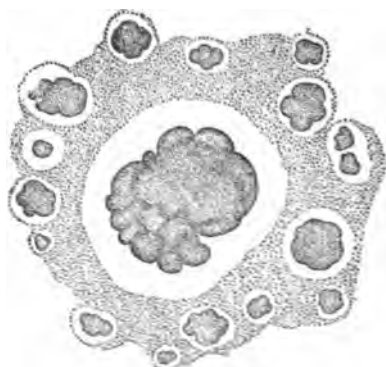


FIG. 309.—*Ascococcus Billrothii*.

Found by Billroth in putrefying flesh infusion.

Morphology.—Small cocci, united in peculiar colonies. Upon the surface of liquid media form a cream-like layer in which numerous small, spherical or oval masses are seen. Under the microscope these are seen to consist of a jelly-like, cartilaginous, extremely resistant envelope from ten to fifteen μ thick; in the interior of this one or more spherical or oval masses of cocci are enclosed, which are from twenty to seventy μ or more in diameter; the cocci are closely crowded and united by an unusually firm and scanty intercellular substance.

Biological Characters.—*Aërobic*. Grows at the room temperature. Produces a strongly alkaline reaction in culture media, due to the development of ammonia. According to Cohn, a greenish-white, slimy mass is developed upon slices of beet, and in the juice of the sugar beet it produces a slimy fermentation.

236. *LEUCONOSTOC MESENTEROIDES* (Cienkowski).

Synonyms.—Froschlaichpilz; Pilz der Dextrangärung.

Found upon the beet juice and the molasses of sugar factories, where it develops as large, jelly-like masses resembling frog spawn; also upon raw or cooked carrots or sugar beets.

Morphology.—The reproductive elements are spherical or oval, and from 1.8 to $2\ \mu$ in diameter; they consist of a firm, membranous envelope with shining contents. When these germinate—according to Van Tieghem—the external layer of the cell wall is ruptured in an irregular way and a middle layer is extruded to form a thick, jelly-like envelope, while the inner layer remains in contact with the protoplasmic contents of the cell; the cell and its envelope then become elongated and binary division of the reproductive element occurs; this process is repeated in the segments, and as a result a chain of spherical elements enclosed in a sausage-shaped mass of jelly is developed; later the chains become curved and break up into longer or shorter fragments, which are enclosed in an irregular gelatinous mass; these zoöglœa crowded together form small masses having a parenchymatous structure; these adhere to each other when they come in contact, and when distributed in a liquid may be brought together by shaking to form a large, jelly-like mass which has apparently been developed by the process of shaking, as the smaller masses suspended in the fluid could not readily be seen. The zoöglœa masses have a cartilage-like consistence and can be cut into sections with a razor. The jelly is hyaline, but in beet juice often has a grayish color from the presence of impurities attached to its surface. When treated with a solution of hæmatoxylon it acquires a brown color. After some time the jelly is dissolved, and the cocci are set free and in a suitable medium again develop zoöglœa. Van Tieghem has demonstrated the formation of spores. These are formed when conditions are no longer favorable for the development of the vegetative cells; here and there a cell in a chain is seen to increase in dimensions, and in the interior of this mother cell a considerable number of refractive spores are developed.

Biological Characters.—This coccus is *aërobic*. It is readily cultivated in liquid media containing glucose. According to Van Tieghem, it appropriates grape sugar directly and cane sugar indirectly—*i.e.*, after it has been changed into grape sugar by a ferment produced by the coccus. Development is very rapid under favorable conditions. Thus Durin reports that a wooden tub in which beet juice had been kept, and upon the walls of which a thin, slimy mass of this microörganism remained attached, was filled with a neutral solution of molasses containing ten per cent of sugar. At the end of twelve hours the entire contents of the tub had been changed into a compact, jelly-like mass by the development of *Leuconostoc mesenteroides*. The chemical formula for the jelly is $C_{12}H_{10}O_{12}$.

IX.

NON-PATHOGENIC BACILLI.

A. Chromogenic, Non-Liquefying Bacilli.

237. BACTERIUM LUTEUM (List).

Found in water.

Morphology.—Elliptical cells from 1.1 to 1.3 μ long.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* bacillus. Produces an orange-yellow pigment soluble in water, alcohol, and ether; not changed by alkalies, but at once destroyed by acids. Grows at the room temperature in the usual culture media. Upon *gelatin plates* forms irregular colonies upon the surface, of slimy consistence and orange-yellow in color; under a low power these are seen to consist of many club-shaped, coarsely granular, zoöglæa masses, each one of which is made up of several segments. In *gelatin stick cultures* development occurs slowly along the line of puncture and an orange-yellow layer forms on the surface. In *milk*, at 30° C., a layer forms upon the surface in from twenty-four to thirty hours, and the milk below has a pale-yellow color; later coagulation of the casein occurs.

238. BACILLUS AURANTIACUS (Frankland).

Found in well water.

Morphology.—Short, thick bacilli which vary greatly in their dimensions; sometimes associated in pairs; often grow out into long filaments.

Biological Characters.—An *aërobic, non-liquefying, slightly motile, chromogenic* bacillus. Forms an orange-colored pigment. Grows at the room temperature in the usual culture media. Upon *gelatin plates* the superficial colonies are homogeneous, opaque, elevated masses of a pale-orange color; the deep colonies spherical and granular. In *gelatin stick cultures* scarcely any development occurs along the line of puncture; upon the surface a shining, orange-colored layer is formed. Upon *agar* a similar development occurs. Upon *potato* the growth is limited to the line of inoculation and is of shining, orange-red color.

239. BACILLUS BRUNNEUS (Adametz).

Found in water.

Morphology.—Small, slender bacilli.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, non-motile* bacillus. Forms a brown pigment. Forms spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the superficial colonies appear as little drops of slimy consistence, but not viscid; under a low power they are at first white and opaque, with circular contour; later, in ten to fourteen days, they are gray, and a brown color is given off from the lower surface. In *gelatin stick cultures* development

occurs chiefly along the line of puncture; upon the surface a milk-white, slimy growth is seen about the point of puncture; this becomes about one millimetre thick and changes to a gray color, while a brown pigment is given off from its lower surface and also along the line of inoculation.

240. BACILLUS AUREUS (Adametz).

Found in water and also upon the surface of the body in cases of eczema seborrhœicum—by Unna and Tommasoli.

Morphology.—Slender bacilli, straight or slightly curved, from 1.5 to 4 μ long and 0.5 μ broad; often arranged in groups in which the bacilli lie parallel to each other; frequently in pairs or in long filaments.

Biological Characters.—An *aërobic, non-liquefying, slightly motile, chromogenic* bacillus. Forms a golden-yellow pigment. Spore production not observed. Grows slowly at the room temperature in the usual culture media. Upon *gelatin plates* forms small superficial colonies of irregular form, which at the end of eight days still appear as small white points; these later become pale-yellow and finally chrome-yellow in color; they are opaque, have well-defined outlines, and may attain a diameter of one to four millimetres; they vary in form, being round or elliptical, and later sometimes sausage-shaped. In *gelatin stick cultures* development occurs slowly upon the surface in the form of small, hemispherical colonies crowded together to form an irregular, dull-shining, dark chrome-yellow layer; very scanty growth occurs along the line of puncture. Upon *potato* broad, glistening, hemispherical colonies are formed, which gradually become confluent and form a dark chrome-yellow layer which in old cultures acquires a deep reddish-brown color.

241. BACILLUS FLAVOCORIACEUS (Eisenberg).

Synonym.—Sulphur-yellow bacillus (Adametz).

Found in water.

Morphology.—Very small bacilli.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* bacillus. Produces a sulphur-yellow pigment. Spore formation not observed. Grows at the room temperature. Upon *gelatin plates* forms small, round, sulphur-yellow colonies, which under a low power are seen to be coarsely granular and have a brownish-yellow centre surrounded by a pale-yellow marginal zone; older colonies are irregular in outline. In *gelatin stick cultures* a scanty, granular, grape-like growth occurs on the surface and along the line of puncture.

242. BACILLUS BEROLINENSIS INDICUS (Clässen).

Found in water—of the Spree.

Morphology.—Slender bacilli with round ends, resembling the typhoid bacillus in form and dimensions; usually solitary, sometimes united in pairs or in chains of three. The rods are surrounded by a delicate, transparent envelope.

Biological Characters.—An *aërobic, non-liquefying, insoluble, chromogenic* bacillus. Forms an indigo-blue pigment which is insoluble in water, alcohol, or chloroform, soluble in cold concentrated hydrochloric acid; is decolorized by ammonia. Spore formation not demonstrated. Grows rapidly at the room temperature in the usual culture media. Upon *gelatin plates*, at the end of three days, grayish-white colonies the size of a pin's head are developed; by the fourth day these commence to acquire an indigo-blue color, the deep colonies around the margin and the superficial colonies more in the centre; the uncolored margins of the superficial colonies are irregularly bulged and have a pearly lustre. In *gelatin stick cultures* a small, indigo-blue mass develops at the point of puncture at the end of twenty-

four hours; in the course of three or four days this becomes a regular wall, and then extends slowly over the surface of the gelatin, forming a layer with irregular outlines. Upon the *surface of agar* the growth is very characteristic; at the room temperature an indigo-blue streak is seen at the end of twenty-four hours, and in four or five days this becomes a tolerably thick layer, which gradually thins out at the edges and which has a deep indigo-blue color; the surface is covered up to within two or three millimetres of the margin with a thin, shining layer of pigment similar to that seen upon a saturated solution of gentian violet. Upon *potato*, at the end of three or four days, an abundant layer, thickest in the middle and with well-defined margins, is formed; this has a deep indigo-blue color, and also has a thin film of pigment on the surface when the potato has an acid reaction; when alkaline a thin, dirty-green layer is formed without the film of pigment on the surface.

243. BACILLUS CONSTRICTUS (Zimmermann).

Found in water.

Morphology.—Bacilli with round ends, from 1.5 to 6.5 μ in length and 0.75 μ thick; the rods are constricted at intervals so as to form two to six or more segments, which in stained preparations are separated by an unstained interval.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile*, *chromogenic* bacillus. Forms a Naples-yellow pigment. Spore formation not observed. Grows slowly at the room temperature in the usual culture media—no development at 30° C. and above. Upon *gelatin plates* forms, in four to five days, small, shining drops of a Naples-yellow color; the deep colonies under a low power are spherical, granular, and yellowish-gray in color; they have well-defined outlines, but the margins are irregularly dentate. In *gelatin stick cultures* development occurs along the line of puncture, and upon the surface a layer is developed which consists of irregular masses closely crowded together and which have a Naples-yellow color. Upon the *surface of agar* a tolerably thick, shining, Naples-yellow streak is developed along the *impfstrich*. Upon *potato* after a time a thin layer of a cadmium-yellow color is developed.

244. BACILLUS FLUORESCENS AUREUS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Short bacilli, about 1.9 μ long and 0.74 μ broad, usually in pairs and associated in longer or shorter groups; have long terminal flagella.

Biological Characters.—An *aërobic*, *non-liquefying*, *actively motile*, *chromogenic* bacillus. Forms an ochre-yellow pigment. Spore formation not observed. Grows in the usual culture media—best at the room temperature. Upon *gelatin plates* the deep colonies are small, yellowish-white, and spherical; the superficial colonies are glistening, yellowish-gray, and have ill-defined margins; under the microscope they are seen to be thickest in the middle and have irregular outlines, while the deep colonies are spherical, pale-yellow, and granular. In *gelatin stick cultures* a scanty development occurs along the line of puncture, which after a time acquires a brownish color; upon the surface a thin layer is developed, which later becomes thicker at the centre and extends to the walls of the tube; it has a yellow color. Upon the *surface of agar* an abundant ochrous or golden-yellow layer is developed. Upon *potato* the growth is similar but more scanty.

245. BACILLUS FLUORESCENS LONGUS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Straight or slightly curved bacilli, about 0.83 μ broad, and varying greatly in length—from 1.45 to 14 μ and more.

Biological Characters.—An *aërobic, non-liquefying, motile, chromogenic* bacillus. Forms a grayish-yellow pigment. Spore formation not positively determined, but unstained bodies resembling spores are seen in the rods. The short bacilli only are motile, the longer filaments not. Upon *gelatin plates* the deep colonies are small, spherical, and white with a greenish lustre; under a low power they are seen to be yellowish, sharply defined, and the interior is marked by contorted stripes. The superficial colonies are at first quite thin, nearly circular in form, and with a pearly lustre; they increase rapidly in size, and at the end of three days appear as yellowish-green patches which may be as much as nine millimetres in diameter; the margins are very thin and appear to be permeated with yellowish-white threads; under a low power the superficial colonies are seen to be striped by broad, convoluted bands resembling the intestine of a small animal. In *gelatin stick cultures* a very thin layer first forms, which later becomes thicker at the centre and soon reaches the walls of the test tube; at first it has a blue and later bluish-green fluorescence. Upon the *surface of agar* a rather thin layer is developed and the agar acquires a greenish-yellow color. Upon *potato* a thin, gray layer with a yellowish reflection and moist, shining surface extends over the surface.

246. BACILLUS FLUORESCENS TENUIS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Short, thick bacilli with round ends, from 1 to 1.85 μ long and about 0.8 μ thick; one or both ends are somewhat pointed; the rods are associated in irregular groups, and in old cultures often in chains containing four to six elements.

Biological Characters.—An *aërobic, non-liquefying, motile, chromogenic* bacillus. Forms a greenish-yellow, fluorescent pigment. Spore formation not determined. Grows rather rapidly in the usual culture media at the room temperature. Upon *gelatin plates* thin, shining superficial colonies are developed, which are irregularly circular in outline and have a radiate-cleft margin; the gelatin acquires a green color far beyond their boundary. In *gelatin stick cultures* a thin layer is formed upon the surface, which afterwards becomes thicker, of a grayish-white color, and extends to the walls of the test tube in about four days; the gelatin acquires a beautiful yellow color near the surface; by reflected light the surface growth is at first blue and later bluish-green; the line of puncture is marked by a scanty development which does not become any more pronounced. Upon the *surface of agar* a smooth, gray, shining, and rather scanty layer is developed; the agar gradually acquires a greenish color. Upon *potato* a thin, grayish-yellow, shining layer is developed along the impfstrich; later this acquires a reddish-brown color.

247. BACILLUS FLUORESCENS NON-LIQUEFACIENS.

Found in water.

Morphology.—Short, slender rods with round ends.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* bacillus. Forms a greenish-yellow, fluorescent pigment. Spore formation not observed. Grows in the usual culture media at the room temperature—not in the incubating oven. Upon *gelatin plates* forms fern-shaped superficial colonies, around which the gelatin has a pearly lustre. In *gelatin stick cultures* very scanty growth occurs along the line of puncture; the superficial growth has a fluorescent shimmer. Upon the *surface of agar* a layer is formed having a greenish color. Upon *potato* a rapidly developed, diffuse, brownish layer is formed, and the surface of the potato acquires a grayish-blue color.

248. *BACILLUS FLUORESCENS PUTIDUS* (Flügge).

Found in water.

Morphology.—Short bacilli with round ends.

Biological Characters.—An *aërobic, non-liquefying, actively motile, chromogenic* bacillus. Produces a greenish, fluorescent pigment. Does not form spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the superficial colonies are small, and under a low power they are seen to be finely granular discs with a dark centre surrounded by a yellow zone, and a pale-gray margin which is serpentine in outline; older colonies are dentate and have a greenish shimmer; the gelatin around acquires a fluorescent green color; an odor of trimethylamin is given off. In *gelatin stick cultures* development occurs upon the surface only, as a thin, dirty-white layer; in the course of a few days the gelatin commences to acquire a greenish, fluorescent color, most intense near the surface and gradually fading out below. Upon *potato* a thin, slimy, gray or brownish layer is developed.

249. *BACILLUS ERYTHROSPORUS* (Eidam).

Found in water, in putrefying flesh infusion, etc.

Morphology.—Slender bacilli with round ends; often grow out into short filaments.

Biological Characters.—An *aërobic, non-liquefying, motile, chromogenic* bacillus. Forms a greenish-yellow, fluorescent pigment. Forms oval spores—from two to eight in each filament. Grows at the room temperature in the usual culture media—not in the incubating oven. Upon *gelatin plates* forms whitish colonies, which spread out over the surface of the gelatin as a wrinkled and furrowed layer, around which the gelatin acquires a greenish-yellow, fluorescent color; under a low power the colonies are seen to have an opaque, brownish centre surrounded by a transparent, greenish-yellow marginal zone, an irregular outline, and the surface is marked by indistinct, radiating lines. In *gelatin stick cultures* an abundant growth occurs along the line of puncture and upon the surface; the gelatin throughout gradually acquires a fluorescent green color by transmitted, and a yellow color by reflected light. Upon *potato* a layer of limited dimensions is formed, which is at first reddish in color and later nut-brown.

250. *BACILLUS VIRIDIS PALLESCENS* (Frick).

Morphology.—Bacilli which are somewhat longer and more slender than the typhoid bacillus; form long filaments.

Biological Characters.—An *aërobic, non-liquefying, actively motile, chromogenic* bacillus. Produces, in presence of oxygen, a pale bluish-green pigment, which later becomes yellowish-brown with a green fluorescence. Grows best at the room temperature. Spore formation not observed. Upon *gelatin plates* the deep colonies are small and spherical; superficial colonies flat, with irregular, well-defined margins and a granular surface—similar to *Bacillus virescens*, but larger and of more rapid development; the colonies are green in color at first, but fade out to a pale bluish-green; they are frequently slightly iridescent. In *gelatin stick cultures* growth occurs upon the surface only, of a green color which quickly fades out to a bluish-green. Upon the *surface of agar* the growth is the same as on gelatin. Upon *potato* a nut-brown, moist layer is formed, and the potato around it acquires a dirty-violet color.

251. *BACILLUS VIRESCENS* (Frick).

Found in green sputum.

Morphology.—Bacilli of about the size of the typhoid bacillus, three to four times as long as broad; frequently grow out into long filaments.

Biological Characters.—An *aërobic, non-liquefying, actively motile, chromogenic* bacillus. Forms a green pigment, which after several weeks becomes yellowish-brown and then dark-green and fluorescent. Spore formation not observed. Grows best at the room temperature—rather slowly. Upon *gelatin plates* the deep colonies are small and spherical; the superficial colonies are flat, with irregular but well-defined margins and a finely granular surface; at the end of two to three days the colony and the surrounding gelatin have an intensely green color; later the colony becomes thicker, softer, and of a still deeper green color. In *gelatin stick cultures* growth occurs on the surface only; the gelatin acquires a decided green color. Upon *agar* an abundant development occurs upon the surface. Upon *potato* a nut-brown, moist layer is formed, and the potato around it acquires a dirty-violet color. In *bouillon* forms a layer upon the surface, which is with difficulty made to sink to the bottom, and not until it is broken up by shaking; the upper portion of the bouillon has a green color.

252. BACILLUS IRIS (Frick).

Morphology.—Very small, slender bacilli.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* bacillus. Produces a fluorescent green color, which later becomes yellowish-brown, and finally dark-green and fluorescent. Spore formation not observed. Grows best at the room temperature. Upon *gelatin plates* forms prominent, whitish, round, sharply defined superficial colonies with a smooth, shining surface; a green color is slowly developed. In *gelatin stick cultures* a prominent mass develops about the point of puncture; no growth along the line of puncture. Upon *potato* a dry, pale-brown layer is formed. In *bouillon* no layer is formed upon the surface, and the bouillon is not colored.

253. BACILLUS FUSCUS (Zimmermann).

Found in water.

Morphology.—Straight or curved bacilli with round ends, an irregular contour, here and there slightly bulging; about 0.63μ in diameter and of various lengths.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* bacillus. Forms a dark chrome-yellow pigment. Spore formation not observed. Grows slowly at the room temperature in the usual culture media—best at 30°C . Upon *gelatin plates* the deep colonies are punctiform and yellowish-brown in color; later they project from the surface in button form, and often have an irregular, knobby form; under the microscope such colonies are seen to be made up of smaller, spherical masses. The deep colonies, under a low power, are spherical or irregular in form, granular, and grayish-yellow or brownish-yellow in color; the superficial colonies have a brownish-yellow central portion surrounded by a highly refractive marginal zone. In *gelatin stick cultures* a button-like mass develops at the point of puncture; later the growth extends over the surface, forming a thick, wrinkled layer, at first pale-yellow and then chrome-yellow in color. The growth upon *agar* is similar to that upon gelatin. Upon *potato* a dark chrome-yellow, friable layer is developed.

254. BACILLUS RUBEFACIENS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli with round ends, from 0.75 to 1.65μ long and about 0.32μ thick; united in pairs, or in chains consisting of several elements.

Biological Characters.—An *aërobic, non-liquefying, actively motile, chromogenic* bacillus. Produces a pale-pink pigment. Grows best at the room temperature. Upon *gelatin plates* the deep colonies are spherical or lenticular in form, and white with a shade of yellowish-red; the superficial

colonies are flat, gray with a reddish tint; under the microscope the deep colonies are seen to be granular, spherical, and yellowish or brownish in color. In *gelatin stick cultures* development occurs along the line of puncture, and upon the surface a tolerably thick, grayish-white layer is formed, which later has a yellowish tint, while the gelatin around it has a bluish-white lustre. In old cultures the gelatin frequently acquires a pale wine color. Upon *agar* a smooth, rather thick, bluish-gray layer is formed. Upon *potato* an abundant, yellowish-gray layer is developed, which later has a reddish-brown color, while the potato around it after forty-eight hours acquires a pink color.

255. BACILLUS STRIATUS FLAVUS (Von Besser).

Found in nasal mucus—rare.

Morphology.—Small, thick rods, often curved, of about the dimensions of the diphtheria bacillus. In preparations stained with methylene blue, has a striated appearance. In old preparations various involution forms are seen.

Biological Characters.—An *aërobic, non-liquefying, chromogenic* bacillus. Forms a sulphur-yellow pigment. Grows at the room temperature in the usual culture media. Spore formation not observed. Upon *gelatin plates* forms thick, dry, granular colonies of a yellowish color. Upon *agar plates* projecting milk-white colonies of about 0.5 centimetre in diameter are developed; later these acquire a sulphur-yellow color. Upon the *surface of agar* a white, thick layer develops along the *impfstrich*; after some days this acquires a sulphur-yellow color. Upon *potato* a narrow, yellow stripe is developed along the *impfstrich*.

256. BACILLUS SUBFLAVUS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli with round ends, from 1.5 to 3 μ long and about 0.77 μ broad; united in chains of several elements.

Biological Characters.—An *aërobic, non-liquefying, motile, chromogenic* bacillus. Forms a pale-yellow pigment. Grows best at the room temperature. Spore formation not observed. Upon *gelatin plates* the deep colonies are small, yellowish-white, and spherical; these break through the surface and form hemispherical, yellowish-white, shining masses, which later extend over the surface and have an irregular outline; under a low power the surface is seen to have a pearly lustre; they gradually acquire a dirty-yellow color. In *gelatin stick cultures* a thin, yellowish-gray layer with finely dentate margins forms about the point of puncture. Upon the *surface of agar* a pale-yellow layer gradually extends over the entire surface; the color gradually becomes darker and is finally between a pale chrome-yellow and yellow ochre. Upon *potato* a scanty, dull, clay-yellow layer is formed.

257. BACILLUS CYANOGENUS (Hueppe).

Synonyms.—*Bacterium syncyanum*; *Bacillus lactis cyanogenus*; *Bacillus* of blue milk.

Found in milk.

Morphology.—Bacilli with slightly rounded corners, from 1.4 to 4 μ long and from 0.3 to 0.5 μ broad.

Biological Characters.—An *aërobic, non-liquefying, actively motile, chromogenic* bacillus. Produces a grayish-blue pigment. Forms spores, which are located at the extremities of the rods, giving them a club shape. Grows rapidly at the room temperature or in the incubating oven. Upon *gelatin plates*, at the end of two days, small, punctiform, grayish-white colonies are developed; the superficial colonies appear as slimy drops which

attain a diameter of one to two millimetres; they are finely granular, have a dirty-white color and circular, smooth outline; the gelatin around them acquires a steely grayish-blue shimmer. Under a low power the deep colonies are seen as round discs with a dark centre and a brownish, granular marginal zone with a sharply defined dark contour; the superficial colonies have a dark-brown centre surrounded by a grayish-brown, and outside of this a narrow, yellowish, finely granular marginal zone with a sharply defined contour. In *gelatin stick cultures* a white mass develops around the point of puncture, and around it the gelatin acquires a dark steel-blue color. Upon *agar* a grayish layer is developed and the culture medium acquires a dark-brown color near the surface. Upon *potato* a slimy, yellowish layer is developed, limited to the vicinity of the impfstrich; around this the potato acquires a diffused grayish-blue color. Upon *blood serum* development occurs without the formation of pigment. In *milk* a slightly alkaline reaction is produced by the growth of this bacillus; the casein is not coagulated; at the surface and gradually extending downward a slate-blue color is developed, and upon the addition of an acid this changes to an intense blue. In milk which has not been sterilized and which contains acid-forming bacteria a sky-blue color is produced without the addition of an acid. The pigment is produced most abundantly at a temperature of 15° to 18° C.; at 25° C. it is less abundant, and at 37° C. is not formed at all.

Note.—Jordan gives an account of *Bacillus cyanogenus* which differs materially from that above given by Flügge, and which leads to the belief that two or more different bacilli have been described under this name. Jordan's description agrees tolerably well, however, with that of Heim. The bacillus described by him was quite frequently found in Lawrence sewers and is described as follows:

"Morphology.—Small bacilli with rounded ends, often oval in form. Occur in chains in all media, isolated individuals being quite the exception. The chain is usually long and its members cohere quite firmly. On no medium has there been observed anything resembling spore formation. The individuals are about 1.3 μ long and 0.8 μ broad. *Motility:* There is a slight independent movement to be observed in hanging drops. We have certainly not found this species to be 'very motile.' *Temperature:* Does not grow as well at 37° as at 21° C. *Need of oxygen:* Grows very scantily under the mica plate. *Plate cultures:* The young colonies below the surface of the gelatin are usually slightly oval, with a coarsely granular interior and an even, regular edge. Often, however, the colonies have a frayed, irregular appearance to the naked eye, and with the microscope show fine branchings from the centre. On coming to surface the colonies always spread out into a dull, dry expansion with irregularly hacked edges. Sometimes the colonies are surrounded by a light blue-green haze, which soon changes to a faint brown; and this becomes deeper and deeper till the whole plate is colored an intense dark-brown. More often, however, in our experience, the brown color comes without a previous development of the blue. In slightly acid gelatin the blue color comes more surely and constantly than in the ordinary alkaline medium. The gelatin is not liquefied. *Tubes—gelatin:* In about three days there is a thin surface growth, smooth and faintly lustrous. The contour is at first quite regular, with the edges only slightly toothed. There is only a slight growth along the inoculation line. The gelatin near the surface soon takes on a brown tint, and eventually the whole tube of gelatin is colored dark-brown. The blue coloration is not observed as well in tubes as on plates. This species grows fairly well in acid gelatin, but not so well as in alkaline. In the former the brown color invariably comes more slowly. *Tubes—agar:* In three days there is a good surface growth, white and lustrous. The agar is colored dark-brown. The growth itself

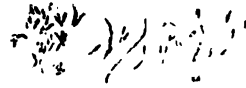


FIG. 210.—*Bacillus cyanogenus*; at a the rods contain spores. $\times 700$. (Flügge.)

also assumes a brownish cast. *Potato culture*: A very rapid growth on potato. In twenty-four hours the potato is colored a deep-brown color over nearly its whole surface. We have in no case observed a previous blue coloration. The growth is brownish, thin, spreading, and dry. *Milk*: The milk very slowly becomes a light-chocolate color. The reaction is slightly alkaline. *Bouillon*: In three days the bouillon has become slightly turbid. Month-old cultures are a deep brown, with a tough, thick skin on the surface. *Effect upon nitrates*: Nitrates are slowly reduced."

Jordan remarks: "This is undoubtedly the 'bacillus of blue milk' described originally by Hueppe."

258. BACILLUS FUSCUS LIMBATUS (Scheibenzuber).

Obtained from rotten eggs.

Morphology.—Short rods, occasionally united into filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *actively motile*, *chromogenic* bacillus. Produces a brown pigment. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates* colonies are developed in the form of small brown masses of circular outline and often surrounded by a paler marginal zone which is two to three times as broad as the brown interior. In *gelatin stick cultures* a branching growth is seen along the line of puncture, and the short branches are beset with small projections; the gelatin in the vicinity of the line of growth acquires a brown color. Upon the surface a scanty development occurs. Upon the *surface of agar* a superficial layer, beneath which the medium acquires a dark-brown color. Upon *potato* a brown layer is developed.

259. BACILLUS LATERICEUS (Eisenberg).

Synonym.—Ziegelroter bacillus (Adametz).

Found in water.

Morphology.—Bacilli, from three to five times as long as broad; grow out into short curved filaments.

Biological Characters.—An *aërobic*, *non-liquefying*, *non-motile*, *chromogenic* bacillus. Forms a brick-red pigment. Grows very slowly at the room temperature. Spore formation not observed. Upon *gelatin plates* forms punctiform, brick-red colonies; under a low power these are seen to be spherical, finely granular, and brownish-red in color; the opaque centre is surrounded by a more transparent marginal zone. In *gelatin stick cultures* a tolerably thick, slimy, brick-red layer is developed; very scanty growth along the line of puncture. Upon *potato* a brick-red layer.

260. BACILLUS SPINIFERUS (Unna).

Found upon the surface of the body in cases of *eczema seborrhœicum*.

Morphology.—Straight or slightly curved bacilli, $2\ \mu$ long and 0.8 to $1\ \mu$ broad; solitary or in pairs, frequently seen in irregular groups or lying parallel to each other in bundles.

Biological Characters.—An *aërobic*, *non-liquefying*, *chromogenic* bacillus. Forms a grayish-yellow pigment. Spore formation not observed. Grows rather slowly at the room temperature. Upon *gelatin plates*, at the end of eight days, the superficial colonies are prominent, shining, skin-colored, and circular in outline, from one to two millimetres in diameter; under a low power they are seen to be irregular in outline, covered with coarse projections; later finely granular, with a margin surrounded by small, thorn-like projections, which after a time form a radiating aureole and give the colony a porcupine-like appearance. In *gelatin stick cultures*, at the end of six days, an irregular, wrinkled layer about two millimetres broad is formed upon the surface; this is grayish-yellow upon the folds and bluish

in the furrows; as this extends the folds become more prominent and more decidedly yellow in color, the margins are irregular, dentate, and thin; along the line of inoculation very small, yellowish, punctiform colonies are developed. Upon the *surface of agar* the growth is similar to that upon gelatin. Upon *potato* development is very slow; first as a glistening, yellowish-white line along the *impfstrich*; at the end of three weeks here and there along this line small, grayish-yellow, glistening masses are seen.

261. BACILLUS RUBESCENS (Jordan).

Found in sewage at Lawrence, Mass.

Morphology.—Bacilli with round ends, about $4\ \mu$ long and $0.9\ \mu$ broad, often slightly curved; solitary, in pairs, or in short chains.

Biological Characters.—An *aërobic, non-liquefying, motile, chromogenic* bacillus. Forms a pale-pink pigment. Spore formation not observed. Movements slow. Grows best at the room temperature. Upon *gelatin plates* the deep colonies are spherical or oval; on coming to the surface they form a projecting, porcelain-white drop; they slowly increase in size, and after a time have a slight brownish color. In *gelatin stick cultures* a prominent porcelain-white mass is formed upon the surface at the point of inoculation; very scanty growth along line of puncture; grows in slightly acid gelatin. Upon the *surface of agar* a white and lustrous layer is quickly developed, which is at first smooth, but later becomes wrinkled; in cultures about three weeks old a slight pinkish tinge is seen. Upon *potato* the growth is rapid and abundant; it is at first light-brown in color, and this gradually changes to pink; at the end of three weeks there is a projecting growth which has a delicate flesh-pink color; the potato itself is not colored. *Milk* is not coagulated and has an alkaline reaction; in old cultures a slight pinkish tinge is observed near the surface. *Bouillon* becomes slightly turbid and a heavy white precipitate is formed. At the end of several weeks a thick, tenacious film forms upon the surface.

262. BACILLUS ALLII (Griffiths).

Synonym.—*Bacterium allii*.

Found upon the surface of putrefying onions.

Morphology.—Bacilli with round ends, from 5 to $7\ \mu$ long and about $2.5\ \mu$ broad; solitary or in pairs; form *zoöglœa*.

Biological Characters.—An *aërobic, chromogenic* bacillus. Produces a green pigment which is soluble in alcohol. Grows tolerably well on nutrient agar, and produces a bright green pellicle on the surface of this medium. Sulphuretted hydrogen is given off by the cultures in small quantities.

B. Chromogenic, Liquefying Bacilli.

263. BACILLUS FULVUS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Short bacilli with round ends, from 0.88 to $1.3\ \mu$ in length and $0.77\ \mu$ broad; solitary, in pairs, or in chains containing several elements.

Biological Characters.—An *aërobic, liquefying, non-motile, chromogenic* bacillus. Grows slowly at the room temperature—best at 30°C . Spore formation not observed. Forms a gamboge-yellow-colored pigment. Upon *gelatin plates* the deep colonies are irregularly spherical, oval, or elliptical, granular, and yellowish-gray in color; they are usually surrounded by pale-yellow patches. The superficial colonies are reddish-yellow, drop-like, and at the end of eight days measure about one millimetre in diameter. In *gelatin stick cultures* a prominent, arched mass of irregularly circular contour and of the color of gamboge is formed about the point of puncture; a scanty

growth occurs along the line of inoculation, which has a slightly yellowish color; liquefaction commences after some weeks. Upon the *surface of agar* an abundant, glistening layer of a gamboge-yellow color. Upon *potato* development occurs slowly along the *impfstrich*; the growth has at first an india-yellow color, which later becomes ochre yellow.

264. *BACILLUS HELVOLUS* (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli from 1.5 to 4.5 μ in length and about 0.5 μ broad; usually in pairs, but also in chains of four or more elements and in long filaments.

Biological Characters.—An *aërobic, liquefying, motile* (rotary motion only), *chromogenic* bacillus. Forms a Naples-yellow pigment. Spore formation not observed. Grows rather rapidly at the room temperature. Upon *gelatin plates* the deep colonies are small, spherical, and of a pale-yellow color; when they come to the surface they form a prominent pale-yellow drop, which lies in a shallow cavity. The superficial colonies for some time have a sharply defined contour and an irregular outline; by transmitted light they have a dark-brown color. In *gelatin stick cultures* a button-like mass of a Naples-yellow color forms about the point of puncture; this gradually extends until it nearly reaches the walls of the test tube; in the meantime the gelatin below is slowly liquefied and a saucer-shaped depression is formed; liquefaction progresses very slowly. Upon the *surface of agar* an abundant layer of a Naples-yellow color is developed. Upon *potato* a tolerably thick and abundant, shining, yellow layer, which frequently has a shade of green.

265. *BACILLUS OCHRACEUS* (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli with round ends, from 1.25 to 4.5 μ long and from 0.65 to 0.75 μ broad; usually in pairs, or in chains containing several elements, also in long jointed filaments. In stained preparations a capsule-like envelope may often be seen.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Forms an ochre-yellow pigment. Spore formation not observed. Motions slow and serpentine. Grows rather slowly—best at the room temperature. Upon *gelatin plates* forms small, pale-yellow, spherical colonies, which gradually increase in size and cause the gelatin above them to become liquefied, so that they lie at the bottom of a saucer-shaped cavity; here the color is more intense and becomes a golden ochrous yellow; later the yellow mass extends in diameter, and is of a paler yellow color at the margin than at the centre. Under a low power the colonies are seen at first as granular, yellowish-brown discs; later they are covered with wart-like projections. In *gelatin stick cultures* liquefaction occurs at first in funnel form, and when it extends to the walls of the tube it gradually progresses downward; a pale-yellow deposit is seen at the bottom of the liquefied gelatin, which later acquires an ochrous-yellow tint. Upon the *surface of agar* an ochrous-yellow layer is formed, which at the end of four to six weeks has a breadth of about six millimetres at the lower portion of the *impfstich*. Upon *potato* a thin, ochrous-yellow layer is developed.

266. *BACILLUS PLICATUS* (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Small, thin bacilli with rounded ends, about 0.48 μ (?) in length and 0.45 μ broad; usually in pairs, but also in chains of four or more elements.

Biological Characters.—An *aërobic, liquefying, non-motile, chromogenic* bacillus. Forms a grayish-yellow pigment. Spore formation on

observed. Grows best at the room temperature. Upon *gelatin plates* small, yellowish-white colonies of irregular form are developed beneath the surface, which under a low power are seen to present hemispherical projections; when they break through the surface they have a mulberry-like appearance and yellowish-white color. In *gelatin stick cultures* a wrinkled, yellowish-white layer with irregular contour is developed; at the end of one or two weeks it is depressed and the gelatin gradually undergoes liquefaction; an abundant development of small, granular, yellowish-white colonies occurs along the line of puncture. Upon *potato* a thin layer is formed, which soon becomes dry and friable and has a grayish-yellow color.

267. BACILLUS JANTHINUS (Zopf).

Synonym.—Violet bacillus.

Found in water and in sewage (at Lawrence, Mass.).

Morphology.—Very small, slender bacilli, often associated in short filaments; about $2\ \mu$ long and 0.5 to $0.6\ \mu$ broad.

Biological Characters.—An *aërobic*, *liquefying*, *motile*, *chromogenic* bacillus. Forms a bluish-violet pigment. Spore formation not observed. Grows best at the room temperature. Upon *gelatin plates* the deep colonies are spherical or oval, with an even contour; upon coming to the surface they spread out as a broad, irregular expansion with deeply notched edges; these superficial colonies are at first thin, but later increase in thickness; a deep-violet color soon appears, sometimes near the centre of the colony, sometimes around its edges; the gelatin is very slowly liquefied. In *gelatin stick cultures* a scanty growth, without color, is developed along the line of puncture, and a thin, violet-colored layer upon the surface. The gelatin is slowly liquefied, and an abundant violet-colored precipitate is seen at the bottom of the liquefied gelatin in old cultures. Upon the *surface of agar* a rather tough and coherent layer is developed, which has a dark-violet color. Upon *potato* the development is rapid and covers the entire surface; the growth frequently has a beaded appearance, and the membranous growth is with difficulty detached from the surface; it has a black-violet color. In *milk* an abundant development occurs without producing coagulation, and causing it to acquire a violet color. Reduces nitrate to nitrite very rapidly and completely. (Above characters given by Jordan.)

268. BACILLUS VIOLACEUS LAURENTIUS (Jordan).

"Found in large numbers in the effluent of Tank 1" at Lawrence, Mass.

Morphology.—Bacilli with round ends, from 3 to $3.6\ \mu$ long and $0.7\ \mu$ broad; often in pairs, and sometimes in chains of four or five elements.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *actively motile* bacillus. Produces a dark-violet pigment. Spore formation not observed. Grows best at the room temperature. Upon *gelatin plates* the deep colonies, at the end of two days, are small, spherical, and coarsely granular; they usually have a radiating margin and a dark centre; upon coming to the surface a thin, irregular expansion occurs and the gelatin is quickly liquefied in the vicinity of the colony; at the centre a violet-colored spot is seen, and around this a zone of slightly clouded liquid gelatin; liquefaction progresses rapidly, but the central violet mass does not materially increase in size. In *gelatin stick cultures* liquefaction occurs rapidly along the line of inoculation; the liquefied gelatin is clouded and of a violet color, and an abundant dark-violet precipitate accumulates at the bottom of the tube. Upon *agar* an abundant development occurs, which at first has a dark-violet color and later becomes jet-black. Upon *potato* an abundant growth of a dark-violet color spreads over the entire surface; this soon becomes jet-black, except near the edge. In *milk* the bacillus develops rapidly and abundantly, causing rapid coagulation of the casein and an acid reaction; the milk acquires a deep blue-violet color. In ordinary *bouillon* the

violet color is not developed, but when nitrates are added the growth is more luxuriant and a rich violet color is produced. Nitrates are reduced to nitrites by this bacillus, rather slowly.

269. BACILLUS TREMELLOIDES (Schottelius).

Found in the Freiburg water supply by Tils.

Morphology.—Bacilli with round ends, from 0.75 to $1\ \mu$ long and $0.25\ \mu$ broad, associated in friable masses.

Biological Characters.—An *aërobic, liquefying, chromogenic* bacillus. Forms a golden-yellow pigment. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies appear as yellow points; at the end of forty-eight hours the superficial colonies appear as prominent yellow masses and the plate looks as if it were covered with coarse sand; at the end of a few days the colonies measure several millimetres in diameter and do not subsequently increase in size; under a low power these are seen to be made up of cloud-like masses and have a yellow or yellowish-brown color, the contour is smooth or irregularly bulging; at the end of eight days the margin is surrounded by a golden-yellow, slimy layer and the colony commences slowly to sink into the gelatin. In *gelatin stick cultures* isolated, punctiform, yellow colonies are developed along the line of inoculation, and upon the surface a colony which resembles those upon gelatin plates; at the end of ten to fourteen days the superficial layer is slowly depressed as a result of liquefaction of the underlying gelatin. Upon the *surface of agar* a layer is developed which is at first dry and granular, later slimy and of a golden-yellow color. Upon *potato* a yellow layer is developed which may attain a thickness of several millimetres; after a long time the growth is surrounded by a golden-yellow, slimy zone, and no further extension occurs upon the surface of the potato. In *milk*, at the end of thirty-six hours, a strongly acid reaction is produced and the fluid becomes viscid.

270. BACILLUS CUTICULARIS (Tils).

Found in water.

Morphology.—Bacilli from 2 to $3\ \mu$ long and 0.3 to $0.5\ \mu$ broad; may grow out into short filaments.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Spore formation not observed. Forms a yellow pigment. The shorter rods are slightly motile, the longer filaments not. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies appear under the microscope as brownish discs with an irregular but smooth contour. The superficial colonies are yellowish-brown in color, with well-defined contour; after several days the centre commences to be depressed and the gelatin is quickly liquefied; the colony then consists of a membranous film surrounded by a well-defined zone of liquefied gelatin; finally the gelatin of the plate is entirely liquefied and these membranous colonies float upon the surface. In *gelatin stick cultures*, at the end of two days, liquefaction commences near the surface and progresses rapidly; a membranous layer is formed on the surface. Upon *potato* development is slow, in the form of a pale-yellow layer, which later becomes slimy and dark-yellow. In *milk* a pale-yellow, membranous layer is formed upon the surface in from twenty-four to thirty-six hours, and an odor of sulphuretted hydrogen is developed.

271. FLESH-COLORED BACILLUS (Tils).

Found in water.

Morphology.—Bacilli of $2\ \mu$ in length and $0.5\ \mu$ broad; in hanging-drop cultures always seen solitary and in active motion.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Forms a dark flesh-colored pigment. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates*, at the end of two days, circular cavities with well-defined margins and filled with liquefied gelatin are seen; under a low power the centre of the colonies is seen to be more opaque and is surrounded by concentric rings, alternately of lighter and darker color, while the marginal zone is colorless and appears finely granular. In *gelatin stick cultures* liquefaction occurs rapidly along the line of puncture, in funnel shape; at the lower part of the funnel a deposit of a pale-pink color accumulates and the liquefaction ceases to extend. Upon the *surface of agar* it grows rapidly and forms a thick, slimy, pale-pink layer. Upon *potato* a layer is formed which is first pale and later dark flesh-colored.

272. BACILLUS ARBORESCENS (Frankland).

Found in the London water supply.

Morphology.—Bacilli with round ends, about 2.5μ long and 0.5μ broad; often united in pairs, or in chains of three or four elements; also form long, flexible filaments.

Biological Characters.—An *aërobic, liquefying, chromogenic* bacillus. Spore formation not observed. Oscillating movements only. Grows in the usual culture media at the room temperature.

Upon *gelatin plates* the colonies, at the end of twenty-four hours, consist of a thin axial trunk with root-like offshoots at both ends; later the body becomes thicker and the branching extremities are so strongly developed that the whole has the appearance of a sheaf of wheat; the naked-eye appearance, in this stage, is also peculiar, and the colony is seen as an iridescent bundle, constricted in the middle and with the ends striped in a radial direction; later the gelatin is



FIG. 211.—*Bacillus arborescens*, from a gelatin culture. $\times 1,000$. (Frankland.)

liquefied slowly and the central part of the colony acquires a yellow color, while the periphery is beautifully iridescent. In *gelatin stick cultures*, by the second day, an iridescent layer is seen on the surface; in the middle of this the gelatin is slightly depressed and filled with a semi-fluid, yellowish mass; along the line of puncture a transparent, grayish cloudiness is seen; liquefaction progresses slowly at the surface, and a funnel is formed, at the bottom of which the yellow deposit gradually increases; along the line of puncture no further changes occur. Upon the *surface of agar* a rather thin, dirty orange-colored layer is formed, the margins of which are slightly iridescent and striped in a radial direction. Upon *potato* a thick, glistening stripe of a deep orange-red color is formed along the line of inoculation; the surface of this is covered with irregular protuberances.



FIG. 212.—Colony of *Bacillus arborescens*. $\times 100$. (Frankland.)

273. BACILLUS CITREUS CADAVERIS (Strassmann).

Found in a cadaver fifty hours after death from accidental shooting—in blood from a vein.

Morphology.—Oval bacilli, 0.9μ long and 0.6μ broad, usually united in chains.

Biological Characters.—An *aërobic, liquefying, non-motile, chromogenic* bacillus. Forms a yellow pigment. Spore formation not observed. Grows slowly at the room temperature. Upon *gelatin plates* forms small, pale-

yellow colonies, around which the gelatin is liquefied in circular form. In *gelatin stick cultures* liquefaction occurs at the surface, with formation of an air bubble at the surface, a scanty, yellow deposit at the bottom of this, clear liquefied gelatin below, and a yellow deposit on the concave floor of the liquefied gelatin; below this, along the line of puncture, small colonies are seen. The cultures give off an odor of sulphuretted hydrogen. Upon the *surface of agar* a narrow, yellow layer forms along the *impfstrich*. Upon *potato* a dry, lemon-yellow, slightly granular layer is developed.

274. *BACILLUS MEMBRANACEUS* AMETHYSTINUS (Eisenberg).

Found in a specimen of well water from Spalato.

Morphology.—Short bacilli with round ends, from 1 to 1.4 μ long and 0.5 to 0.8 μ broad; in irregular groups; some of the rods in stained preparations show end staining.

Biological Characters.—An *aërobic, liquefying, non-motile, chromogenic* bacillus. Produces a dark-violet pigment which has a metallic lustre. Spore formation not determined. Grows at the room temperature in the usual culture media—no development at 37.5° C. Upon *gelatin plates*, at the end of two to three days, colonies are developed the size of a poppy-seed, which have a homogeneous, dark-violet color; later a ring of liquefied gelatin forms around each colony, which gradually extends, and at the end of one to two weeks the entire gelatin is liquefied and the colonies are seen floating upon its surface, not much larger than when liquefaction commenced. When the colonies are not so closely crowded superficial colonies are seen at the end of three to four days, which resemble those of the typhoid bacillus; they have a yellowish-white color and a dentate margin; at the end of ten to fourteen days the gelatin around these colonies commences to soften and a dark-violet color extends from the centre to the periphery; at the end of three to four weeks the colonies are seen floating upon the surface of the scarcely liquefied gelatin, as large violet layers. In *gelatin stick cultures* a yellowish-white layer with dentate borders develops upon the surface, which at the end of ten to fourteen days begins to acquire a violet color at the centre, which gradually extends to the periphery; liquefaction occurs slowly, so that at the end of about four weeks a thick, violet-colored layer rests upon the softened and depressed surface of the culture medium; at the end of three to four months the gelatin in the tube is completely liquefied. Upon the *surface of agar* a yellowish-white, milky, thick layer is formed, which commences to acquire a violet color at the end of eight to ten days, and at the end of three to four weeks is seen as a wrinkled, dark-violet layer with a metallic lustre, which is easily lifted entire from the surface of the culture medium. Upon *potato* a dirty-yellow or olive-green layer is slowly formed and gradually extends from the line of inoculation. In *bouillon* development is very slow; at the end of several weeks the dark-brown *bouillon* is seen to have a violet film upon the surface and a deposit of the same color at the bottom of the tube.

275. *ASCOBACILLUS CITREUS* (Unna and Tommasoli).

Found upon the surface of the body of individuals with eczema seborrhœicum.

Morphology.—Straight or curved bacilli, 1.3 μ long and 0.3 μ broad, solitary, in pairs, or in irregular groups.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Produces a lemon-yellow pigment. Spore formation not observed. Grows in the usual media at the room temperature—slowly in gelatin, rapidly upon agar and potato. Upon *gelatin plates*, at the end of two weeks, prominent, opaque, yellow, punctiform colonies are developed upon the surface, while the deep colonies are scarcely visible with the naked eye; under a low power they are seen to be grayish-yellow, opaque, sometimes like little

drops of oil and sometimes a conglomeration of minute spherical masses. Later the deep colonies are oval, dark, sharply defined, and as large as a pea; those nearer the surface are conglomerate; those upon the surface are partly homogeneous, pale-yellow, and round; some show an opaque, conglomerate mass in the centre, with a more transparent, yellowish-green marginal zone; some have a form resembling that of Saturn with its rings. In *gelatin stick cultures* a slimy, thick, lemon-yellow layer develops upon the surface; this is gradually depressed in the middle, while the margins remain elevated and granular; along the line of puncture small colonies are developed which form a funnel above; at the end of five to six weeks the gelatin in the funnel, when shaken, appears pap-like, and the layer floats upon its surface, while some greenish flocculi are seen below. Upon the *surface of agar* development is rapid and the entire surface is covered within a few days; below, the layer consists of an abundant jelly-like growth, covered with protuberances resembling drops of honey; above, it has an orange color and creamy consistence, and is covered with numerous small, spherical or oval masses. Upon *potato* a slimy, lemon-yellow layer is quickly developed and extends over the entire surface; at the margins this is more transparent and albuminous in appearance; at the end of two weeks the central portion has a greenish-yellow color, distributed like the veins of a grape leaf, with smaller, pale-yellow veins.

276. BACILLUS CŒRULEUS (Smith).

Found in water of the Schuylkill River.

Morphology.—Bacilli from 2 to 2.5 μ long and 0.5 μ broad, frequently associated in chains.

Biological Characters.—An *aërobic, liquefying, chromogenic* bacillus. Produces a beautiful blue pigment. Spore formation not observed. Grows slowly at the room temperature. Upon gelatin plates forms superficial colonies having a blue color, around which the gelatin is liquefied. In *gelatin stick cultures* cup-shaped liquefaction occurs at the surface and a blue color is developed, while below a scanty, colorless growth occurs along the line of puncture. Upon the *surface of agar* a bluish layer is formed. Upon *potato* at first a beautiful dark-blue layer, which later acquires an intense blue-black color.

277. BACILLUS FLUORESCENS LIQUEFACIENS.

Found in water and in various putrefying infusions—very common.

Morphology.—Short bacilli, in pairs with a constriction at the point of junction.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Forms a greenish-yellow, fluorescent pigment. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* whitish colonies are developed upon the surface, which may attain a diameter of three millimetres; a ring of liquefied gelatin forms around each; under a low power the colonies are seen to have a sharp contour and irregularly circular outline, a dark-brown, finely granular centre surrounded by a finely granular marginal zone of a yellow color, which becomes more transparent and grayish-white toward the edge; the gelatin gradually acquires a greenish tint. In *gelatin stick cultures* a whitish growth occurs along the line of puncture; a small funnel of liquefied gelatin is first seen near the surface, and this has an air bubble above; gradually the liquefaction extends to the walls of the test tube and also in a downward direction, forming a superficial layer of liquefied gelatin, upon the floor of which a thick white deposit is formed; the gelatin below this has a greenish-yellow fluorescence and the liquefied gelatin also, although less pronounced. Upon *potato* an abundant brownish layer is developed along the line of inoculation.

278. *BACILLUS FLUORESCENS LIQUEFACIENS MINUTISSIMUS*
(Unna and Tommasoli).

Found upon the surface of the body in cases of eczema seborrhœicum. Possibly identical with the previously described species.

Morphology.—Bacilli with round ends, usually constricted in the middle, from 1.5 to 2 μ long and 0.3 μ broad; often united to form filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile, chromogenic* bacillus. Forms a greenish-yellow, slightly fluorescent pigment. Forms spherical spores. Grows rapidly in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of three to four days, the colonies consist of an outer yellowish-white zone of transparent, liquefied gelatin and a thick, pale-brown centre which is made up of grayish granular material. In *gelatin stick cultures*, at the end of three days, a broad funnel of liquefied gelatin is formed above, which is about one centimetre deep and five millimetres in diameter at the surface; the liquefied gelatin is clouded, greenish-yellow, and contains some whitish flocculi, while a thick whitish deposit accumulates at the bottom; at the end of eight days the gelatin is entirely liquefied and a thick, opaque, grayish-yellow, fluorescent layer floats upon the surface. Upon the *surface of agar* a slimy, moist, smooth, pale-brown layer is developed. Upon *potato* a broad, compact, flat layer is quickly developed; this has a pale-brown color and elevated, sharply defined margins; the potato acquires a dark color.

279. *BACILLUS FLUORESCENS NIVALIS* (Schmolck).

Found in ice water and snow from Norwegian glaciers. (Probably identical with No. 277.)

Morphology.—Short bacilli, often united in chains.

Biological Characters.—An *aërobic liquefying, motile, chromogenic* bacillus. Forms a bluish-green, fluorescent pigment. Spore formation not observed. Upon *gelatin plates* whitish, punctiform colonies are formed, which spread out upon the surface as round discs and cause liquefaction of the surrounding gelatin; the non-liquefied gelatin in the vicinity acquires a bluish-green fluorescence. In *gelatin stick cultures* liquefaction occurs in funnel form, and the liquefied gelatin acquires a greenish fluorescence like that of *Bacillus liquefaciens fluorescens*. Upon the *surface of agar* a whitish layer is formed and the culture medium acquires a fluorescent color. Upon *potato* a brownish layer is developed.

280. *BACILLUS LACTIS ERYTHROGENES* (Hueppe).

Synonym.—Bacillus of red milk.

Found in red milk, and by Baginsky in the fæces of a child.

Morphology.—Short bacilli with round ends, from 1 to 1.4 μ long and from 0.3 to 0.5 μ thick; in bouillon cultures may grow out into short filaments.

Biological Characters.—An *aërobic, liquefying, non-motile, chromogenic* bacillus. Produces a yellow pigment which is destroyed by acids and is developed either in the presence or absence of light; and a red pigment which is absorbed by the culture medium and is produced most abundantly in an alkaline or neutral medium in the absence of light. Does not form spores. The cultures give off an intense and disagreeable odor. Grows at the room temperature in the usual culture media—more rapidly at 28° to 35° C. Upon *gelatin plates* small, spherical colonies are developed, which are at first grayish-white and later yellow in color; after a time the surrounding gelatin is liquefied in saucer shape and acquires a pale-pink color. In *gelatin stick cultures* a rather thin, round layer is developed upon the surface, which is at first whitish and later yellow in color; the gelatin

around it acquires a pale-pink color, and after a time liquefaction occurs; along the line of puncture the development is scanty; at the end of ten to twelve days a slightly clouded, pink liquid is seen at the upper part of the test tube, in which well-defined yellow colonies are suspended, while the unliquefied gelatin below has a pink color. Upon the *surface of agar* a glistening, yellowish layer is slowly developed. Upon *potato* development is more rapid and an extended layer is formed, which is first grayish-white and later yellow in color; the potato acquires a dark color which later becomes yellowish-red; at 37° C., at the end of six to eight days, an intense golden-yellow color is developed. In *bouillon* development is rapid and yellowish cloudiness of the culture medium is seen. In *milk* the casein is slowly precipitated and later is peptonized, with a neutral or alkaline reaction of the medium; a stratum of blood-red serum is seen above the precipitated casein, and above this a yellowish-white layer of cream.

281. BACILLUS GLAUCUS (Maschek).

Found in water.

Morphology.—Slender bacilli of various lengths.

Biological Characters.—An *aërobic, liquefying, non-motile, chromogenic* bacillus. Produces a gray pigment. Spore formation not observed. Grows rapidly in the usual culture media at the room temperature. Upon *gelatin plates* forms round, gray colonies with sharply defined outlines; on the fourth day the centre becomes intensely gray, the margin brown and folded in a radial direction; on the eighth day liquefaction has occurred and the colony sinks beneath the surface. In *gelatin stick cultures* development is rapid both upon the surface and along the line of puncture; the entire gelatin is quickly liquefied and a gray deposit is seen at the bottom of the tube. Upon the *surface of agar* a gray layer is quickly developed. Upon *potato* the growth is at first of a dirty-white; later a slimy, dark-gray layer is formed.

282. BACILLUS LIVIDUS (Plagge and Proskauer).

Found in the Berlin water supply.

Morphology.—Slender bacilli of medium size.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, motile, chromogenic* bacillus. Produces an intense blue-black pigment. Spore formation not observed. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* the colonies at first resemble drops of ink; the gelatin around them is slowly liquefied and a bluish-violet deposit is seen at the bottom of the liquefied gelatin. In *gelatin stick cultures* a colorless growth is seen along the line of puncture and a violet layer upon the surface; liquefaction occurs very slowly. Upon the *surface of agar* a beautiful blue-black layer is developed. Upon *potato* an abundant layer of a violet color is formed along the line of inoculation.

283. BACILLUS INDICUS (Koch).

Found in the contents of the intestine of a monkey, by Koch, while pursuing his cholera investigations in India.

Morphology.—A short and slender bacillus with round ends.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, motile, chromogenic* bacillus. Spore formation not observed. Produces a brick-red pigment. Grows rapidly in the usual culture media—best in the incubating oven. Upon *gelatin plates* the deep colonies are white and punctiform; under a low power they are seen to be granular, irregular in form, and of a greenish-brown color. The superficial colonies quickly cause liquefaction of the gelatin and form circular depressions with a well-defined outline and grayish contents, which under the microscope appear as dense, finely granular, grayish-yellow masses, the edges of which appear to be

fringed with short fibres; later the gelatin acquires a pale-red color, which gradually becomes more intense. In *gelatin stick cultures* liquefaction is rapid along the entire line of inoculation; a wrinkled, red film forms upon the surface and grayish-white flocculi accumulate at the bottom of the liquefied medium. Upon the *surface of agar* an abundant layer, covering the entire surface, is quickly developed in the incubator; this usually acquires a brick-red color, but the margins of the layer, or even the entire growth, may remain colorless, especially in cultures kept in the incubating oven. Upon *potato* development occurs very rapidly along the line of inoculation; this soon acquires the characteristic color. *Blood serum* is liquefied by this bacillus. Large quantities (twenty cubic centimetres) of a pure culture injected into a vein or into the peritoneal cavity of a rabbit or guinea-pig cause the death of the animal in from three to twenty hours; at the autopsy an intense gastro-enteritis is found.

284. BACILLUS PRODIGIOSUS.

Synonyms.—*Micrococcus prodigiosus*; *Monas prodigiosa*.

This bacillus has long been known, having attracted attention because of the blood-red stains which it causes upon farinaceous substances, such as boiled potatoes, moist bread, etc. It was described by Ehrenberg under the name of *Monas prodigiosa*. At times, in certain parts of Europe, it has been exceptionally abundant, and the bloody-looking patches produced by its rapid development upon favorable media have been regarded with apprehension by the superstitious.

Morphology.—Short bacilli with rounded ends, which are sometimes so short as to be scarcely distinguishable from micrococci, but also occur as rod-shaped cells and short filaments; frequently in pairs and occasionally in chains containing ten or more elements—especially in acid media.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, usually *non-motile*, *chromogenic* bacillus. Although usually described as non-motile, this bacillus is said under certain circumstances to be capable of spontaneous movements. According to Schottelius, these are best seen when it is cultivated in strongly diluted liquid media and under unfavorable conditions of growth. Forms a red pigment which is soluble in alcohol and ether, but not in water; this is only formed in presence of oxygen; it is changed to a pale-red color by the action of acids, and the deep-red color is restored by ammonia and other strong alkalies. The pigment is not seen in the interior of the bacterial cells, but the chromogenic substance formed by them develops the color outside of the cells, where it is seen in the form of granules. The formation of pigment is influenced not only by the presence of oxygen, which is essential to its production, but also by conditions relating to temperature, constitution of the culture medium, etc. By continuous cultivation in the incubating oven a non-chromogenic variety may be obtained, and the same result is obtained by continuous cultivation in acid bouillon. But under favorable conditions color production again returns after a few successive transplantations upon potato or nutrient agar, if the cultures are kept at the room temperature and freely exposed to the air. Spore formation has not been observed, but this bacillus retains its vitality for a long time in a desiccated condition. Cultures give off a strong odor of trimethylamin; and in culture media containing sugar, fermentation occurs with production of alcohol and carbon dioxide. This bacillus grows rapidly in the ordinary culture media—best at the room temperature or a little above (25° C.). Upon *gelatin plates* small, white, punctiform colonies are developed below the surface, and upon the surface round, granular colonies which quickly cause liquefaction of the gelatin; saucer-like depressions are produced, at the bottom of which the colony is seen as a whitish mass which later acquires a deep-red color, first appearing at the centre. In *gelatin stick cultures* liquefaction quickly occurs along the entire line of inoculation and rapidly extends until the medium is completely liquefied; pigment for-

mation occurs at the surface only, but after a time the entire medium is colored red by the deposition of granular, colored masses from the surface growth. Upon the *surface of agar* an abundant purplish-red layer is formed, but the color is not absorbed by the culture medium. Upon *potato* very rapid and abundant development occurs at the room temperature, forming a thick, purplish-red layer which after some days has the color of undissolved fuchsin and a metallic lustre. *Blood serum* is liquefied by this bacillus. In *milk* the development of *Bacillus prodigiosus* causes a precipitation of the casein and a deep-red color of the medium. When cultivated for some time in acid media the peptonizing (liquefying) power of the bacillus is greatly reduced, as well as its chromogenic power. It has been shown by Roger that animals which are not susceptible to the disease known as malignant oedema, become infected and die when inoculated with the malignant-oedema bacillus and at the same time with one or two cubic centimetres of a culture of *Bacillus prodigiosus*. But this bacillus alone has no decided pathogenic power. An interesting discovery made by Pawlowsky is the fact that when rabbits are inoculated simultaneously with a virulent culture of the anthrax bacillus and with a culture of *Bacillus prodigiosus* they recover from the inoculation, the chemical products of one bacillus having apparently the power to neutralize the toxic substances to which the other owes its pathogenic potency.

285. BACILLUS MESENERICUS RUBER.

Synonym.—Rothen Kartoffelbacillus (Globig).

Found upon potatoes.

Morphology.—Slender bacilli with round ends, united in pairs, in chains of four, or in long filaments.

Biological Characters.—An *aërobic*, *liquefying*, *motile*, *chromogenic* bacillus. Produces a reddish-yellow or pink pigment. Forms oval spores which have a great resistance against high temperatures and germicidal agents. Grows rapidly, especially at a temperature of 45° C.; also at the room temperature in the usual culture media. Upon *gelatin plates*, at 15° to 20° C., at the end of two days deep colonies are formed which are spherical and of a yellow color; when these come to the surface they spread out as a fine network, around the margins of which projecting points are seen; liquefaction commences on the fourth day and this network vanishes, leaving a grayish-brown, friable mass at the bottom of the liquefied medium. In *gelatin stick cultures*, at the end of three or four days, a cloudy white growth is developed along the line of puncture, and liquefaction occurs in funnel shape near the surface; this soon extends to the walls of the tube and downward, and a thin film is formed on the surface. Upon *potato*, at 15° C., at the end of three days the surface is covered by a thin, viscid, slimy, yellowish, finely wrinkled layer; at 37° C. the entire surface is covered in twenty-four hours with a reddish-yellow or pink layer; in forty-eight hours this extends over the lower surface of the potato also, except where it is in contact with the receptacle in which it is placed.

286. BACILLUS PYOCYANUS β (Ernst).

Found in pus from bandages colored green.

Morphology.—Slender bacilli from 2 to 4 μ long—occasionally 5 to 6 μ —and from 0.5 to 0.75 μ broad; sometimes united in pairs, or chains of three elements.

Biological Characters.—An *aërobic*, *liquefying*, *actively motile*, *chromogenic* bacillus. Produces a yellowish-green pigment; when old cultures are shaken up with chloroform and this is allowed to stand, three layers are formed—an upper, clouded, dirty-yellow layer; below this is a milky, pale-green layer; and at the bottom a transparent, azure-blue layer. Spore formation has not been demonstrated. Grows in the usual culture media at the

room temperature—more rapidly at 35° C. Upon *gelatin plates* colonies are formed resembling those of the well-known *Bacillus pyocyaneus*, but liquefaction is more rapid. In *gelatin stick cultures* funnel-shaped liquefaction occurs at the upper part of the line of puncture by the third day, and progresses more rapidly than is the case with *Bacillus pyocyaneus* under the same circumstances; on the fifth day a bluish-green color is developed; by the twelfth day liquefaction has obliterated the entire line of growth and extends to the margins of the tube; the liquefied gelatin for a depth of about one centimetre has a dark emerald-green color, and a film consisting of bacilli is seen upon the surface. Upon the *surface of agar* a flat, greenish-white, dry layer is formed along the line of inoculation, and the agar around, at the end of a week, acquires a bluish-green color. Upon *potato*, at the end of three days, an abundant dry layer of a fawn-brown color has developed; this is surrounded by a pale-green coloration of the potato, and at points where the surface is fissured an intense dark-green color is developed; the growth on potato has a more or less wrinkled appearance; when one of the fawn-colored colonies is touched with the platinum needle the point touched, at the end of two to five minutes, acquires an intense dark leaf-green color, which reaches its maximum intensity in about ten minutes, and has faded out again at the end of half an hour. Ernst considers this “chameleon phenomenon” the most characteristic distinction between the bacillus under consideration and *Bacillus pyocyaneus*. In *milk* a green color is developed at the surface, the casein is precipitated and subsequently peptonized.

287. BACILLUS MYCOIDES ROSEUS (Scholl).

Found in the soil.

Morphology.—Resembles the anthrax bacillus.

Biological Characters.—An *aërobic, liquefying, chromogenic* bacillus. Produces a red pigment when cultivated in the absence of light. Spore formation not reported. Grows rapidly at the room temperature. Upon *gelatin plates* forms colonies of interlaced filaments which cause liquefaction of the surrounding gelatin. In *gelatin stick cultures* liquefaction rapidly occurs; a red layer is formed upon the surface, and a sediment of the same color is seen at the bottom of the liquefied medium, but the gelatin itself is not colored. Upon the *surface of agar*, in the dark, a pink layer is developed, while in the light it is white.

288. BACILLUS ROSACEUM METALLOIDES (Dowdeswell).

Morphology.—Bacilli from 0.6 to 0.8 μ broad and about twice as long.

Biological Characters.—An *aërobic, liquefying, motile* (usually not motile), *chromogenic* bacillus. Forms a magenta-red pigment which has a metallic lustre. Spore formation not observed. Grows best at 15° C.; no development at 35° C.; is destroyed in five minutes by a temperature of 55° C. Upon *gelatin plates*, at 15° C., superficial colonies are developed, which in the course of a few days are elevated, colorless discs about two millimetres in diameter; under a low power the centre appears dark, the margin transparent and granular; later the colony acquires a red color and liquefaction of the surrounding gelatin occurs. In *gelatin stick cultures* a red layer is developed upon the surface, and later a broad funnel of liquefied gelatin is slowly developed. Upon the *surface of agar* a pale-red layer is formed. Upon *potato*, at 15° C., a thick layer quickly covers the entire surface; this has a beautiful red color, especially near the margins.

289. BACILLUS VISCOSUS (Frankland).

Resembles very closely, and is perhaps identical with, *Bacillus fluorescens liquefaciens*.

Found in unfiltered river water.

Morphology.—Bacilli with round ends, 1.5 to 2 μ long and about three or four times as long as broad; usually united in pairs.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Produces a fluorescent green pigment. Spore formation not observed. Grows rapidly at the room temperature in the usual culture media. Upon *gelatin plates* the deep colonies appear under a low power as finely granular discs with a smooth contour; when they come to the surface the margin is fringed and hair-like offshoots extend into the gelatin; at the same time liquefaction occurs around the colonies and rapidly extends; each liquefying colony is surrounded by a fluorescent green zone, and the liquefied gelatin has the same color. In *gelatin stick cultures* liquefaction in funnel shape is already seen on the second day, and the liquefied gelatin is filled with whitish flocculi, while a slight green fluorescence is seen near the surface; liquefaction progresses rapidly, and a viscid layer of a greenish-white color forms upon the surface, while the liquefied gelatin below is more or less clouded and has a fluorescent green color; an abundant flocculent deposit is seen at the bottom of the tube. Upon the *surface of agar* a smooth, glistening, greenish-white layer is formed along the line of inoculation and the agar quickly acquires a green color. In *bouillon*, at the end of two days, the liquid is clouded, and later a thin, greenish-white layer forms upon the surface, while the bouillon acquires a green color. Upon *potato* a chocolate-colored, moist-shining layer quickly extends over the entire surface.

290. BACILLUS VIOLACEUS.

First found in the water of the Spree at Berlin, and since by Frankland in the water of the Thames and of the Lea.

Morphology.—Bacilli with round ends, about 1.7 μ long and 0.8 μ broad; usually in pairs; may grow out into long filaments.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, motile, chromogenic* bacillus. Produces a dark-violet pigment. Forms oval spores, which are located in the centre of the rods. Upon *gelatin plates*, at the end of two days, the colonies are seen to be irregular in outline and granular; on the fourth day a funnel of liquefied gelatin is formed by each colony, and under a low power an opaque mass surrounded by convoluted, filamentous offshoots is seen at the bottom of this; later the funnel of liquefied gelatin increases in dimensions and the colony acquires a deep-violet color. In *gelatin stick cultures* liquefaction occurs rapidly along the line of inoculation as a funnel-shaped sac; the liquefied gelatin is clouded, and a violet-colored deposit is seen at the bottom of the tube. Upon the *surface of agar* a smooth, shining layer of a deep-violet color is quickly developed. Upon *potato* growth is limited to the line of inoculation and a dark-violet stripe is slowly developed. *Blood serum* is liquefied by this bacillus.

291. BACILLUS SULFUREUM (Holschewnikoff).

Morphology.—Bacilli with round ends, from 1.6 to 2.4 μ long and 0.5 μ broad.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, motile, chromogenic* bacillus. Forms, in the absence of oxygen, a reddish-brown or red pigment. Produces sulphuretted hydrogen in sterilized urine and certain other media when cultivated in the absence of oxygen. Spore formation not observed. Grows in the usual culture media at the room temperature—also in the incubating oven. Upon *gelatin plates* small, punctiform colonies are developed within forty-eight hours, which, when they reach the surface, cause a funnel-shaped liquefaction; as the liquefaction progresses very slowly the liquefied gelatin is dried and the funnel-shaped cavities are filled with air. In *gelatin stick cultures* small colonies are developed along the line of puncture, and liquefaction in funnel shape occurs

very slowly; in contact with the air the colonies are white; in the absence of oxygen liquefaction does not occur and the colonies have a reddish-brown or red color. Upon the *surface of agar*, in the incubating oven, a slimy, gray layer is quickly developed. Upon *potato* no growth occurs in the presence of oxygen; when it is excluded a reddish-brown layer is developed. In *milk*, at the end of ten days, solution of the casein commences without previous coagulation.

292. BACILLUS RUBIDUS (Eisenberg).

Found in water.

Morphology.—Bacilli of medium size, with blunt ends, often united in long filaments.

Biological Characters.—An *aërobic*, *liquefying*, *motile*, *chromogenic* bacillus. Forms a shining, brownish-red pigment. Spore formation not observed. Grows very slowly in the usual culture media at the room temperature—not in the incubator at 37° C. Upon *gelatin plates* form spherical, finely granular colonies, which are of a reddish color at the centre and around which the gelatin is slowly liquefied. In *gelatin stick cultures* liquefaction occurs slowly and a brownish-red pigment is formed. Upon the *surface of agar* a brownish-red layer is developed, which quickly extends over the surface. A similar development occurs upon *potato*—not limited to line of inoculation. *Blood serum* is liquefied by this bacillus.



FIG. 213.



FIG. 214.

FIG. 213.—*Bacterium termo* of Vignal, from a bouillon culture. $\times 1,500$. (Vignal.)

FIG. 214.—The same from a culture fifteen days old. $\times 1500$. (Vignal.)

293. BACTERIUM TERMO OF VIGNAL.

Found in the salivary secretions of healthy persons by Vignal, and described under the name of *Bacterium termo*, which was formerly given to various motile bacilli encountered in putrefying infusions, many of which have been differentiated by modern methods and are described under different names.

Morphology.—Bacilli from 1.5 to 2 μ long, constricted in the middle, and 0.5 to 0.7 μ broad; never united in chains or filaments; possess terminal flagella.

Biological Characters.—An *aërobic*, *liquefying*, *motile*, *chromogenic* bacillus. Forms in gelatin cultures a fluorescent yellowish-gray pigment. Spore formation not determined. Grows at the room temperature in the usual culture media—better at a higher temperature. Gelatin cultures give off a putrefactive odor. Upon *gelatin plates*, at the end of twenty-four hours, small, white colonies are seen; at the end of forty-eight hours these have a diameter of two to five millimetres; the centre of the colony is white and opaque, and it is surrounded by a zone of liquefied gelatin which is

clouded and more or less granular in appearance; the area of liquefaction increases and the opaque central colony disappears, while the margins of the liquefied gelatin are clouded and whitish. In *gelatin stick cultures*, at the end of two days, liquefaction has occurred all along the line of puncture and a broad funnel is formed above, while below a mass of white flocculi fills the narrow tube; at the end of three days the gelatin is completely liquefied, it is opalescent, fluorescent, and greenish; on the fifth or sixth day it has a yellowish-green color and a strong odor of putrefaction. Upon the *surface of agar*, at a temperature of 36° C., it forms circular, grayish-white, almost transparent colonies; these rapidly coalesce to form a layer of uniform thickness, which is easily broken up. In *bouillon* the liquid is at first clouded throughout, and at the end of a week has a green color, while the bacilli are seen at the bottom as a pulverulent white deposit. *Blood serum* is slowly liquefied and gives off a strong putrefactive odor. Upon *potato*, at the end of forty-eight hours in the incubating oven, a glairy, grayish layer is formed the size of a five-franc piece; later this acquires a pale-yellow color.

294. BACILLUS BUCCALIS MINUTUS.

Synonym.—*Bacillus g.* Vignal.

Found by Vignal in the salivary secretions of healthy persons.

Morphology.—A very short bacillus, with round ends, almost as broad as long; in cultures upon agar the length is from 0.5 to 1 μ —usually about 0.7 μ ; in neutral bouillon it is from 1 to 1.7 μ long; in old cultures involution forms are common; in stained preparations the two ends are more deeply stained than the central portion.

Biological Characters.—An *aërobic, liquefying, chromogenic* bacillus. Produces a yellow pigment. Spore formation not observed. Motility not mentioned. Grows slowly at the room temperature. Upon *gelatin plates*, at the end of forty-eight hours, the colonies are round, with refractive contour and of a mastic-yellow color; they are but slightly elevated and the gelatin commences to liquefy around them. In *gelatin stick cultures*, at the end of forty-eight hours, a yellowish-white growth is seen along the line of puncture, and upon the surface a layer having the same color and several millimetres in diameter has developed; by the fourth day the surface growth has increased to twice the size and is yellow at the centre, while the periphery is white; the growth along the line of puncture is abundant and consists of small, closely crowded colonies; below the surface growth a cup-shaped cavity filled with clouded liquefied gelatin is seen; by the sixth day a small funnel of liquefaction has formed, the liquefied gelatin is clear and contains some white flocculi in suspension; by the twelfth day the gelatin in the tube is completely liquefied, an abundant yellow deposit is seen at the bottom and the liquefied gelatin has the same color. Upon the *surface of agar* golden-yellow plaques are developed, which are easily removed with the platinum needle. In *bouillon* a thin, iridescent pellicle is formed upon the surface and the fluid below is clouded, while an abundant yellow deposit accumulates at the bottom. Does not grow well in acid bouillon. Upon *potato*, at the end of forty-eight hours, a thin and extended layer is formed of a yellow color, which later has a brownish tint.

295. BACILLUS OF CANESTRINI.

Found in larvæ and bees from infected hives in Italy (1891).

Morphology.—Bacilli with rounded ends, from 4 to 6 μ long and about 2 μ broad; the isolated elements are somewhat longer than those in chains; solitary, in pairs, or in chains which may contain numerous segments.

Biological Characters.—An *aërobic, liquefying, motile, chromogenic* bacillus. Forms a pink pigment. The movements are slow and oscillating. Forms oval spores 3 μ long and 1.5 μ broad. Grows at the room tempera-

ture—better at 37° C. *Stains* with the aniline colors and by Gram's method. In *gelatin stick cultures* causes liquefaction of the gelatin; after some days the upper portion of the liquefied gelatin has a pink color, while at the bottom an abundant white sediment collects. Upon the *surface of agar* forms a white layer which contains numerous spores. Liquefies *blood serum*; in this medium the bacilli are surrounded by a capsule, and are frequently seen in long chains containing fifteen to twenty elements, all enclosed in a single capsular envelope. Upon *potato* development is rapid at 37° C.; at the end of twenty-four hours a wine-colored layer is formed. Not pathogenic for mice or guinea-pigs. Experiments made with pure cultures show that it is pathogenic for bees and their larvæ, and that it is the cause of an infectious malady which is destructive to these insects in certain localities (in Italy).

C. Non-chromogenic, Non-liquefying Bacilli.

296. BACILLUS UBIQUITUS (Jordan).

Found in sewage at Lawrence, Mass.; also in water and in the air—"apparently abundant everywhere" (Jordan).

Morphology.—Bacilli from 1.1 to 2 μ long and about 1 μ broad—resemble micrococci; quite variable in form; in bouillon short filaments are sometimes formed.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, non-motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media—also at 37° C. Upon *gelatin plates* forms small, spherical or oval colonies, which have a yellowish tinge; at the end of two days the superficial colonies are prominent, white, and glistening, resembling a drop of milk; they gradually increase in diameter, become somewhat irregular in outline, and acquire a dull brownish tint. Under a low power the young colonies are seen to be finely granular and to have a smooth contour. In *gelatin stick cultures* development occurs upon the surface and along the line of puncture, producing a "nail-shaped" growth at the end of a week; the color is at first a lustrous porcelain-white, which later changes to a dull brownish-gray; grows well in slightly acid gelatin. Upon the *surface of agar* a whitish-gray layer is developed which has a slightly metallic lustre. Upon *potato* a shining, white

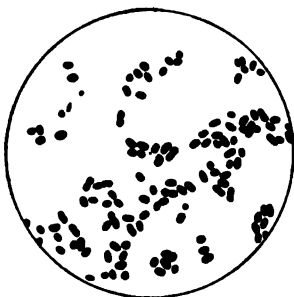


FIG. 215. — *Bacillus ubiquitus*.
× 1,000. From a photomicrograph.
(Jordan.)

growth of limited extent. In *milk* coagulation occurs quickly at 37° C., and the milk acquires a strongly acid reaction. Reduces nitrates vigorously. "This species apparently resembles quite closely the *Bacillus candicans* described by the Franklands" (*Zeit. für Hyg.*, Bd. vi., page 397). "It differs from that, however, among other respects, in its capacity for reducing nitrates and in its mode of growth upon agar and potato" (Jordan).

297. BACILLUS CANDICANS (Frankland).

Found in the soil.

Morphology.—Short, thick bacilli, resembling micrococci; often form short filaments.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows slowly at the room temperature in the usual culture media. Upon *gelatin plates* the superficial colonies resemble drops of milk; the deep colonies under a low power are seen to be spherical, slightly

granular at the margins, and have a smooth contour. In *gelatin stick cultures* the superficial growth is like a drop of milk in appearance; very scanty growth along the line of puncture at first, later a row of spherical colonies is seen; the surface growth in old cultures has a slightly reddish tint. Upon the *surface of agar* a thin, transparent, grayish-white layer with smooth margins. Upon *potato* an abundant layer is developed.

298. BACILLUS ALBUS (Eisenberg).

Found in water.

Morphology.—Short bacilli with blunt ends, often united in short chains.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Spore formation not observed. Grows slowly at the room temperature—not in the incubator at 37° C. Upon *gelatin plates* forms round, white colonies the size of a pin's head. In *gelatin stick cultures* grows slowly, forming a white line along the puncture and a small, button-like, white mass at the point of entrance. Upon the *surface of agar* forms a milk-white layer. Upon *potato* a dirty yellowish-white growth, limited to the line of inoculation.

299. BACILLUS ACIDI LACTICI (Hueppe).

Found in sour milk.

Morphology.—Bacilli from 1 to 1.7 μ long and from 0.3 to 0.4 μ broad; usually in pairs, sometimes in chains of four.

Biological Characters.—An *aërobic and facultative anaërobic, non-liquefying, non-motile* bacillus. Forms spherical spores, which are located at the ends of the rods. Grows slowly at the room temperature in the usual culture media. Upon *gelatin plates* forms small, white, punctiform colonies, which later develop into shining, porcelain-colored discs with a transparent margin; under a low power they have a yellowish tint in the centre and thin, irregular margins. In *gelatin stick cultures* small colonies are developed along the line of puncture, and later a dry, glistening, soft, grayish-white, and tolerably thick layer is developed upon the surface. Upon *potato* an extended, yellowish-brown layer is formed. In *milk* lactic acid is produced, the casein is precipitated, and carbon dioxide is given off.

300. BACILLUS LIMBATUS ACIDI LACTICI (Marpmann).

Found in fresh cow's milk.

Morphology.—Short, thick bacilli, usually in pairs; every rod is surrounded by a capsule which is not stained by the aniline colors.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows slowly in the usual culture media at the room temperature—also in the incubator. Upon *gelatin plates*, at the end of twenty-four hours, forms milk-white, punctiform colonies. In *gelatin stick cultures* scanty development occurs along the line of puncture, and upon the surface a flat, irregular layer of a white, pus-like color is formed. In *milk*, at the end of twelve hours, a slightly reddish color is seen; at the end of twenty-four hours coagulation of the casein and a strongly acid reaction—lactic acid; does not produce gas.

301. BACILLUS LACTIS PITUITOSI.

Synonym.—Bacillus der schleimigen Milch (Löffler).

Found in milk.

Morphology.—Tolerably thick, slightly curved bacilli, which very quickly break up into small segments resembling micrococci.

Biological Characters.—An *aërobic, non-liquefying* bacillus. Grows

rather rapidly at the room temperature. Spore formation not determined. Upon *gelatin plates* forms white colonies with well-defined contour, from one-quarter to one-half millimetre in diameter; by transmitted light these are grayish-brown in color and present a slightly radial striped appearance. Upon the *surface of agar* a dirty-white layer is developed. Upon *potato* a grayish-white, tolerably dry layer. In *milk* a slightly acid reaction is produced, and a very viscid substance having a peculiar odor is formed, especially in the lower portion of the liquid; this can be drawn out into long threads.

302. BACILLUS AËROGENES (Miller).

Found in the alimentary tract of healthy persons.

Morphology.—Small bacilli of various lengths.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Spore formation not observed. Grows at the room temperature. Upon *gelatin plates* forms spherical, homogeneous, transparent, white or slightly yellowish colonies; older colonies sometimes appear to be formed of concentric rings. In *gelatin stick cultures* development occurs along the line of puncture and the growth has a yellowish color; upon the surface a thin, pearl-gray layer with dentate margins is formed; in old cultures the line of inoculation acquires a dark-brown color and is surrounded by a pale-brown halo. Upon *potato* a dry layer of a dirty bluish-yellow color, and with irregular outlines, is slowly developed.

303. BACTERIUM AËROGENES (Miller).

Found in the alimentary tract of healthy individuals.

Morphology.—Short rods, solitary or in pairs.

Biological Characters.—An *aërobic and facultative anaërobic, non-liquefying, motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates* forms sharply defined, yellowish colonies, which are marked by dark lines radiating from the centre toward the margin. In *gelatin stick cultures* development occurs along the entire line of puncture and the growth has a brownish-yellow color; upon the surface a soft, flat, grayish-white mass is formed about the point of puncture. Upon the *surface of agar* a soft, grayish-white layer. Upon *potato* a soft layer with irregular margins and of a slightly yellowish-white color.

304. HELICOBACTERIUM AËROGENES (Miller).

Found in the alimentary tract of healthy persons.

Morphology.—Slender bacilli, solitary or in chains; grow out into long, undulating or spiral threads.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms transparent, white or slightly yellowish colonies of various forms—spherical, oval, snail-shaped, spindle-shaped, spiral, etc. In *gelatin stick cultures* grows upon the surface as a thin, bluish, scarcely visible, dry layer, which covers the entire surface at the end of forty-eight hours; along the line of puncture a uniform, light-yellowish growth. Upon *agar* the growth is not characteristic. Upon *potato* a layer is formed which has a dry surface, indented margins, and a yellowish-brown color.

305. BACILLUS AQUATILIS SULCATUS NO. I. (Weichselbaum).

Found in the Vienna water supply.

Morphology.—Bacilli resembling the bacillus of typhoid fever in form and dimensions.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature—not so well in the incubating oven. Growth occurs at a temperature as low as 5° to 7° C. Upon *gelatin plates*, at the end of two days, the superficial colonies are seen as flat discs with a thicker and whitish centre and very thin, bluish, notched margins; under a low power the surface is seen to be covered with fine lines or furrows which cross each other at various angles; the color is white, with a yellowish tint at the centre; later very numerous lines are seen crossing each other in all directions at the centre, while the periphery is still white and is marked by more delicate lines. The deep colonies are round and yellowish. In *gelatin stick cultures* a flat, white layer with notched edges is seen at the end of twenty-four hours; this becomes thicker and is of less diameter than the surface growth of the typhoid bacillus under similar conditions. Upon the *surface of agar*, at 37.5° C., a tolerably thick, white layer is developed; this has an odor like that of milk. Upon *potato*, at 37.5° C., the growth is invisible, the line of inoculation having only a moist appearance; at the room temperature this is also the case at first, but later a very thin, moist-shining, often cream-colored layer with raised edges is seen, and the potato around this acquires a bluish-gray color, which again fades out.

306. BACILLUS AQUATILIS SULCATUS NO. II. (Weichselbaum).

Found in the Vienna water supply.

Morphology.—Short bacilli with round ends, of the form and dimensions of the shorter typhoid bacilli.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—not so well in the incubating oven. Grows at a lower temperature than the typhoid bacillus—5° to 7° C. Upon *gelatin plates*, at the end of two days, the superficial colonies are similar to those of the typhoid bacillus and of the preceding species, but somewhat thicker and not visibly notched; under the microscope they are seen to be indistinctly notched and marked by lines, although not so distinctly as are the colonies of *Bacillus aquatilis sulcatus* No. I.; the centre of the disc-shaped colonies is yellowish, the periphery white; after three days they become thicker and the notching of the margins and lines upon the surface are no longer seen, while the entire colony, with the exception of the outer margin, has a yellowish color. In *gelatin stick cultures* a whitish, rather thick layer of limited dimensions is formed upon the surface. Upon the *surface of agar*, in the incubator, a grayish-white layer is developed in twenty-four hours. Upon *potato*, at the room temperature, a bluish-gray color is first seen along the line of inoculation, and a yellowish-gray or yellowish-brown layer is subsequently developed; this may become tolerably abundant, while the original color disappears; potato cultures give off a slight urinous odor.

307. BACILLUS AQUATILIS SULCATUS NO. III. (Weichselbaum).

Found in the Vienna water supply.

Morphology.—Very short bacilli, frequently resembling micrococci.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows at the room temperature and at 5° to 7° C.—not so well in the incubating oven. Old cultures give off a disagreeable odor. Upon *gelatin plates*, at the end of two or three days, the superficial colonies are disc-shaped, with a thicker, white centre and a very thin, bluish periphery; the margin is notched; under the microscope the surface is seen to be marked with lines; later the colonies increase in thickness and diameter and lose the bluish color; the system of fine lines also disappears, and the surface is covered with numerous short lines and furrows; the yellow color extends from the centre nearer to

the periphery. In *gelatin stick cultures*, at the end of twenty-four hours, a very thin, white layer with notched edges is developed; this extends rapidly to the margins of the tube. Upon the *surface of agar* an abundant grayish-white layer is quickly developed in the culture oven; this has an odor resembling that of milk. Upon *potato*, at the room temperature, a discoloration of the line of inoculation is seen at the end of twenty-four hours; later an abundant pale-yellow layer with raised margins is developed which has an odor of herring brine; at the end of nine days the potato around the growth has a bluish-green color.

308. BACILLUS AQUATILIS SULCATUS NO. IV. (Weichselbaum).

Found in the Vienna water supply.

Morphology.—Bacilli of various lengths; often grow out into filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* (the short rods only) bacillus. Spore formation not demonstrated. Grows slowly at the room temperature—still more so in the incubating oven. Upon *gelatin plates* the colonies are first visible on the fourth day; the superficial colonies are thin, bluish discs with notched margins and a somewhat thicker, whitish centre; under a low power the surface is seen to be covered with fine lines, and the larger colonies have a yellowish color in the centre; later they increase in thickness and diameter and acquire a yellow color throughout, while the system of lines is replaced by more numerous and shorter lines and furrows. In *gelatin stick cultures* development is very slow, but at the end of several days a grayish-white layer with notched margins is developed, which gradually extends to the walls of the test tube. Upon the *surface of agar*, at 37° C., the growth is very scanty at the end of six days; at the room temperature a grayish-white layer of moderate thickness is formed within two days. Upon *potato* no growth occurs either in the incubator or at the room temperature.

309. BACILLUS AQUATILIS SULCATUS NO. V. (Weichselbaum).

Found in the Vienna water supply.

Morphology.—Bacilli with round or pointed ends, of various lengths; somewhat thicker than the typhoid bacillus.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media—not in the incubating oven. Upon *gelatin plates* forms colonies resembling those of *Bacillus aquatilis sulcatus* No. I., which become visible in two or three days. In *gelatin stick cultures* a layer of moderate thickness is formed, which gradually extends over the surface; this is grayish-white at first and later has a yellow color like that of the yolk of an egg. Upon the *surface of agar* no growth occurs in the incubator, but at the room temperature an abundant, viscid, yellowish layer is developed. Upon *potato*, at the room temperature, a bluish-yellow layer is formed and the potato around it acquires a dark-gray color, which disappears later, while the vegetation after a time has a honey-yellow color.

310. BACILLUS MULTIPEDICULUS (Flügge).

Found in the air and in water.

Morphology.—Long slender bacilli.

Biological Characters.—An *aërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates* forms spherical or oval opaque colonies, which under a low power are seen to give off at certain points of the periphery broad, segmented outgrowths consisting of round zoöglæa

masses; at the end of two or three days the oval, white, superficial colonies are seen with the naked eye to be surrounded with these outgrowths, which resemble the feet and antennæ of certain insects. In *gelatin stick cultures* a white layer is developed upon the surface which gives off short, isolated outgrowths. Upon *potato* a smooth, dirty-yellow layer of limited extent is developed.

311. BACILLUS CYSTIFORMIS (Clado).

Found in the urine of a patient with cystitis.

Morphology.—Very short and slender bacilli.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Grows slowly at the room temperature. Upon *gelatin plates* forms transparent, yellowish colonies, first round and later oval in form; from the fourth to the seventh day a granular elevation appears at the centre, around which a finely granular, yellowish zone is seen, and outside of this a broad, transparent zone with double contour. In *gelatin stick cultures* a scanty development occurs along the line of puncture, and on the surface a whitish layer is developed. Upon *agar* a yellowish-white layer.

312. BACILLUS HEPATICUS FORTUITUS (Sternberg).

Obtained, by inoculation in a guinea-pig, from the liver of a yellow-fever cadaver.

Morphology.—Resembles *Bacillus coli communis* in its morphology, but differs from this bacillus in being strictly *aërobic*.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows at the room temperature in the usual culture media. Upon *gelatin plates* the deep colonies are spherical, homogeneous or finely granular, and light-brown in color; at the end of four days they are more or less lobate. The superficial colonies are shaped like a mamma, with striations radiating from the centre, and are of a dark-brown color under the microscope. In *gelatin stick cultures* no growth occurs along the line of puncture, except to a slight extent near the surface; on the surface a white, button-like mass is formed about the point of puncture. Upon the *surface of glycerin-agar* the development is quite rapid at 35° C., the entire surface being nearly covered with a soft, milk-white growth within twenty-four hours. Upon *potato*, at the end of forty-eight hours, a rather dry and thick, cream-white growth forms along the line of inoculation; the potato has a bluish discoloration, which subsequently disappears; at the end of two weeks a rather thin, semi-fluid, light-brown layer covers the entire surface. Not pathogenic for rabbits—single experiment.

313. BACILLUS INTESTINUS MOTILIS (Sternberg).

Obtained from the contents of the intestine of yellow-fever cadavers.

Morphology.—Resembles *Bacillus coli communis* in its morphology, but differs from this bacillus in being very actively motile, in its colonies upon gelatin plates, etc.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, actively motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of twenty-four hours, the deep colonies are spherical, homogeneous, and of a pale-straw color; the superficial colonies resemble little drops of water and are of a pale-brown color by transmitted light. In *gelatin stick cultures* pale straw-colored colonies are developed all along the line of puncture, and a rather thin, translucent, whitish layer forms upon the surface; sometimes a nebulous outgrowth occurs from the line of puncture, and tufted outlying colonies are formed throughout the gelatin; at other times, in old cultures, a few feathery tufts sprout out from the line of puncture.

ture. Upon *potato* the growth is rather thin and of a pale-yellow color, not extending far from the line of inoculation. Does not form gas in a saccharine solution—agua coco.

314. BACILLUS CAVIÆ FORTUITUS (Sternberg).

Obtained, by inoculation in a guinea-pig, from the liver of a yellow-fever cadaver, preserved for forty-eight hours in an antiseptic wrapping.

Morphology.—Bacilli with round ends, from 1 to 4 μ long and 0.5 to 0.8 μ broad; often in pairs.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *actively motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of three days, the colonies are small, spherical, and under the microscope light-brown in color; later opaque, or sometimes with an opaque centre surrounded by a transparent zone. In *gelatin stick cultures* there is a scanty growth about the point of puncture; growth occurs all along the line of puncture in the form of spherical, translucent, straw-colored colonies, which have a pearly lustre by reflected light. Upon *potato*, at the end of a week, development has occurred in the form of small, dirty-yellow masses. Does not form gas in a saccharine liquid—agua coco.

315. BACILLUS COLI SIMILIS (Sternberg).

Obtained from a piece of liver—of man—kept in an antiseptic wrapping for forty-eight hours.

Morphology.—A bacillus with round ends, from 1 to 3 μ long and from 0.4 to 0.5 μ thick; solitary or in pairs.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media—better at 37° C. Upon *gelatin plates*, at the end of two days, the deep colonies are spherical and pale-brown in color; later they become opaque. The superficial colonies are at first translucent, homogeneous, drop-like elevations; later they are quite thin and have a pale-brown color. In *gelatin stick cultures* a translucent growth with irregular margins is developed upon the surface, and a rather scanty line of growth is seen along the track of the inoculating needle. On *potato*, at 22° C., a thick, dirty-white or pale-brown layer is developed along the impfstrich. Not pathogenic for rabbits or guinea-pigs.

316. BACILLUS FILIFORMIS HAVANIENSIS (Sternberg).

Obtained in anaërobic cultures from the liver of a yellow-fever cadaver.

Morphology.—Long and slender bacilli, about 0.3 μ in diameter, and forming long, homogeneous filaments in aërobic cultures, while the bacilli are shorter and thicker in anaërobic cultures in glycerin-agar.

Biological Characters.—An *anaërobic* and *facultative aërobic*, *non-liquefying*, *non-motile* bacillus. Does not form spores. In *gelatin stick cultures* a scanty growth occurs along the line of puncture; no growth on the surface. In anaërobic, glycerin-agar roll tubes colonies are developed which are spherical or irregular in outline, of a pale-brown or straw color by transmitted light, white and opaque by reflected light; the superficial colonies are thin and translucent, and have a bluish lustre by reflected light; later they appear as opaque, cream-like masses with irregular contour. In *nutrient agar* a scanty, milk-white growth occurs upon the surface and an opaque, branching growth along the line of puncture. No growth upon *potato*. Grows in neutral *bouillon*, causing a slight opalescence, and later a scanty white deposit at the bottom of the tube. Not pathogenic for rabbits or guinea-pigs.

317. BACILLUS MARTINEZ (Sternberg).

Obtained from the liver of a yellow-fever cadaver, kept for forty-eight hours in an antiseptic wrapping.

Morphology.—A short, oval bacillus from 1 to 1.2 μ long and from 0.5 to 0.8 μ broad.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are spherical and translucent; superficial colonies shaped like a mamma, with a central nipple-like projection, the surface covered with mosaic markings. In *gelatin stick cultures* a thin, translucent, scanty growth upon the surface, and large, spherical, translucent colonies along the line of puncture. In *glycerin-agar stick cultures* growth to the bottom of the line of puncture, and scanty development on the surface.

318. BACILLUS EPIDERMIDIS (Bizzozero).

Synonym.—*Leptothrix epidermidis*.

Found attached to scales of epidermis from between the toes.

Morphology.—Bacilli from 2.8 to 3 μ long and 0.3 μ broad.

Biological Characters.—An *aërobic*, *non liquefying* bacillus. Forms long, oval spores—at 25° C. in three days. Grows best at a temperature of 15° to 20° C. Very scanty growth in gelatin. Grows upon the *surface of agar*. Upon *potato*, at 15° to 20° C., the development at first is in the form of viscid, transparent, drop-like colonies, which gradually coalesce and form a rather thick layer.

319. BACILLUS NODOSUS PARVUS (Lustgarten).

Found in the healthy urethra of man.

Morphology.—Bacilli from 1.2 to 2.4 μ long and 0.4 μ broad; one extremity often presents an irregular club shape; usually united in pairs, in which the elements lie parallel or are united at an acute angle.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Grows best in the incubating oven. Spore formation not observed. Very slow and scanty growth in nutrient gelatin. Upon the *surface of agar*, at 37° C., at the end of twenty-four hours a white line of growth is seen along the line of inoculation; at the end of two to three days this has a breadth of five to six millimetres, and the central portion of the layer is white, chalky in appearance, porous, and lustreless; around this is a smooth, flat, glistening, grayish-white marginal zone one to two millimetres broad. In *agar stick cultures*, at the end of five to eight days, growth is seen along the line of puncture as a white stripe made up of confluent spherical colonies, while at the point of puncture a small, stearin-like drop is seen.

320. BACILLUS HYACINTHI SEPTICUS (Heinz).

Found in diseased hyacinths.

Morphology.—Bacilli with round ends, 4 to 6 μ long and about 1 μ broad; always solitary.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows at the room temperature. Old cultures have a strong putrefactive odor. Upon *gelatin plates* the superficial colonies are flat, shining, bluish-white in color with a somewhat darker centre, transparent, and about two millimetres in diameter; the deep colonies are oval with rather sharp poles, yellowish-white, and lustreless. In *gelatin stick cultures* growth occurs all along the

line of puncture, and on the surface as a shining layer of moderate extent. Upon *agar* the growth is similar to that upon gelatin. Upon *potato*, at the end of thirty-six hours, a dirty-yellow, slimy, granular layer.

321. BACTERIUM GLISCROGENUM (Malerba).

Found in urine which was viscid and acid in reaction.

Morphology.—Oval bacilli, from 0.57 to 1.14μ long and 0.41μ broad.

Biological Characters.—An *aërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media—better at 30° to 37° C. Upon *gelatin plates*, at the end of two days, punctiform colonies are seen, which gradually increase in size, are perfectly round, granular, and lenticular in form; later there is a wavy depression of the surface of the gelatin and gas bubbles are formed in the interior. In *gelatin stick cultures* growth occurs along the line of puncture and upon the surface in nail shape; the growth along the line of inoculation consists of disc-shaped, isolated colonies closely piled one upon another. Upon the *surface of agar*, at the room temperature, a granular, opalescent stripe is developed in from three to five days; at 37° C. an abundant development occurs in twenty-four hours; a white, viscid film forms upon the surface of the condensation water. Upon *potato* a yellowish or yellowish-brown layer is developed, and in the course of a few days numerous gas bubbles are formed, which become confluent; later the growth becomes viscid and extends over the entire surface. In *bouillon* diffuse cloudiness is seen at the end of twenty-four hours, and the fluid becomes viscid; at the end of four to five days a whitish layer forms upon the surface.

322. BACILLUS OVATUS MINUTISSIMUS (Unna).

Found upon the skin in cases of eczema seborrhœicum.

Morphology.—Short oval bacilli with pointed ends, 0.6 to 0.8μ long and 0.4μ broad; associated in irregular groups.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media. Gelatin and potato cultures give off a strong and disagreeable odor. Upon *gelatin plates*, at the end of eight days, the superficial colonies are the size of a mustard seed, prominent, spherical, grayish-white, shining, and resemble a drop of a solution of gum tragacanth; the deep colonies are punctiform and grayish-white in color; later the superficial colonies become flat, or occasionally preserve the form of a small pearl; these are almost one centimetre in diameter, round, finely granular, yellowish-gray, with a darker centre and a more transparent peripheral zone; the margins are notched. The deep colonies, under a low power, are seen to be spherical or oval, opaque, finely granular, dark-yellowish in the centre and paler at the periphery; they may attain the size of a pea. In *gelatin stick cultures* growth is rather rapid upon the surface in form of an abundant, slimy, grayish-white layer with irregular outlines; this later becomes dry and opaque, and presents small, spherical protuberances which are more transparent; along the line of puncture numerous grayish-white, closely crowded, punctiform colonies. Upon *agar* the growth is similar to that upon gelatin. Upon *potato* an abundant, grayish-white, dull-glistening layer.

323. CAPSULE BACILLI OF SMITH.

Theobald Smith has described three species or varieties (?) of capsule bacilli, resembling Friedländer's bacillus, obtained by him from the intestine of swine. These he designates by the letters *a*, *b*, and *c*.

Morphology.—Slight morphological differences were detected by cultivating all under the same circumstances in peptone-bouillon.

a. $1.2\ \mu$ long and 0.8 to $0.9\ \mu$ broad; capsule not visible in hanging drop.

b. 1.6 to $1.8\ \mu$ long and 0.8 to $0.9\ \mu$ thick; capsule clearly visible in hanging drop; usually in pairs; both ends somewhat thickened as in *a*.

c. Bacilli somewhat thicker than *b*; capsule not visible.

Friedländer's bacillus, 1 to $2\ \mu$ long and $1\ \mu$ thick. A second examination of all four, made at the end of forty-eight hours, showed no apparent difference in *a*, *b*, and Friedländer's bacillus, while *c* remained shorter than the others. None of the species retained their color when treated by Gram's method.

Biological Characters.—*Aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacilli.

Upon gelatin plates—

a forms colonies resembling those of *Bacillus coli communis*; they spread out in a thin layer, the margins of which are very thin and bluish by transmitted light; the contour is irregular; later the colonies become thicker, white, and opaque, and flap-like processes may be given off from the margins which double the original diameter of the colony—of four to five millimetres; in the centre a small, button-like elevation is seen.

b. The colonies are at first thicker and more opaque than those of *a*; they reach a diameter of four millimetres, and are thinner toward the margin, which is irregular in outline; a small central prominence is usually observed.

c. The colonies are thicker, circular in outline, with smooth margins, and attain a diameter of five millimetres; the central projection is strikingly large.

In peptone-bouillon, at 36°C ., at the end of five hours *a*, *b*, and *c* all cause the culture medium to be decidedly clouded, while Friedländer's bacillus only causes a slight cloudiness at the end of twenty-four hours. At the end of a week the cultures of *a* show a dense clouding and an abundant deposit at the bottom of the tube, but no layer on the surface; the cultures of *b* a similar cloudiness and deposit, and also a thick, gelatinous layer on the surface; the cultures of *c* a densely clouded medium with a thin film upon the surface. Friedländer's bacillus, cultivated in the same medium, showed a slight cloudiness and a few fragments of a mycoderma floating upon the surface. In the cultures of *b* the whole fluid becomes very viscid and can be drawn out into long threads; the same character is developed later and to a less extent in cultures of *a*; and in *c* the superficial film is somewhat viscid—cultures of Friedländer's bacillus do not exhibit this character. All of the cultures have an alkaline reaction, and those of *a*, *b*, and *c* after a time have a disagreeable odor.

In milk, at 37°C ., at the end of a week coagulation is not complete, although the fluid is very thick; *b* causes milk to be quite thick in two days, and to be completely coagulated in four days; *c*, at the end of a few days, causes the lower half of the milk to coagulate, and at the end of a week the whole is firmly coagulated; Friedländer's bacillus in milk did not produce any apparent change. The milk cultures of *a*, *b*, and *c* had an acid reaction; at the end of two weeks some viscid serum was seen above the coagulum in *a*, and the culture smelled like sour dough; at the same time the whole coagulum was viscid in the culture of *b* and a similar odor was perceived; the culture of *c* showed a superficial layer of serum which was not viscid, and the odor was that of cheese.

Upon potato *a* and *b* developed a thin, shining, grayish, transparent layer; the growth of *c* upon potato was thick and cream-white, resembling that of Friedländer's bacillus. In cultures containing glucose, and in mashed potato, *c* produced considerable quantities of gas—equal parts of CO_2 and of H_2 . Upon agar the growth of all is similar in appearance, but that of *a* and *b* is very viscid, that of *c* less so, while Friedländer's bacillus was destitute of this character.

Smith concludes his description of these bacilli by the remark that they may be identical with *Bacillus lactis aërogenes* of Escherich.

324. *BACILLUS PUTRIFICUS COLI* (Bienstock).

Found in human faeces.

Morphology.—Slender bacilli, about $3\ \mu$ long, often shorter, frequently united in long filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *actively motile* bacillus. Forms large spherical end spores, which give the bacilli the form of a drumstick; the motile, spore-bearing rods always advance with the spore in front. Grows at the room temperature. Upon nutrient gelatin a layer having a pearly lustre is developed; later this has a yellowish color and is homogeneous in appearance. This bacillus was supposed by Bienstock to be constantly present in faeces, and to be especially concerned in the decomposition of albuminous substances. Its characters of growth have not been determined with precision.

325. *BACILLUS SUBTILIS SIMULANS NO I.* (Bienstock).

Found in human faeces.

Morphology.—Bacilli with round ends, resembling *Bacillus subtilis*; grows out into long filaments, which become segmented and form short chains of two to five elements; or, more commonly, separate into single rods.

Biological Characters.—An *aërobic*, *non-liquefying*, *non-motile* bacillus. Grows best at 37° to 39° C. Forms oval spores, in which germination occurs at the two poles simultaneously, leaving the central portion of the new-formed bacillus bulging from the presence of the spore membrane. Upon *nutrient agar* this bacillus grows out in the form of a mesentery, yellowish-white "veins" running in all directions, which are united with each other smaller anastomosing branches.

326. *BACILLUS SUBTILIS SIMULANS NO. II.* (Bienstock).

Found in human faeces.

Morphology.—The same as the preceding species.

Biological Characters.—An *aërobic*, *non-liquefying* bacillus. Grows very rapidly—best at 37° to 39° C. Upon the *surface of agar* forms a glistening, white layer, which is at first smooth and later has a somewhat uneven surface, while the margins are surrounded by grape-like outgrowths. Imperfectly described.

327. *BACILLUS STRIATUS ALBUS* (Von Besser).

Found in nasal mucus from healthy individuals.

Morphology.—Small, thick bacilli, of about the size of the diphtheria bacillus; often more or less curved; in preparations stained with methylene blue the bacilli have a striped appearance; club-shaped involution forms may be seen.

Biological Characters.—An *aërobic*, *non-liquefying* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates* forms small, dry, superficial colonies. Upon *agar plates* forms prominent, milk-white colonies, one-half centimetre in diameter, which under the microscope have a brown nucleus surrounded by a paler brown marginal zone. Upon the *surface of agar* forms a flat, shining, grayish-white layer. Upon *potato* a scanty, transparent, jelly-like layer.

328. *BACILLUS STOLONATUS* (Adametz).

Found in water.

Morphology.—Bacilli two and one half times as long as thick.

Biological Characters.—An *aërobic*, *non-liquefying*, *actively motile* bacillus. Spore formation not observed. Grows rather quickly in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are small, spherical or oval, finely granular, whitish or yellowish-brown; the superficial colonies are whitish or brownish, elevated, hemispherical, with sharply defined outlines, and about one μ in diameter. In *gelatin stick cultures* a granular growth is slowly developed along the line of puncture and a white layer about the point of inoculation; at the end of two to three weeks the upper part of the line of puncture forms a saucer or flask-shaped cavity and the walls are covered with a white layer; there is, however, no liquefaction of the gelatin. Upon *agar plates* the growth is very characteristic: branches are given off from a central point; these are variously bent and give off numerous smaller, wavy branches; these colonies extend only upon the surface and may have a diameter of two to three centimetres; under a low power they are seen as very thin, finely granular, yellowish layers with club-shaped branches. Upon *potato* forms a dirty-white layer.

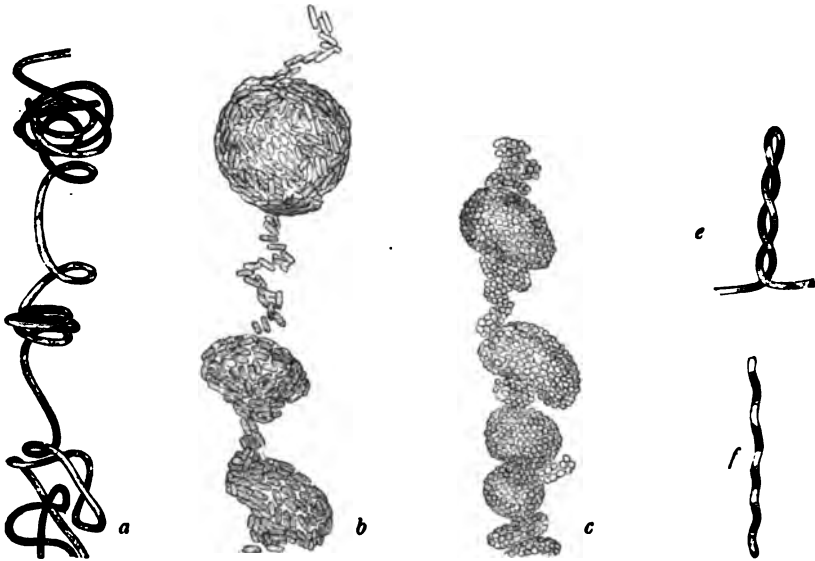


FIG. 216.—*Bacterium Zopfii*; a, long filament with commencement of "ball formation"; b shows the breaking up into short rods; c, further breaking up into spherical elements; e, f, spirilla-like filaments. $\times 740$. (Kurth.)

329. *BACILLUS VENTRICULI* (Raczyński).

Found in the stomach of dogs fed exclusively on meat.

Morphology—Bacilli from 1.5 to 3 μ long and 1 μ thick; united in pairs or in chains of four.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Does not form spores. Grows slowly at the room temperature. Upon *gelatin plates* forms round colonies, which are opaque at the centre and become more transparent toward the margin, which is surrounded by a dark contour. In *gelatin stick cultures* small, white, punctiform colonies are developed along the line of puncture. Upon *agar* a whitish layer is formed. This bacillus is said to cause the peptonization of albuminous substances when the reaction is acid or neutral.

330. BACTERIUM ZOPFII (Kurth).

Found in the intestine of chickens.

Morphology.—Bacilli from 2 to 5 μ long and 0.75 to 1 μ broad; form long filaments, which in gelatin cultures are folded and coiled in a peculiar manner. In liquid media straight filaments only are seen. The coiled filaments in gelatin cultures form tangled balls, in which they subsequently break up into short rods and finally into spherical bodies which appear to be reproductive elements.

Biological Characters.—An *aërobic, non-liquefying, actively motile* bacillus. Forms spherical spores (?), which are not highly refractive and stain deeply with the aniline colors. Grows best at the room temperature: at 30° to 37° C. the bacillus is no longer motile; at 37° to 40° C. involution forms are developed. Upon *gelatin plates*, at the end of twenty-eight hours, white, punctiform colonies are developed, from which as a centre a mass of fine, radiating filaments is given off; among these numerous small, white points are distributed, which under the microscope are seen to be spherical zoöglæa masses of a brownish-yellow color; the centre of the colonies consists of bundles of interlaced or parallel filaments. In *gelatin stick cultures*, at the end of twenty-four hours, a thick, white layer is developed along the upper portion of the line of puncture, and later radiating lines of growth are given off from this; these cross each other in various directions and resemble the mycelium of a fungus. No development occurs on blood serum.

331. BACTERIUM ZÜRNIANUM (List).

Found in water.

Morphology.—Short rods with slightly pointed ends, from 1.2 to 1.5 μ long and 0.6 to 0.8 μ broad; in stained preparations the ends are most deeply stained, giving the rods the appearance of diplococci.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not form spores. Grows in the usual culture media at the room temperature—better at 25° to 30° C. Upon *gelatin plates* forms spherical, dirty-white or gray, extremely viscid colonies, which develop into slimy, grape-like masses. In *gelatin stick cultures* a grape-like mass upon the surface and scanty growth along the line of puncture. Upon *potato*, at 25 to 30° C., a slimy, translucent, gray or yellowish-white layer extends over the surface within forty-eight hours.

332. BACILLUS OF COLOMIATTI.

Obtained from xerotic masses from the eye of a child and in certain forms of conjunctivitis.

Morphology.—Bacilli which correspond with the bacillus of mouse septicæmia in length, associated in irregular masses.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Does not grow at the room temperature. Forms spherical spores, which lie at the ends of the rods. Does not grow in nutrient gelatin or on potato. Upon the *surface of agar* a thin film is developed at 34° to 39° C., which gives to the surface a fatty lustre. Upon *blood serum*, at 35° to 39° C., development occurs along the line of inoculation as a dull-gray layer with a fatty lustre, which has a breadth of two to three millimetres and a rosette-like form.

333. BACILLUS SCISSUS (Frankland).

Found in the soil.

Morphology.—A short, thick bacillus of variable dimensions; usually about 1 μ broad and 1 to 2 μ long; resembles *Bacillus prodigiosus*, and, like

this, may easily be mistaken for a micrococcus; sometimes in pairs or in short chains.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Spore formation not observed. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are yellowish and punctiform; under a low power they are seen to be opaque in the middle and have dentate margins; when they break through to the surface they form small, prominent masses, and from these superficial colonies are developed which have a pale-greenish tint, a deeply dentate margin, and are seen under the microscope to be finely granular. In *gelatin stick cultures* a very thin, smooth, shining layer is developed upon the surface; this has an irregular and deeply dentate margin; the gelatin acquires a greenish tint; no development occurs along the line of puncture. Upon the *surface of agar* a smooth, shining layer with wrinkled margins is formed; the agar acquires a green tint. Upon *potato* a shining, flesh-colored layer extends over a considerable portion of the surface.



FIG. 217.

FIG. 217.—*Bacillus scissus*. $\times 1,000$. (Frankland.)



FIG. 218.

FIG. 218.—*Bacillus scissus*; superficial colony on gelatin plate. $\times 100$. (Frankland.)

334. BACILLUS NO. I. OF FULLES.

Found in the soil.

Morphology.—Bacilli from 1 to 1.2 μ long and 0.6 μ broad.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Spore formation not observed. Upon *gelatin plates* forms spherical, finely granular colonies; under the microscope these have a yellowish-brown centre surrounded by a yellowish marginal zone; later these differences in color are not seen, and to the naked eye the colonies are bluish-white. In *gelatin stick cultures* a thin layer forms upon the surface which is not characteristic. In *bouillon*, at the room temperature, the liquid becomes densely clouded and white flocculi are deposited at the bottom of the tube. Upon *potato* a moist, dirty-yellow layer is developed. Not pathogenic for mice or for guinea-pigs.

335. BACILLUS NO. II. OF FULLES.

Found in the soil.

Morphology.—Small bacilli with round ends; resemble cocci.

Biological Characters.—An *aërobic, non-liquefying* bacillus. Spore formation not observed. Presents oscillatory movements only. Upon *gelatin plates* somewhat elevated, round, grayish-white colonies are developed upon the surface; the deep colonies are small, yellowish in color, and under a low power are seen to be finely granular, brownish-yellow, and with sharp contours. The superficial colonies attain a diameter of two millimetres and have a brownish-yellow color. In *gelatin stick cultures* the growth is at

first nail-shaped; later it spreads out upon the surface. Upon *agar* a thin, white layer with irregular outlines is developed. In *bouillon*, at the room temperature, a dense cloudiness is developed and a thick, slimy deposit accumulates at the bottom of the tube. Upon *potato* a moist, granular, yellowish-white layer is developed. In *milk* an acid reaction is not produced.

336. BACILLUS PHOSPHORESCENS GELIDUS (Forster).

Found in phosphorescent sea fish.

Morphology.—In recent cultures the bacilli appear as small rods with slightly rounded ends and about three times as long as broad; in cultures more than twenty-four hours old they are thicker and nearly oval in form.

Biological Characters.—An *aërobic, non-liquefying* bacillus. Spore formation not positively determined. Grows slowly at the room temperature, and even as low as 0° C.; is killed in a few hours by exposure to a temperature of 35° to 37° C. Cultures freely exposed to the air are phosphorescent in the dark when kept at a temperature between 0° and 20° C.—phosphorescence ceases at 32° C. Upon *gelatin plates*, at the end of forty-eight hours, small, punctiform colonies are developed; under the microscope these are seen to be spherical, grayish-white in color with a greenish shimmer; later granular, yellowish, and with somewhat irregular outlines. In *gelatin stick cultures* a white layer is developed upon the surface; very scanty growth along the line of puncture. Upon *agar* the growth is similar to that on gelatin. Upon *potato* a broad, white layer is developed.

337. BACILLUS SMARAGDINO-PHOSPHORESCENS (Katz).

Obtained from a herring from the fish market at Sydney, New South Wales.

Resembles *Photobacterium phosphorescens* (Cohn) described by Beyerinck, and *Photobacterium Pflügeri* (Ludw.).

Morphology.—Bacilli with somewhat pointed ends, about 2 μ long and 1 μ broad; solitary or in pairs; in recent cultures closely resemble cocci.

Biological Characters.—An *aërobic, non-liquefying, non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of eighteen hours, the deep colonies are seen as grayish points, and the superficial as thin, grayish-white drops; under a low power these are pale-gray with a yellowish tint, finely granular, and transparent near the margins, which are finely dentate; diameter from 0.3 to 0.45 millimetre. The deep colonies, under the microscope, are seen to be oval or lemon-shaped, with a smooth, well-defined contour, and about 0.15 millimetre in diameter; they consist of a broad central portion surrounded by a narrow central and still narrower marginal zone; zone formation not observed in colonies in eight-per-cent gelatin. At the end of twenty days the superficial colonies attain a diameter of two millimetres; they are flat, have irregular outlines, and are composed of a relatively small central portion of a yellowish color, surrounded by a slate-colored marginal zone; the deep colonies at this time (twenty days) are about 0.6 millimetre in diameter and yellowish-white in color—under the microscope straw-yellow. In *gelatin stick cultures* (six-per-cent gelatin) a thin, white line of growth is seen along the line of puncture, and a flat, round, grayish-white layer, with a stearin lustre, is developed upon the surface; this acquires a diameter of about five millimetres. Cultures made by Katz for a year in six-per-cent gelatin gave no evidence of liquefaction, but subsequently the same bacillus, cultivated in six-per-cent gelatin containing 2.7 per cent of sodium chloride, caused liquefaction of the gelatin beneath the surface growth, which gradually extended downward. Growth occurs upon the *surface of agar*, but this is not a very favorable medium. In neutral *bouillon* development occurs, and a diffuse cloudiness is seen, but no growth occurs in simple flesh infusion; when, however, 2.5 per cent of sodium

chloride is added to this it constitutes a favorable medium. Upon the surface of *sterilized milk* a tolerably abundant, sticky, glistening layer of a cream-white color is developed. Upon *cooked egg* a thin, grayish-white, phosphorescent layer is formed. No growth on *potato* which has an acid reaction, but when the acidity is neutralized with a solution of sodium phosphate a thin, brownish-yellow layer is developed. Small quantities of a pure culture added to sea water cause it to exhibit a very decided phosphorescence, and the addition of sodium chloride to culture media favors the growth of the bacillus and its phosphorescent power. Free access of oxygen is essential for the growth and phosphorescence of this species. The color of the light given off by fresh cultures is emerald-green; it is less intense in agar cultures than in cultures in nutrient gelatin, bouillon, or upon cooked fish.

338. BACILLUS ARGENTEO-PHOSPHORESCENS NO. I. (Katz).

Obtained from sea water at Elisabeth Bay, Sydney, New South Wales.

Morphology.—Slender, slightly curved bacilli with pointed ends, about $2.5\ \mu$ long and one-third as thick as they are long; solitary or in pairs; occasionally in long, wavy filaments.

Biological Characters.—An *aërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—not in the incubator at 35°C . Upon *gelatin plates* (six-per-cent gelatin), at the end of twenty hours, the superficial colonies appear as flat, shining, transparent drops from 0.4 to 0.6 millimetre in diameter; under the microscope they are seen to be homogeneous and usually circular in outline, with a dentate margin. The deep colonies are spherical or short oval; they have a smooth, well-defined contour and pale-yellow color. At the end of forty-eight hours the superficial colonies are granular, pale-yellow, with an undulating contour, and about 1.25 millimetres in diameter; at the same time the deep colonies are pea-yellow and uniformly granular; later the deep colonies present three well-defined zones; the superficial colonies also, at the end of twenty days, present two or three distinct zones; they attain a diameter of about three millimetres. In eight-per-cent gelatin the deep colonies, at the end of two days, are oval, and under the microscope show three well-defined zones; the superficial colonies present the same appearance in from four to seven days, at which time they have a diameter of three millimetres—later as much as seven millimetres. In *gelatin stick cultures* (six per cent) development occurs upon the surface as a flat, shining, usually circular layer, about one cubic centimetre in diameter, and of a greenish-yellow or wax-like color. No growth occurs in acid gelatin. In old gelatin cultures containing 2.7 per cent of sodium chloride liquefaction sometimes occurs at a temperature approaching that at which the gelatin would become liquid. In *bouillon* a diffuse cloudiness is produced and a film is formed upon the surface; no growth occurs in flesh infusion without the addition of peptone and salt, but the addition of 2.5 per cent of sodium chloride to neutral flesh infusion makes it a favorable medium. Upon the surface of *sterilized fish* a pale-yellow, glistening, sticky layer is developed. Upon *cooked egg* a thin, grayish-white layer. No growth on *potato*. Phosphorescence depends upon the presence of certain salts—especially sodium chloride—in the culture medium, and upon the free access of oxygen. In bouillon cultures, when a mycoderma has formed upon the surface this shows phosphorescence, while the liquid below does not. The light given off is of a silver-white color, and a recent culture upon the surface of gelatin gives sufficient light to enable one to determine the time from a watch in a dark room.

339. BACILLUS ARGENTEO-PHOSPHORESCENS NO. II. (Katz).

Obtained from phosphorescent pieces of fish found in the market at Sydney, New South Wales.

Morphology.—Bacilli with round ends, about $2.7\ \mu$ long and $0.67\ \mu$ broad; occasionally form short filaments.

Biological Characters.—An *aërobie*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—not so well at 32° to 34° C. Upon *gelatin plates* (six per cent), at the end of twenty-four hours at 18° to 20° C., the superficial colonies have a diameter of 0.5 millimetre and resemble little drops of stearin; they have a circular and sharply defined contour, are homogeneous, and of a pale yellowish-gray color; at the end of forty-eight hours they are grayish-yellow, finely granular, with irregular contour, and about one millimetre in diameter. The deep colonies are much smaller, have a greenish-yellow color, granular contents, and a smooth, well-defined contour. The deep colonies which come to the surface spread out to form a bluish-gray, shining disc, which may attain a diameter of six millimetres. In *gelatin stick cultures* scanty development along the line of puncture, and a grayish-white, glistening layer of limited extent upon the surface. Upon the surface of ten-per-cent gelatin a bluish-gray band with cloudy margins along the impfstrich. In *bouillon cultures* the liquid is diffusely clouded, but no mycoderma is formed upon the surface. Upon *sterilized fish* a shining, sticky layer of a yellowish color—here and there lemon-yellow; the cultures have a silver-white phosphorescence in the dark, and a small quantity added to a considerable quantity of sea water causes this to be phosphorescent. The presence of certain salts, and especially of sodium chloride, and free contact with oxygen, is favorable to the growth of this bacillus and to the development of phosphorescence.

340. BACILLUS ARGENTEO-PHOSPHORESCENS NO. III. (Katz).

Obtained from a fragment of cuttlefish which was phosphorescent, at Sydney, New South Wales.

Morphology.—Resemble *Bacillus argenteo-phosphorescens* No. II., but the rods are a little thinner and are motile; frequently united in pairs and occasionally form short filaments.

Biological Characters.—An *aërobie*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media—not so well at 32° to 34° C. Upon *gelatin plates*, at the end of twenty-four hours at 18° to 20° C., the superficial colonies appear as white scales with irregular outlines, sometimes wrinkled, and marked with fine lines or furrows, about 0.45 millimetre in diameter. The deep colonies are spherical, oval, or lemon-shaped, homogeneous, and greenish-yellow in color; at the end of forty-eight hours they appear finely granular and divided into two zones. At the end of a week the superficial colonies attain a diameter of about three millimetres; they are bluish-gray and cloudy in appearance, quite thin, and have a notched or dentate margin. Upon the surface of *gelatin stick cultures* a layer is developed which extends nearly to the walls of the tube, and which becomes very thin at the margins. Development occurs upon the *surface of agar*, but this is not a very favorable medium. In *bouillon* a diffuse cloudiness is produced and a mycoderma forms upon the surface. The addition of 2.5 per cent of sodium chloride to bouillon or other media is favorable to the growth and phosphorescence of this bacillus, as of those previously described. Upon *sterilized fish* a viscid, glistening, yellowish layer is developed. No growth upon acid *potato*. The cultures give off a silver-white, phosphorescent light. Bouillon cultures commence to give off light from the surface at the end of four or five days, and at the end of eight days the floating mycoderma may give a light by which the time can be discerned, in a dark room, from a watch; the light is a "bluish-greenish white."

*D. Non-chromogenic, Liquefying Bacilli.*341. *BACILLUS CYANEO-PHOSPHORESCENS* (Katz).

Obtained from sea water at Little Bay, near Sydney, New South Wales. Very nearly related to *Bacillus phosphorescens* of Fischer (Katz).

Morphology.—Bacilli with round ends, about $2.6\ \mu$ long and $1\ \mu$ thick; solitary or in pairs; occasionally grow out into long filaments.

Stains by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation not observed. The cultures give off a bluish phosphorescence which has a faint greenish tint. Grows in the usual culture media at the room temperature—best at 26°C .; very scanty growth upon nutrient agar at 32° to 34°C . Upon *gelatin plates*, at the end of eighteen hours, colonies are already visible; those upon the surface and those in the interior of the gelatin are of about the same dimensions— 0.25 to 0.4 millimetre; under the microscope they are seen to be finely granular, have a sharply defined, smooth contour and a dark-gray color; the superficial colonies are finely granular and pale yellowish-gray in color. At the end of forty-eight hours the superficial colonies are surrounded by a broad girdle of liquefied gelatin, at the bottom of which they lie in contact with the glass plate; they are of a dirty brownish-yellow color and irregular in outline; the liquefied gelatin is pale-gray or yellowish-gray by transmitted light, and contains here and there granular masses scattered through the more finely granular structure. At this time the deep colonies are yet well defined, have irregular outlines, and are of a dirty yellowish-brown color; they have a diameter of about 0.3 to 0.5 millimetre, and are surrounded by a zone of liquefied gelatin about 0.05 to 0.1 millimetre in diameter; this is finely granular, light-brown or light-gray in color, and marked by delicate radial striations. When the colonies are crowded upon a plate liquefaction may be complete at the end of eighteen hours; the liquefied gelatin gives off a peculiar odor. In *gelatin stick cultures* (six-per-cent gelatin) liquefaction commences beneath the surface growth, and at the end of forty-eight hours a shallow cavity of watch-glass form is seen which has a diameter of about five millimetres—temperature of 20° to 22°C .; at the bottom of this a grayish-white layer is seen, and below this a line of development along the track of the inoculating needle; this is surrounded by a narrow zone of liquefaction. At the end of three or four days liquefaction at the surface reaches the walls of the test tube; when liquefaction is complete a yellowish, viscid mass is seen at the bottom of the tube and a mycoderma upon the surface; the gelatin is at first diffusely clouded and later becomes transparent; it has a yellowish color, which gradually changes to reddish-brown. In six-per-cent gelatin containing 2.7 per cent of sodium chloride development is especially abundant and rapid, and short, radiating processes are given off into the gelatin in advance of liquefaction. In *bouillon* diffuse cloudiness occurs and a mycoderma forms upon the surface. No development occurs in simple flesh infusion, but the addition of 0.5 per cent of sodium chloride to this constitutes a medium in which growth occurs—still better 2.5 per cent. Upon *sterilized fish* a glistening, sticky layer of a yellowish or yellowish-brown color, in the thicker places, is developed. Phosphorescence depends upon the free access of oxygen and the presence of certain salts, especially sodium chloride. A very minute quantity of a culture added to sea water causes it to exhibit phosphorescence. Cultures upon the surface of agar give sufficient light to enable one to distinguish printed letters in a dark room. No growth upon *potato*.

342. *BACILLUS ARGENTEO-PHOSPHORESCENS LIQUEFACIENS* (Katz).

Obtained from sea water at Bondi Bay, near Sydney, New South Wales. Resembles *Photobacterium luminosum* of Beyerinck.

Morphology.—Straight or slightly curved bacilli with round ends; about two μ long and one-third as broad; grow out into filaments of various lengths.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation not observed. Cultures give off a silvery phosphorescence, which is less intense than with the previously described species. Grows at the room temperature in the usual culture media—best at 25° C.; does not grow in the incubating oven at 34° C. Upon *gelatin plates*, at the end of twenty-four hours at the room temperature, small, hyaline discs are developed, which under the microscope are seen to be finely granular and light-brown in color; they are irregularly circular in outline and about 0.7 millimetre in diameter; the deep colonies are considerably smaller, mulberry-like in structure, and straw-yellow in color. At the end of forty-eight hours shallow liquefaction has occurred beneath the superficial colonies, in watch-glass form, and about two millimetres in diameter; under the microscope a central mass of a straw-yellow color is seen, around this a narrow, light-brown zone with granular contents, and outside of this a broader peripheral zone, from which fine, radiating outgrowths are given off into the non-liquefied gelatin. At the same time (forty-eight hours) the deep colonies have a diameter of 0.3 to 0.45 millimetre and a more or less polygonal contour; they are straw-yellow in color and consist of a finely granular central mass, surrounded by a slender, marginal zone which is marked by radial striations. After complete liquefaction of the gelatin the colonies, which remain attached to the glass plate, have a lemon-yellow color. In *gelatin stick cultures* (six per cent) liquefaction occurs beneath the superficial layer which is developed, in form of a shallow watch glass, and gradually extends in diameter and depth; growth also occurs along the line of puncture, and the cultures resemble those of *Bacillus cyaneo-phosphorescens*, but with less rapid development and liquefaction of the gelatin; also without the formation of hair-like outgrowths into the non-liquefied gelatin. The addition of 2.7 per cent of sodium chloride is favorable for the development of this as for the previously described species of phosphorescent bacilli; on the other hand, the addition of two per cent of glucose exercises a restraining influence upon the growth of all the species studied by Katz. In *bouillon* a diffuse cloudiness is produced by the growth of this bacillus, and a mycoderma is formed upon the surface. No growth occurs in simple meat infusion, but an abundant development when 2.5 per cent of sodium chloride is added to this. Upon *sterilized fish* a shining, sticky, yellowish-gray layer is developed. No growth upon *potato*.

343. *BACILLUS PHOSPHORESCENS INDICUS* (Fischer).

Found in sea water from the Gulf of Mexico.

Morphology.—Bacilli with rounded and pointed ends, from two to three times as long as broad; length from one-sixth to one-quarter the diameter of a red blood corpuscle; solitary or in pairs; also in short filaments.

Stains readily with the aniline colors, but unstained places are often seen in the interior of the rods.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Cultures, especially upon animal substances and in presence of certain soda salts, exhibit a decided phosphorescence in the dark; this depends upon free access of air, and is most marked at a temperature of 25° to 30° C. It is no longer manifested at a temperature of 0° C., and is neutralized by putrefaction. Grows in the usual culture media at the room temperature—not so well in the incubating oven. Upon *gelatin plates*, at the end of thirty-six hours, small, round, grayish-white, punctiform colonies are developed; under a low power these are seen to be spherical, with well-defined outlines, and have a sea-green color with a pink shimmer; later they become granular, have a wavy outline and a dirty-yellow color. In *gelatin stick cultures*, at the end of four days, a grayish-white line of

growth is seen along the track of the inoculating needle, and at the surface a cup-shaped depression, the size of a hempseed, which contains air; in older cultures the gelatin is liquefied near the surface and a thin, dirty-yellow film swims upon it. Upon the *surface of agar* a grayish-white layer is developed. Upon *potato*, at 15° to 20° C., a thin and broad white layer. Upon *blood serum* a narrow, grayish-white stripe, which extends to a tolerably deep channel, with irregular margins, from 0.5 to 1 centimetre wide; this is lined with a slimy, grayish-white growth. *Cooked fish* or flesh constitutes a favorable medium for the growth of this bacillus. By means of the phosphorescent light given off by cultures of this bacillus, Fischer has succeeded in making photographs not only of the cultures, but of a watch dial placed between two cultures—an exposure of twenty-four hours' duration and a very sensitive dry plate were required to accomplish this.

344. BACILLUS PHOSPHORESCENS INDIGENUS (Fischer).

Found in sea water from the harbor at Kiel and upon phosphorescent herring.

Morphology.—Bacilli with round and slightly pointed ends; somewhat shorter than *Bacillus phosphorescens Indicus*, but of the same thickness—from 1.3 to 2.1 μ long and 0.4 to 0.7 μ broad; solitary or in pairs; may grow out into filaments.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Cultures give off a bluish-white, phosphorescent light—not so intense as that from *Bacillus phosphorescens Indicus*; phosphorescence depends upon free access of oxygen. Sea water to which a small amount of a culture is added is phosphorescent in the dark. Spore formation not observed. Grows at the room temperature in the usual culture media—more slowly than *Bacillus phosphorescens Indicus*. Grows at 5° to 10° C., and even below; at a temperature of 32° C. development still occurs, but the cultures do not exhibit phosphorescence. Upon *gelatin plates* the gelatin is depressed about the small spherical colonies, and at the end of a week cylindrical cavities filled with air, and not more than one millimetre in diameter, are formed in the gelatin; at the bottom of these, on the surface of the plate, the colonies are seen; these are the size of a pin's head, thin, disc-formed, and dirty yellow in color; under a low power very young colonies are seen to be circular, with well-defined margins, and of a pale sea-green color; here and there reddish-shimmering granules are seen in the otherwise homogeneous contents; the older colonies are made up of irregular, dirty yellowish-gray masses. In *gelatin stick cultures*, at the end of a week, a conical cavity forms near the surface, which is filled with air and is lined with a thin, friable growth; at the surface the mouth of the cone measures about two millimetres in diameter; this cavity increases in dimensions without containing any liquefied gelatin; in old cultures it may be three to five millimetres in diameter and two to three centimetres deep, the walls being covered with a thin layer of bacilli, and a mass of the same accumulating at the bottom. No growth occurs upon *potato* or upon *blood serum*.

345. BACILLUS CIRCULANS (Jordan).

Found occasionally in water from the Merrimac River.

Morphology.—Bacilli with round ends, from 2 to 5 μ long and about 1 μ broad; usually solitary, but sometimes in loosely connected chains of three or four elements.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, motile* bacillus. Forms oval spores, which are located at the ends of the rods and are of about the same diameter as these. Grows in the usual culture media at the room temperature—better at 37° C. Upon *gelatin plates*, at the end of two days, round, brownish colonies become visible; under a low power the liquid contents of these colonies are seen to be in mo-

tion, owing to the active movements of the individual bacteria; the motion may be compared to the circulation of protoplasm in a cell; this is seen after forty-eight hours of growth, but ceases, as a rule, on the third day, at which time the contents are seen to be coarsely granular; a round, deep, even depression is formed in the gelatin, which at the end of several weeks may have a diameter of a centimetre; the contour of the colonies is usually smooth, but may be somewhat lobate and irregular. In *gelatin stick cultures* development is slow along the upper part of the line of puncture, and



FIG. 219. — *Bacillus circulans*, from an agar culture five days old. $\times 1,000$. (Jordan.)

a conical-shaped cavity is formed, at the bottom of which a precipitate accumulates, while above the conical cavity is empty owing to the slowness of growth and drying-up of the liquefied gelatin. The upper part of the cone often has a somewhat ringed appearance. This bacillus grows well in a slightly acid medium. Upon the *surface of agar* development occurs along the line of puncture, and a very thin, translucent layer is formed upon the surface. Upon *potato* a slow and scanty growth having about the same color as the cut surface of the potato. In *milk* a slightly acid reaction occurs and the casein is slowly precipitated; after cultivation for several months the bacillus no longer caused coagulation of milk. In *bouillon* a cloudiness is seen at the end of three or four days, and a considerable slimy precipitate is formed; no film forms upon the surface. Nitrates are slowly reduced to nitrites by this bacillus.

346. *BACILLUS SUPERFICIALIS* (Jordan).

Found frequently in sewage at Lawrence, Mass.

Morphology.—Bacilli with round ends, about 2.2μ long and 1μ broad; solitary or in pairs.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media—better at 37°C . Upon *gelatin plates* colonies become visible at the end of forty-eight hours; under a low power they are seen to be divided by irregular lines into angular lumps, giving a cracked appearance to the whole colony, which is irregularly spherical in form. Upon coming to the surface the colonies spread out to form a round, finely granular disc, which to the naked eye looks like a projecting, translucent drop; this slowly increases in dimensions, and the surrounding gelatin is slowly liquefied; after some days the colony has an opaque, yellowish-brown centre and a translucent edge. In *gelatin stick cultures* there is a very scanty growth along the line of puncture and more abundant development upon the surface; liquefaction proceeds slowly, and by the end of ten days may reach the walls of the test tube. This bacillus grows well in acid gelatin. Upon the *surface of agar* a moist, lustrous, gray, translucent layer is developed; at the end of several weeks this growth is still smooth and glistening, and has a light-brown tint. Does not grow upon *potato*. Does not cause coagulation of *milk*, which, however, acquires a slightly acid reaction. In *bouillon* a diffuse cloudiness is slowly developed, and after some time a scanty white sediment is seen; no film forms upon the surface.

347. *BACILLUS RETICULARIS* (Jordan).

Found in water at Lawrence, Mass.

Morphology.—Bacilli with slightly rounded ends, about 5μ long and 1μ broad; often united in chains of eight to ten elements.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Motion slow and sinuous. Grows at the room temperature in the usual culture media—much better at 37° C. Upon *gelatin plates* the deep colonies send out long, spiral filaments, which give a hazy appearance to the colony; under a low power the colony, surrounded by radiating filaments, resembles a jelly-fish with streaming tentacles; the surrounding gelatin is slowly liquefied. On coming to the surface the colony spreads out as an irregular expansion, under which the gelatin is slowly liquefied, and at the same time dried out so as to form shallow, cup-shaped depressions; the surface of these cavities is mottled in appearance, as if covered with a fine, irregular reticulation. In *gelatin stick cultures*, at the end of two days, the growth at the surface resembles a cup with flaring edges; liquefaction occurs slowly, and the liquefied gelatin is dried by evaporation, so that the cup-shaped cavity gradually increases in dimensions; it is lined with a reticulated growth similar to that seen in the colonies upon gelatin plates; at the end of three days fine filaments begin to grow out from the line of puncture, but these do not reach any considerable length. This bacillus exhibits a scanty development under a mica plate. Upon the *surface of agar* a prominent, dull, dry layer is slowly developed. Upon *potato* a white, dull, dry layer is developed at the end of two days, and at the end of five days this has a characteristic woolly appearance. *Milk* acquires an acid reaction and is slowly coagulated—fifteen to twenty days at the room temperature. *Bouillon* slowly becomes turbid and a slight viscid sediment is formed. Nitrates are rapidly reduced to nitrites by this bacillus.

348. BACILLUS HYALINUS (Jordan).

Found in large numbers in the sand of Tank 13 at Lawrence, Mass., "at a time when the tank was nitrifying well."

Morphology.—Bacilli with round ends, from 3.6 to 4 μ long and 1.5 μ broad; usually united in short chains.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, actively motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—better at 37° C. Upon *gelatin plates* the colonies are visible at the end of twenty-four hours; they are seen to consist of a dark central nucleus surrounded by a broad, translucent zone, which gives the colonies a hazy appearance; under a low power the interior is seen to present a coarsely fibrillar appearance, with short fibrils radiating from the edge. In two days the colonies attain a diameter of about one and one-half centimetres; they are round in contour, with a distinct, opaque, yellowish margin, from which radiating fibrils are given off; the interior is slightly translucent. In *gelatin stick cultures*, at the end of two days, a long and narrow, funnel-shaped growth is seen; the gelatin is rapidly liquefied and is at first clouded, with a white deposit at the bottom of the funnel; later a lustrous and tenacious white layer is seen upon the surface and a slight flocculent deposit at the bottom, while the liquefied gelatin between is entirely transparent. This bacillus grows well in acid gelatin. Upon the *surface of agar* a dry, grayish, spreading growth is rapidly developed; when four or five days old small, warty projections are seen; growth also occurs along the line of puncture in agar stick cultures. Upon *potato*, at the end of two days, the growth is just visible; at the end of four days a dry, whitish-gray, spreading layer is developed; later small protuberances are seen upon the surface of this. In *milk* a strongly acid reaction with coagulation of the casein is produced in seven days. In *bouillon* a diffuse cloudiness is quickly produced; a viscid sediment is formed and a mycoderma forms upon the surface. This bacillus reduces nitrates vigorously and rapidly.

349. BACILLUS CLOACÆ (Jordan).

Isolated from sewage at Lawrence, Mass.—"one of the most common bacteria in sewage."

Morphology.—Short oval bacilli with round ends, from 0.8 to 1.9 μ long and from 0.7 to 1 μ broad; frequently united in pairs.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *actively motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—better at 37° C. Upon *gelatin plates*, at the end of one or two days, spherical, yellowish colonies are seen; upon coming to the surface these form a slight bluish expansion with irregularly notched edges, and liquefaction of the gelatin quickly occurs; under the microscope the colonies are seen to have an opaque centre surrounded by a translucent zone and a darker margin; the interior is finely granular; liquefaction of the entire plate occurs within three or four days. In *gelatin stick cultures* development occurs along the line of puncture, and liquefaction rapidly occurs; later an iridescent film is seen upon the surface of the gelatin and an abundant flocculent, whitish sediment at the bottom of the tube. This bacillus grows well in slightly acid gelatin. Upon the *surface of agar* a moist, slimy, porcelain-white layer is developed; an abundant growth also occurs along the line of puncture in agar stick cultures. Upon *potato* a prominent, yellowish-white layer is quickly formed. In *milk* coagulation occurs in about four days, with a strongly acid reaction. In *bouillon* a diffuse cloudiness is seen at the end of two days, and at the end of ten to fourteen days an abundant whitish sediment is seen; a slight film forms upon the surface; this falls to the bottom when the tube is disturbed; the bouillon is still clouded at the end of two or three weeks. This bacillus reduces nitrates, in bouillon, vigorously.

350. BACILLUS DELICATULUS (Jordan).

Found in the water supply at Lawrence, Mass.

Morphology.—Bacilli about 2 μ long and 1 μ broad; often united in pairs or in short chains.

Biological Characters.—An *aërobic*, *liquefying*, *actively motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—somewhat better at 37° C. Upon *gelatin plates* the young colonies are seen as whitish, homogeneous spheres with a regular, radiating margin; at the end of two days liquefaction of the surrounding gelatin occurs. In *gelatin stick cultures* liquefaction progresses rapidly along the line of puncture and is complete in about seven days; a thick, whitish layer is then seen upon the surface, and an abundant flocculent, brownish deposit at the bottom of the tube; the liquefied gelatin remains clouded. Grows well in slightly acid gelatin. Upon the *surface of agar* a wrinkled, grayish layer is developed, which later becomes porcelain-white and glistening; development also occurs along the line of puncture in *agar stick cultures*. Upon *potato* a thin, spreading, gray layer is developed. *Milk* acquires a strongly acid reaction and is coagulated. *Bouillon* quickly becomes clouded; a white film forms upon the surface and a white deposit is seen at the bottom of the tube. Nitrates are quickly and completely reduced to nitrites by this bacillus. Does not grow at a temperature below 15° C., and quickly dies out in artificial culture media—in a few weeks.

351. BACILLUS AQUATILIS (Frankland).

Found in water—often almost the only microorganism in water from a deep well in the chalk formation at Kent.

Morphology.—Bacilli about 2.5 μ in length; grow out into filaments of 17 μ or more in length—resembles *Bacillus arborescens* (Frankland).

Biological Characters.—An *aërobic*, *liquefying*, *motile* bacillus. Exhibits oscillatory movements only. Spore formation not observed. Grows very slowly in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are at first smooth in outline, later the mar-

gin is irregular; when the colony reaches the surface a very slow liquefaction of the gelatin commences and the appearance of the colonies becomes very characteristic. From the yellowish-brown centre twisted bundles of filaments are given off which are at first also of a yellowish-brown color, but gradually become colorless toward the periphery. In *gelatin stick cultures* development is extremely slow; upon the surface a small, yellowish mass is formed at the point of puncture, while the line of inoculation is scarcely visible; liquefaction occurs later and the liquefied gelatin is clouded; when liquefaction has once commenced it progresses more rapidly. Upon the *surface of agar* a glistening, yellowish layer is developed which extends but little beyond the *impfstrich*. In *bouillon* a diffuse cloudiness is produced and a whitish sediment is seen at the bottom of the tube; no film is formed upon the surface. Upon *potato* scarcely any development occurs—a faint yellowish stripe along the *impfstrich* only. Reduces nitrates with formation of ammonia.

352. BACILLUS DIFFUSUS (Frankland).

Found in the soil.

Morphology.—Bacilli about 1.7μ long and 0.5μ broad; solitary or in pairs; also grow out into long, flexible filaments.

Biological Characters.—An *aërobic, liquefying* bacillus. Exhibits rotatory and oscillatory movements only. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the superficial colonies after a time have a very characteristic appearance; they extend from the original centre as a broad, thin, bluish-green layer. Under a low power the deep colonies resemble colonies of the cholera spirillum; they are nearly spherical, coarsely granular, and have somewhat jagged margins; later the margins are still more irregular and finely dentate, while the surface near the margin appears coarsely granular; when the colonies come to the surface the centre is no longer well defined, but remains granular, while about it a very characteristic surface growth occurs. In *gelatin stick cultures* the development is almost limited to the surface, upon which a smooth, thin, shining, somewhat greenish-yellow layer is formed; liquefaction of the gelatin beneath this progresses very slowly. Upon the *surface of agar* a very thin, shining, smooth layer of a feebly yellow or cream color is developed. In *bouillon* a diffuse cloudiness is developed and a greenish-yellow sediment is formed, while some flocculi may float upon the surface, which, however, do not constitute a film. Upon *potato* a thin, smooth, shining, faintly greenish-yellow layer is formed.

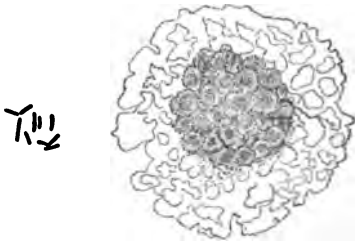


FIG. 220.

FIG. 221.

FIG. 220.—*Bacillus diffusus*, from a gelatin culture. $\times 1,000$. (Frankland.)

FIG. 221.—*Bacillus diffusus*; superficial colonies in nutrient gelatin. $\times 100$. (Frankland.)

353. BACILLUS LIQUIDUS (Frankland).

Found in river water from the Thames; very common.

Morphology.—Short and thick bacilli with round ends; usually in pairs, of which the length varies from 1.5 to 3.5μ ; differ greatly in dimensions.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates* the deep colonies, under a low power, are seen to be spherical, with smooth outlines; when liquefaction commences

the margin becomes somewhat granular and jagged, while the centre remains opaque; development and liquefaction of the surrounding gelatin progress rapidly, so that the colonies soon coalesce. In *gelatin stick cultures* development occurs along the entire line of puncture, and a broad funnel of liquefied gelatin is formed in the course of a few days; this is clouded and contains numerous flocculi; later the surface is covered with a thin film, which sinks to the bottom when the test tube is shaken. Upon the *surface of agar* development also occurs rapidly, forming a smooth, shining layer on both sides of the impfstrich. In *bouillon* a diffuse cloudiness is developed; an abundant deposit collects at the bottom of the tube, and a film is formed upon the surface. Upon *potato* a thick, flesh-colored layer covered with protuberances and having a dull, moist surface. This bacillus reduces nitrates with the production of nitric acid

354. BACILLUS VERMICULARIS (Frankland).

Found in water from the river Lea.

Morphology.—Bacilli with round ends, from 2 to 3 μ long and 1 μ broad; may grow out into "worm-shaped" filaments. Upon potato oval spores are formed; these are about 1.5 μ long and 1 μ broad; they are formed in the centre of the rods, and remain attached to each other in chains of considerable length.

Biological Characters.—An *aërobie*, *liquefying* bacillus. Exhibits oscillatory movements only. Forms oval spores. Grows slowly in the usual culture media at the room temperature.



FIG. 222.—*Bacillus vermicularis*; a, from a gelatin culture; b, spore-bearing filaments from a potato culture. $\times 1,000$. (Frankland.)

Upon *gelatin plates* the deep colonies have a somewhat irregular contour; upon the surface flat colonies are developed, which have irregular margins composed of wavy bundles of bacilli closely crowded together; the centre of the superficial colonies is rough and wrinkled; later liquefaction slowly occurs and the colonies sink beneath the surface of the gelatin. In *gelatin stick cultures* a moist, shining, gray layer with dentate margins is developed

upon the surface; this does not extend far from the point of puncture; a scanty growth is seen along the line of puncture; after some time the gelatin commences to liquefy beneath the surface growth. Upon the *surface of agar* a smooth, shining layer of a gray color is slowly developed. In *bouillon* the liquid remains clear, and a white, flocculent growth is seen at the bottom of the tub. Upon *potato* a thick, flesh-colored layer with irregular outlines. Reduces nitrates to nitrites.

355. BACILLUS NUBILUS (Frankland).

Found in filtered London water.

Morphology.—Bacilli about 3 μ long and 0.3 μ broad; solitary or in short chains; grow out into long filaments in bouillon cultures—these may be twisted in spiral form; upon potato the short bacilli are often curved, and the long filaments may have a spiral form.

Biological Characters.—An *aërobie* and *facultative anaërobie*, *liquefying* bacillus. Exhibits rotatory movements without change of location. Spore formation not observed. Grows slowly at the room temperature in the usual culture media. Upon *gelatin plates* the colonies are very characteristic; at the end of forty-eight hours small clouded spots are seen, which are not sharply defined; under the microscope these are seen with difficulty by transmitted light; on the third day, when the gelatin commences to be

softened, these colonies are seen to consist of a network of interlaced filaments; at the centre a thicker portion is often seen in the cloud-like colonies of interlaced threads; liquefaction progresses rapidly, the colonies become confluent, and soon the plate is destroyed. In *gelatin stick cultures* the gelatin is liquefied upon the surface, but liquefaction does not progress rapidly; the liquefied gelatin is clouded, and a yellowish deposit accumulates at the bottom of the liquefied gelatin; along the line of puncture, almost to the surface, no growth is seen, but around it a row of flat, horizontal rings are developed, the diameter of which increases from above downward; the lower end of the line of puncture is uniformly clouded; the rings described are made up of delicate, cloud-like masses; the culture somewhat resembles that of the bacillus of mouse septicæmia; liquefaction progresses slowly, but finally the entire amount of gelatin is liquefied. Upon the *surface of agar* a thin, opalescent, bluish-white layer is formed, the fringed margins of which show a violet fluorescence. In *bouillon* a diffuse cloudiness is developed, and a dirty-white deposit collects at the bottom of the tube, while a very thin film collects upon the surface; this falls to the bottom when the tube is shaken. Upon *potato* the growth is almost invisible and of a very faint yellow color; it extends, however, over a great portion of the surface.

356. BACILLUS PESTIFER (Frankland).

Found in the air.

Morphology.—Bacilli with round ends, $2.3\ \mu$ long and $1\ \mu$ broad; often grow out into filaments.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Grows slowly at the room temperature. Upon *gelatin plates* forms colonies resembling those of *Bacillus vermicularis*, but the deep colonies are more regular and the superficial colonies have a smooth centre. Upon the *surface of agar* a shining, transparent layer with dentate margins. Upon *potato* a thick, irregular, flesh-colored layer.

357. BACILLUS FILIFORMIS (Tils).

Found in water.

Morphology.—Bacilli with round ends, $4\ \mu$ long and $1\ \mu$ broad; usually united in chains, which may consist of as many as ten segments which are indistinctly separated from each other.

Biological Characters.—An *aërobic, liquefying* bacillus. Exhibits oscillatory movements only. Forms, in potato cultures, large oval spores. Upon *gelatin plates* grayish-white colonies are developed which have a marbled structure; under a low power the deep colonies are seen to be finely granular and irregular in outline; the superficial colonies have a dentate margin, which is composed of many bundles of bacilli closely crowded together; the centre is somewhat rough, elevated, and granular; the colonies are colorless at the margin, but have a yellowish color at the centre; at the end of eight to ten days liquefaction commences and the colonies sink slowly beneath the surface. In *gelatin stick cultures* a moist layer with deeply dentate margins forms around the point of puncture; the gelatin is slowly liquefied and thick, flocculent masses of bacilli accumulate at the bottom. In *bouillon* growth occurs chiefly upon the surface in the form of a firm mycoderma. Upon the *surface of agar* a white layer is developed like that upon gelatin. Upon *potato* a thick, slimy, dirty-white layer, which later becomes dry and acquires a gray or brownish color. In *milk* coagulation occurs at the end of thirty-six hours, and an odor of putrefaction is perceived.

358. BACILLUS DEVORANS (Zimmermann).

Found in well water.

Morphology.—Bacilli with round ends, from 0.99 to $1.2\ \mu$ long and about $0.74\ \mu$ thick; solitary, in pairs, or occasionally in short chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *actively motile* bacillus. Spore formation not observed. Grows at the room temperature in the usual culture media. Upon *gelatin plates* the deep colonies appear as small, white spheres; the superficial colonies are seen as round, white, not homogeneous masses at the bottom of a funnel of liquefied gelatin; under a low power they have a granular-thready structure and yellowish-gray color, and the margins are surrounded by thread-like processes of various lengths. In *gelatin stick cultures* growth is visible along the line of puncture at the end of twenty-four hours; on the second or third day an air bubble is seen at the upper portion and a whitish growth below; the funnel-shaped cavity gradually extends in dimensions without a trace of liquid being seen, and the growth is distributed upon the walls and at the bottom of this cavity; often liquefaction occurs. Upon the *surface of agar* a thin, uniform, gray layer is developed, which covers the entire surface at the end of two or three days. No growth upon *potato*.

359. BACILLUS GRACILIS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Long bacilli with round ends, usually more or less curved, from 2.4 to 3.6 μ long and about 0.77 μ broad; grow out into long filaments, which are bent at an angle or present several wave-like curves.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* bacillus. Forms oval spores 1.88 μ long and 1.3 μ broad. Exhibits rotatory and oscillatory movements only. Grows slowly at the room temperature—not in the incubating oven. Upon *gelatin plates* the deep colonies appear as small, grayish-white spheres, which at first are well defined, but later have very indistinct outlines; under a low power they are first seen as sharply defined, pale-yellow discs, which are gradually surrounded by numerous thread-like outgrowths, which after a time form a thick network. Upon the surface there is no development or small, yellowish-gray, drop-like colonies are seen; at the end of five days these attain a diameter of four to six millimetres and are circular in outline; the centre is round, nebulous, and bluish-gray, and outside of this one or two concentric, nebulous rings are seen. In *gelatin stick cultures* there is a scanty growth upon the surface, often appearing as an opalescent film, or the superficial layer of the gelatin is penetrated by radiating lines given off from centres of growth which do not extend above the surface. Along the line of puncture a row of whitish discs is developed which are largest above; at the end of three to five weeks the upper portion of the gelatin is liquefied to some extent. Upon the *surface of agar* a thin, irregular layer of a bluish-white color is developed; an abundant growth occurs along the line of puncture in agar stick cultures. Upon *potato* the development is very scanty.

360. BACILLUS GUTTATUS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli with round ends, from 1 to 1.13 μ long and 0.93 μ broad; at first solitary or in pairs, later united in chains containing several elements.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *actively motile* bacillus. Appears to form spherical spores (?). Grows best at the room temperature. Upon *gelatin plates* the deep colonies are small, grayish-white spheres; under a low power these are seen to be finely granular and have a gray or bluish-brown color. The superficial colonies appear as bluish-gray drops; under the microscope a brownish shimmer is observed at the centre, while the sharply defined margins are colorless and are distinguished from the surrounding gelatin only by being less transparent; later the outline sometimes becomes irregular. In *gelatin stick cultures* an irregular, bluish-white, shiuing layer is developed, the surface of

which is frequently opalescent; an abundant development occurs along the line of puncture, consisting of a series of spherical colonies; liquefaction occurs after a considerable time—four weeks. Upon the *surface of agar* a thin, grayish-white layer is developed along the impfstrich. Upon *potato* a tolerably abundant slimy, yellowish-green layer.

361. BACILLUS IMPLEXUS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli with blunt ends, about $2.5\ \mu$ long and $1.15\ \mu$ broad; form long, jointed filaments.

Biological Characters.—An *aërobic, liquefying, non-motile* bacillus. Forms oval spores about $1.6\ \mu$ long and $0.95\ \mu$ thick. Grows at the room temperature—more quickly at 30°C . Upon *gelatin stick cultures*, at the end of twenty-four to thirty-six hours, whitish, punctiform colonies become visible; under a low power these are seen to be granular, opaque, and irregular in outline—more transparent toward the margin; under a higher power ($1:175$) filaments are seen which are interwoven; these grow out from the margins and again become contorted and interlaced; at the end of three days the round colony sinks into the liquefied gelatin and appears then as an interlaced mass of fine white filaments. In *gelatin stick cultures* growth is visible along the line of puncture at the end of twenty-four hours, and at the end of seventy-two hours bundles of short threads radiate into the gelatin in all directions; liquefaction soon occurs, which quickly reaches the walls of the test tube at the surface; a thick, white layer forms upon the surface of the liquefied gelatin, and it is filled below with white flocculi. Upon the *surface of agar* a tolerably thick, white layer is formed which has a dull, shagreen-like surface and soon becomes wrinkled. Upon *potato* a yellowish or greenish-white, felt-like layer is formed.

362. BACILLUS PUNCTATUS (Zimmermann).

Common in the Chemnitz water supply.

Morphology.—Bacilli from 1 to $1.60\ \mu$ long—when in rapid multiplication—and $0.77\ \mu$ broad; solitary, in pairs, or in chains containing several elements. Do not stain readily with the usual aniline colors.

Biological Characters.—An *aërobic, liquefying, actively motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature—still better at 30°C . Upon *gelatin plates* the colonies, on the third day, already have a diameter of twelve millimetres and have caused a saucer-shaped liquefaction of the gelatin; in the grayish-blue liquid whitish, punctiform collections of bacteria are seen, which are often united with each other by whitish strings. In *gelatin stick cultures* liquefaction occurs in stocking shape; the liquefied gelatin is uniformly clouded, and an abundant white deposit accumulates at the bottom of the tube. Upon the *surface of agar* a delicate, gray, glistening layer with a perfectly smooth surface is developed. Upon *potato* an abundant brownish or flesh-colored layer soon extends over the entire surface; gradually this acquires a darker hue.

363. BACILLUS RADIATUS AQUATILIS (Zimmermann).

Found in the Chemnitz water supply.

Morphology.—Bacilli about $0.65\ \mu$ broad and from 1 to $6.5\ \mu$ long.

Biological Characters.—An *aërobic, liquefying* bacillus. The shortest rods only exhibit slight movements. Spore formation not observed. Grows best at the room temperature. Upon *gelatin plates*, at the end of two days, irregular, bluish-white colonies are seen, which often have a white point in the middle; under the microscope the colonies have a root-like appearance, being surrounded by simple and branching mycelial-like offshoots; at the

end of three days these colonies are nearly spherical, and under the microscope resemble little balls of wool, from the margins of which fine filaments are given off; finally they break through the gelatin and appear upon the surface as one or more small, transparent droplets. At the end of four days the gelatin is liquefied in saucer shape; in the middle is seen a yellowish-white or cream-colored mass; around this a ring of a still deeper color, and from this delicate offshoots radiate toward the periphery of the saucer-shaped cavity. In *gelatin stick cultures* a thin, round layer appears upon the surface, which is often delicately wrinkled in a radial direction; below development occurs along the line of puncture in the form of a slender funnel, and on the third day liquefaction commences; a yellowish deposit collects at the bottom of the liquefied gelatin, which is clouded throughout and contains numerous flocculi of various dimensions. Upon the *surface of agar* a smooth, glistening layer is developed, which is yellowish-brown by transmitted light and pale bluish-green by reflected light. Upon *potato* an ochrous-yellow layer is formed, which may have a reddish-brown tint.

364. BACILLUS VERMICULOSUS (Zimmermann).

Found in water.

Morphology.—Bacilli with round ends, $1.5\ \mu$ long and about $0.85\ \mu$ broad; united in pairs or chains of three, or in long, worm-like filament in which segmentation is very indistinct; the bacilli are surrounded by a slimy envelope.

Biological Characters.—An *aërobic, liquefying* bacillus. The smaller rods exhibit a rotatory or oscillating motion. Spore formation not observed. Grows slowly at the room temperature—better at 25° to 30° C. Upon *gelatin plates* the deep colonies appear as small, white spheres; under a low power they are seen to be nearly round, well defined, gray, and granular. The superficial colonies are flat, gray, drop-like discs; under a low power they are seen to be irregular in outline, with a wavy or bulging contour; the interior is marked with paler lines, which cross each other in various directions, dividing the colony into mesh-like fields; later this appearance is only seen at the margin. In *gelatin stick cultures* a pale-gray, viscid layer with finely notched margins is developed upon the surface; after the fourth day, when this has attained a diameter of about seven millimetres, liquefaction commences and the superficial growth becomes depressed; liquefaction extends slowly toward the walls of the tube, and very slowly in a downward direction, until about one-third of the gelatin is liquefied; at the bottom of this clouded liquid an abundant reddish-gray sediment is formed. Upon the *surface of agar* a flat, smooth, glistening layer is developed; later the surface of this is opalescent. Upon *potato* an abundant yellowish-gray, shining layer is formed.

365. BACILLUS AËROPHILUS (Liborius).

Found as an accidental contamination, probably from the air.

Morphology.—Slender rods of various lengths, about two-thirds as thick as *Bacillus subtilis*; frequently united in jointed filaments.

Biological Characters.—A strictly *aërobic, liquefying, non-motile* bacillus. Forms oval spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* small, punctiform colonies are developed at the end of forty hours; under a low power these are seen to be oval or pear-shaped, well defined, and of a yellowish-gray color; liquefaction quickly occurs and the colonies do not increase materially in size. In *gelatin stick cultures* a broad, sac-like channel of liquefaction is formed, the upper part of the liquefied gelatin is opaque and yellowish-gray, while the lower portion is clear or simply contains suspended flocculi. Upon *potato* a yellowish layer is formed; this has a dull, smooth surface and a paraffin-like lustre;

later it becomes drier at the periphery, slightly granular, and has a striped appearance.

366. BACILLUS MYCOIDES (Flügge).

Found in the soil and in water—common.

Morphology.—Bacilli from 1.6 to 2.4 μ in length and about 0.9 μ thick; usually in long filaments, which when stained are seen to be made up of separate elements; as a rule, the long filaments are united in tangled bundles.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms elliptical spores from 1.3 to 1.48 μ long and from 0.74 to 0.9 μ broad. Grows very rapidly—best at the room temperature. Upon *gelatin plates* the colonies first appear as cloudy, white spots, in which fine, white, interlaced threads are soon developed; very soon a mycelial-like branching occurs, giving the colony the appearance of the commencing growth of a microscopic fungus; as long as the bundles of filaments remain beneath the surface of the gelatin they are delicate and slender, but when they reach the surface they spread out, lose their sharply defined outlines, and liquefy the gelatin. In *gelatin stick cultures*, at the end of eighteen hours, a superficial layer of about four millimetres in diameter has formed and already commences to sink in the gelatin; on the third day liquefaction has reached the walls of



FIG. 223.

FIG. 223.—*Bacillus mesentericus vulgaris*, from a culture in bouillon. $\times 1,000$. (Vignal.)



FIG. 224.

FIG. 224.—*Bacillus mesentericus vulgaris*, from an agar culture. $\times 1,400$. (Vignal.)

the test tube, and from the line of puncture a branching, filamentous growth is given off; liquefaction extends downward from the surface, and after the tenth day the bacterial growth is seen suspended in the liquefied gelatin. Upon the *surface of agar* a mycelial-like, branching growth develops along the line of inoculation. Upon *potato*, at the end of twenty-four hours, a whitish-gray, shining layer, about three millimetres broad, is developed; at the end of forty-eight hours this has extended over the entire surface.

367. BACILLUS MESENTERICUS VULGATUS.

Synonym.—Potato bacillus.

First found upon potatoes—a common and widely distributed species; found in milk by Löffler, in the Freiburg water supply by Tils, in the alimentary tract of man by Vignal, etc.

Morphology.—Thick bacilli with round ends, from 1.2 to 3.5 μ long; often united in pairs, or in chains containing several elements.

Biological Characters.—An *aërobic, liquefying* bacillus. Forms spherical spores. Grows rapidly at the room temperature—also in the incubating oven. Upon *gelatin plates* the colonies are at first almost transparent, bluish-white, later with an opaque, white centre; the superficial colonies may attain a diameter of nearly one centimetre; they are somewhat sunken in the liquefied gelatin; under a low power they are seen to be granular and have rough margins; liquefaction of the gelatin is rapidly induced. In

gelatin stick cultures liquefaction occurs at first in funnel form along the line of inoculation, and rapidly progresses until the gelatin is entirely liquefied; numerous grayish flocculi are seen in the liquefied gelatin, and a delicate, grayish-white, wrinkled layer forms upon the surface; an abundant flocculent deposit collects at the bottom of the tube. Upon the *surface of agar* a dirty-white layer is developed. Upon *potato* a thick, wrinkled, white layer, extending over the entire surface, is quickly developed: this penetrates the substance of the potato and is extremely viscid, stringing out into long threads when touched with the platinum needle. Spores are formed in the bacilli cultivated upon potato. Upon *blood serum* a white layer is developed and liquefaction of the medium occurs. Grows in *bouillon* containing one part in two hundred of hydrochloric acid (Vignal). In *milk* causes coagulation of the casein, which is subsequently dissolved and floats upon the surface as a slimy layer (Flügge).

368. BACILLUS MESENTERICUS FUSCUS (Flügge).

Found in the air, in hay dust, upon potato, and in water—a common and widely distributed species.

Morphology.—Slender and short bacilli, often in pairs or in chains of four.

Biological Characters.—An *aërobic, liquefying, actively motile* bacillus. Forms small, shining spores which are irregularly distributed in the rods. Grows at the room temperature. Upon *gelatin plates* forms spherical, whitish colonies, which under a low power are seen to have a well-defined contour; later these are surrounded by delicate offshoots, have a yellowish-brown color and a finely granular surface; liquefaction of the gelatin quickly occurs. In *gelatin stick cultures* a whitish cloudiness is first seen along the line of puncture, and at the same time liquefaction commences near the surface in funnel form; this extends to the walls of the tube in from four to six days, and the liquefied gelatin contains numerous grayish-white flocculi. Upon *potato*, at the end of twenty-four hours, a smooth, yellowish layer is developed; the surface of this soon becomes wrinkled and brown in color; the growth is comparatively thin and does not penetrate deeply into the potato, as is the case with *Bacillus mesentericus vulgaris*; it extends rapidly over the entire surface.

369. BACILLUS MEGATHERIUM (De Bary).

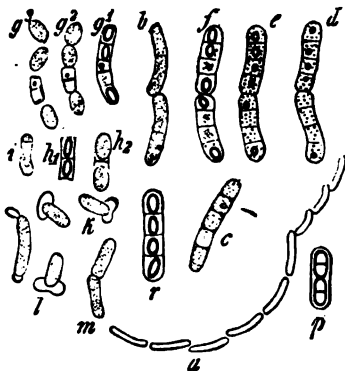


FIG. 235.—*Bacillus megatherium*; a, chain of bacilli ($\times 250$); b, bacilli ($\times 600$); c-f shows the development of spores; h-m shows the germination of spores; p, bacilli stained with solution of iodine. (De Bary.)

First found upon the leaves of boiled cabbage.

Morphology.—Bacilli with round ends, about 2.5μ thick and three to four times as long as broad; often somewhat curved; forms chains containing as many as ten elements; the protoplasm of the cells is granular; involution forms are common.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. The movements are peculiar, being slow and amoeboid in character. Forms long-oval spores which are nearly as long as the cells containing them, but not so broad. Grows best at the room temperature—also in the incubating oven. Upon *gelatin plates* forms whitish, punctiform colonies, which under the microscope are yellowish and irregular in form. The superficial colonies are sometimes kidney-shaped or crescentic; they cause the gelatin to be slowly liquefied. In *gelatin stick cultures* lique-

faction occurs in funnel form along the upper portion of the line of inoculation and gradually extends downward; an abundant deposit is seen at the bottom of the tube, and the liquefied gelatin above is but slightly clouded; no mycoderma forms upon the surface. Upon the surface of agar a whitish layer is developed which is easily separated from the culture medium. Upon potato a thick, cheesy, yellowish-white layer is quickly developed along the line of inoculation; in potato cultures an abundant development of spores occurs and involution forms are common.

370. BACILLUS ALBUS PUTIDUS (De Bary).

Found in water.

Morphology.—Small bacilli, which grow out into filaments.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature. Upon *gelatin plates* forms thin, round colonies upon the surface, which under a low power are light-brown in color and are surrounded by a transparent aureole which at the end of four days has a diameter of five millimetres. In *gelatin stick cultures* development occurs both on the surface and along the line of puncture, producing rapid liquefaction of the gelatin; gelatin cultures give off an intense and disagreeable odor, like that of liquid manure. Upon the surface of agar a sneary layer is developed. Upon potato a slimy growth.

371. BACILLUS BRASSICÆ (Pommer).

Obtained from an infusion of cabbage leaves.

Morphology.—Bacilli from 1.9 to 5.4 μ long and 0.91 to 1.2 μ thick; differ greatly in different culture media, forming sometimes twisted and tangled filaments, often spiral in form, and producing at times a network similar to that of *Bacterium Zopfii*; the filaments are often segmented and slightly notched and bent at the points where the segments join.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, non-motile* bacillus. Forms spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* forms colonies which resemble the mycelium of a mucus and cause liquefaction of the gelatin. In *gelatin stick cultures* a branching, mycelial-like growth is seen along the line of puncture, in funnel form, and liquefaction of the gelatin quickly occurs. Upon the surface of agar a layer is formed consisting of spots surrounded by a dull cloudy appearance; later these have a whitish or yellowish color; under the microscope they are seen to consist of closely lying parallel filaments, which may run in a straight or serpentine direction, or may form circular and ellipsoidal figures. Along the line of puncture in agar cultures small, white colonies are developed, which are seen under the microscope to be made up of a confused mass of straight or curved, short filaments, and a network of filaments is given off from these.

372. BACILLUS BUTYRICUS OF HUEPPE.

Found in imperfectly sterilized milk.

Morphology.—Bacilli, often slightly curved, about 2.1 μ long and 0.38 μ thick; may grow out into filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, actively motile* bacillus. Forms, at 30° C., oval spores which are located in the centre of the rods. Grows at the room temperature—more rapidly at 35° to 40° C. Upon *gelatin plates* deep-lying colonies of yellow color are developed, which cause rapid liquefaction of the gelatin and unite into coarsely granular, brown masses. In *gelatin stick cultures* liquefaction rapidly occurs along the entire line of puncture; upon the surface of the

liquefied gelatin a thin, whitish-gray, slightly wrinkled film is formed, and below this it is densely clouded and of a yellowish color. Upon the surface of agar a thin, smeary, yellowish layer is formed. Upon potato a fawn-colored, transparent layer, which is sometimes slightly wrinkled; later the surface loses its transparency and becomes clouded. In milk coagulation occurs and the precipitated casein is subsequently dissolved; the milk acquires a bitter taste; produces butyric acid in milk. *Bouillon* cultures to which sulphuric acid has been added give off an acid distillate which has the odor of butyric acid.

373. BACILLUS GASOFORMANS (Eisenberg).

Found in water.

Morphology.—Small bacilli.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *actively motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature—not in the incubating oven at 37° C. Upon *gelatin plates* forms tolerably large, saucer-shaped cavities of liquefied gelatin, the contents of which are finely granular and may contain gas bubbles. In *gelatin stick cultures* liquefaction rapidly occurs all along the line of puncture and gas bubbles form in the non liquefied gelatin.

374. BACILLUS CARABIFORMIS (Kaczynsky).

Found in the stomach of dogs which had been fed exclusively on meat for three days.

Morphology.—Short and slender bacilli.

Biological Characters.—An *aërobic*, *liquefying*, *actively motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature. Upon *gelatin plates* forms small colonies, from the centre of which are given off long processes with jagged outlines. In *gelatin stick cultures* liquefaction occurs along the line of puncture, and the liquefied gelatin has a greenish-yellow color, while a whitish deposit accumulates at the bottom. Upon the surface of agar a yellowish-white layer.

375. BACILLUS GRAVEOLENS (Bordoni-Uffreduzzi).

Found attached to scales of epidermis from between the toes of man.

Morphology.—Bacilli 0.8 μ long and of about the same breadth (micrococci ?)

Biological Characters.—An *aërobic*, *liquefying* bacillus (?). Grows at the room temperature. Cultures have a disagreeable odor. Upon *gelatin plates* forms irregular, grayish-white spots which cause rapid liquefaction of the gelatin and give off a disagreeable odor like that from the feet; later the gelatin has a greenish-yellow color. Upon *potato* forms a grayish, stinking layer. Liquefies *blood serum*.

376. BACILLUS CAROTARUM (A. Koch).

Obtained upon cooked carrots and sugar beets.

Morphology.—Bacilli from 0.97 to 1.05 μ long; grows out into long, flexible filaments—resembles *Bacillus brassicæ*.

Biological Characters.—An *aërobic*, *liquefying*, *non-motile* bacillus. Forms large oval spores. Grows best at 40° C. Upon *gelatin plates* forms round, white colonies upon the surface, which under a low power appear to be perforated with holes at the centre and are marked by fine lines. The deep colonies in the liquefied gelatin are spherical, with a sharply defined, smooth outline. In *gelatin stick cultures* a considerable growth occurs upon the surface; very scanty development along the line of puncture.

Upon the *surface of agar* a white layer is formed. Upon *potato* a light-brown, circular layer is quickly developed; this at first has a dull and later a shining surface.

377. BACILLUS INFLATUS (A. Koch).

Found as an accidental impurity—from the air?

Morphology.—Bacilli from 4.6 to 5.5 μ long and 0.6 to 0.8 μ broad; often grow out into filaments.

Biological Characters.—An *aërobic, liquefying, actively motile* bacillus. Forms two large spores in each rod. Grows at the room temperature. Upon *gelatin plates* forms spherical white colonies with folded margins, from which an outgrowth of delicate filaments is seen, similar to *Bacillus alvei*. In *gelatin stick cultures* development occurs all along the line of puncture, and short, hair-like filaments radiate into the gelatin, which is very slowly liquefied. Upon the *surface of agar* forms a thin, slimy, light-brown layer. In *bouillon* a thin, slimy, whitish film forms upon the surface, which sinks to the bottom when the tube is shaken.

378. BACILLUS RAMOSUS.

Synonym.—Wurtzel bacillus.

Found in the soil and in water—common.

Morphology.—Bacilli with round ends, about three times as long as broad—"about as long as *Bacillus subtilis*, but thicker" (Fränkel); frequently united in long chains, or may grow out into filaments.

Biological Characters.—An *aërobic, liquefying* bacillus. Exhibits slight movements. Forms large oval spores which are located at the centre of the rods. Grows rapidly at the room temperature—also in the incubating oven. Upon *gelatin plates*, at the end of two days, veil-like, rapidly extending, whitish colonies with ill-defined margins are developed; these resemble the mycelium of a fungus; under a low power the colonies are seen to consist of a network of twisted and interwoven filaments; liquefaction occurs in the course of a few days. In *gelatin stick cultures* an outgrowth of branching filaments occurs along the line of puncture, looking "like a small fir tree turned upside down"; upon the surface a moist and shining, white layer is developed; liquefaction soon occurs at the surface, and progresses until the gelatin is entirely liquefied; in old cultures a mycoderma is seen upon the surface, the gelatin below this is transparent, and at the bottom there is a deposit of crumbling, whitish flocculi. Upon the *surface of agar* a development of branching filaments occurs along the line of inoculation; these form a moist-looking, grayish-white layer, which later becomes thicker at the centre, and the root-like growth is only seen at the edges. Upon *potato* a whitish, smeary streak is developed along the line of inoculation; the bacilli form numerous large oval spores when cultivated on potato.

379. BACILLUS SUBTILIS (Ehrenberg).

Synonym.—Hay bacillus.

A widely distributed species; found in hay infusion, in water, in the soil, etc.

Morphology.—Bacilli with slightly rounded corners, from 4.5 to 6 μ long, and about three times as long as broad; usually in chains consisting of several elements; often grows out into very long filaments; possesses a terminal flagellum at each end of a single rod or at the two extremities of a chain.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms large oval spores, which are located at the centre of the rods; these are about 1.2 long and 0.6 μ broad. The movements are of a waddling charac-

ter and not very rapid. Grows rapidly at the room temperature—better at 30° C. Development may occur at any temperature between 10° and 45° C. Spore formation is favored by a temperature of 30° C. The spores germinate most readily at 30° to 40° C. The exosporium is ruptured at one side of the long-oval spore, and the newly formed bacillus takes its exit from this opening in a direction perpendicular to the long axis of the reproductive element. The spores have great resistance to heat and to chemical agents. By boiling a hay infusion for a short time a pure culture may often be obtained, as other microorganisms present are killed, while the spores of *Bacillus subtilis* survive and subsequently germinate. Upon *gelatin plates* small, white colonies are first developed, which under the microscope are seen to be slightly granular, somewhat irregular in outline, and of a greenish tint; development progresses very rapidly, and liquefaction of the surrounding gelatin is quickly induced, forming saucer-like cavities with grayish, translucent contents; the central portion is white and opaque; frequently

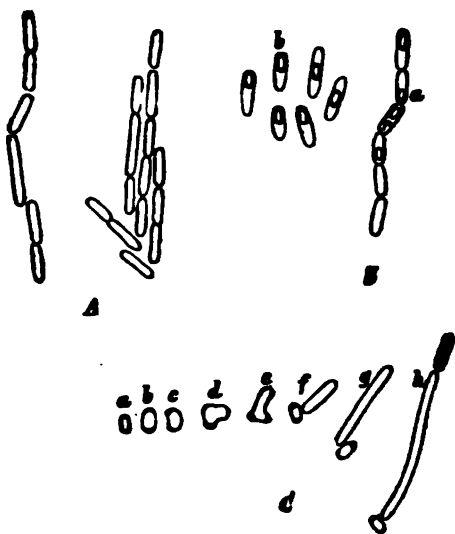


FIG. 236.—*Bacillus subtilis*: A, bacilli; B shows formation of spores; C shows the germination of a spore, *a*, and development of a short rod, *f*, and subsequently of a longer filament, *h*. $\times 1,020$. (Prazmowski.)

a radiate appearance of the bacterial growth is observed; under the microscope a dense, grayish-yellow central mass is seen, and around this a tangled network of filaments and of rods undergoing the characteristic waddling movements; at the margins the filaments are seen to radiate into the non-liquefied gelatin, forming a crown-like aureole. In *gelatin stick cultures* liquefaction quickly occurs along the entire line of puncture; later a dense, dry, and friable mycoderma forms upon the surface, and the gelatin below, which was at first filled with whitish flocculi, becomes clear as a result of their deposition at the bottom of the tube. Upon the *surface of agar* a wrinkled, white layer is developed which is easily lifted entire from the culture medium. *Blood serum* is liquefied by this bacillus, and a wrinkled mycoderma forms upon the surface. Upon *potato* the entire surface is soon covered with a cream-like, white layer, which in a short time contains an abundance of spores.

380. *BACILLUS SUBTILIS SIMILIS* (Sternberg).

Obtained in cultures from the liver of a yellow-fever cadaver in Havana, 1889.

Morphology.—Bacilli with slightly rounded ends, from 2 to 4 μ long and about 1 μ thick; grow out into jointed filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Forms long-oval spores, which are centrally located and nearly as long as the cells in which they are developed. The motion is like that of *Bacillus subtilis*, viz.: a slow, to-and-fro, progressive movement. Upon *gelatin plates* the deep colonies, at the end of thirty-six hours at the room temperature, are spherical, finely granular, and pearl-like by reflected light; the superficial colonies have commenced to liquefy the gelatin at this time, and have a granular, white mass at the centre surrounded by a saucer-shaped cavity containing liquefied gelatin. In *gelatin stick cultures* liquefaction does not occur as rapidly as with *Bacillus subtilis*; at the end of ten days at the room temperature the upper half of the gelatin is liquefied and small, pearl-like colonies are scattered along the line of puncture below; on the floor of the liquefied gelatin is a flocculent, white deposit and a thin mycoderma is seen upon the surface. On *potato* a dry, yellowish-white layer the size of a dime is formed at the end of forty-eight hours at 30° C., and the bacilli, which grow out into long, jointed filaments, contain spores. Upon the *surface of agar* a thick, cream-white layer is formed in four or five days at the room temperature. In *agar stick cultures* there is a branching growth along the upper portion of the line of puncture; in old agar cultures variously contorted involution forms are seen in the surface growth and but few spores are present.

381. *BACILLUS LEPTOSPORUS* (L. Klein).

Obtained as an accidental contamination of a pure culture—from the air?

Morphology.—Resembles *Bacillus subtilis*; when cultivated at 35° C. forms short chains; at 18° to 20° C. grows out into long, twisted and interlaced filaments. Forms spores which are 0.6 μ thick and 1.5 μ long; in vegetating, these first increase in thickness to 1 to 1.2 μ —the thickness of the vegetative cells; the spores have a membranous envelope consisting of two layers, and are surrounded by a jelly-like substance having a dull silvery lustre; vegetation occurs at the same time from both poles, and the membranous envelope is not ruptured and left intact after the emergence of the vegetative cell, as is the case with *Bacillus subtilis*, but is gradually dissolved, or serves as the cell wall of the newly formed bacillus (?).

Biological Characters.—Characters of growth in solid culture media not given. The motions are said to be peculiar, especially in filaments made up of four, eight, or sixteen elements; one end of the chain is jerked hither and thither with a whip-like, convulsive motion, by which the elements are thrown into various and constantly changing angular figures.

382. *BACILLUS SESSILIS* (L. Klein).

Found in the blood of a cow supposed to have died of anthrax.

Morphology.—Resembles *Bacillus subtilis*, but is distinguished from this species by the germination of the spores.

Biological Characters.—An *aërobic, non-motile* bacillus. In alkaline *bouillon* at 28° C. a diffuse cloudiness is seen on the second day, and on the fourth day a mycoderma is developed upon the surface, which contains spore-bearing bacilli. The spores resemble those of *Bacillus subtilis*, but the germination of these reproductive elements is quite different and resembles more that of *Bacillus butyricus*. The vegetative cell emerges from a rupture at one of the poles of the exosporium, and a second rod appears to fol-

low the first, pushing it before it; this appearance is, however, due to binary division of the vegetative cell before it has completely emerged from the spore membrane. Characters of growth in solid media not given. Not pathogenic for guinea-pigs.

383. BACILLUS ALLANTOIDES (L. Klein).

Obtained as an accidental contamination from a culture of *Bacillus megatherium*—from the air?

Morphology.—Bacilli about $0.5\ \mu$ thick and three to four times as long as thick; form chains of four to eight elements, the members of which are rather widely separated from each other, but are firmly bound together by a jelly-like membrane which is somewhat narrower than the rods. The rods exhibit an intermittent to-and-fro movement; subsequently they undergo segmentation into spherical elements which are surrounded by a jelly-like material and form sausage-shaped zoöglöea masses. Characters of growth in solid media not determined.

384. BACILLUS OF SCHEURLLEN.

Found in cancerous tissues by Scheurlen; upon the skin of healthy persons by Bordoni-Uffreduzzi (*Bacillus epidermidis*); in scales of epidermis from the nipple and in the mammae of healthy women by Rosenthal.

Morphology.—Bacilli from 1.5 to $2.5\ \mu$ long and $0.5\ \mu$ broad.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms long-oval spores. Grows very slowly at the room temperature—best at 29°C . In *gelatin stick cultures*, at the end of eight to fourteen days, a funnel-shaped cavity is formed near the surface, which is covered by a wrinkled, membranous layer, while but little liquid gelatin is contained in it. Upon the *surface of agar*, at 39°C ., at the end of twelve hours a dull, fissured, colorless layer is developed. Upon *potato*, at the end of twelve to twenty-four hours in the incubating oven, the whole surface is covered with a yellow, wrinkled layer; the potato beneath this has a dirty-pink color.

385. BACILLUS LACTIS ALBUS (Löffler).

Found in milk.

Morphology.—Bacilli which average $3.4\ \mu$ in length and $0.96\ \mu$ in thickness; in milk grow out into long filaments; resembles the bacillus of anthrax.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms large spores. Grows slowly in the usual culture media at the room temperature. In *gelatin stick cultures* liquefaction occurs slowly, and upon the surface of the liquefied medium a whitish layer, made up of interlaced filaments, is seen. Upon the *surface of agar* forms a tolerably thick layer with thinner margins. Upon *potato* a thin, dry, white layer. In *milk* causes coagulation and subsequent solution of the casein, and produces leucin and tyrosin.

386. BACILLUS LIODERMOS (Löffler).

Synonym.—Gummibacillus.

Found in milk

Morphology.—Resembles *Bacillus mesentericus vulgatus*—small, thick rods with round ends, in pairs or in jointed filaments.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms spores. Grows rapidly at the room temperature. In *gelatin stick cultures* causes rapid liquefaction in funnel form; the liquefied gelatin is slightly clouded and is soon covered with a whitish mycoderma. Upon the *surface*

of agar a whitish, rosette-like layer is developed. Upon potato forms a transparent, gum-like layer, which later is thrown into thick, soft folds, similar to the growth of *Bacillus mesentericus vulgaris*. In milk causes coagulation of the casein, which at 30° C. is precipitated as a whitish, cloudy sediment, above which a clear serum is seen; the casein is subsequently peptonized.

387. *BACILLUS ULNA* (Cohn).

First described by Cohn, and subsequently found by Prazmowski in a solution of egg albumin.

Morphology.—Bacilli of 1.5 to 2.2 μ in breadth and of various lengths—at least 3 μ ; forms long, jointed filaments; forms spores which are 2.5 to 2.8 μ long and 1 μ broad.

Biological Characters.—An *aërobie*, *motile* bacillus which is said to grow only in albuminous solutions (?), in which it develops as cloudy masses which collect at the surface and form a thick, dry mycoderma consisting of long, interlaced filaments in bundles and irregular aggregations; does not give off a putrefactive odor. Imperfectly described.

388. *BACILLUS ULNA* OF VIGNAL.

Found by Vignal in the salivary secretions of healthy persons, and supposed to correspond with *Bacillus ulna* of Cohn.

Morphology.—Straight bacilli with round ends, about 2 μ long; often united in pairs, in which the elements are strongly adherent; rarely in chains containing more than two segments.

Biological Characters.—An *aërobie*, *liquefying* bacillus. Spore formation not observed. Motility not mentioned. Upon *gelatin plates*, at the end of twenty-four hours, small, grayish superficial colonies are formed; the centre of these is thicker than the periphery; under the microscope the peripheral zone is seen to be made up of fine interlaced filaments; by the second or third day the colonies have increased considerably in size, and a small, grayish-white, opaque mass is seen, which is surrounded by an extended zone of liquefied gelatin; this is seen to consist of four secondary zones: next the central mass the gelatin is almost transparent, outside of this is a granular zone, outside of this a grayish, less granular zone, and finally an outer zone which is almost transparent. In *gelatin stick cultures*, at the end of forty-eight hours, liquefaction in funnel form has occurred along the line of puncture; the liquefied gelatin is transparent and contains suspended opaque, white flocculi, which accumulate at the bottom; by the fourth day the gelatin is completely liquefied as far as the inoculating needle penetrated, and a whitish film is seen upon the surface. Upon the *surface of agar* a very adherent, white layer is developed which presents punctiform and linear depressions; the agar below acquires a slightly brownish color. In *bouillon* the liquid remains transparent and acquires a yellowish tint, while a tolerably thick, fragile, smooth, white mycoderma is formed upon the surface; rather a scanty white deposit accumulates at the bottom. Grows in acid bouillon—1 : 2,000 of hydrochloric acid. Upon *potato* a thin, grayish-white layer is formed at the end of forty-eight hours, upon the surface of which fine, slightly elevated lines cross each other in all directions; sometimes this forms quite regular hexagonal figures. *Blood serum* is liquefied by this bacillus. All the cultures have a disagreeable odor, similar to that given off by *Bacillus pyogenes foetidus*.



FIG. 287. — *Bacillus ulna* of Vignal, from a culture in nutrient agar. $\times 1,500$. (Vignal.)

389. *BACILLUS LIQUEFACIENS* (Eisenberg).

Found in water.

Morphology.—Short, rather thick rods with round ends.

Biological Characters.—An *aerobic, liquefying, actively motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature—not at 37° C. Upon *gelatin plates* forms round colonies with a smooth margin, which in the middle are white and slimy; liquefaction commences in saucer shape and progresses rapidly; after a time a putrefactive odor is given off. In *gelatin stick cultures* development is rapid and liquefaction occurs in the form of a funnel, often like an air bubble; the line of puncture is filled with a whitish, granular mass. Upon the *surface of agar* forms a dirty-white layer. Upon *potato* a pale-yellow growth.

390. *BACILLUS MAÏDIS* (Cuboni).

Obtained from corn which had been soaked in water for eight hours at 30° C., and in the *faeces* of individuals suffering from pellagra.

Morphology.—Bacilli with square ends, from 2 to 3 μ long; solitary, in pairs, or in chains of three elements—seldom more; resembles *Bacillus mesentericus fuscus* in morphological and biological characters.

Biological Characters.—An *aerobic, liquefying, actively motile* bacillus. Forms large oval spores located at the centre of the rods. Grows in the usual culture media at the room temperature—best at 26° to 30° C. Produces in saccharine solutions acetic and butyric acids. Upon *gelatin plates*, at the end of twenty-four to thirty-six hours, grayish-white, punctiform colonies are developed below the surface, which have a yellowish color. The superficial colonies are thin and veil-like; under a low power they are seen to be finely granular and have an irregularly folded margin; later liquefaction commences and they have a radiate, finely striped margin; the liquefaction progresses rapidly, forming shallow, saucer-like cavities. In *gelatin stick cultures* liquefaction occurs along the line of puncture within twenty-four hours in the form of a funnel or cylindrical tube, and rapidly extends to the walls of the test tube at the surface. Upon the *surface of agar*, at 34 to 36° C., a thin, dry, wrinkled layer covers the entire surface within twenty-four hours; this is white or yellowish-white and easily detached. Upon *potato* a white, somewhat granular, and later finely wrinkled layer, which acquires a yellowish-brown color. *Blood serum* is liquefied by this bacillus.

391. *PROTEUS SULFUREUS* (Lindenborn).

Found in water.

Morphology.—Bacilli of various lengths, the average length being about 1.6 μ and the breadth 0.8 μ ; often in long chains or filaments. Resembles *Proteus vulgaris*, and is perhaps identical with this species (Eisenberg).

Biological Characters.—An *aerobic, liquefying, motile* bacillus. Spore formation not observed. Grows rapidly at the room temperature. Forms sulphuretted hydrogen. Upon *gelatin plates* forms white colonies, from which filamentous outgrowths upon the surface of the gelatin are given off; these form "swimming islands"; later liquefaction occurs in the form of broad funnels with grayish-white contents. In *gelatin stick cultures* development occurs along the line of puncture, and liquefaction in funnel shape near the surface. Upon the *surface of agar* a thick, grayish-white layer is developed. Upon *potato* a slimy, grayish-white layer, which later acquires a brownish color. In *milk* an alkaline reaction is produced in the course of several weeks, and the casein is peptonized without previous precipitation; the milk acquires a bitter taste and a yellowish color.

392. *BACILLUS THERMOPHILUS* (Miquel).

Found in the contents of sewers, in the alimentary tract of man and animals, and in the soil.

Morphology.—Bacilli about $1\ \mu$ thick, which form filaments of various lengths.

Biological Characters.—An *aërobic, non-motile* bacillus. Forms spores, which are located at the extremities of the rods. No growth in gelatin at the room temperature. Grows at temperatures between 42° and 72° C., but not below 42° or above 72° ; grows best at 65° to 70° C. Upon the *surface of agar*, at 42° to 45° C., a white, disc shaped, prominent layer is formed—more abundant growth at 50° to 65° C. In *bouillon*, at 50° C., development occurs upon the surface in the form of a mycoderma which is easily broken up, and the liquid below is diffusely clouded.

393. BACILLUS TUMESCENS (Zopf).

Found upon beet.

Morphology.—Short bacilli, about $1.17\ \mu$ broad; grow out into irregularly bent and twisted filaments; resembles *Bacillus megatherium*.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Movements slow. Forms oval spores. Grows rapidly at temperatures above 20° C. Upon *gelatin plates* forms round, superficial colonies which have a

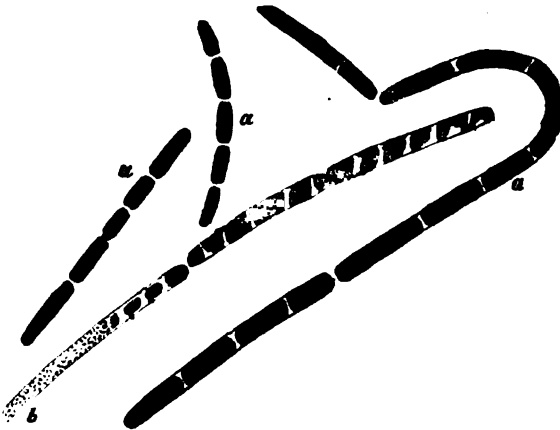


FIG. 228.—*Bacillus buccalis maximus*, after treatment with iodine solution. $\times 1,800$. (Miller.)

brownish-yellow color; after several days the margins are no longer well defined, but have a fringed appearance, and liquefaction commences. Upon *potato* a thick, white, viscid layer, with somewhat folded margins, is developed; later this extends over the entire surface.

394. BACILLUS BUCCALIS MAXIMUS (Miller).

Found in the mouth of man—common.

Morphology.—"Isolated bacilli or threads, but much oftener tufts of threads, parallel to or crossing each other, from 30 to $150\ \mu$ long, and distinctly articulated. The rods are from 2 to $10\ \mu$ long, sometimes even longer, and from 1 to $1.3\ \mu$ broad. This bacterium is therefore the largest occurring in the mouth; it has a very regular contour and usually the same thickness throughout. Not all of the cells of this bacterium show the iodine reaction—a statement which applies equally well to most bacteria that turn blue on the addition of iodine. The majority of them, however, respond very distinctly to the test, becoming stained brown-violet, either throughout or only in isolated places."

Biological Characters not determined.

395. *LEPTOTHRIX BUCCALIS* OF VIGNAL.

Found by Vignal in the healthy human mouth—rare. Supposed by Vignal to be identical with *Leptothrix buccalis* of Robin; but as the characters of the microorganism receiving this name were not definitely determined by Robin, such identification is impossible. It seems more probable that the large bacillus described by Miller (*Bacillus buccalis maximus*), which is found very commonly in the human mouth, but has not been cultivated, is the *leptothrix* of Robin.

Morphology.—Bacilli from 1 to 1.5 μ in breadth and varying greatly in length—from 1.6 to 30 μ ; often united in long chains, or in filaments which are seen to be segmented when treated with staining agents. This *leptothrix* is characterized by the presence of transverse partitions in the interior of the rods, seen only in stained preparations; these are best seen in old rods, which are less deeply stained than those of more recent development; the partitions in these are more deeply stained than the remaining portion of the filament.

Biological Characters.—An *aërobic, liquefying* bacillus. Spore formation not observed. Grows very slowly in the usual culture media at temperatures above 20° C. Upon *gelatin plates*, by the third or fourth day, small, round, grayish-white, projecting colonies are formed; by the fifth or sixth day an irregular border of rounded festoons is developed; this is semi-transparent and presents depressions and ridges which are radial or parallel with the margin; the prominent centre remains opaque and of a grayish-white color; later this border extends considerably and the gelatin below it becomes liquefied. In *gelatin stick cultures*, at the end of forty-eight hours, a small mass is developed at the point of puncture, about two millimetres in diameter, and a slender line of growth is visible along the track of the needle below; by the fourth day the surface growth has a diameter of five to six millimetres, and below it a little transparent, liquefied gelatin is seen in a cup-shaped cavity; this increases slowly in dimensions and the liquefied gelatin remains clear, while a bluish mycoderma is seen upon the surface; the growth along the line of puncture remains scanty, but is a little more abundant above in contact with the cup shaped cavity; by the eighth day this extends to the walls of the tube; by the twelfth day the cup form has disappeared and the gelatin is liquefied to a depth of about one centimetre; it is still covered with a bluish membranous layer, and a scanty whitish deposit of *leptothrix* filaments is seen at the bottom of the liquefied gelatin. Upon the *surface of agar*, at the end of twenty-four hours at 36° to 38° C., a slightly wrinkled, transparent, white layer is developed; later this is still more wrinkled, dry, and of a transparent yellow color; it is easily broken up. In neutral *bouillon* a slight cloudiness is produced and a scanty white precipitate is formed at the bottom of the tube; no mycoderma develops upon the surface. Upon *potato* a layer is developed which is of a dirty-white color at the centre and of a purer white near the margins.

396. *BACILLUS b* OF VIGNAL.

Obtained quite frequently by Vignal in cultures from healthy buccal secretions.

Morphology.—Bacilli with square ends, straight or slightly curved, about 0.5 μ in diameter and varying greatly in length—from 1.5 to 6.5 μ ; often united in chains.

Biological Characters.—An *aërobic, liquefying* bacillus. Spore formation not observed. Motility not mentioned. Grows rather slowly at the room temperature—more abundantly at 37° C. Upon *gelatin plates*, at the end of twenty-four hours at a temperature of 18° to 20° C., prominent, small, grayish-white colonies are developed upon the surface; at the end of forty-eight hours a collarette with irregularly festooned margins is developed around this central mass; this is thinner and much more transparent

than the central portion of the colony; under a low power it is seen to be formed of an innumerable series of skein-like bundles, arranged side by side and more or less twisted, which proceed from the central mass. In *gelatin stick cultures*, at the end of forty-eight hours, a small, flat mass is developed at the point of puncture, and a scanty growth is seen along the line of inoculation. On the fourth day the superficial growth covers the entire surface, it is translucent by transmitted light and white by reflected light; by the sixth day the gelatin is liquefied to a depth of one centimetre below the superficial growth; the liquefied gelatin remains transparent; upon the surface is seen a white, membranous layer, and at the bottom a rather scanty white deposit; liquefaction slowly extends downward, and by the twelfth day has reached a level corresponding with the bottom of the line of puncture. Upon the *surface of agar*, at the end of twenty-four hours at 36° to 38° C., a dull-white layer, having a thickness of about one millimetre, is developed; this is easily broken up with the platinum needle. In neutral *bouillon* a slight cloudiness is quickly produced, a thin film forms upon the surface, and a scanty white precipitate at the bottom of the tube. Upon *potato*, at the end of forty-eight hours, a layer the size of a five-franc piece is developed, which has a pale-pink color and a rough surface—resembling a lichen. *Blood serum* is liquefied rather rapidly, and acquires a brownish color, while an abundant white precipitate accumulates at the bottom of the tube.

397. BACILLUS *f* OF VIGNAL.

Found by Vignal in the salivary secretions of healthy persons.

Morphology.—Bacilli with slightly rounded ends, from 0.8 to 1.2 μ in length when cultivated upon agar, and from 1.4 to 2.4 μ when cultivated in neutral bouillon; usually solitary, occasionally united in short chains.

Biological Characters.—An *aërobic, liquefying* bacillus. Spore formation not observed. Motility not mentioned. Grows rather slowly at the room temperature—more rapidly at 37° C. Upon *gelatin plates*, at the end of forty-eight hours, small, projecting, opaque, white colonies are developed; at the end of four days the colonies are seen as conical, opaque, white masses, divided into about twenty segments by grooves which start from the summit. In *gelatin stick cultures* a small but prominent white mass is seen at the point of puncture, and a scanty line of development along the track of the inoculating needle; on the fourth day the surface growth has extended nearly to the walls of the tube, and just below this some fine branches are given off from the line of growth; the sixth day the entire surface is covered and the gelatin below is liquefied for a short distance; the eighth day the liquefaction has extended downward, and the solid gelatin below has a clouded appearance owing to the development of a quantity of small, white colonies; by the twelfth day the liquefied gelatin has a depth of about two centimetres, a shining, white mycoderma is seen upon the surface, a white deposit at the bottom, and below this numerous small colonies in the solid gelatin. Upon the *surface of agar* very adherent, white colonies are formed, which later extend to form a transparent white membrane. In neutral *bouillon* a slight cloudiness is produced, and a very scanty, whitish deposit is seen at the bottom of the tube. Does not develop well in acid bouillon. Upon *blood serum* a whitish layer is formed, which later becomes semi-transparent and causes a slow liquefaction of the medium. Upon *potato*, at the end of forty-eight hours, a layer is developed which has a velvety appearance in the centre, and a yellowish or brownish-white color; by the end of forty-eight hours this layer is as large as a five-franc piece.

398. BACILLUS BUCCALIS FORTUITUS.

Synonym.—Bacillus *j*, Vignal.

Found by Vignal in the salivary secretions of healthy persons.

Morphology.—Bacilli with square ends, from 1.4 to 3 μ long; often united in pairs, the elements of which may be joined at an angle of greater or less degree.

Biological Characters.—An *aërobic, liquefying* bacillus. Spore formation not observed. Motility not mentioned. Grows at the room temperature in the usual culture media. Upon *gelatin plates*, at the end of forty-eight hours, small, round colonies are developed, which increase considerably in thickness and diameter, and by the fourth or fifth day have caused liquefaction of the surrounding gelatin. In *gelatin stick cultures*, at the end of forty-eight hours, a small mass has formed at the point of inoculation, and a scanty line of development is seen along the track of the inoculating needle: by the fourth day the growth has extended over the entire surface and presents a decided prominence at the centre; the gelatin below is liquefied and remains transparent, with some opaque, white flocculi in suspension; by the twelfth day the liquefaction extends to a depth of two centimetres, and a yellowish-white, abundant deposit is seen at the bottom. Upon the *surface of agar* at 36° to 38° C., small, white, opaque colonies are developed, which present a small, nipple-like projection at the centre. In *neutral bouillon* a diffuse cloudiness is produced; a thick, dull-white layer forms upon the surface, and an abundant dull-white deposit is seen at the bottom of the tube. Does not grow well in acid bouillon. Upon *potato* a rather thick growth is developed, which extends slowly and acquires a slightly pinkish tint.

399. BACILLUS HAVANIENSIS LIQUEFACIENS (Sternberg).

Obtained in cultures from the surface of the body of patients in the charity hospital at Havana.

Morphology.—Bacilli with round ends, about 0.8 μ thick and varying greatly in length—from 1.2 to 5 μ ; solitary, in pairs, or may grow out into long filaments.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. Grows at the room temperature—better at 37° C. Upon *gelatin plates*, at the end of twenty-four hours at 22° C., round colonies are developed which have a milky opacity and are surrounded by a transparent marginal zone with irregular margins; under the microscope these colonies are seen to be finely granular; at the end of twenty-four hours liquefaction commences. In *gelatin stick cultures* liquefaction occurs along the entire line of puncture; at the end of four days the liquefied gelatin is clouded throughout; in old cultures it is quite transparent, and a slight flocculent deposit is seen at the bottom of the tube. Upon the *surface of agar*, at the end of two weeks, a thin, pale-brown layer is developed. No growth upon *potato*. Not pathogenic for rabbits.

400. BACILLUS LIQUEFACIENS COMMUNIS (Sternberg).

Obtained in cultures from the fæces of yellow-fever patients at Decatur, Ala. (1888).

Morphology.—Bacilli with round ends, from 1 to 2 μ long and about 0.7 μ thick; solitary or in pairs.

Biological Characters.—An *aërobic and facultative anaërobic, liquefying, actively motile* bacillus. Grows in the usual culture media at the room temperature, also at comparatively low temperatures, and in the incubating oven at 37° C. Spore formation not observed. Grows in an acid medium—1 : 500 of hydrochloric acid. In *gelatin stick cultures* liquefaction occurs rapidly in the form of a purse. On *potato*, at the end of two weeks, an irregular, corrugated layer is developed which has a pinkish color. Not pathogenic for rabbits.

*E. Strictly Anaërobic Bacilli.*401. *BACILLUS MUSCOIDES* (Liborius).

Found in the soil by inoculations in mice, also in old cheese, and in the excrement of cattle.

Morphology.—Bacilli about $1\ \mu$ thick, with slight inclination to form filaments.

Biological Characters.—An *anaërobic, non-liquefying, motile* bacillus. Forms short-oval spores, which are located at the ends of the rods. In *gelatin* and in *agar* forms colonies which give off delicate, branching, moss-like offshoots. In *gelatin stick cultures* a delicate, branching growth is given off from the lower two-thirds of the line of puncture.



FIG. 229.—*Bacillus muscoides*; colony in nutrient gelatin. $\times 80$. (Liborius.)

402. *BACILLUS SOLIDUS* (Lüderitz).

Obtained from garden earth by inoculation into mice and guinea-pigs.

Morphology.—Bacilli of from 1 to $10\ \mu$ in length—average $4.5\ \mu$ —and $0.5\ \mu$ thick: the longer bacilli consist of two segments, but regular filaments are not formed.

Biological Characters.—An *anaërobic, non-liquefying, motile* bacillus. Movements tolerably active, pendulous and progressive. In *old gelatin cultures* some of the bacilli contain, at one or both ends, small refractive bodies which are probably spores. Grows at the room temperature. In nutrient *gelatin* containing grape sugar, at the end of two days, punctiform colonies are developed which later attain the size of a poppy seed; they are spherical and have smooth outlines; the gelatin is not liquefied, but gas bubbles are formed, and a disagreeable odor is given off by the cultures, resembling decomposing perspiration from the feet. In the absence of grape sugar the development is scanty and there is no development of gas. In nutrient *agar* the colonies are but little larger, they are transparent, and under a low power resemble little flocculi of cotton. In *blood serum* development occurs only in the middle and lower part of the line of puncture.

In *bouillon*, when oxygen is excluded, an abundant development occurs at 37° C. at the end of twenty-four hours; the *bouillon* becomes clouded and gives off stinking gases.

403. *BACILLUS POLYPIFORMIS* (Liborius).

Found in the soil by inoculations in mice, in the excrement of cows, etc.

Morphology.—Slender bacilli of various lengths, a little more than 1 μ thick; do not form filaments.

Biological Characters.—An *anaërobic, non-liquefying* bacillus: exhibits very slight independent motion. Forms long-oval spores, which occupy one-half to two-thirds of the length of the rods, and the diameter of which is a little more than that of the bacilli. The colonies in *gelatin* consist of small, yellowish masses with irregular, broad, flap-like projections: under the microscope these are seen to consist of variously twisted and bent outgrowths, varying in thickness, which are given off in all directions like the tentacles of a polyp; later these outgrowths increase in thickness. In *agar* white colonies of irregular form, the size of a pin's head, are developed: under a low power these appear as finely granular, brownish, mulberry-like

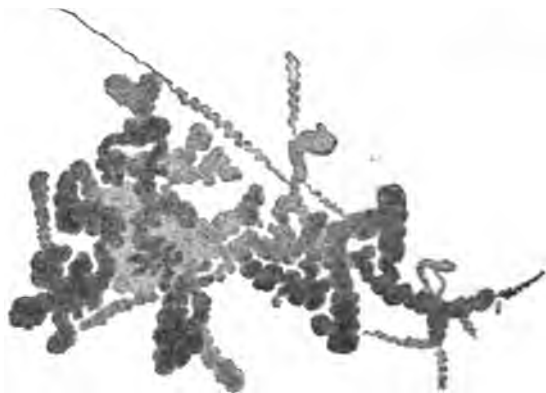


FIG. 230.—*Bacillus polypiformis*; colony in nutrient gelatin. $\times 80$. (Liborius.)

masses. In *blood serum* a diffuse cloudiness is developed at the bottom of the line of puncture. The addition of two per cent of sugar to culture media favors the growth of this bacillus; no gas is developed as a result of its growth in such a medium.

404. *BACILLUS BUTYRICUS* (Prazmowski).

Synonyms.—*Bacillus amylobacter*; *Clostridium butyricum*.

Found in putrefying vegetable infusions, in old cheese and milk, in the soil, etc.

Morphology.—Bacilli with round ends, from 3 to 10 μ long and about 1 μ thick; frequently associated in chains; also in filaments not apparently segmented. The rods assume a spindle or tadpole form when spore formation is about to occur, and may then be from 1.8 to 2.6 μ thick; the spores are oval, about 2 to 2.5 μ long and 1 μ broad, and are located centrally or at one extremity. When the spore vegetates a rupture of the exosporium occurs at one of the poles, and the bacillus grows out in the direction of the long axis of the reproductive element; the empty spore case retains its form after the vegetative cell is extruded.

Biological Characters.—An *anaërobic, motile* bacillus. Forms large oval spores. Grows at the room temperature, in the absence of oxygen. The

spores are destroyed by a temperature of 100° C. maintained for five minutes. In solutions containing starch, sugar, dextrin, or salts of lactic acid this bacillus produces a considerable quantity of butyric acid, and at the same time carbon dioxide and hydrogen are given off. The most favorable temperature for its development is from 35° to 40° C. According to Fitz, it does not cause the coagulation of sterilized milk, but the casein is slowly peptonized. This bacillus also causes the decomposition of cellulose, producing hydrogen and carbon dioxide, or methane, carbon dioxide, and sulphuretted hydrogen, according to the composition of the culture medium (Tappeiner). It appears to be the bacillus which usually gives rise to butyric acid fermentation in milk which has been kept for some time, and also in cheese; this occurs in milk after the lactic acid fermentation, which is due to aërobic bacilli, and especially to *Bacillus acidi lactici*. The characters of growth in solid media have not been determined. A peculiar staining reaction occurs when this

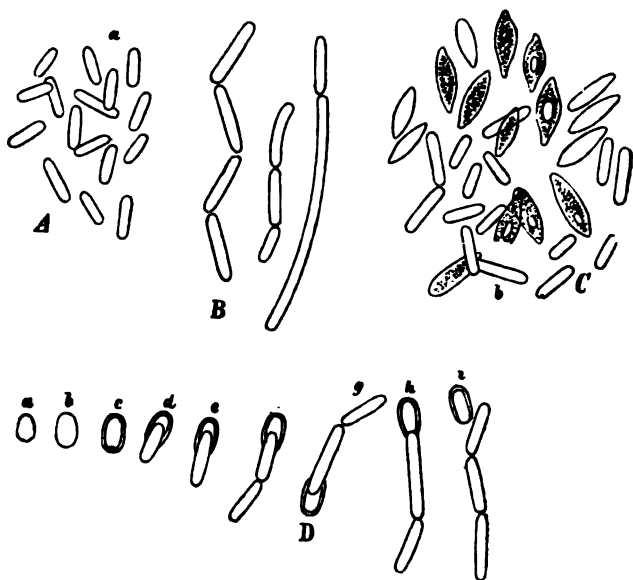


FIG. 281.—*Bacillus butyricus*; A, single bacilli; B, chains and filaments; C, spore formation, showing "clostridium form"; D, germination of a spore—a to i. $\times 1,000$. (Prasnowski.)

bacillus from cultures containing starch or cellulose is treated with iodine solution; the protoplasm of the cells acquires a blue or dark-violet color, the younger cells being of a pure blue; in some cases an oblique zone of blue only is seen, in others the entire cell is stained.

405. *CLOSTRIDIUM FETIDUM* (Liborius).

Obtained from garden earth by inoculations in mice, etc.

Morphology.—Bacilli of various lengths and about 1 μ thick; sometimes grow out into filaments. Forms large oval spores, located centrally or at one end of the rod; these are of greater diameter than the bacilli before spore formation, and cause the rods to have a spindle (clostridium) form, or occasionally an expanded extremity; resembles *Bacillus butyricus*.

Biological Characters.—An *anaërobic*, *liquefying*, *actively motile* bacillus. Forms spores. Grows in the usual culture media, in the absence of

oxygen, at the room temperature. Upon *agar plates*, in the absence of air, small, yellowish-white colonies are developed which are irregular in form and vary considerably in size; these are surrounded by outgrowths which are more compact and less branched than similar colonies of the bacillus of malignant oedema. In nutrient *gelatin* irregular, spherical colonies are developed, which rapidly cause liquefaction of the surrounding gelatin, and spherical cavities filled with a deeply clouded liquid are formed. In *blood serum* a homogeneous cloudiness is seen in the vicinity of the line of puncture, and at the lower part of this a few isolated, irregularly branching colonies are developed. A considerable amount of gas is formed in the cultures in various media; this appears as scattered bubbles, and also causes a splitting-up of the culture medium; in gelatin cultures liquefaction gradually extends upward until it reaches the surface. The gases evolved have an extremely disagreeable odor; they are produced more abundantly in culture media containing sugar.

406. BACILLUS LIQUEFACIENS MAGNUS (Lüderitz).

Found in garden earth by inoculations in mice and guinea-pigs.

Morphology.—Bacilli with slightly rounded ends, straight or slightly curved, from 3 to 6 μ long and from 0.8 to 1.1 μ thick; may grow out into long, flexible filaments.



FIG. 232. — *Bacillus liquefaciens magnus*; young colonies in nutrient gelatin. $\times 60$. (Lüderitz.)

Biological Characters.—An *anaerobic*, *liquefying*, *motile* bacillus. Grows rapidly at the room temperature, in the absence of oxygen, in the usual culture media. Forms long-oval spores, from 1 to 2 μ long and 0.8 μ broad; these are located at or near the middle of bacilli from 4 to 6 μ long—not in the long filaments unless these are segmented. When cultivated in a medium containing grape sugar, the spore-bearing bacilli are stained violet with iodine solution—sometimes pale and in places only, at others throughout. In *gelatin cultures* development is already evident, at the end of twenty-four hours, one or two centimetres below the surface, as scattered punctiform colonies; at the end of two days these are one to two millimetres in diameter and have smooth margins, with transparent, dull-white contents; at the end of three or four days the gelatin is usually

entirely liquefied; the liquefaction extends upward, and the liquefied gelatin, which at first is clouded, becomes clear, while a slimy, whitish deposit is seen at the bottom of the tube. In cultures containing grape sugar some gas is developed; this has a disagreeable odor like that of old cheese. In *gelatin stick cultures* liquefaction begins along the line of puncture, from 1 to 1.5 centimetres below the surface, within forty-eight hours; and a sausage-shaped collection of liquefied gelatin, of a dull-white, or silver-gray color, is seen. In long *agar cultures* development is rapid, and colonies are seen nearer the surface than in similar gelatin cultures—often within five millimetres; the young colonies have a delicately branched, moss-like appearance; in older colonies the branching is coarser. In stick cultures in *blood serum*, in the incubating oven, at the end of twenty four hours a simple line of growth is seen along the lower portion of the track of the inoculating needle; later numerous lateral offshoots give the growth a brush-like appearance; the blood serum is soon liquefied and foul-smelling gases are given off. Not pathogenic for mice or for guinea-pigs.

407. BACILLUS LIQUEFACIENS PARVUS (Lüderitz).

Obtained from garden earth by inoculations in mice and guinea-pigs.

Morphology.—Bacilli from 2 to 5 μ long and 0.5 to 0.7 μ thick; often grow out into long, flexible filaments.

Biological Characters.—An *anaërobic, liquefying, non-motile* bacillus. Grows rapidly at the room temperature, in the absence of oxygen, in the usual culture media. In bouillon cultures the thickness of many of the rods is increased to 1 or 1.2 μ , and in these small, round, refractive bodies are seen at the ends, or in a linear series, which are probably spores. The addition of two per cent of grape sugar to culture media is favorable to the growth of this bacillus. In *gelatin* cultures development occurs at a distance of one to two centimetres below the surface; at the end of two days, at 20° C., punctiform colonies are developed; these, when not too close together, attain a diameter of 2 to 2.5 millimetres; they are filled above with transparent liquid gelatin and below with a whitish mass of bacteria. In *gelatin stick* cultures a row of spherical colonies is developed along the line of puncture; these increase in diameter from above downward. In nutrient *agar* opaque colonies are developed within 0.5 to 1 millimetre of the surface; these are tabular, almond-shaped, or whetstone-shaped, and have a tolerably smooth contour at first; later they are irregular in outline. *Blood serum* is slowly liquefied by this bacillus. In *agar* cultures a few gas bubbles are developed, and also in blood serum. In *bouillon* a decided putrefactive odor is developed. Not pathogenic for mice.



FIG. 233.—*Bacillus liquefaciens parvus*; colony in nutrient agar. $\times 60$. (Lüderitz.)

408. BACILLUS RADIATUS (Lüderitz).

Obtained from garden earth by inoculations in mice and guinea-pigs.

Morphology.—Bacilli with round ends, from 4 to 7 μ long and about 0.8 μ thick; often grow out into long filaments, which are seen to be composed of separate segments.

Biological Characters.—An *anaërobic, liquefying, motile* bacillus. Movements are less active than in *Bacillus liquefaciens magnus*, and only observed in specimens from a recent culture. Spores are developed in the single rods only—not in the filaments; they are from 1.2 to 2 μ long and 0.8 to 0.9 μ thick, and are centrally located in the rods; the spore-bearing bacilli are somewhat thicker than others, but do not acquire a spindle shape. Grows in the usual culture media at the room temperature when oxygen is excluded. The addition of two per cent of grape sugar is favorable to the development of this bacillus. In test-tube cultures in nutrient *gelatin*, at 22° C., colonies are developed to within one or two centimetres of the surface; when these are numerous the gelatin is penetrated throughout, below the limit mentioned, with numerous glistening threads, and within two or three days is liquefied; some gas accumulates beneath the layer of solid gelatin above; later liquefaction extends to the surface, and the liquefied gelatin, which at first is clouded, gradually becomes transparent from

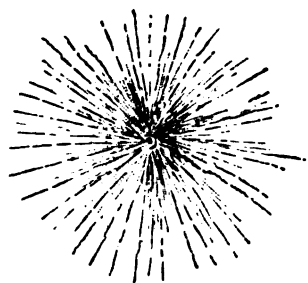


FIG. 234.—*Bacillus radiatus*; young colony in nutrient gelatin. $\times 60$. (Lüderitz.)

a deposition of the suspended bacilli. When only a few colonies are developed the growth resembles that of the mycelium of a fungus, the margin consisting of interlaced filaments, while the centre shows commencing liquefaction; new centres of development are formed by the filaments which penetrate the gelatin, and this is soon completely permeated and at the same

time liquefied; a similar appearance is observed when gelatin plates are prepared and kept in an atmosphere of hydrogen. When successive cultures are made in gelatin tubes the bacillus gradually loses its vigor of growth and the colonies have a different appearance; the thread-like outgrowths are no longer seen, or are developed to a slighter extent, and the spherical or balloon-shaped colonies are filled with a clouded liquid, while a thick sediment is formed. In *gelatin stick cultures* development occurs along the line of puncture to within one or two centimetres of the surface; at the end of two days numerous branching filaments are given off, which give the growth the appearance of a hairy caterpillar. In nutrient *agar stick cul-*

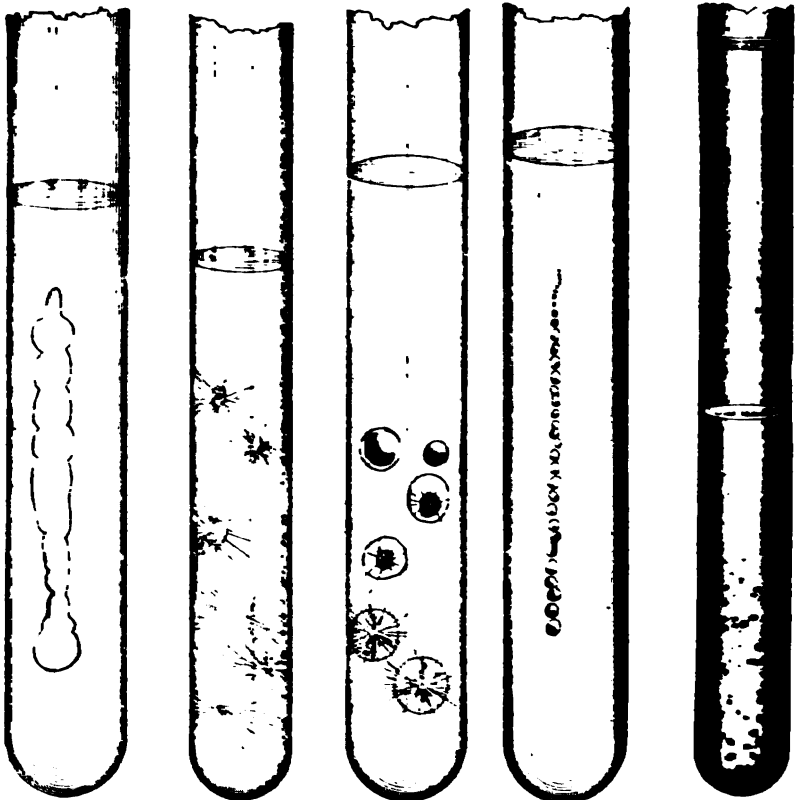


FIG. 235.

FIG. 236.

FIG. 237.

FIG. 238.

FIG. 239.

FIG. 235.—*Bacillus liquefaciens magnus*; stick culture in nutrient gelatin. (Löderitz.)

FIG. 236.—*Bacillus radiatus*; stick culture in nutrient gelatin. (Löderitz.)

FIG. 237.—*Bacillus sponosus*; stick culture in nutrient gelatin. (Löderitz.)

FIG. 238.—*Bacillus liquefaciens parvus*; stick culture in nutrient gelatin. (Löderitz.)

FIG. 239.—*Clostridium foetidum*; culture in nutrient agar, under oil. (Liborius.)

tures growth occurs to within one centimetre of the surface; it is branching in character and resembles that of *Bacillus liquefaciens magnus*, but the filaments are more delicate; later the principal branches are thicker, and they are surrounded by a denser mass of fine filaments, among which some thick outgrowths are also seen. *Blood serum* is rapidly liquefied by this

bacillus. The gases which are developed in cultures containing sugar have a disagreeable odor like that of cheese and onions; cultures in blood serum give off a putrefactive odor. Not pathogenic for mice.

409. BACILLUS SPINOSUS (Lüderitz).

Obtained from garden earth by inoculations into mice and guinea-pigs.

Morphology.—Bacilli with round ends, straight or curved, about $0.6\ \mu$ thick and of various lengths—usually from 3 to $8\ \mu$; may grow out into long, segmented, flexible filaments. Spores are formed in the single rods only and are long-oval in form; they are located toward one or the other extremity of the rod, which is from 1 to $1.2\ \mu$ broad at the point where they are developed, and has more strongly rounded or pointed ends than those not containing spores.

Biological Characters.—An *anaërobic, liquefying, motile* bacillus. Grows in the usual culture media, with the addition of two per cent of grape sugar, at the room temperature, in the absence of oxygen. In *gelatin* tube cultures, at the end of two days at 20°C ., colonies are formed in the deepest portion of the gelatin to within 3 or 3.5 centimetres of the surface; these are irregular in form and the size of a poppy- or hemp-seed; they contain liquefied gelatin, and as they increase in size the gray, shining contents are seen to contain a radiating, whitish growth; this is most marked in the deep-lying colonies, in which the radiating growth extends to the non-liquefied gelatin around the colony; later the colonies become confluent and the liquefaction slowly extends upward. In *agar* tubes opaque colonies are developed which may attain a diameter of four millimetres; under the microscope they are seen to be made up of numerous interlaced filaments which in old colonies are only seen in the marginal zone; or they may be composed of thick, knotty masses. *Blood serum* is liquefied by this bacillus. Gas is formed in all of the cultures, and in those containing sugar it has an odor which is compared to a mixture of Swiss cheese and fermented raspberry juice. Not pathogenic for mice or guinea-pigs.

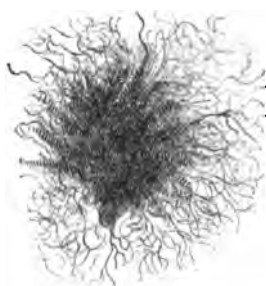


FIG. 240.—*Bacillus spinosus*; colony in nutrient agar, one day old. $\times 60$. (Lüderitz.)

410. BACILLUS ANAËROBICUS LIQUEFACIENS (Sternberg).

Obtained in anaërobic cultures from the contents of the intestine of a yellow-fever cadaver.

Morphology.—Slender bacilli, about $0.6\ \mu$ in diameter and three to five times as long as broad; often in pairs; grows out into long filaments.

Biological Characters.—An *anaërobic, liquefying, non-motile* bacillus. Forms spores. Grows in the usual culture media at the room temperature, when oxygen is excluded. In *gelatin* roll tubes (Esmarch's) filled with hydrogen, granular, white colonies are developed, around which the gelatin is liquefied. In a long stick culture in nutrient *agar* it grows along the line of puncture nearly to the surface. Pathogenic power not tested.

X.

NON-PATHOGENIC SPIRILLA.

411. SPIRILLUM SPUTIGENUM (Miller).

Found in the mouths of healthy individuals, especially along the margin of inflamed gums in neglected mouths—common.

Morphology.—Curved rods which resemble the “commas” of *Spirillum cholerae Asiaticæ*; these are frequently united in S-form or in short, spiral filaments—actively motile.

This spirillum was supposed by Lewis to be identical with Koch's “comma bacillus,” but Miller has shown that it differs essentially from this, inasmuch as it fails to grow in the usual culture media. He says: “Although I obtained nearly pure culture material in several cases and used all possible kinds of nutrient media, I did not succeed in producing the slightest growth of this fungus.”

412. SPIRILLUM DENTIUM.

Synonyms.—*Spirochæte denticola*; *Spirochæte dentium*.

Found in the mouths of healthy individuals, “under the margins of the

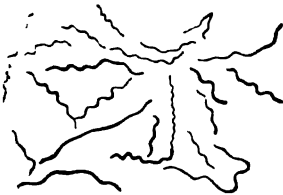


FIG. 241.

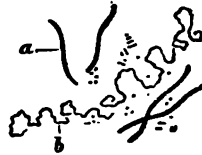


FIG. 242.



FIG. 243.

FIG. 241.—“*Spirochæte dentium*.” $\times 1,100$. (Miller.)

FIG. 242.—“*Spirochæte plicatilis* (b), *Vibrio rugula* (a), and other bacteria.” $\times 500$. (Flügge.)

FIG. 243.—“*Spirillum dentium*.” $\times 500$. (Flügge.)

gums, when they are covered with a dirty deposit and slightly inflamed” (Miller).

Morphology.—Long, flexible, spiral filaments of unequal thickness and irregular spiral windings; from eight to twenty-five μ long.

Biological Characters.—Not known, as all attempts to cultivate this spirillum have been unsuccessful.

413. SPIRILLUM PLICATILE.

Synonym.—*Spirochæte plicatilis*.

Found in swamp water containing decomposing algæ.

Morphology.—Long, flexible, spiral filaments. The spiral curves are close and regular, but the extremely long filaments present secondary, wave-like curves which are not uniform. The ends of the filaments are blunt. They may attain a length of one hundred to two hundred μ ; the movements are extremely active.

Biological Characters not determined.

414. *VIBRIO RUGULA* (Müller).

Found in swamp water, in faeces, and in tartar from the teeth.

Morphology.—Rod-shaped cells from 6 to 8 μ long and from 0.5 to 2.5 μ thick, slightly bent or with a flat spiral curve; sometimes united in long chains. Spores are developed at one extremity of the slightly curved rods; they are spherical, and the end of the rods containing them presents a spherical enlargement, giving them a comma-like appearance; terminal flagella have been demonstrated by Koch.

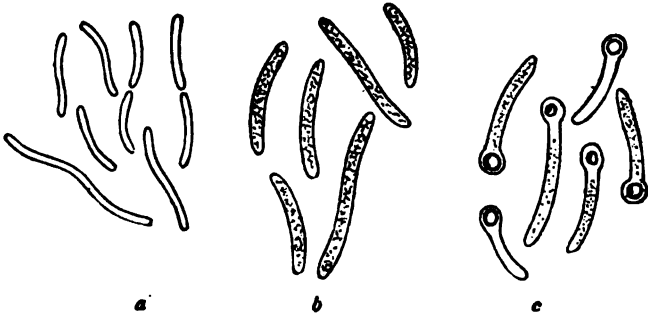


FIG. 244.—*Vibrio rugula*; a, young rods; b, thicker rods; c, spore-bearing rods. $\times 1,020$. (Prazmowski.)

Biological Characters.—The earlier investigators did not determine the characters of growth of *Vibrio rugula*, but Vignal (1886) has succeeded in cultivating a *strictly anaërobic* microorganism, obtained by him from the human mouth, which he has described under the same name, although he is not entirely satisfied that it is identical with *Vibrio rugula* of Prazmowski and previous observers. The biological characters, as determined by the author named, are as follows: An *anaërobic, liquefying, motile* "vibrio." Movements rotary and progressive. Forms terminal spores, which are rather pear-shaped than round. Upon *gelatin plates*, in the absence of oxygen, it forms spherical, opaque, yellowish colonies; about the third day the original colony is surrounded by a zone of transparent, liquefied gelatin. In *gelatin stick cultures*, in an atmosphere of hydrogen, development occurs along the line of puncture within twenty-four hours, and a small mass is formed at the point of puncture; at the end of forty-eight hours liquefaction occurs in funnel form; the liquefied gelatin is clouded, white, and opaque; later it becomes transparent. In long stick cultures in nutrient gelatin, without exclusion of the air, development occurs only at the bottom of the line of puncture. Upon the *surface of agar* a white, slightly wrinkled pellicle, covering the entire surface, is formed in an atmosphere of hydrogen. Development occurs in neutral or in acid bouillon, which becomes diffusely clouded, and from which an abundant white sediment is deposited. Upon the *surface of blood serum* a white layer is developed and liquefaction of the medium occurs. Upon *potato* a wrinkled, white layer is rapidly developed;

later this acquires a yellowish tint; the growth penetrates the potato to a considerable depth. Much gas is formed in all of the cultures, and they give off a strong fæcal odor. According to Prazmowski, *Vibrio rugula* causes an energetic decomposition of cellulose.

415. SPIRILLUM VOLUTANS (Ehrenberg).

Found in swamp water, etc.

Morphology.—Spiral filaments with round and somewhat pointed ends, from 1.5 to 2 μ thick and 25 to 30 μ long; each filament consists of two and one-half to three and one-half—rarely six or seven—spiral turns which are from 9 to 13 μ long; a single long, whip-like flagellum at each extremity; the protoplasm contains numerous opaque granules. Exhibits active rotary and progressive movements—sometimes motionless.

Biological Characters not determined.

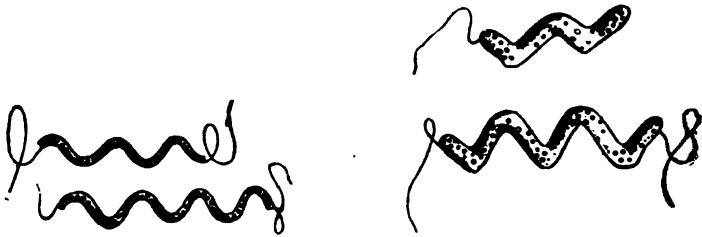


FIG. 245.

FIG. 246.

FIG. 245.—*Spirillum volutans*. $\times 600$. (Cohn.)

FIG. 246.—*Spirillum sanguineum*. $\times 600$. (Cohn.)

416. SPIRILLUM SANGUINEUM.

Synonym.—*Ophidomonas sanguinea*.

Found in brackish water containing putrefying marine algæ.

Morphology.—Rigid spiral filaments with round ends, 3 μ or more thick, and having two to two and one-half spiral turns, each 9 to 12 μ long; the protoplasm contains numerous refractive red granules.



FIG. 247.

FIG. 248.

FIG. 249.

FIG. 247.—*Spirillum serpens*. $\times 650$. (Flügge.)

FIG. 248.—*Spirillum tenue*. $\times 650$. (Flügge.)

FIG. 249.—*Spirillum undula*. $\times 650$. (Flügge.)

417. SPIRILLUM SERPENS (Müller).

Synonym.—*Vibrio serpens*.

Found in stagnant water, vegetable infusions, etc.

Morphology.—Rigid filaments having two or three wave-like undulations, from 11 to 28 μ long and 0.8 to 1.1 μ thick; sometimes united in chains. Actively motile and often associated in closely crowded swarms.

Biological Characters not determined.

418. *SPIRILLUM UNDULA* (Ehrenberg).

Found in putrefying animal and vegetable infusions.

Morphology.—Rigid, spiral filaments, from 8 to 12 μ long and from 1.1 to 1.4 μ thick. Each spiral turn has a length of 4 to 5 μ and each filament from one-half to three turns. A long, whip-like flagellum may be demonstrated at each extremity. The movements are rotary and rapidly progressive—darting.

Biological Characters not determined.

419. *SPIRILLUM TENUE* (Ehrenberg).

Found in putrefying vegetable infusions, etc.

Morphology.—Slender spiral filaments, from 4 to 15 μ long. The height and length of a single turn are from 2 to 3 μ , and each filament has from one and a half to five turns. Often associated in closely crowded swarms. Motions rotary and progressive—extremely rapid.

Biological Characters not determined.

420. *SPIRILLUM LINGUÆ*.

Synonyms.—*Vibrio lingualis*; *Zungenbelagvibrio* (Weibel).

Obtained from deposit upon the tongue, by inoculation in a mouse.

Morphology.—Curved rods of the size of the cholera spirillum; sometimes in S-shape. Grow out into longer or shorter wavy filaments, the extremities of which are sometimes enlarged—button-like. Involution forms are common.

Stains by Gram's method, and is differentiated by this from other "vibrios" described by Weibel.

Biological Characters.—An *aërobic* and *facultative anaërobic*, non-liquefying, non-motile spirillum ("vibrio").

Spore formation not determined. Grows at the room temperature in the usual culture media. Upon *gelatin plates* forms dirty-white colonies, which at the end of a week may attain a diameter of one millimetre. Under a low power the margin of the deep colonies is seen to consist of fine, white, interlaced filaments, with irregular offshoots. The margin of the superficial colonies has a greenish-yellow shimmer, and thread-like offshoots are given off in a tangential direction; the contour is round. In *gelatin stick cultures* a delicate, white, veil-like stripe is developed along the line of puncture, and no growth occurs upon the surface. Upon the *surface of agar* a dirty-white, finely granular—not slimy—layer is developed. In *bouillon* a flocculent deposit collects at the bottom of the tube, and the liquid above is slightly clouded. The flocculi in bouillon cultures consist of closely interlaced filaments, and frequently shorter rods or fragments are so placed as to give the impression that they are bud-like offshoots from the longer filaments.

Not pathogenic for white mice.



FIG. 360.—*Spirillum linguae*. (Weibel.)

421. *SPIRILLUM NASALE*.

Synonyms.—*Vibrio nasalis*; *Nasenschleimvibrio* (Weibel).

Found in nasal mucus.

Morphology.—Curved rods with rounded ends, about as thick as the an-

thrax bacillus, and two to five times as long as thick; the amount of curvature varies considerably, from nearly straight to semicircular, and short rods may be quite straight. In preparations from agar or gelatin cultures long, closely wound spiral filaments are sometimes seen, or the filaments may be wavy or made up of curved segments.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* spirillum. Grows slowly at the room temperature—better at 37° C. Spore formation not determined. Upon *gelatin plates*, at the room temperature, grows very slowly; by the fifth day the colonies may have a diameter of 0.3 millimetre, and finally of 0.6 millimetre at the outside; under a low power they are seen to be finely granular, and yellowish-brown by transmitted light; they are spherical in form, with sharply defined margins. In *gelatin stick cultures* a delicate, veil-like growth is seen along the track of the needle; no growth upon the surface. Upon the *surface of agar* a dirty-white, slimy layer is developed at 37° C. No growth upon *potato*.

422. SPIRILLUM α OF WEIBEL.

Synonym.—*Vibrio saprophiles* α (Weibel).

Found in putrefying hay infusion and in slime from sewers.

Morphology.—Curved rods, with pointed ends, about 0.6 μ thick at the centre, and averaging 3 μ in length; often two are united in S-form—longer



FIG. 251.

FIG. 251.—*Spirillum* α of Weibel. (Weibel.)



FIG. 252.

FIG. 252.—*Spirillum* β of Weibel. (Weibel.)

chains are not common; grow out into spiral filaments; involution forms common.

Biological Characters.—An *aërobic*, *non-liquefying*, *actively motile* spirillum. Grows rather slowly at the room temperature. Spore formation not determined. Potato cultures give off a strong odor of ammonia. Upon *gelatin plates* the deep colonies, by the third day, are from 0.2 to 0.3 millimetre in diameter; later they may attain a diameter of 0.6 millimetre; under a low power they are seen to be spherical and yellowish-brown in color; the opaque centre is surrounded by concentric rings. The superficial colonies are flat white or yellowish discs, irregular in outline, with a finely granular structure; the centre is greenish-yellow and opaque, and the color fades out toward the periphery; at the end of a week they may attain a diameter of two millimetres. In *gelatin stick cultures* a veil-like, white growth is seen along the line of puncture; later this has a dirty yellowish-red color; upon the surface a whitish layer gradually extends from the point of puncture, and beyond this a transparent, whitish film covers the surface. Upon the *surface of agar* a cream-like, yellowish-white layer is developed, and the agar beneath is clouded to a depth of one to two millimetres. Upon *potato*, at the end of two days, an abundant slimy layer is developed; this has a yellowish-red to chocolate-brown color, and resembles the growth of the bacillus of glanders.

423. *SPIRILLUM* β OF WEIBEL.

Synonym.—*Vibrio saprophiles* β (Weibel).

Found in putrefying hay infusion.

Morphology.—Slender, curved rods, of about the thickness of the tubercle bacillus, and averaging 2μ in length; the ends are blunt; frequently two elements are united in S-form, but long filaments do not occur; involution forms common.

Biological Characters.—An *aërobic*, *non-liquefying*, *actively motile* spirillum ("vibrio"). Spore formation not determined. Grows rather slowly at the room temperature. Upon *gelatin plates* the colonies never exceed 0.3 millimetre in diameter; they are spherical, and by transmitted light have a yellowish-brown color. In *gelatin stick cultures* the growth is similar to that of the preceding species—*Spirillum* α . In *agar stick cultures* no growth occurs along the line of puncture; on the surface a cream-like, yellowish-white, viscid layer is formed, which cannot be raised without bringing away some of the culture medium. Upon *potato* a thin,

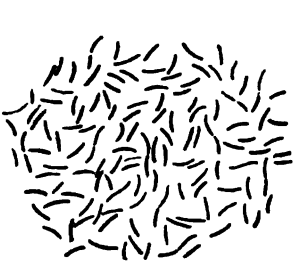


FIG. 253.

FIG. 253.—*Spirillum* γ of Weibel. (Weibel.)



FIG. 254.

FIG. 254.—*Spirillum aureum*. (Weibel.)

shining, varnish-like layer of a dirty brownish-green color and a viscid, dry consistence, which is with difficulty removed by the platinum needle.

424. *SPIRILLUM* γ OF WEIBEL.

Synonym.—*Vibrio saprophiles* γ (Weibel).

Found in the slime deposited in sewers.

Morphology.—Curved rods with round ends, one-half larger than "*Vibrio saprophiles* α " of Weibel; S-shaped forms are seen, but spiral filaments are rare; involution forms very common in old cultures.

Biological Characters.—An *aërobic*, *non-liquefying*, *actively motile* spirillum ("vibrio"). Spore formation not determined. Grows rather quickly at the room temperature. Upon *gelatin plates* the deep colonies are white, and at the end of a week about 0.5 millimetre in diameter; under a low power they are seen to be spherical, granular, and orange-colored at the centre, with a sharply defined, pale-yellow marginal zone. The superficial colonies are flat, dirty-white, slightly opalescent, with a more prominent white centre; under a low power the margin is irregular and notched, the marginal zone white and marked by numerous fine, anastomosing furrows, next to this a light ochre-yellow zone marked by darker lines, and at the centre a golden-brown nucleus marked by delicate, dark, interlacing lines; at the end of a week the superficial colonies may attain a diameter of five millimetres. In *gelatin stick cultures* development occurs along the line of

puncture and to a moderate extent upon the surface. In *agar stick cultures* no growth occurs along the track of the inoculating needle; on the surface a dirty-white, pap-like layer is developed. Upon *potato* the growth is sometimes dry, viscid, and dark-brown in color, or it may be of a mahogany-brown with a moist lustre.

425. SPIRILLUM AUREUM.

Synonym.—*Vibrio aureus* (Weibel).

Found in the air and in the slimy deposit in sewers.

Morphology.—Curved rods with blunt ends, about one-half thicker than the *Spirillum cholerae Asiaticae*, and varying greatly in length; typical "commas" and S-forms are seen; also filaments of various lengths and sometimes regular spiral filaments which are extremely slender; also involution forms.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* spirillum. Produces a golden or orange-yellow pigment. Spore formation not determined. Grows rather rapidly at the room temperature—also in the incubating oven. Upon *gelatin plates* the deep colonies, at the end of three days, have a diameter of 0.3 millimetre (in ten days of one millimetre); under a low power they are seen to be coarsely granular, spherical or whetstone-shaped, golden-yellow in the centre and later brown or black, with more transparent, golden-yellow margins. The superficial colonies are round, with well-defined margins, granular, and of a pure golden-yellow color; at the end of three days they have a diameter of one millimetre and in ten days of three to four millimetres. In *gelatin stick cultures* a tolerably abundant, finely granular growth is seen along the line of puncture; upon the surface a rounded mass of a yellow-ochre color is developed about the point of puncture. Upon the *surface of agar* a dirty-white layer extends over the entire surface; later round, elevated, golden-yellow islands are seen, and finally a uniform pap-like layer two millimetres thick. Upon *potato* an abundant, thick, pap-like growth of a golden-yellow or orange-yellow color.

426. SPIRILLUM FLAVESCENS.

Synonym.—*Vibrio flavescens* (Weibel).

Found in the slimy deposit of sewers.

Morphology.—The same as *Spirillum aureum*.

Biological Characters.—An *aërobic, non-liquefying, non-motile, chromogenic* spirillum. Produces a dirty yellowish-green pigment. Grows rather rapidly at the room temperature. Upon *gelatin plates* the colonies resemble those of *Spirillum aureum*, except that the yellow color is developed later and the shade is paler, duller, and less pure—upon a transparent background a dirty yellowish-green. In *gelatin stick cultures* a finely granular line of growth is seen along the track of the needle, and along the margin of this in old cultures are coarser granular masses; upon the surface a pale-yellow, flat layer with flap-like margins slowly extends from the point of puncture. Upon *agar* a dirty-white color is developed, which gradually extends to the walls of the test tube; elevated, round, yellow masses are developed in this, which increase in diameter, become confluent, and finally form a thick and uniform pap-like layer. Upon *potato* an abundant dull-yellow, pap-like layer.

427. SPIRILLUM FLAVUM.

Synonym.—*Vibrio flavus* (Weibel).

Found in the slimy deposit in sewers.

Morphology.—The same as *Spirillum aureum*.

Biological Characters.—The same as *Spirillum aureum*, but the pigment produced is ochre-yellow. Upon a dark background the colonies appear grayish-yellow, on a white ground straw-yellow; under a low power the deep colonies are first pale-yellow then golden-yellow, and of the same tint throughout, without being darker in the centre; they are finely granular, with a net-like marking. The superficial colonies under the microscope are pale-yellow with dull-gray spots; on the margin the color usually remains white; upon the *surface of agar* an ochre-yellow layer; the same on *potato*.

428. SPIRILLUM CONCENTRICUM (Kitasato).

Found in putrefying blood.

Morphology.—Short spirilla with pointed ends, with two to three spiral turns which are 3.5 to 4 μ in length and 2 to 2.5 μ in diameter—i.e., each complete spiral; the filaments are a little thicker than the cholera spirillum; in bouillon they grow out into long spirals having from five to twenty turns.

Biological Characters.—An *aërobic, non-liquefying, actively motile* spirillum. Grows best at a temperature of 20° to 23° C. Spore formation not observed. Upon *gelatin plates* the colonies, by transmitted light, are seen, to be made up of concentric rings, which from the centre to the periphery are alternately opaque and transparent; the contour is round, and from the margin numerous small, spiral outgrowths are given off. In *gelatin stick cultures* development occurs principally upon the surface as a cloudy layer penetrating the medium to a depth of one millimetre. Upon the *surface of agar* a diffuse growth which adheres firmly to the culture medium. Upon *potato* no growth occurs. In *bouillon* a diffuse cloudiness is slowly developed; later the medium becomes clear and an abundant, slimy deposit is seen at the bottom of the tube.

429. SPIRILLUM RUBRUM (Von Esmarch).

Obtained from the putrefying cadaver of a mouse.

Morphology.—Spirilla about twice as thick as the cholera spirillum and having from one to three spiral turns; in bouillon grows out into very long, spiral filaments.

Biological Characters.—An *aërobic and facultative anaërobic, non-liquefying, actively motile, chromogenic* spirillum. Produces a wine-red pigment in the absence of oxygen only; on the surface of culture media the growth is colorless. According to Löffler, this spirillum has numerous short flagella. In old cultures unstained, slightly refractive bodies are seen in the filaments, which appear to be spores. Grows very slowly—best at 37° C. Upon *gelatin plates* small, slightly granular colonies with a tolerably smooth contour are developed at the room temperature; these are at first gray and bluish-red, later wine-red in color. In *gelatin stick cultures* closely crowded, isolated, spherical colonies are developed along the line of puncture; the growth, except upon and near the surface, has from the outset a beautiful wine-red color. Upon the *surface of agar* a grayish-white layer of limited extent is developed; later this has a pink color. Upon *potato* deep-red colonies the size of a hempseed are developed.

430. SPIRILLUM OF SMITH.

Found in the intestine of swine.

Morphology.—Comma-shaped rods and spiral filaments of one and a half to two or even as many as ten spiral turns; have terminal flagella.

Biological Characters.—An *aërobic, non-liquefying, motile* spirillum. Grows at the room temperature. Upon *gelatin plates*, at the end of thirty-six to forty-eight hours, small, spherical, finely granular colonies are developed, which have a brownish color; and upon the surface small, round colonies with a somewhat irregular contour. In roll tubes, when the colo-

nies are not crowded, they may have a diameter of 0.3 to 0.5 millimetre, and at the end of several days appear to be made up of concentric zones; at the end of several weeks they have the appearance of the end of a tree trunk which has been sawed off and shows many concentric rings of growth, which are not very clearly defined. Under certain circumstances outgrowths occur around some of the deep colonies, which give them the appearance of a raspberry. The superficial colonies may attain a diameter of three to five millimetres; they remain flat and circular in outline and acquire a slight yellowish color. Upon *agar plates* the deep colonies have a diameter of 0.5 to 0.7 millimetre; those upon the surface are flat, round discs of a gray color and smooth appearance; they may attain a diameter of five millimetres. In neutral *bouillon* containing one-fourth to one per cent of peptone, at 36° C., development is abundant, and the culture liquid in a few days is densely clouded; examined in a hanging-drop culture, they appear a little larger than the spirillum of cholera and exhibit very active movements. They grow in *milk* without producing any perceptible change in this fluid. The cultures acquire a slightly alkaline reaction. Upon *potato*, at 37° C., a thin, yellow layer is developed in the course of a few days.

Not pathogenic for guinea-pigs or pigeons.

431. SPIRILLUM OF MILLER.

Synonym.—Miller's bacillus.

Obtained by Miller (1884) from carious teeth.

Morphology.—Straight or slightly curved rods, frequently in pairs in form of a letter S or of an O; also in homogeneous or segmental spiral filaments.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *motile* spirillum. Spore formation not observed. Grows at the room temperature in the usual culture media. No growth upon the *surface of gelatin cultures*, which are liquefied. Upon *agar* the growth is similar to that of the *spirillum* of Finkler and Prior. Growth upon *potato* not characteristic.

XI.

LEPTOTRICHEÆ AND CLADOTRICHEÆ.

Zopf and other systematic botanists place among the bacteria certain microorganisms which are more interesting to the student of general biology than to the pathologist, but which require description in a manual of bacteriology. In our descriptions of the species included by Zopf in the Leptotricheæ and Cladotricheæ we shall follow the author named. Four genera are included in the LEPTOTRICHEÆ, viz.: *Crenothrix*, *Beggiatoa*, *Phragmidiothrix*, and *Leptothrix*. The CLADOTRICHEÆ are included in a single genus: *Cladothrix*.

432. CRENOTHRIX KÜHNIANA (Rabenhorst).

Synonym.—Brunnenfaden.

Very common in running or in stagnant water. It sometimes develops so abundantly in reservoirs and conduits of water that the water supply is unfit for drinking or for certain industrial purposes.

Morphology.—In different stages of development appears as cocci, short rods, and long filaments. The cocci are small spheres of from 1 to 6 μ in diameter; the cell wall of these becomes gelatinous and they multiply by binary division, the gelatinous capsule of the daughter cell remaining enclosed in that of the mother cell; later they are set free by the solution of this gelatinous envelope; the zoöglœa formed by these cocci are irregular in form and may attain a diameter of one centimetre or more. These zoöglœa sometimes accumulate in enormous masses in reservoirs of water; at first they are colorless, but later they are colored by hydrated oxide of iron and appear brick-red, olive green, dark-brown, or brownish-black. When cultivated in swamp water these cocci grow out into rods, which by binary division produce filaments; these are seen projecting in all directions from the zoöglœa masses. When these reach a certain age they become segmented, and the segments, enclosed in a common sheath, acquire a rusty-red or dark-brown color from being impregnated with oxide of iron; the rod-shaped segments break up into spherical bodies which are comparatively large ("macrococci"); some broad filaments, however, contain disc-like segments, which break up into smaller cocci. The rod-shaped and spherical segments escape from the ruptured extremity of the common sheath. Sometimes the sheath becomes prematurely gelatinous, and the cocci and rods remain *in situ* and germinate; in this case they break through the gelatinous walls, and the original filament is seen to be surrounded by a brush-like outgrowth of filaments. In these secondary filaments, as well as in the primary, there is a distinct differentiation of the two extremities, growth occurring at the free extremity and not at the base. The filaments are sometimes wavy or even spiral in form.

Biological Characters not well determined, owing to the difficulty of cul-

tivating this microorganism in artificial media. It is *aërobie*. Genuine spore formation has not been demonstrated.

The *BEGGIATO*A are distinguished by the presence of grains of sulphur in the vegetative cells; these are seen as highly refractive granules with a dark contour. They are widely distributed, and are found both in salt and fresh water containing decomposing animal or vegetable material; in sulphurous waters they are especially abundant, and accumulate upon the muddy bottom, or upon organic substances undergoing decomposition, as a milk-white, gray, pink, or purple layer; the bottom of ponds or of small bays is often colored red by an extended and abundant growth of these water bacteria. Meyer has shown that they are able to decompose sulphate of

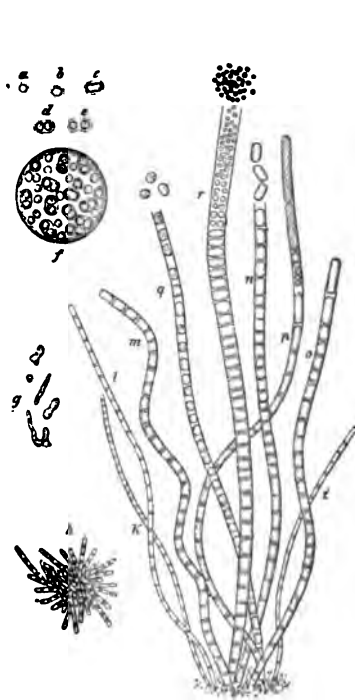


FIG. 255.

FIG. 255.—*Crenothrix Kühniana*; *a-e*, cocci in various stages of division; *f*, zoëgloea of cocci ($\times 400$); *g*, zoëgloea of various forms (natural size); *A*, colony of short filaments, composed of rod-shaped cells, and developed from a group of cocci; *i-r*, filamentous forms, partly spirally bent and of different thicknesses. (Zopf.)

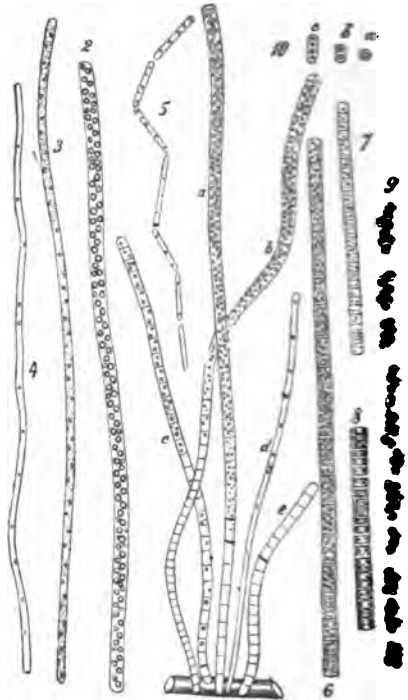


FIG. 256.

FIG. 256.—*Beggiatoa alba* ($\times 400$); *a* and *b*, filaments having an evident difference between base and extremity, being segmented and free from sulphur grains below; 2-5, fragments of filaments of various thicknesses and containing a greater or less number of sulphur grains; 6-8, filaments stained with methyl violet, showing segmentation into rods and cocci-like elements; 9, cocci; 10, development of a rod, *c*, from a coccus, *a* ($\times 800$). (Zopf.)

soda in organic solutions suitable for their growth. Like the previously described genus (*Crenothrix*), spherical, rod-shaped, filamentous, and spiral forms are included in the life history of the species. The filaments show a differentiation as to base and free, growing extremity, but, unlike the *Crenothrix*, the segments into which the filaments divide are not included in an external sheath. The filaments are flexible and exhibit a gliding movement;

they are able to multiply abundantly in thermal sulphur waters having a temperature of 55° C. and above.

433. BEGGIATOA ALBA (Vauch.).

This is an extremely common and widely distributed species; found especially in thermal waters and in the refuse waters from sugar refineries and factories of different kinds.

Morphology.—The filaments differ greatly in diameter as well as in length—from 1 to 5 μ in diameter; the young and slender filaments often contain but few grains of sulphur or none at all; the older filaments usually contain a considerable number of fine or coarser granules of sulphur. The filaments, which are attached to some substance, are usually seen to be segmented, even without the use of reagents, at least near the base where the grains of sulphur are less numerous or absent; the free ends, containing numerous grains of sulphur, may not appear to be segmented, but when they are stained with one of the aniline colors, or treated with hot glycerin, segmentation becomes apparent. In the thicker threads the segments divide into thin discs, and these again, under certain circumstances, divide, in a direction parallel with the long axis of the filament, into quadrants, each of which later becomes a spherical or ellipsoidal coccus; these usually contain one or several large grains of sulphur. These cocci remain attached for a time, then under favorable conditions become separated and enter upon the "swarm stage" of their existence, during which they are endowed with active movements. They come to rest upon some filament of an alga or other substance, which may be completely covered, and has a dark color as a result of their presence. They multiply by binary division and form zoöglœa masses of irregular form. Under certain circumstances they form straight or curved rods, which may also exhibit active movements. When these come to rest they grow out into long filaments, which may be straight or spirally curved. Fragments of the spiral filaments may, under certain circumstances, become actively motile and resemble genuine spirilla; their movements are due to the presence of terminal flagella. The diameter, length, and the height of the spiral turns vary greatly. The straight filaments also show a decided tendency to break up into longer or shorter fragments, but these do not exhibit active motions; they, in common with the fragments of spiral filaments, are, however, flexible and exhibit "crawling" motions.

434. BEGGIATOA ROSEO-PERSICINA (Zopf).

Synonyms.—*Clathrocystis roseo-persicina* (Cohn); *Ophidomonas sanguinea* (Ehrenberg); *Bacterium rubescens* (Lankester).

Found upon the surface of putrefying animal or vegetable material in fresh or salt water.

Morphology.—Presents the same developmental forms as *Beggiatoa alba*, viz. cocci, rods, filaments, and spiral threads. The filaments correspond with those of *Beggiatoa alba*, but are distinguished by their red or violet color. The cocci, formed in the filaments, when set free undergo binary division and form characteristic zoöglœa of very various forms; these were formerly described by Cohn under the name of *Clathrocystis roseo-persicina*. These zoöglœa may be spherical, oval, irregularly branched, etc.; at one time they may contain but a little, and at another considerable gelatinous intercellular material. Rods are developed from the cocci under certain circumstances, which may be straight or curved. In suitable media both the cocci and the rods, after the intercellular substance is dissolved, may become actively motile—"swarm stage." The short rods grow out into filaments, and these may, under certain circumstances, become spiral in form, either partially or entirely; the spiral filaments may break up into motile fragments which correspond with *Ophidomonas sanguinea* of Ehren-

berg (*Spirillum sanguineum*, No. 416). The red pigment produced by this species is known as bacterio-purpurin. It is insoluble in water, alcohol, chloroform, ammonia, or acetic acid, and is changed by hot alcohol into a brown, and by chloroform into an orange-brown, substance.

435. BEGGIATOA MIRABILIS (Cohn).

Found in sea water, upon the surface of putrefying animal and vegetable substances, as a white layer; common upon the coasts of Denmark and Norway (Warming).

Morphology.—This species is distinguished by the considerable thickness of the filaments, which may be as much as $30\ \mu$; these undergo segmentation into cylindrical masses, which again break up into thin discs, and these are supposed to divide into cocci-like elements such as Zopf has described in the other species of this genus. The complete life history of this species has not been determined. Like the other species described, the cells contain granules of sulphur.

436. PHRAGMIDIOTHRIX MULTISEPTATA (Engler).

Found in sea water, attached to crabs

Morphology.—Filaments from 3 to $6\ \mu$ in diameter, made up of thin disc-like segments, the diameter of which is four to six times less than the thickness; these cylindrical segments undergo segmentation into halves and quadrants, and finally into still smaller fragments which become rounded and resemble cocci; these probably are set free, but this has not been observed; apparently they grow out first into very thin filaments which subsequently increase in diameter. The genus to which this species belongs is distinguished from *Beggiatoa* by the absence of sulphur grains, and from *Crenothrix* by the fact that the segments are not enveloped in an exterior sheath, as well as by the comparative thinness of the cylindrical segments.

LEPTOTHRIX BUCCALIS.—According to Zopf, the leptothrix so common in the human mouth presents the same variety of forms as has been ascribed to the *Beggiatoa* and *Crenothrix*. We cannot accept this as established, and have described *Leptothrix buccalis* among the bacilli (No. 395). Vignal claims to have cultivated this species, but, in view of the failure of Miller and others to obtain it in cultures, it appears doubtful whether the microorganism described by him under this name corresponds with the *Leptothrix buccalis* of Robin (see page 683).

437. CLADOTHRIX DICHOTOMA (Cohn).

Found in stagnant and running water—very common. It is frequently associated with *Beggiatoa* and is common in the refuse water of factories, especially of sugar factories. In Russia it is often found in abundance in the water supply of towns. It may readily be obtained from the surface of putrefying algæ or animal substances immersed in river or swamp water.

Morphology.—According to Zopf, the cocci-like reproductive elements grow out into rods, and these into fine filaments, from which later pseudo-branches are given off. This apparent branching of the filaments is the distinguishing generic character of *Cladotrix*. Under a sufficiently high power the branched appearance is seen not to be a true dichotomous ramification, but to result from the growing out in a lateral direction of a detached segment. These long filaments break up again into long rods, these into shorter rods, and finally into cocci. The sheath of the filaments is often colored yellow, rusty-red, olive-green, or dark-brown by oxide of iron. The segments are forced from the free extremity of the common sheath by the

growth and binary division of those lying deeper, or by their own motility—"swarm stage"; they may emerge from the sheath either solitary or in chains of several elements. Sometimes the cocci-like forms germinate within the common sheath and grow out through its walls into filaments. Fragments of the filaments are sometimes seen to exhibit peculiar gliding movements; again they may exhibit very active movements as a result of the development of terminal flagella. The filaments are sometimes straight and sometimes twisted in spiral form, the spiral turns being sometimes quite flat and at others well-developed corkscrew-like convolutions. When the spiral filaments break up into motile fragments provided with flagella they

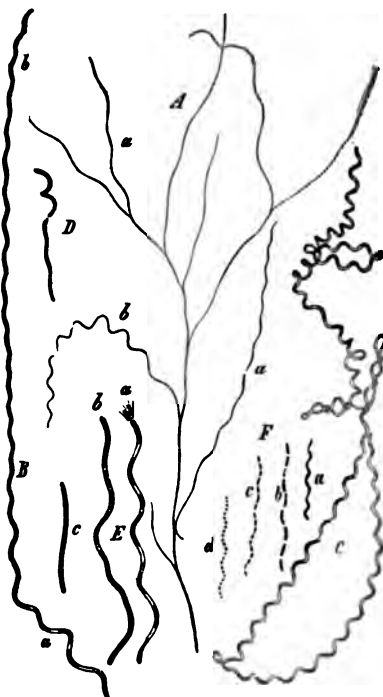


FIG. 437.—*Cladotrix dichotoma*; A, branching plant with wavy (a) or spiral (b) filaments; B, spiral filament more highly magnified; C, long spirochæte-like filament; D, fragment with one extremity spiral; E, spiral filaments segmenting into rods (b) and cocci (c); F, spirochæte form, a undivided, b dividing into long rods, c into short rods, d into cocci-like elements. (Zopf.)

resemble genuine spirilla. According to Zopf, the so-called zoöglœa ramigera is one form of development of *Cladotrix dichotoma*.

Biological Characters not determined.

438. CLADOTRICH FOERSTERI.

Synonym.—*Streptothrix Foersteri* (Cohn).

Found by Gräfe in the lachrymal ducts of the human eye.

Morphology.—Forms cocci-like masses, rods, and leptothrix filaments, which may be spirally curved. According to Zopf, this species closely resembles *Cladotrix dichotoma*.

Biological Characters not determined.

439. CLADOTHRIX INTRICATA (Russell).

Obtained by Russell (1891) from mud at the bottom of the Gulf of Naples.

Morphology.—Differs considerably, according to the age of the culture and the culture medium. In gelatin it forms long filaments made up of long and slender cells having homogeneous contents. In potato cultures the cells are shorter and with rounded ends; these elements divide into several short and thick separate cells, and many of these finally contain a slender, oblong spore. The filaments in agar and potato cultures may present the appearance characteristic of the genus *Cladotrix*, viz., a false branching.

Stains with the usual aniline colors.

Biological Characters.—An *aërobie*, *liquefying*, *slightly motile* cladotrix. Forms long-oval spores. Grows in the usual culture media at the room temperature. Upon *gelatin plates* colonies are developed in from twenty-four to thirty-six hours; these resemble, to the naked eye, colonies of mould fungi; under a low power the interior of the colony is seen to be made up of a thick network of filaments, from which a quantity of curled and intertwined filaments extend in all directions. The outside filaments are often tolerably straight, but they soon become more or less spiral and intertwined, or are united into interwoven, braid-like masses which extend in various directions from the principal colony. Liquefaction of the gelatin quickly occurs, and the filaments form in the liquefied gelatin a felt-like mass. When a cover glass is placed over a young colony and a microscopical examination is made with a tolerably high power, the straight filaments at the margin of the colony are seen to present pseudo-branches. In *gelatin stick cultures* development is rapid; at the end of twenty-four hours the line of inoculation is marked by finely curled filaments, which extend horizontally into the gelatin in all directions, the growth being most abundant near the surface. The gelatin near the surface is soon liquefied, and the liquefaction gradually extends downward. Upon *potato* an irregular, dull-white mass is quickly developed which is not especially characteristic; this ceases to extend after three days. Upon *agar* a tolerably abundant but thin, dull white layer is developed, and fine filaments extend from this into the culture medium, giving the culture a characteristic appearance. An abundant development occurs in *bouillon* and a jelly-like mass accumulates at the bottom of the tube; this is readily broken up by shaking the tube.

XII.

ADDITIONAL SPECIES OF BACTERIA, NOT CLASSIFIED.

440. NITROMONAS OF WINOGRADSKY.

Obtained from the soil at Zurich by Winogradsky (1890), who says: "In speaking of a nitrifying ferment I do not wish to affirm that there is but a single species capable of exercising this function over the whole surface of the globe. That appears to me not at all probable. . . . But it is probable that there are but few. At Zurich, for example, up to the present time I have found but a single one."

Morphology.—Ellipsoidal cells, more or less elongated, the youngest cells often nearly spherical; 0.9 to $1\ \mu$ broad and from 1.1 to $1.8\ \mu$ long; the longer cells already show the central constriction which precedes binary division; sometimes among the oval cells spindle-shaped cells are seen, and occasionally this is the prevailing form. As a rule, the cells do not remain associated after binary division has occurred, but occasionally a chain of three or four elements is seen. Irregular masses are formed in cultures, which are held together loosely by a gelatinous material.

Biological Characters.—Does not grow in the usual culture media, but was cultivated by Winogradsky in a solution containing one gramme of potassium phosphate and one gramme of ammonium sulphate in one thousand grammes of pure water. To this solution he adds half a gramme to a gramme of basic magnesium carbonate.

In testing the nitrifying power of the ferment the ammonium sulphate is added in separate portions, a standard solution of ten grammes in five hundred cubic centimetres of water being used. From two to five cubic centimetres of this solution are added at intervals of twenty-four to forty-eight hours, according to the rapidity of the ferment action, and the absence of ammonia as a result of this action is determined by the use of Nessler's solution. The cultures are kept at the room temperature.

The nitromonas of Winogradsky does not form spores. It is sometimes seen to exhibit active movements, but is usually at rest. The researches of the author named show that it is capable of growing and of exercising its ferment action in solutions from which all organic matter has been excluded, and the conclusion is reached that it is able to assimilate the carbon required for its development from the carbonic acid set free in the culture liquid as a result of the action of the nitric acid formed by the nitrifying ferment upon the magnesium carbonate in suspension.

Recently (1891) Winogradsky has succeeded in cultivating this nitrifying ferment in a solid medium containing soluble silicates, which form a gelatinous mass (see page 47). The bacillo-cocci of G. and P. F. Frankland appear to be identical with the nitromonas of Winogradsky. The authors named, finding that the nitrifying ferment did not grow in nutrient gelatin, succeeded in isolating it in liquid cultures by the method of dilution. They describe the nitrifying organism obtained by this method as a bacillus which is but little longer than it is broad, and which they designate a "bacillo-coccus." In this country E. O. Jordan and Ellen H. Richards have also

succeeded in isolating a nitrifying ferment by the method of dilution and by cultivation in a liquid containing certain salts dissolved in distilled water. Their culture medium was constituted as follows:

Ammonium chloride (resublimed),	1.9070 grammes.
Sodium carbonate,	3.7842 "
Sodium phosphate,	0.2000 "
Potassium sulphate,	0.2000 "

"These salts were dissolved in such a quantity of redistilled water that the solution contained one hundred parts of nitrogen per one hundred thousand, and two equivalents of alkali. Ten cubic centimetres of this solution were mixed with one litre of redistilled water and then inoculated as desired."

The nitrifying ferment obtained by Jordan and Richards is described as a short bacillus of a slightly oval shape, about 8 to 9 μ broad and from 1.1 to 1.7 μ long. These bacilli are associated in irregular zoöglæa by a jelly-like material. The masses are found chiefly at the bottom of the flasks, as was the case with the nitrifying ferment isolated by Winogradsky. No independent movements were observed by Jordan and Richards, who state that they have not been able to determine in a definite manner whether the nitrifying organism isolated by them is identical with that of Winogradsky and of the Franklands. They remark that their bacillus stains with some difficulty with the usual aniline dyes.

441. NITRIFYING BACILLUS OF WINOGRADSKY.

Obtained by Winogradsky (1891) from the soil by cultivation in a gelatinous medium containing soluble silicates.

Morphology.—Small bacilli of somewhat irregular form, often pyriform; the average length does not usually exceed 0.5 μ , and the breadth is quite variable, even in the same cell. The bacilli are grouped in irregular masses and united into a membranous pellicle by a gelatinous material. This pellicle in old cultures is seen attached to the bottom of the tube as a transparent membrane, which may be detached by agitation; in more recent cultures it is firmly adherent and so transparent as to be seen with difficulty; it gives to the glass a feeble bluish-gray tint, and is not detached by repeatedly rinsing the tube with distilled water. The bacilli are somewhat difficult to recognize in stained preparations, owing to the staining of the gelatinous intercellular substance. Winogradsky recommends washing the stained preparation with water at 50° to 60° C. to remove the color from the gelatinous material.

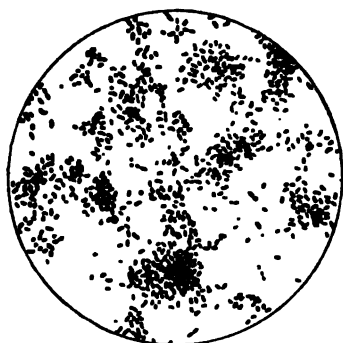


FIG. 258.—Nitrifying bacillus of Winogradsky. $\times 1,000$. From a photomicrograph. (Winogradsky.)

The *Biological Characters* of this bacillus have not been fully determined. In a gelatinous medium made by the addition of silicic acid to a culture liquid containing certain salts, yellowish-gray, lenticular colonies were developed, from which liquid cultures were subsequently made by Winogradsky. In these the liquid remained transparent, and no pellicle formed upon the surface, but a very transparent, gelatinous film was found attached to the bottom of the flask; the bacillus above described was found to be present in this and to be the cause of the active oxidation of nitrites present in the culture medium. With reference to its ferment action Winogradsky says: "The oxidizing power of this imponderable quantity of a living ferment is astonishing." When this bacillus is associated with the microorganism previously

described (No. 440) in a solution containing salts of ammonia, the "nitromonas" takes the precedence; and, according to Winogradsky, when the oxidation of the ammonia is completed this ferment ceases to absorb oxygen and enters into a state of repose, while the nitrifying bacillus commences to multiply and to exercise its special ferment action.

442. STREPTOCOCCUS CONGLOMERATUS (Kurth).

Obtained by Kurth (1890) from cases of scarlet fever.

Morphology.—As obtained from bouillon cultures it consists of masses made up of chains of cocci; free chains are only occasionally seen.

Biological Characters.—This streptococcus is said to differ from *Streptococcus pyogenes* and various other previously described streptococci by the fact that in *bouillon cultures*, at a temperature of 37° C., it forms at the bottom of the tube smooth, round, and very firm white scales, or a single flat layer which is not disintegrated when the tube is slightly agitated; other streptococci are said to form a loose deposit which is either entirely broken up or forms viscid threads when the tube is gently rotated.

Pathogenesis.—Very pathogenic for mice.

443. BACILLUS THALASSOPHILUS (Russell).

Obtained by Russell (1891) from mud at the bottom of the Gulf of Naples.

Morphology.—A slender bacillus, varying greatly in length, which grows out into filaments which are not visibly segmented.

Stains with Ziehl's solution when obtained from a recent culture, but not by Löffler's solution or fuchsin.

Biological Characters.—An *anaërobic*, *liquefying*, *motile* bacillus. Forms spores, which are very small and are located at the middle or the ends of the rods. Grows at the room temperature in the absence of oxygen. In *nutrient gelatin*, prepared with sea water, at the end of two or three days colonies appear at the bottom of the line of puncture in the form of small, clouded bubbles; later other colonies develop above these, forming finally a long, irregular, grayish, semi-transparent, liquid, sac-like mass. At the upper portion of this sac gas accumulates. The gelatin is now rapidly liquefied, except above, the liquefaction extending to the walls of the tube. Finally the entire amount of gelatin is liquefied, and becomes clear above from the deposition of the bacilli at the bottom of the tube, where they form a thick, sticky mass. The cultures give off a penetrating, disagreeable odor. In gelatin between double plates colonies first appear at the end of two or three days; under a low power these show a very thin network of filaments, which penetrate the gelatin in all directions. Much gas is developed in the colonies, and when the upper plate is lifted an odor of skatol is given off. The colonies soon become confluent and the gelatin is entirely liquefied. In *agar* a scanty development occurs along the line of puncture to within two centimetres of the surface.

444. BACILLUS GRANULOSUS (Russell).

Obtained by Russell (1891) from the mud at the bottom of the Gulf of Naples; common, even at a depth of 1,100 metres.

Morphology.—In hanging-drop cultures this bacillus grows out into long, slender filaments, made up of tolerably large bacilli having a thick cell wall and finely granular protoplasmic contents. In older cultures the long filaments are broken up into shorter, irregular fragments, and the separate segments are seen as short, thick rods with coarsely granular contents. Upon *agar* and potato the development of filaments is irregular and they break up into a grape-like mass. These masses, which might be taken for involution

forms or degenerative products, are capable of forming spores. In potato cultures these masses are seen to consist of short, plump cells, each one of which contains a highly refractive spore. The segments of a filament divide as in *Bacillus megatherium*, without any external appearance of division, the new cells growing in breadth rather than in length; as a result of unequal growth irregular masses are formed, in which later spores are developed. In anaërobic cultures the cells are usually solitary, or, at most, united in pairs. The protoplasm of young cells is fine and granular; in older cells, and especially in unfavorable media, the contents consist of large, shining granules.

Stains with the usual aniline colors and by Gram's method; the granules are deeply stained by Kühne's carbol-methyl-blue solution.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, usually *non-motile* bacillus; in bouillon cultures kept at 37° C. a slight to-and-fro movement may be observed. Forms spores. Grows in the

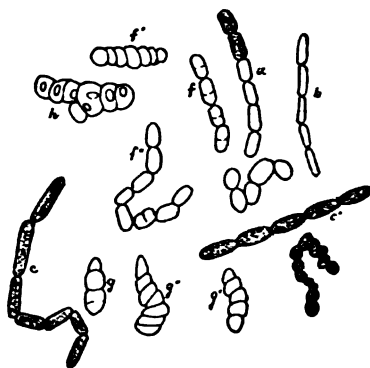
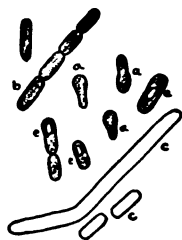


FIG. 230.

FIG. 230.

FIG. 230.—*Bacillus thalassophilus*; culture in nutrient gelatin at end of fourth day; a, drum-stick forms from gelatin culture; c, bacilli containing spores. (Russell.)

FIG. 230.—*Bacillus granulosus*; young surface colony upon gelatin plate; a, b, c, normal forms from recent gelatin culture; f, f', from an old gelatin culture; g-h, abnormal forms from potato culture—at h, spores. (Russell.)

usual culture media at the room temperature. Upon *gelatin plates* the colonies differ considerably in appearance, especially those upon the surface. At first they are usually thin, almost transparent, and leaf-like; under a low power these colonies are seen to be covered with irregular, concentric lines, which are formed by the parallel arrangement of the filaments, and under the microscope resemble the anastomosing "nerves" upon a leaf. Liquefaction of the gelatin quickly occurs. The deep colonies are not characteristic, and remain as small, round, shining, opaque masses. The colonies are viscid, drawing out into threads, or, when touched with the platinum needle, the entire colony may be picked up. In *gelatin stick cultures* liquefaction occurs near the surface, forming a shallow cavity, at the bottom of which

the bacilli accumulate in zoöglæa masses; later the liquefaction extends to the deeper layers of the gelatin medium. Upon *agar*, in the course of two or three days at the room temperature, or in twenty-four hours in the incubating oven, rather scanty and thin, whitish or yellowish colonies are developed, which remain separated where the culture medium is scanty, as at the upper part of oblique cultures, while below, where the culture medium is moist, development is more abundant. In *bouillon* an abundant development occurs, which causes a diffused cloudiness of the liquid and a considerable deposit at the bottom of the tube. The growth upon *potato* is quite characteristic. At the end of twenty-four hours at the room temperature a moist, white, shining patch is seen; instead of extending over the surface, this increases in thickness, forming a thick, viscid, shining-white mass; later this loses its lustre, becomes dull and waxy in appearance, and acquires a brownish color.

445. *BACILLUS LIMOSUS* (Russell).

Obtained by Russell (1891) from mud from the bottom of the Gulf of Naples; very abundant in all of the specimens examined.



FIG. 261.

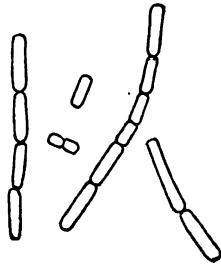


FIG. 262.

FIG. 261.—*Bacillus limosus*; culture in nutrient gelatin made with sea water, at end of two days, and bacilli from a gelatin culture. (Russell.)

FIG. 262.—*Spirillum marinum*; culture in nutrient gelatin made with sea water, at end of two days, and spirilla. (Russell.)

Morphology.—Long and slender bacilli, $1.25\ \mu$ broad and 3 to $4\ \mu$ long; usually united in pairs, or in chains containing several elements. In hanging-drop preparations from potato cultures the cells are shorter and thicker than in gelatin cultures; the ends are rounded, and the cell contents often appear finely granular.

Biological Characters.—An *aërobic, liquefying, motile* bacillus; exhibits slow to-and-fro movements. Forms spores, which are located at one extremity of the rods. Grows in the usual media at the room temperature—

more luxuriantly in nutrient gelatin made with sea water. Upon *gelatin plates* colonies are developed in from twenty-four to thirty hours, which at first are almost transparent; under a low power these are seen to be surrounded by long, slender filaments which extend far out into the gelatin; in its further development the central portion of the colony extends and includes the slender filaments, forming a round mass surrounded by little, thorn-like projections; liquefaction commences at the centre, causing a depression of the colony; in old colonies the liquefied gelatin has a grayish color and a thin pellicle forms upon the surface; below this swim flocculent masses, and around the margins the thorny appearance is preserved. In *gelatin stick cultures* containing sea water development is very rapid, and at the end of twenty-four hours a funnel-shaped cavity, containing liquefied gelatin which is clouded throughout, has been formed; in the course of seventy hours the gelatin is entirely liquefied and light, flocculent masses collect in the lower portion of the tube, while upon the surface a thin pellicle is formed which is easily broken up. Upon *agar* the development is abundant, forming a moist, shining-white layer. In *bouillon* a dense cloudiness is developed, and a thick and very resistant layer is formed upon the surface, while a considerable deposit accumulates at the bottom of the tube. Upon *potato* a thin, grayish-white layer is formed, which covers the greater part of the surface.

446. SPIRILLUM MARINUM (Russell).

Obtained by Russell from mud at the bottom of the Gulf of Naples, and from water from the same source; not very abundant.

Morphology.—Rods which are usually more or less curved, like the cholera spirillum; usually in pairs, but several elements are sometimes united to form a spiral filament; the rods are sometimes straight.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, liquefying, actively motile* spirillum. Spore formation not observed. The movements are sometimes rotary, and sometimes progressive without rotation. Grows in the usual culture media at the room temperature, but not in the incubating oven; grows better in media made without the addition of sea water. Upon *gelatin plates* the colonies are first seen as round, granular masses, often presenting radial striations. When liquefaction of the gelatin commences the colonies have a rougher appearance, and flocculent masses float in the shallow cavities containing liquefied gelatin. In *gelatin stick cultures* development is very rapid at first; the gelatin is liquefied and clouded, and a thin, semi-transparent membrane forms upon the surface; later a stratum of liquefied gelatin is seen above, while below the culture medium remains unaltered. Upon *agar* the growth is luxuriant and forms a moist, white, pus-like layer. Upon *potato* the growth is very characteristic; at the end of twenty-four hours a reddish-brown, sharply defined layer is developed, and the potato changes its color in the vicinity of the line of inoculation; the growth increases in dimensions and forms a thick, wax-like mass, which after a time covers the greater portion of the surface; it remains soft and does not penetrate the potato, which gradually acquires a dark greenish-gray color. In *bouillon* made with sea water development is abundant and causes the culture liquid to be densely clouded; a smooth, white layer forms upon the surface.

447. BACILLUS LITORALIS (Russell).

Obtained by Russell (1891) from mud at the bottom of the Gulf of Naples, near the shore.

Morphology.—Bacilli two to four times as long as broad.

Stains with Löffler's solution, but not by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation not observed. Movements very

irregular. Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* deep colonies are seen at the end of three days as well-defined, small, brown points. The superficial colonies are at first shining and opalescent; under the microscope they are seen to be finely granular and have smooth margins; at the end of five to eight days the colonies in contact with the air commence to cause liquefaction of the gelatin; liquefaction progresses so slowly that evaporation keeps pace with it, and the colonies slowly sink into the gelatin; when the layer of gelatin is quite thin a ring of liquefied gelatin may surround the colony. In *gelatin stick cultures* a scanty growth is seen along the line of puncture at the end of twenty-four hours. Upon the surface a thin, irregular, whitish layer commences to form around the point of inoculation at the end of three or four days; the gelatin is slowly liquefied below this, and as a result of evaporation a small cavity is formed which is lined with a thin layer of bacilli. But little de-

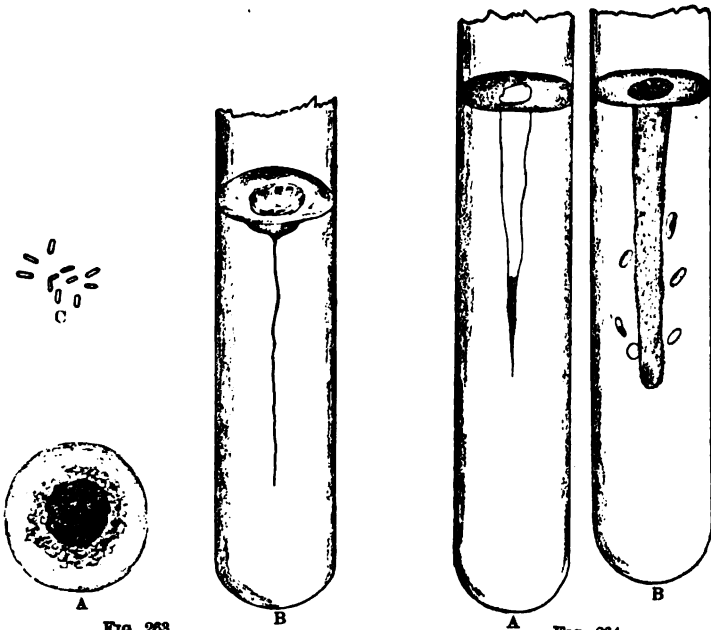


FIG. 263.

FIG. 264.

FIG. 263.—*Bacillus litoralis*; A, colony upon gelatin plate, ten days old; B, gelatin stick culture ten days old; C, bacilli from hanging-drop culture. (Russell.)

FIG. 264.—*Bacillus halophilus*; A, culture in nutrient gelatin at end of twenty-four hours; B, culture in sea-water gelatin at end of twenty-four hours. (Russell.)

velopment occurs along the line of inoculation below, but the growth often acquires a reddish-brown color, and the gelatin around it is stained brown—this color is only developed in the absence of oxygen. In *streak cultures* white, semi-transparent colonies are formed along the line of inoculation; these become visible after the second day, extend slowly, and finally coalesce; in the course of five to seven days the thicker white colonies commence to sink below the surface as a result of liquefaction and evaporation; liquefaction now progresses more rapidly, and after a time the deposit at the bottom of the liquefied gelatin acquires a reddish-brown tint. *No growth occurs upon potato*. Upon *agar* a scanty, thin, moist-looking, grayish-white layer is formed. In *bouillon* a uniform cloudiness is developed and no pellicle forms upon the surface.

448. *BACILLUS HALOPHILUS* (Russell).

Obtained by Russell (1891) from water of the Gulf of Naples and from mud from the bottom.

Morphology.—Recent cultures in nutrient gelatin made with sea water contain bacilli of from 1.5 to 3.5 μ in length and 0.7 μ broad; these are often united in pairs. In older cultures the form quickly changes, and at the end of two days cells resembling those of a torula—"yeast-like"—are seen: these abnormal forms increase in number and variety as the culture becomes older; some contain a granular protoplasm, and some are so transparent as to be easily overlooked. These cells, however, are motile and resemble the so-called monads. The variability of form is still greater in ordinary nutrient gelatin, which is a less favorable medium than gelatin made with sea water.

Stains with difficulty, and not at all with Löffler's solution; does not stain by Gram's method; the protoplasm is irregularly stained by Ziehl's solution, while fuchsin solution stains it feebly but homogeneously.

Biological Characters.—An *aërobic, liquefying, actively motile* bacillus. Spore formation not observed. Grows slowly in the usual culture media at the room temperature. In stick cultures in *nutrient gelatin* made with sea water, at the end of twenty-four to thirty-six hours, punctiform colonies are developed, which quickly coalesce; liquefaction occurs along the line of growth and gas is developed; sometimes this is so abundant that the liquefied gelatin is forced up over the surface of the culture as a foamy mass; later the liquefied gelatin becomes transparent, and a fine deposit is seen at the bottom of the tube. In nutrient gelatin not made with sea water the growth is considerably slower; at first a white line is seen, extending only along a portion of the puncture; in the course of seventy hours a slender cavity is formed as a result of slow liquefaction and evaporation; this gradually increases in length and gives to the cultures a characteristic appearance. In *plates* made with sea-water gelatin, spherical, grayish-white, semi-transparent colonies are developed; these cause liquefaction, and by evaporation sharply defined, deep funnels are formed in the gelatin. The cultures have a strongly alkaline reaction.

449. *BACILLUS CAPSULATUS MUCOSUS* (Fasching).

Obtained from the nasal secretion in two cases of influenza.

Morphology.—Bacilli from 3 to 4 μ long and 0.75 to 1 μ thick, enveloped in a capsule containing one to four individuals.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-motile, non-liquefying* bacillus. Does not form spores. Grows in the usual culture media at 18° to 35° C. Upon *gelatin plates* circular, milk-white colonies are developed; these have a faint aromatic odor and are cupped upon the upper surface; they resemble drops of mucus about the size of a pin's head. In stick cultures in gelatin a nail-like growth, like that of Friedländer's bacillus, is seen, and there is a formation of gas.

Stains with the usual aniline colors, but not by Gram's method.

Pathogenesis.—White mice and field mice die from general infection in from thirty-six to forty-eight hours after inoculation; they also suffer from conjunctivitis. Not pathogenic for rabbits or for pigeons.

450. *BACILLUS* OF POTATO ROT (Kramer).

Obtained by Kramer (1891) from potatoes affected with wet rot—"Nassfäule."

Morphology.—Bacilli with round ends, from 2.5 to 4 μ long and 0.7 to

0.8 μ broad; often united in chains; grow out into filaments; in old cultures thicker rods of an ellipsoidal form are seen, and spores are formed which entirely fill the cell containing them.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms large oval spores. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* liquefaction occurs rapidly, forming a funnel, at the bottom of which the bacilli accumulate. In streak cultures upon gelatin, at the end of twelve hours, an elevated, dirty-white line of growth is seen along the impfstrich; this extends laterally, and the margins are scalloped so that the growth resembles a leaf. Upon *nutrient agar* small, dirty-white, slimy drops, with sharply defined margins, are developed. Gelatin cultures containing litmus or carmine are decolorized by this bacillus. In solutions containing dextrose it multiplies abundantly, producing carbon dioxide and butyric acid. In starch paste containing ammonium tartrate the starch is only dissolved to a slight extent, and no butyric acid is developed. It has but slight action upon cellulose. Potatoes sterilized by steam and inoculated with a pure culture of the bacillus undergo changes corresponding with "Nassfäule" in these tubers. These consist of a decomposition of soluble carbohydrates (sugar), with a formation of carbon dioxide and butyric acid; then of the intercellular substance and the cell membranes, producing an acid reaction in the contents of the tuber. The starch is not changed. The albuminous substances undergo putrefactive decomposition, with production of ammonia, methylamin, and trimethylamin. These bases first neutralize the butyric acid, and later cause an alkaline reaction of the affected portions of the potato. In *milk* coagulation of the casein occurs, but no putrefactive change is induced, even at the end of several weeks, as is the case with *Bacillus butyricus* (Hueppe).

451. BACILLUS VACUOLOSIS (Sternberg).

Obtained by Sternberg (1888) in cultures from the intestine and stomach of yellow-fever cadavers.

Morphology.—Bacilli with round ends, which vary considerably in the r dimensions and are sometimes slightly curved; length from 1.5 to 5 μ , breadth about 1 μ . In stained preparations unstained places—vacuoles?—are seen in the rods. In surface cultures upon agar various involution forms are seen, which also present vacuoles and which are usually considerably larger than the normal bacilli. The bacilli sometimes grow out into long, jointed filaments.

Stains with the usual aniline colors.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms large oval spores. Grows in the usual culture media at the room temperature. In *gelatin stick cultures* liquefaction occurs slowly, near the surface, forming a cup-shaped cavity. The liquefied gelatin is quite viscid, and a cream-white layer of bacilli forms upon the surface of the liquefied medium. In *nutrient agar* the development along the line of puncture is scanty; on the surface a cream-white layer is formed and the bacilli are united in long, jointed filaments. It is not always seen to be motile, but under certain circumstances exhibits slowly progressive, to-and-fro movements, as if propelled by a flagellum. Does not grow in an acid medium. On *potato* a thin, cream-white layer is formed.

Pathogenesis.—Not pathogenic for rabbits. Not tested upon other animals.

452. BACILLUS OF DANTEC.

Synonym.—Bacille du rouge de morue.

Obtained by Dantec (1891), in association with other microorganisms, from salted codfish, to which it imparts a red color.

Morphology.—Bacilli with round ends, from 4 to 12 μ long, and usually containing a spore at one extremity. Resembles the bacillus of tetanus, but is considerably thicker.

Biological Characters.—An *aërobie*, *liquefying*, *motile*, *chromogenic* bacillus. Forms spores. Grows in the usual culture media at the room temperature. Produces a red pigment. In *gelatin stick cultures* liquefaction in funnel form occurs along the line of puncture. In old gelatin liquefaction may not occur. Upon the surface of nutrient gelatin a red streak is developed along the line of inoculation, and the gelatin below this is very slowly liquefied. The colonies in gelatin plates may attain a diameter of two millimetres: they have a pale-red centre with a deeper red periphery, and are disc-shaped. In *bouillon* development is abundant, but without the formation of pigment. Does not grow well upon *potato*. Pigment is formed more abundantly at 10° to 15° C. than at a higher temperature. Upon dried codfish it grows readily, forming a red pigment, especially upon the side which has been exposed to the salt.

Not pathogenic.



FIG. 235.—Bacillus of Dantec.

453. BACILLUS (MICROCOCOCCUS ?) HAVANIENSIS (Sternberg).

Morphology.—Short-oval bacilli, usually in pairs, about 0.4 to 0.5 μ in diameter. The cells are so nearly spherical that the writer has been in doubt whether to describe this microorganism as a bacillus or as a micrococcus.

Biological Characters.—An *aërobie*, *non-liquefying*, *chromogenic* bacillus(?). Grows slowly in the usual culture media at the room temperature. Upon *gelatin plates* forms small, spherical, translucent colonies of a beautiful blood-red color. In *gelatin stick cultures* a thick, opaque, carmine layer develops about the point of inoculation and slowly increases in thickness and circumference: very scanty growth, without color, at the upper part of the line of puncture. Upon the *surface of agar* the growth is slow but continuous, and forms, at the room temperature, a thick, carmine layer along the line of inoculation: this has wavy outlines and a glistening, varnished-like surface; it gradually extends in thickness and breadth, which at the end of a month may be five to six millimetres. Frequently this micrococcus fails to grow on *potato*, perhaps because of an acid reaction. But upon old and rather dry potato it sometimes develops, as it does on nutrient agar, forming a thick, irregular mass of a carmine color. The pigment is only formed in the free presence of oxygen.



FIG. 236.—Bacillus Havanien-sis. $\times 1,000$. (Sternberg.)

454. BACILLUS AMYLOZYMA (Perdrix).

Obtained by Perdrix (1891) from the hydrant water of Paris—best from the deposit left in a Chamberlain filter through which this water has been passed.

Morphology.—Bacilli with round ends, from 2 to 3 μ long and 0.5 μ broad; usually in pairs, or in chains of several elements.

Stains with the usual aniline colors.

Biological Characters.—A strictly *anaërobic, non-liquefying, motile* bacillus. Forms spores. Grows in the usual culture media in an atmosphere of hydrogen at the room temperature. In *nutrient gelatin* colonies are developed at the end of five or six days; these are small, white plaques surrounded by gas bubbles. Upon *potato*, in an atmosphere of hydrogen, whitish colonies are developed, around which the potato appears excavated; at the same time it is partly liquefied by the ferment action of the bacillus, and the liquid collects at the bottom of the tube. When these potato tubes are opened there is a little explosion, due to escaping gas. The development of the bacillus is most rapid and its ferment action most energetic at a temperature of about 35° C. It causes fermentation of sugar and of starch, but not of cellulose. It ceases to grow in cultures containing 0.10 to 0.12 per cent of acid (estimated in sulphuric acid), and, as it produces an acid reaction of the culture medium, it is necessary to add carbonate of lime to this to insure its continuous development. The acids formed from the fermentation of sugar are acetic (at the outset) and butyric. In culture media containing starch, ethylic and amylic alcohol are produced, as well as butyric acid; a considerable portion of the starch is converted into sugar, and hydrogen and carbon dioxide are given off freely as a result of the fermentation.

455. BACILLUS RUBELLUS (Okada).

Obtained by Okada (1892) from dust by inoculations in guinea-pigs. Not pathogenic, but found in association with pathogenic bacteria in the bloody serum effused about the point of inoculation — with dust from the streets.

Morphology.—Resembles *Bacillus œdematis maligni* in its form and dimensions. Bacilli with slightly rounded ends, usually in pairs or chains of three; in old bouillon cultures grows out into filaments 10 to 15 μ long; these are often surrounded by a capsule-like envelope. The rods show a slight swelling when spore formation occurs, becoming spindle-shaped or having an expanded extremity, according as the spore is formed in the middle or at one extremity. Flagella may be demonstrated by Löffler's method of staining; these are seen at one or at both extremities.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *anaërobic, chromogenic, liquefying, motile* bacillus. Forms large oval spores, which are located at one extremity or in the centre of the rods. In *gelatin plates* kept in an atmosphere of hydrogen, dull-white, punctiform colonies are developed at the end of ten days at 15° to 18° C; under a low power these are seen to be long-oval in form and surrounded by fine offshoots; after a time the gelatin is liquefied and acquires a reddish color. In deep *gelatin stick cultures* development commences at the bottom of the line of puncture, where, at the end of ten days, small, dull-white, spherical or oval colonies are developed; these are seen to be surrounded by radiating offshoots; the colonies increase in size, and the gelatin is gradually liquefied in the lower two-thirds of the tube, while above it remains solid; a flocculent deposit is now seen at the bottom of the tube, which has a reddish color; finally the upper portion of the gelatin is also liquefied and the whole of the fluid has a red color. Upon *agar plates*, at 37° C., development is more abundant and rapid; colonies are developed in twenty-four hours, which subsequently increase in size and acquire a red color. In deep stick cultures in agar, in the incubating oven, development occurs below at the end of twenty-four hours and gradually extends upward to near the surface. Gradually the upper portion of the growth acquires a red color, which increases in intensity, and the upper portion of the culture medium is after a time diffusely colored. In *bouillon* in an atmosphere of hydrogen, at 37° C., development is abundant and rapid, and the medium acquires a red color.

Not pathogenic.

456. *BACTERIUM UREÆ* (Jaksch).

Found in ammoniacal urine.

Morphology.—Bacilli with round ends, about $2\ \mu$ long and $1\ \mu$ thick.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying* bacillus. Spore formation not observed. Grows very slowly at the room temperature. Changes urea into carbonate of ammonia. Upon *gelatin plates* forms small, almost transparent colonies, which at the end of ten days may have the diameter of a "pfennig." In *gelatin stick cultures* a thin, gray, branching growth is seen along the line of puncture. Old cultures have the odor of herring-brine. Imperfectly described.

457. *SARCINA MOBILIS* (Maurea).

Obtained by Maurea (1892), from ascitic fluid which had been preserved a long time in a test tube.

Morphology.—Micrococci having a diameter of $1.5\ \mu$ and associated in pairs or in tetrads.

Stains with the usual aniline colors and also by Gram's method.

Biological Characters.—An *aërobic*, *liquefying*, *motile*, *chromogenic* sarcina. Does not grow in the incubating oven at 37°C . Motions progressive, serpentine, or rotatory (?). Upon *gelatin plates*, at 15° to 20°C ., punctiform, white colonies are seen on the third day. By the seventh day liquefaction of the gelatin commences around the colonies and a brick-red pigment is formed. In *gelatin stick cultures*, at the end of five days, a scanty development is seen along the line of puncture and a more abundant growth upon the surface; later the surface growth acquires a brick-red color; in from fifteen to twenty days liquefaction has occurred in the form of a small funnel, and by the end of thirty days one-half of the contents of the tube is liquefied. In *bouillon* the fluid becomes clouded in two or three days, and later a yellowish-red deposit is seen at the bottom of the tube. In *agar stick cultures* a whitish layer is developed on the surface, which later acquires a brick-red color. In *milk* growth occurs without producing coagulation. No growth upon *potato*. In hay infusion sarcina-like packets are developed in abundance, as well as tetrads and diplococci.

458. *BACILLUS STOLONIFERUS* (Pohl).

Obtained from swamp water (1892).

Morphology.—Bacilli $1.2\ \mu$ long and $0.8\ \mu$ broad.

Biological Characters.—An *aërobic*, *liquefying*, *motile* bacillus. Spore formation not observed. In *gelatin stick cultures* liquefaction, in funnel form, commences at the end of twenty-four hours and progresses rapidly. Upon the *surface of agar* a thick, white mass develops along the track of the inoculating needle. Upon *potato* small colonies the size of a pin's head are developed along the line of inoculation and extend over the entire surface. In *milk* a scanty development occurs; the milk is not coagulated and no acid is formed.

459. *BACILLUS INCANTUS* (Pohl).

Obtained from swamp water.

Morphology.—Bacilli $1.7\ \mu$ long and $0.4\ \mu$ broad.

Biological Characters.—An *aërobic*, *liquefying*, *slightly motile* bacillus. In *gelatin stick cultures* growth occurs along the line of puncture and upon the surface as a grayish-white, elevated mass. At the end of forty-eight hours slight liquefaction is observed; this progresses very slowly. Upon the *surface of agar* a grayish-white, granular mass is developed along the track of the inoculating needle. Upon *potato* a gray, viscid layer is formed, which

quickly extends over the entire surface. Does not produce an acid reaction in milk.

460. BACILLUS INUNCTUS (Pohl).

Obtained from swamp water.

Morphology.—Bacilli $3.5\ \mu$ long and 0.8 to $0.9\ \mu$ broad.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation not observed. Upon *gelatin plates* forms oval or round colonies with a smooth margin and oily, shining appearance. In *gelatin stick cultures* grows along the line of puncture, and below the growth has a radiating appearance; upon the surface a thick, white, shining layer is quickly formed; later liquefaction commences and progresses slowly. Upon the *surface of agar* white, cloud-like masses are formed along the streak. Upon *potato* a white, slimy mass is formed which soon covers the entire surface. In *milk* an acid reaction is produced by the growth of this bacillus, but no coagulation occurs. In Pasteur's solution containing cane sugar or starch an abundant development of alcohol occurs.

461. BACILLUS FLAVESCENS (Pohl).

Obtained from swamp water (1892).

Morphology.—Bacilli 2.1 to $2.2\ \mu$ long and $0.8\ \mu$ broad.

Biological Characters.—An *aërobic, non-liquefying, chromogenic, slightly motile* bacillus. Spore formation not determined. Forms a yellow pigment. Upon *gelatin plates*, at the end of four days, colonies are visible; these are yellow, granular, and attain the size of a pin's head. In *gelatin stick cultures* it grows along the line of puncture, and slowly extends over the entire surface. Upon the *surface of agar* small, solitary, yellow colonies are developed along the track of the needle. Upon *potato* development is somewhat more rapid, and at the end of four days the entire surface is covered with a slimy, yellow layer. Gelatin colored blue with litmus is decolorized without the previous change of color to red. In *milk* an acid reaction is produced without coagulation.

462. BACILLUS BUTYRICUS OF BOTKIN.

Obtained by Botkin in anaërobic cultures from milk, from hydrant water, well water, garden earth, and dust.

Morphology.—In agar and gelatin cultures containing one and one-half per cent of grape sugar the bacilli are $0.5\ \mu$ long and 1 to $3\ \mu$ in diameter, with round ends; they are often united in pairs or in chains of three; in liquid media they are not so thick, and often attain a length of $10\ \mu$ or more, without segmentation; long chains may also be observed. The spores are located in the centre of the rods, or occasionally at one extremity; they vary greatly in their dimensions; the diameter is usually about $1\ \mu$ and the length 2 to $3\ \mu$. In old cultures various involution forms are seen.

Stains with the usual aniline colors.

Biological Characters.—An *anaërobic, liquefying, slightly motile* bacillus. Forms large oval spores, which have a high resisting power for heat. To obtain pure cultures Botkin subjects milk containing this bacillus to the boiling temperature—in the steam sterilizer—for half an hour. The spores resist this temperature, and the flask containing the milk is hermetically sealed and placed in an incubating oven at 37°C . At the end of twelve hours, as a rule, fermentation commences; the casein is coagulated and collects, together with the fat, at the upper part of the flask, while a clear, yellowish serum is seen below; there is an abundant development of gas. The most favorable temperature for growth is 37° to 38°C ., but development may occur at temperatures ranging from 18° to 42°C .; at 18° development is very slow and usually no gas is formed. In culture media containing

starch many of the bacilli, at the end of two or three days, contain granules which acquire a deep-blue color when treated with iodine solution. Upon *agar plates* (with 1.5 per cent of grape sugar), in an atmosphere of hydrogen, colonies are developed in fifteen to eighteen hours, which under the microscope are seen to be round or elliptical in form and to consist of closely interwoven filaments; as a rule, the margins are not well defined, and radiating filaments are given off from the felt-like colony. In deep stick cultures in agar, growth commences at a depth of one and one-half centimetres below the surface; later, when the oxygen is displaced by the gases produced, it approaches the surface. The growth along the line of puncture has an irregular outline with wave-like projections. In *gelatin* (with 1.5 per cent of grape sugar) growth begins, in deep stick cultures, about two centimetres below the surface; much gas is developed and the gelatin is quickly liquefied. In *bouillon* (with 1.5 per cent of grape sugar) an abundant development occurs within twenty-four hours in an atmosphere of hydrogen. The fluid becomes clouded and much gas is developed; at the end of three days the fermentation ceases, the bouillon becomes clear, and a thick, white deposit is seen at the bottom of the tube. In *milk*, contained in flasks completely full, and from which the oxygen has been expelled by boiling, growth commences near the bottom, where at the end of fifteen hours a transparent layer of serum is seen, from which numerous gas bubbles are given off; the fermentation progresses rapidly, and at the end of eighteen hours the coagulated casein and particles of fat have accumulated at the upper part of the flask; the pressure of gas is so great that many of the flasks are blown into fragments; at the end of a week development has ceased and the casein is almost entirely dissolved, the contents of the flask consisting of a transparent, yellowish fluid, with spongy masses of fatty material upon the surface and a flocculent, white deposit at the bottom. Upon *potato*, in an atmosphere of hydrogen, development occurs in the interior of the potato, but not upon the surface.

Not pathogenic.

Note.—Botkin is not able to identify the bacillus described by him with butyric-acid bacilli described by Pasteur, Cohn, Hueppe, and other authors, which for the most part have been cultivated only in liquid media. He remarks that it is differentiated from *Clostridium butyricum* of Prazmowski (*Vibrio butyrique* of Pasteur) by the fact that it does not decompose cellulose or salts of lactic acid. It closely resembles a butyric-acid ferment described by Perdriz, but is differentiated from it by the fact that the bacillus of Perdriz does not liquefy gelatin.

463. *UROBACILLUS PASTEURI* (Miquel).

Obtained by Miquel from decomposed urine.

Morphology.—Bacilli differing greatly in dimensions in different media—the diameter may be as much as $1.2\ \mu$. In urine to which two per cent of urea has been added it is usually seen in the form of short rods united in chains of two to six elements. In gelatin cultures containing urea the bacilli are from 4 to $6\ \mu$ long and are usually in pairs.

Biological Characters.—An *aerobic, liquefying, motile bacillus*. Forms spherical spores, usually solitary, and situated at one extremity of the rod. Grows at the room temperature in the usual culture media when these are made alkaline by the addition of ammonia, or when urea is added. In *gelatin plates* containing urea minute colonies are developed within twenty-four hours and an ammoniacal odor is given off; under a low power the colonies are seen to be spherical or oval, yellowish, and surrounded by dumbbell-shaped crystals; at the end of eight days the gelatin commences to liquefy, and after a time has the consistence of castor oil. In ordinary flesh-peptone-gelatin no development occurs, unless the medium has a distinctly alkaline reaction. No growth occurs in the usual agar-agar medium, but when urea

is added an abundant development occurs, and numerous very minute crystals are scattered through the culture medium. In *urine* alkaline fermentation is quickly induced, and an abundant deposit of crystals of ammonio-magnesian phosphate and of alkaline urates, together with the bacilli, accumulates at the bottom of the test tube; this deposit acquires a blackish color. The most favorable temperature for the growth of this bacillus is 30° to 40° C.

464. UROBACILLUS DUCLAUXI (Miquel).

Obtained by Miquel (1879) in the water of sewers, and subsequently in river water—very common and widely distributed.

Morphology.—Slender filaments, from 0.6 to 0.8 in diameter and from 2 to 10 μ long. In alkaline bouillon it may attain a diameter of 1 μ ; the bacilli are frequently united in chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *motile*, *liquefying* bacillus. The movements are comparatively slow. Forms spores located at the centre of the rods, which then are spindle-shaped. Develops most rapidly at 40° C. No development occurs at 5° C., but at 8° to 10° the fermentation of a culture medium containing urea is accomplished in about a month, and at 20° in two or three days. Does not grow in ordinary bouillon or nutrient gelatin, but grows in the usual culture media when they are rendered alkaline by the addition of ammonia, or when urea is added. In *nutrient gelatin containing urea* numerous small, white colonies are developed along the track of the needle, and a quantity of minute crystals are scattered through the culture medium. At the end of three or four months the gelatin medium is transformed into a transparent, ammoniacal, syrupy liquid. In alkaline gelatin not containing urea liquefaction does not occur. In *bouillon* made alkaline by the addition of ammonia, growth occurs, causing the liquid to become clouded within twenty-four hours; later an abundant sediment accumulates at the bottom of the tube, the liquid becomes viscid and gives off a disagreeable odor.

465. UROBACILLUS FREUDENREICHI (Miquel).

Obtained by Miquel from the air, from dust, from sewer water, etc.

Morphology.—Closely resembles *Urobacillus Pasteuri*, but forms longer chains and has more active movements. The rods are from 1 to 1.3 μ thick and have rounded extremities; the length varies considerably, but in recent cultures is usually 5 to 6 μ . Under favorable conditions of temperature, 30° C., actively motile filaments, consisting of six to ten elements, are developed in alkaline bouillon; upon solid media long filaments, composed of comparatively short elements and quite motionless, are developed.

Biological Characters.—An *aërobic*, *liquefying*, *motile* bacillus. Forms spores. The most favorable temperature for the development of this bacillus is from 33° to 35° C. As a ferment of urea it is ten times less active than *Urobacillus Pasteuri*. In *nutrient gelatin*, at 20° C., development occurs upon the surface as a milk-white growth, which is visible at the end of two days, and later forms a layer with irregular outlines having a diameter of three to four millimetres; but little development occurs along the track of the inoculating needle; liquefaction commences at the surface at the end of eight to ten days and progresses slowly; at the end of thirty to forty days the gelatin is completely liquefied, it is of a pale-yellow color and quite viscid; an abundant white deposit accumulates at the bottom of the tube. Upon *gelatin plates* (containing urea) small, spherical, white colonies are developed in two or three days; these gradually increase in dimensions and are surrounded by an aureole of minute crystals; later the development ceases and the bacilli are killed by an excess of carbonate of ammonia resulting from the decomposition of the urea. In neutral *bouillon*, at the end of two or three days, a slight cloudiness is developed, and later a scanty white deposit is seen at the bottom of the tube.

466. *UROBACILLUS MADDOXI* (Miquel).

Obtained from sewer water and from river water—relatively rare.

Morphology.—Bacilli with round ends, $1\ \mu$ thick and 3 to $6\ \mu$ long; in old cultures the bacilli are often seen as oval or spherical cells having a diameter of 6 to $8\ \mu$; in solid media the length is usually from 2 to $3\ \mu$.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms spores. Grows best at 38°C ., but causes fermentation of urine at the end of several days at a temperature as low as 10°C . In *nutrient gelatin* it usually fails to grow, but occasionally a scanty development occurs along the track of the needle in the form of small, spherical colonies. In nutrient gelatin containing urea the line of inoculation is marked by numerous crystals, which are seen at the end of twenty hours, although the growth of the bacilli is scarcely apparent. In gelatin plates containing urea very small, spherical, opaque, whitish colonies, surrounded by crystals, may be seen under a low power. In nutrient agar containing urea, at 30° to 35°C ., a layer is formed upon the surface, which is at first white and later of a grayish-yellow color. In *bouillon* which is slightly alkaline it grows rapidly, and produces, at the end of two days, a dense turbidity of the culture liquid; later an abundant glairy sediment accumulates at the bottom of the tube. In one litre of bouillon an amount of soluble ferment is produced by this bacillus which is capable of decomposing sixty to eighty grammes of urea in the course of two or three hours.

467. *UROBACILLUS SCHÜTZENBERGI* (Miquel).

Obtained by Miquel from river water and from the water of sewers.

Morphology.—Small oval bacilli about $0.5\ \mu$ thick and $1\ \mu$ long; usually united in pairs.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Spore formation not observed. In *nutrient gelatin*, at 20°C ., liquefaction in cup shape occurs at the surface and progresses rapidly, being complete by the end of ten days. In gelatin containing two per cent of urea liquefaction commences at the surface and numerous crystals are formed in the solid portion of the culture medium; growth is soon arrested by the antiseptic action of the products developed. In *gelatin plates* small, translucent, spherical colonies are developed, which increase rapidly in size and acquire a milky appearance; when the colonies reach the surface liquefaction rapidly occurs; when the gelatin contains urea the colonies cease growing after attaining a diameter of one to two millimetres, and they are surrounded by a zone of crystals. Upon *nutrient agar*, at 28° to 30°C ., development occurs in the form of a whitish layer with a slightly greenish tint. In *bouillon*, at the end of twenty-four hours, the medium becomes clouded; later a thin pellicle forms upon the surface and extends upward for a short distance upon the walls of the test tube; the bouillon remains clouded for several weeks. When urea is added to peptonized bouillon development is still more abundant, but ceases at the end of four or five days and the liquid becomes entirely transparent.

468. *BACILLUS OF BOVET*.

Obtained by Bovet (1891) from the intestine of a woman who died of an acute enteritis with choleraic symptoms.

Morphology.—Bacilli from 1 to $1.5\ \mu$ thick and 2 to $4\ \mu$ long, isolated or in pairs.

Stains with carbol-fuchsin solution.

Biological Characters.—An *aërobic, non-liquefying, motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature—more rapidly at 37°C . In *nutrient gelatin*, at 15° to 18°C ., a grayish-white, somewhat translucent layer with irregular outlines is

developed in two or three days; along the line of the inoculating needle in stick cultures a granular growth is seen which has a brownish-gray color. Upon *agar* the surface growth is similar to that upon gelatin and quickly covers the entire surface. When the culture medium contains sugar or glycerin lenticular bubbles of gas are formed in it. Upon *potato* a thick layer is developed resembling purée of peas, but not so decidedly yellow in color; in old cultures the color is a dirty-gray. In anaërobic cultures a scanty development occurs.

Pathogenesis.—Subcutaneous injections in rabbits or guinea-pigs gave a negative result. Intraperitoneal injections in guinea-pigs produce peritonitis and death.

469. BACILLUS SCHAFFERI (Freudenreich).

Obtained by Freudenreich from cheese and from fermenting potato—fragments of raw potato in water.

Morphology.—Bacilli about 1 μ thick and from 2 to 3 μ long; in old cultures long filaments are common—20 to 25 μ .

Stains well with the usual aniline colors; in gelatin cultures the rods are frequently only partly stained; generally the central portion is stained while the poles remain unstained, but occasionally one-half is colored, or two stained portions may be separated by a clear space. Does not stain by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *motile* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates*, at the end of two or three days, punctiform, yellowish colonies are developed; under a low power these are seen to be granular, pale-yellow, and spherical or irregular in outline; upon the surface the colonies are elevated, circular in outline, and porcelain-white in color; when widely separated they may finally attain the size of a two-franc piece—with very irregular margins; under a low power these large colonies are seen to be granular, and to have a yellowish centre with pale margins. The colonies are not viscid and are easily detached from the surface with a platinum needle. In *stick cultures* in nutrient gelatin a layer is developed upon the surface which at first is nearly transparent; later this has a grayish color and extends over the entire surface; the growth along the line of puncture extends to the bottom, but is not characteristic. Upon *agar* a grayish layer is developed, which later sometimes acquires a brownish color, especially along the line of puncture. Upon *potato* a moist, yellowish layer is developed; this remains smooth and is without gas bubbles. In *bouillon*, at 37° C., development occurs within five or six hours, causing turbidity; when milk sugar is added to the peptonized bouillon there is a development of gas, and bubbles are given off in abundance when the culture is agitated. Culture media containing sugar acquire an acid reaction; at the end of several days a pellicle is formed upon the surface, which later falls to the bottom. In sterilized *milk* development is not abundant, but milk filtered through porcelain is a favorable culture medium; an acid reaction is produced, but coagulation, as a rule, does not occur—sometimes an imperfect coagulation occurs. This bacillus dies out in cultures in two or three weeks, and does not resist desiccation longer than forty-seven to fifty days. It is killed by a temperature of 70° C. maintained for fifteen minutes. According to Freudenreich, this bacillus closely resembles *Bacillus coli communis*, but is distinguished from it by the fact that it is actively motile, and by its ability to grow in solutions of milk sugar in the absence of oxygen, also by not being pathogenic for guinea-pigs when injected into the peritoneal cavity.

470. BACILLI OF GUILLEBEAU (*a*, *b*, and *c*).

Obtained by Guillebeau from the milk of cows suffering from mastitis,

and found by Freudenreich to produce an abnormal fermentation of cheese, characterized by the presence of large cavities ("boursoufflement") and by a very bad taste.

BACILLUS *a*.

Morphology.—Varies considerably in size, and may resemble a micrococcus in form; usually $1\ \mu$ broad and 1 to $2\ \mu$ long.

Stains with the usual aniline colors, but rather feebly; does not stain by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *slightly motile*, *non-liquefying* bacillus. Spore formation not observed. Grows in the usual culture media at the room temperature. Upon *gelatin plates* the deep colonies are spherical, granular, and yellowish in color; upon the surface they are round and granular at first, later they become opaque and resemble a drop of wax. In *gelatin stick cultures* development occurs all along the line of puncture, and upon the surface a whitish layer is formed. Upon *agar* a grayish-white layer is developed. Upon *potato* a thick, yellowish layer is formed; this is viscid and contains numerous gas bubbles. In *milk* coagulation is produced at the end of twenty-four hours, and an abundance of gas is given off. In *bouillon* containing milk sugar it multiplies abundantly and a large quantity of gas is liberated. Grows best at a temperature of 30° to 35° C. Thermal death-point 60° C.—fifteen minutes' exposure.

BACILLUS *b*.

Morphology.—Resembles bacillus *a*; bacilli from 1 to $2\ \mu$ long and about $1\ \mu$ thick.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying*, *motile* bacillus. Is differentiated from *a* by the fact that it causes liquefaction of nutrient gelatin after an interval of several weeks, and by the fact that the young colonies upon gelatin plates are quite viscid. Spore formation not observed. Thermal death-point 80° C.—five minutes' exposure. An abundance of gas is given off from cultures containing milk sugar.

BACILLUS *c*.

Morphology.—Short bacilli; often oval or even spherical in form; about $1\ \mu$ long.

Biological Characters.—An *aërobic*, *non-liquefying* bacillus. Spore formation not observed. Upon *gelatin plates* colonies are developed which resemble those of bacillus *a*, but are more coarsely granular; the colonies are very adherent and difficult to remove from the culture medium. Upon *agar* a viscous, white layer is developed. Upon *potato* the growth is of a yellowish-white color and similar to that of *a* and *b*, with gas bubbles; it is very adherent. In liquid media the growth of this bacillus causes the culture liquid to become extremely viscous and almost gelatinous in consistence. In *milk* coagulation occurs at the end of sixty hours at 37° C., and the milk then loses its viscosity.

471. MICROCOCCUS FREUDENREICHI (Guillebeau).

Obtained from milk which had undergone viscous fermentation.

Morphology.—Micrococci having a diameter of $2\ \mu$, solitary or in chains.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *liquefying* micrococcus. Grows best at a temperature of 20° C. Upon *gelatin plates* made with milk serum, punctiform, granular colonies are developed at the end of thirty-six hours; when touched with the platinum needle these may be drawn out into long threads; soon after the gelatin commences to liquefy and becomes very viscous. Upon *potato* the growth is sometimes

in the form of a thin layer having numerous vacuoles; sometimes a thick and shining layer is developed which has a pale sulphur-yellow color or is of a dull-brown mixed with yellow. In *bouillon* a cloudy opacity is first developed; later a flocculent deposit is seen and the liquid above is limpid; it is slightly viscid. In sterilized *milk* the viscosity produced is so great that, in old cultures, threads may be drawn out which are several metres in length. Non-sterilized milk becomes viscous at the end of five hours; later it becomes acid, and at the end of several days the casein is coagulated and is seen as a granular precipitate, above which is a limpid and viscous serum; at this time the milk has acquired a disagreeable odor.

472. BACTERIUM HESSII (Guillebeau).

Obtained from the milk of a cow in the Alps, at an altitude of twelve hundred metres.

Morphology.—Bacilli from 3 to 5 μ long and 1.2 μ thick (from potato cultures); shorter, nearly spherical elements are also seen in considerable numbers, and occasionally long filaments; the ends are round and stain more deeply than the central portion.

Biological Characters.—An *aërobic, liquefying, motile* bacillus. Forms spores. Grows in the usual culture media at the room temperature. In *nutrient gelatin* containing serum from milk, colonies are developed upon plates at the end of a few hours; these at first have well-defined outlines, but later are made up of interlacing filaments; after liquefaction of the gelatin the colony floats upon the surface; liquefaction progresses rapidly and the liquefied gelatin may be drawn out into threads. Upon *potato* a shining layer is developed of a dull-white color, which later acquires a brownish tint. *Bouillon* without sugar is rapidly transformed into a viscous mass having an alkaline reaction. In *milk* an acid reaction is produced and the casein is precipitated at the end of two days. The viscosity produced in milk is less than that produced by the species previously described, and disappears at the end of two days in milk kept at a temperature of 35° C. In old cultures in bouillon a disagreeable odor is developed, said to resemble trimethylamin.

473. BACILLUS DENITRIFICANS.

Obtained by Giltay and Aberson from the soil and from the air. Resembles *Bacterium denitrificans* of Gayou and Dupetit.

Morphology.—Bacilli, usually in pairs, 1.5 μ to 3 μ long and 0.5 μ broad.

Biological Characters.—Completely decomposes nitrates, producing nitrogen monoxide as well as pure nitrogen. The reducing action of the ferment is favored by the presence of carbonate of lime in the culture medium. Grows in ordinary nutrient gelatin. Also cultivated in the following medium: Nitrate of potash two grammes, glucose two grammes, sulphate of magnesia two grammes, citric acid five grammes, monophosphate of potash two grammes, calcium chloride 0.2 gramme, two drops of a solution of perchloride of iron, and one litre of water.

474. BACILLUS CYANO-FUSCUS.

Obtained by Beyerinck from size and glue and Edam cheese.

Morphology.—Bacilli from 0.2 to 0.6 μ long and one-half as thick.

Biological Characters.—An *aërobic, chromogenic, liquefying, motile* bacillus. Grows in the usual culture media at the room temperature. Spore formation not observed. When cultivated in a solution containing one-half per cent of peptone the culture medium acquires at first a green color, which later changes to blue, brown, and black; subsequently the color is almost entirely lost. In *nutrient gelatin* containing one-half per cent of peptone circular colonies are developed which are surrounded by a zone of black pigment and in which crystals of lime are formed.

475. *BACILLUS PYOGENES SOLI*.

Obtained by Bolton from garden earth by inoculation into a rat. Found in association with the tetanus bacillus in pus from the inoculation wound.

Morphology.—Closely resembles the bacillus of diphtheria. "It presents the same irregularities of shape, and the transverse, unstained clear spaces in stained preparations, as the diphtheria bacillus. The individual bacilli vary greatly in length and thickness, and many of them are bent and narrower through the middle than at the poles."

Stains readily with the usual aniline colors, but takes the stain irregularly, sometimes showing deeply stained spots which may be perfectly round. Does not stain by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *non-motile* bacillus. Spore formation not observed with certainty—highly refractive ovoid bodies are sometimes met with, but these do not seem to be specially resistant to heat. In *gelatin roll tubes* very small, spherical colonies are developed, which under a low power are seen to be finely granular and to have a lemon-yellow color. Grows best in a slightly acid medium—very slowly at the room temperature. In *gelatin stick cultures* isolated colonies are formed along the line of puncture. Scanty growth on *potato* or *blood serum*. Bolton says: "I have rarely succeeded in getting a growth in agar."

Pathogenesis.—Subcutaneous inoculations in rats, gray mice, rabbits, and usually in white mice produce an abscess at the point of inoculation. Injections into the ear veins of rabbits sometimes give rise to multiple abscesses, especially in the joints and kidneys. "The abscesses following subcutaneous inoculation form very quickly, within twenty-four hours, and run a longer or shorter course, from forty-eight hours to eight or ten days, in direct proportion to the amount of the culture introduced. The animals do not seem to suffer any inconvenience, as a rule, and after the abscess is opened suppuration ceases. The organism is found aggregated in small and large, irregular clumps in the pus, many of them lying in the pus corpuscles. It seems to form metastatic abscesses only under exceptional circumstances, such as when injected directly into the blood. Otherwise the abscess remains strictly confined to the seat of inoculation in rabbits, white rats, and gray mice."

476. *MICROCOCCUS AQUATILIS INVISIBILIS*.

Obtained by Vaughan from water.

Morphology.—Oval cocci.

Biological Characters.—An *aërobic*, *non-liquefying* micrococcus. Grows in the usual culture media at the room temperature, but feebly at 38° C. On *gelatin plates* deep brown colonies with smooth outline, spreading irregularly upon the surface. In *gelatin tubes* there is a scanty growth along the line of puncture and a spreading growth upon the surface. On *agar* a thin, white growth. The growth upon *potato* is invisible.

Not pathogenic.

477. *BACILLUS GRACILIS ANAËROBIESCENS*.

Obtained by Vaughan from water.

Morphology.—"Bacilli three times as long as broad, often growing into long, slender rods."

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-liquefying*, *actively motile* bacillus. Grows rapidly in the usual culture media at the room temperature—feebly at 38° C. Upon *gelatin plates* brownish colonies are developed, "spreading irregularly." In *gelatin tubes* grows abundantly along the line of puncture and also spreads over the surface. On *agar* a thin, white layer is developed. On *potato* an abundant and promi-

ment yellowish-white layer. Grows in Parietti's solution, but not in Uffelmann's gelatin. Forms gas abundantly in gelatin stick cultures.

Not pathogenic.

478. BACILLUS FIGURANS.

Obtained by Vaughan from water. Crookshank has described a *Bacillus figurans* which appears to be identical with *Bacillus mesentericus vulgatus*.

Morphology.—“Bacilli two to three times as long as broad, but showing marked variation in form. Sometimes they appear as very short bacilli, while at other times they grow into long threads.”

Biological Characters.—An *aërobic, liquefying, sluggishly motile* bacillus. Spore formation not mentioned. Grows rapidly in the usual culture media at the room temperature—feebly at 38° C. “On *gelatin plates* the deep colonies are spherical and smooth; the superficial growth forms curved and interlacing lines, often presenting most grotesque figures. Plates may show no liquefaction after some days.” In *gelatin stick cultures* does not develop along the line of puncture; liquefies slowly, and sometimes the fluid is lost by evaporation as fast as it liquefies. After the gelatin has been liquefied half-way down the tube, the bacteria subside to the bottom and further liquefaction is very slow or does not occur. On *agar* a thin, white layer is formed upon the surface and a heavy deposit in the water of condensation. On *potato* an abundant, faintly yellow, mucilaginous layer. Does not grow either in Parietti's solution or in Uffelmann's gelatin.

Not pathogenic.

479. BACILLUS ALBUS ANAËROBIESCENS.

Obtained by Vaughan from water.

Morphology.—“Bacilli two or three times as long as broad.”

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, non-motile* (“only an oscillation”) bacillus. Grows rapidly at the room temperature in the usual culture media—also at 38° C. Spore formation not mentioned. On *gelatin plates* forms smooth, spherical, yellowish or brownish colonies. In *gelatin stick cultures* grows both on the surface and along the line of puncture. On *agar* a thick, milk-white layer is developed. On *potato* a yellowish-white, glistening growth. Grows both in Parietti's solution and in Uffelmann's gelatin.

Not pathogenic.

480. BACILLUS INVISIBILIS.

Obtained by Vaughan from water.

Morphology.—Large bacilli with rounded ends, from two to five times as long as broad.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, motile* bacillus. Spore formation not mentioned. Grows rapidly in the usual culture media at the room temperature—also at 38° C. On *gelatin plates* pale-yellow, burr-like colonies, with irregular outlines and spreading slightly. In *gelatin tubes* grows abundantly along the line of puncture and also upon the surface, spreading slowly. On *agar* a thick, white growth with but little tendency to spread. On *potato* the growth is invisible. Grows both in Parietti's solution and in Uffelmann's gelatin.

Not pathogenic.

481. BACILLUS VENENOSUS.

Obtained by Vaughan from water.

Morphology.—Bacilli with rounded ends, two to four times as long as broad.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, actively motile* bacillus. Spore formation not mentioned. Grows

rapidly in the usual culture media at the room temperature—also at 38° C. On *gelatin plates* small, white, spherical colonies—sometimes slightly yellow; the superficial colonies are elevated above the surface of the gelatin. In *gelatin tubes* an abundant growth occurs along the line of puncture and slowly extends upon the surface. In cultures from the spleen of an inoculated animal the growth upon the surface is less marked. On *agar* a thin, white layer is formed. On *potato* a light-brown, moist growth. In recent cultures from the spleen of an inoculated animal the growth upon potato may be invisible. Grows abundantly both in Parietti's solution and in Uffelmann's gelatin.

Pathogenesis.—Pathogenic for rats, mice, guinea-pigs, and rabbits.

482. BACILLUS VENENOSUS BREVIS.

Obtained by Vaughan from water.

Morphology.—Short, thick bacilli, about twice as long as broad; in old cultures grows out into threads.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, actively motile* bacillus. Spore formation not mentioned. Grows rapidly in the usual culture media at the room temperature—also at 38° C. On *gelatin plates* forms small, round colonies with concentric rings: the deeper colonies are generally yellowish or brown; the surface colonies are elevated and spread but little. In *gelatin tubes* grows along the line of puncture and spreads slowly upon the surface, finally reaching the sides of the tube. Upon *agar* a thin, white layer is formed. On *potato* a thick and moist, light-brown growth. When kept for fourteen days or longer at 40° C. there is an invisible growth upon potato. Grows abundantly in Parietti's solution and slowly in Uffelmann's gelatin.

Pathogenesis.—Pathogenic for rats, mice, guinea-pigs, and rabbits.

483. BACILLUS VENENOSUS INVISIBILIS.

Obtained by Vaughan from water.

Morphology.—A slender bacillus with rounded ends, from two to four times as long as broad.

Biological Characters.—An *aërobic* and *facultative anaërobic, non-liquefying, motile* bacillus. Spore formation not mentioned. Grows slowly in the usual culture media at the room temperature—also at 38° C. On *gelatin plates* small, granular, yellowish colonies are developed; the superficial colonies are coarsely granular and very irregular in size and outline. In *gelatin tubes* grows slowly both on the surface and along the line of puncture; scarcely visible at end of three days. On *agar* a very thin, white growth. On *potato* the growth is sometimes invisible; on some potatoes a light-brown layer may be developed. Grows well both in Parietti's solution and in Uffelmann's gelatin.

Pathogenesis.—Pathogenic, but in less degree than *Bacillus venenosus*.

484. BACILLUS VENENOSUS LIQUEFACIENS.

Obtained by Vaughan from water.

Morphology.—Bacilli with rounded ends, one and one-half to twice as long as broad.

Biological Characters.—An *aërobic* and *facultative anaërobic, liquefying, motile* bacillus. Spore formation not mentioned. Grows rapidly in the usual culture media at the room temperature—also at 38° C. On *gelatin plates* the deep colonies are finely granular, spherical, and yellowish in color; superficial colonies elevated and spread over the surface. In *gelatin tubes* grows abundantly along the line of puncture and spreads slowly over the surface; liquefaction commences in from four to six weeks. On *agar* a

thin, white growth. On *potato* a moist, light-brown or yellowish growth. When kept for fourteen days or longer on spleen tissue it forms an invisible growth on *potato*. Grows abundantly both in Parietti's solution and in Uffelmann's gelatin.

Pathogenesis.—Pathogenic for mice, rats, guinea-pigs, and rabbits.

485. BACILLUS AËROGENES CAPSULATUS.

Found by Welch in the blood vessels of a patient with thoracic aneurism opening externally; autopsy made in cool weather eight hours after death—the vessels found full of gas bubbles.

Morphology.—Straight or slightly curved bacilli with slightly rounded or sometimes square-cut ends; a little thicker than *Bacillus anthracis*, and varying in length—average length 3 to 6 μ ; long threads and chains are occasionally seen. The bacilli, both from cultures and in the animal body, are enclosed in a transparent capsule.

Biological Characters.—An *anaërobic*, *non-motile*, *non-liquefying* bacillus. Does not form spores. Grows in the usual culture media, in the absence of oxygen, at the room temperature, and produces an abundant development of gas in all. In *nutrient gelatin* there is no marked liquefaction, but the gelatin is slightly peptonized. In *agar*, colonies are developed which are usually one to two millimetres in diameter, but may attain a diameter of one centimetre; they are grayish-white in color and in the form of flattened spheres, ovals, or irregular masses, beset with little projections or hair-like processes. *Bouillon* is rendered diffusely cloudy, with an abundant white sediment. *Milk* is coagulated in one or two days. The cultures in *agar* and *bouillon* have a faint odor, comparable to that of stale glue. Upon *potato* a pale grayish-white layer is developed; growth occurs at 18° to 20° C., but is much more rapid at 30° to 37° C. *Bouillon* cultures are sterilized by exposure to a temperature of 58° C. for ten minutes.

Pathogenesis.—"Quantities up to 2.5 cubic centimetres of fresh *bouillon* cultures were injected into the circulation of rabbits without any apparent effect, except in one instance in which a pregnant rabbit was killed, by the injection of one cubic centimetre, in twenty-one hours. If the animal is killed shortly after the injection the bacilli develop rapidly after death, with an abundant formation of gas in the blood vessels and organs, especially the liver. At temperatures of 18° to 20° C. the vessels, organs, and serous cavities may be full of gas in eighteen to twenty-four hours, and at temperatures of 30° to 32° C. in four to six hours, when one cubic centimetre of a *bouillon* culture has been injected into the circulation shortly before death."

It is suggested by Welch and Nuttall that in some of the cases in which death has been attributed to the entrance of air into the veins, the gas found at the autopsy may not have been atmospheric air, but may have been produced by this or some similar microorganism entering the circulation and developing after death.

486. BACILLUS OF CANON AND PIELICKE.

Found by Canon and Pielicke (1892) in the blood of fourteen patients with measles, and supposed to be the etiological agent in this disease.

Morphology.—Bacilli varying greatly in size; sometimes the length is equal to the diameter of a red blood corpuscle, others are quite short and resemble diplococci; often united in pairs.

Stained by Canon, in blood drawn from the finger, by the use of the following solution: Concentrated aqueous solution of methylene blue, forty cubic centimetres; one-quarter-per-cent solution of eosin in seventy-per-cent alcohol, twenty cubic centimetres; distilled water, forty cubic centimetres. The preparations were first placed in absolute alcohol for five to ten minutes, then placed in the staining solution in the incubating oven at 37° C. from

six to twenty hours. Some of the bacilli do not stain uniformly, but present the appearance of stained spots alternating with unstained portions.

Biological Characters not determined. Does not grow in glycerin-agar or in blood serum. In *bouillon* inoculated with blood from the finger of a measles patient, bacilli were obtained in three cultures which resembled the bacillus found in the blood, and which failed to grow when transplanted to glycerin-agar, blood serum, or *bouillon*. At first the *bouillon* remained clear, with a sediment at the bottom partly made up of the inoculated blood; after several days a faint cloudiness was noticed and small flocculi formed. In these *bouillon* cultures the bacilli had various forms and dimensions, some of them exceeding in length those found in stained preparations from the blood. They appeared to have a slight independent motion. The bacilli in these *bouillon* cultures did not stain by Gram's method. The bacilli referred to were found in the blood preparations in varying numbers—sometimes very few, and at others the first field examined was crowded. They were found during the whole course of the disease, and in one case three days after the fever had disappeared. They were also found in the secretions from the nose and conjunctiva of measles patients.

487. *BACILLUS SANGUINIS TYPHI*.

Obtained (1892) by Brannan and Cheesman from the blood of typhus-fever patients. "The blood, obtained under strict antiseptic precautions from the six living patients, was streaked on six-per-cent glycerin-agar plates, and smeared on sterilized cover glasses by Dr. Brannan and brought at once to the laboratory. The cover-glass smears from all the cases, being dried at once in the air, were fixed in alcohol and stained in Czenzynski's solution for eighteen hours at room temperature. Although all of these covers were examined throughout with a one-sixteenth homogeneous immersion lens in the most careful manner, in only about one-half of them a few blue-stained bacilli were found, never more than eight or ten on a cover."

Morphology.—Bacilli with round ends, from 1 to 2.5 μ long and 0.5 to 0.8 μ broad; solitary or in pairs, and occasionally in chains containing six to eight elements; often club-shaped, or ovoid in recent cultures.

Stains with the usual aniline colors and by Gram's method.

Biological Characters.—An *aërobic* and *facultative anaërobic*, *non-motile* bacillus. Does not form spores. Does not grow at a lower temperature than 27° C. Grows best upon blood serum at 37.5° C. Upon glycerin-agar plates colonies are developed which at the end of eighteen hours appear as minute, bluish-gray, translucent spots, the diameter of which does not exceed 0.25 millimetre; later the colonies appear dry and scaly, they are flat, more opaque, and whiter, and do not exceed two millimetres in diameter. Under a low power the recent colonies are seen to be granular, to have a sinuous and sharply defined margin and a pale-brown color which is more intense at the centre and in scattered points upon the surface. When magnified one hundred diameters the surface appears to be coarsely granular, and coarse, irregular spiculæ are seen about the margin. In *glycerin-agar* tubes, at 37.5° C., growth occurs upon the surface and along the line of puncture as small, white, isolated colonies. Upon *blood serum* a slightly elevated, white, shining layer is developed. In *milk* a white deposit is formed at the bottom of the tube and the milk undergoes no apparent change. On *potato* no visible growth was obtained.

Pathogenesis.—"Inoculations of cultures of the bacillus obtained from two of the cases were made in eight rabbits, two guinea-pigs, and two white mice. All the animals showed marked emaciation, and, with the exception of two rabbits, all the animals experimented upon died in from ten to twenty-nine days. The inoculated bacillus was obtained from the heart's blood of two of the rabbits that died."

488. MICROCOCCUS AGILIS CITREUS.

Obtained by Menge (1882) from an infusion of peas—probably from the air.

Morphology.—Micrococci, usually in pairs, but sometimes in short chains or irregular groups. Has a flagellum which is easily demonstrated by Löffler's method of staining, and which is about six times as long as the diameter of the micrococcus.

Biological Characters.—An *aërobie*, *non-liquefying*, *chromogenic*, *motile* micrococcus. Forms a yellow pigment. Grows in the usual culture media at the room temperature. Upon *gelatin plates* a diffuse cloudiness of the gelatin occurs around the superficial colonies and extends over the plate, except at the numerous points where bundles of crystals are developed. In *gelatin stick cultures* a scanty growth occurs along the line of puncture, which is not colored; upon the surface a round layer of an intense yellow color is slowly developed. Upon *agar* a pale and thin layer is developed along the line of inoculation by the end of the third day; this increases in breadth and thickness and acquires a yellow color; the growth upon agar

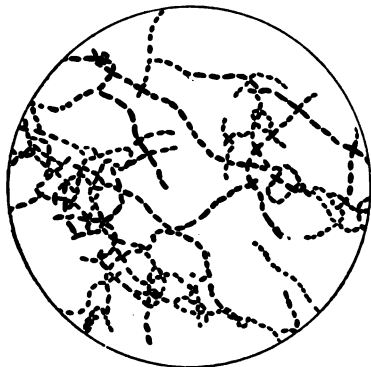


FIG. 267.

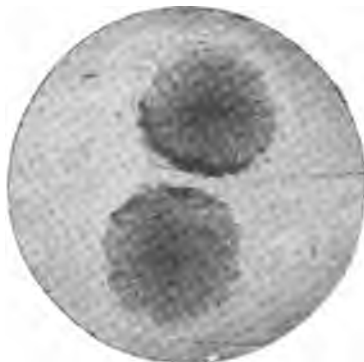


FIG. 268.

FIG. 267.—*Bacillus gracilis cadaveris*, from a gelatin culture. $\times 1,000$. From a photomicrograph. (Sternberg.)

FIG. 268.—*Bacillus gracilis*; colonies in gelatin roll tube, end of forty-eight hours. $\times 12$. From a photograph. (Sternberg.)

is extremely viscid and may be drawn out into long threads when touched with a platinum needle. In *bouillon* a diffuse cloudiness occurs and a yellow, viscid deposit accumulates at the bottom of the tube; there is no film formed upon the surface. Upon *potato* development is very slow, but after a time an abundant bright-yellow layer is formed, and around this the potato acquires a slightly bluish-gray color. Grows in *milk* without producing coagulation. Grows best at a temperature of 20°C .

489. BACILLUS GRACILIS CADAVERIS (Sternberg).

Obtained (1889) from a fragment of liver, of man, kept for forty-eight hours in an antiseptic wrapping.

Morphology.—Bacilli about 1μ broad and 2μ long, associated in long chains.

Biological Characters.—An *aërobie* and *facultative anaërobie*, *non-motile*, *non-liquefying* bacillus. Spore formation not observed. In *gelatin*

roll-tubes the deep colonies are opaque and spherical; superficial colonies circular or slightly irregular in outline, white in color, and opaque or slightly translucent. In *gelatin stick cultures*, at 22° C., at the end of five days a rather thick, white mass at the point of puncture, covering one-third of the surface, and closely crowded, opaque colonies at bottom of line of puncture, with slender, branching outgrowth above. In *nutrient agar*, at the end of five days at 22° C., a milk-white growth upon the surface and opaque growth to bottom of line of puncture. On *potato*, at end of five days at 22° C., rather thick, cream-white growth with irregular margins along the impfstich. Cultures in *bouillon* have a milky opacity and a very disagreeable odor. Grows in *agua coco* without formation of gas.

Pathogenic for rabbits when injected into the cavity of the abdomen.

XIII.

BACTERIOLOGICAL DIAGNOSIS.

THE researches made by bacteriologists during the past ten years show that there is an extensive bacterial flora, especially in water and in the soil, and that many of the species known are widely distributed and may be recognized by their morphological and biological characters wherever they may be found ; but they also show that we cannot depend upon morphology alone for the differentiation of species, and that in many cases a careful study of the mode of growth in various culture media, and of pathogenic power by inoculations in the lower animals, shows slight differences in bacteria which resemble each other so closely in form and in certain biological characters that a less careful study would lead to the belief that they were identical. That there are true species among the bacteria, in the same sense as among the higher plants, is well established; but, as among the higher plants, it is often difficult to determine whether the differences observed should be considered sufficient to justify the description of allied forms as distinct species, or whether they should simply be considered as varieties of a single species. For example, the well-known streptococcus of pus has morphological characters which enable us to distinguish it from many other bacteria, but streptococci have been obtained from various sources which present slight differences as to their growth in certain media and in their pathogenic power. The question arises as to whether these differences are to be considered "specific" or otherwise. If the differences noted are permanent, and enable a bacteriologist to distinguish one streptococcus from the other wherever it may be found, we are justified in considering them as two distinct micro-organisms. And the decision as to whether they are to be considered as different species, or as varieties of a single species, need not detain us. As a matter of fact, nature is continuous, and specific lines are not sharply drawn except in systematic text books of botany, etc. The more complete our knowledge of any class of animal or vegetable organisms, the less sharply differentiated are the so-called species on account of the introduction of intermediate forms in a series having common characters.

When the differences noted are not permanent in character, description under a distinct name only leads to confusion. Among the bacteria, as among higher plants, such differences, constituting more or less permanent varieties, have been developed by cultivation under various conditions, and, without doubt, are constantly being developed under natural conditions as a result of changes in the environment of these minute plants. Thus we have artificial varieties of certain common chromogenic species in which no pigment is produced, non-pathogenic varieties of pathogenic species, and asporogenous varieties of bacilli which usually form spores.

The attempt to classify the bacteria in a systematic manner is attended with especial difficulties, owing to their simple structure, the comparatively slight morphological differences which they present, and also because of the tendency to variation in their biological characters above referred to. But bacteriologists are generally agreed upon the importance of the following characters for their differentiation, viz. : *form*—micrococci, bacilli, spirilla, polymorphous ; *relation to oxygen*—aërobic, facultative anaërobic, strict anaërobic ; *growth in nutrient gelatin*—liquefy, do not liquefy, do not grow in nutrient gelatin at the “room temperature” ; *growth on potato* ; *growth in milk*—coagulate milk, do not coagulate, etc. : *color of growth*—chromogenic, non-chromogenic ; *spore formation* ; *independent movements* ; *pathogenic power*.

It is upon these characters that we must rely chiefly in our bacteriological diagnosis of known species. But the student must remember that the lines are not sharply drawn between the groups formed when we classify bacteria with reference to any one of these characters. Thus we have microorganisms of this class which we find it difficult to classify as regards form, because they are not round, and yet are so slightly elongated in one diameter that it is difficult to consider them bacilli. If we follow Cohn and group these short-oval bacteria under the generic name *Bacterium*, we have not removed the difficulty, but have made two arbitrary and artificial lines instead of one. Again, liquefaction of gelatin is sometimes so slight, or occurs at so late a date, that it may be a question whether a microorganism of this class should be included among the liquefying or non-liquefying bacteria. And our division with reference to the formation of pigment must, to a certain extent, be arbitrary ; for many species which are not decidedly chromogenic present under certain circumstances a slight tint of yellow, gray, brown, or pink. A slight yellow or a decided brown color is often developed in potato cultures of bacteria which upon other culture media present a colorless growth, and which are generally included by bacteriologists among the non-chromogenic species. With reference to independent

movements, it must be remembered that some of the motile bacteria, under certain circumstances, do not exhibit active movements; the question of motility must therefore not be hastily decided in the negative from a single examination. Spore formation also, in many cases, depends upon special conditions, and great care will often be required in determining this character, which, indeed, is still undetermined for some of the best-known species.

We have endeavored in the present volume to include all bacteria which have been described by competent bacteriologists with sufficient detail to permit of their recognition when carefully studied by the usual methods. But we have also included a considerable number which are imperfectly described and which could not be identified by the descriptions given. And no doubt a certain number of those which have been described under different names are in fact identical or simply varieties of a single species. The plan adopted of grouping the different bacteria described with reference to their morphological and biological characters will bring together those micro-organisms which are similar, and will, we trust, be of assistance to working bacteriologists in determining identity or non-identity. Imperfect descriptions, if not completed by future researches, may be eliminated hereafter, but we have thought it best to give them a place in this Manual, as most of them are included in systematic works published abroad—*e.g.*, the micrococci found by Koch in his experimental study of traumatic infectious diseases (1878), *Bienstock's* *fæces bacilli*, etc.

LIST OF BACTERIA DESCRIBED.

PART THIRD, SECTION IV.—PYOGENIC BACTERIA.

1. *Staphylococcus pyogenes aureus* (Rosenbach).
Micrococcus of infectious osteomyelitis (Becker).
2. *Staphylococcus pyogenes albus* (Rosenbach).
Staphylococcus epidermidis albus (Welch).
3. *Staphylococcus pyogenes citreus* (Passet).
4. *Micrococcus pyogenes tenuis* (Rosenbach).
5. *Streptococcus pyogenes* (Rosenbach).
Micrococcus of erysipelas (Fehleisen).
Streptococcus of pus.
Streptococcus longus (Von Lingelsheim).
6. *Micrococcus gonorrhœæ*.
Gonococcus (Neisser).

PART THIRD, SECTION V.—BACTERIA IN CROUPOUS PNEUMONIA.

7. *Bacillus* of Friedländer.

- Pneumococcus (Friedländer).
- Bacillus pneumoniae (Flügge).
- 8. Micrococcus pneumoniae crouposæ.
- Micrococcus Pasteuri (Sternberg).
- Micrococcus of sputum septicæmia (Fränkel).
- Diplococcus pneumoniae (Weichselbaum).
- Bacillus septicus sputigenus (Flügge).
- Bacillus salivarius septicus (Biondi).
- Lancet-shaped micrococcus (Talamon).
- Streptococcus lanceolatus Pasteuri (Gameléia).

PART THIRD, SECTION VI.—PATHOGENIC MICROCOCCI NOT
DESCRIBED IN SECTIONS IV. AND V.

- 9. Diplococcus intercellularis meningitidis (Weichselbaum).
- 10. Staphylococcus salivarius pyogenes (Biondi).
- 11. Micrococcus of progressive tissue necrosis in mice (Koch).
- 12. Micrococcus of progressive abscess formation in rabbits (Koch).
- 13. Micrococcus of pyæmia in rabbits (Koch).
- 14. Micrococcus of septicæmia in rabbits (Koch).
- 15. Micrococcus salivarius septicus (Biondi).
- 16. Micrococcus subflavus (Flügge).
- Yellowish-white micrococcus (Bumm).
- 17. Micrococcus of trachoma ? (Sattler).
- 18. Micrococcus tetragenus (Gaffky, Koch).
- 19. Micrococcus botryogenus (Rabe).
- Micrococcus of "Myko-desmoids" of the horse.
- Micrococcus askoformans (Johne).
- Micrococcus Johnnei (Cohn).
- 20. Micrococcus of Manfredi.
- Micrococcus of progressive granuloma formation (Manfredi).
- 21. Micrococcus of bovine mastitis (Kitt).
- 22. Micrococcus of bovine pneumonia ? (Poels and Nolen).
- 23. Streptococcus septicus (Flügge).
- 24. Streptococcus bombycis.
- Microzyma bombycis (Béchamp).
- 25. Nosema bombycis.
- Micrococcus ovatus.
- Panhistophyton ovatum.
- 26. Micrococcus of Heydenreich.
- Micrococcus of Biskra button—"Clou de Biskra," *Fr.*; "Pendesche Geschwur," *Ger.*
- 27. Micrococcus of Demme.
- Diplococcus of pemphigus acutus (Demme).

28. *Streptococcus* of Manneberg.
29. *Micrococcus endocarditidis rugatus* (Weichselbaum).
30. *Micrococcus* of gangrenous mastitis in sheep (Nocard).
31. *Streptococcus* of mastitis in cows (Nocard and Mollereau).
32. *Diplococcus* of pneumonia in horses (Schütz).
33. *Streptococcus coryzæ contagiosæ equorum* (Schütz).
34. *Hæmatococcus bovis* (Babes).
35. *Micrococcus gingivæ pyogenes* (Miller).
36. *Pseudodiplococcus pneumoniæ* (Bonome).
37. *Streptococcus septicus liquefaciens* (Babes).
38. *Micrococcus* of Kirchner.
39. *Micrococcus* No. II. of Fischel.
40. *Streptococcus* of Bonome.
41. *Micrococcus* of Almquist.
42. *Staphylococcus pyosepticus* (Hericourt and Richet).
43. *Streptococcus perniciosus psittacorum*.
Micrococcus of gray parrot disease (Eberth and Wolff).
44. *Micrococcus* of Forbes.

PART THIRD, SECTION VII.—THE BACILLUS OF ANTHRAX.

45. *Bacillus anthracis*.
Milzbrandbacillus, *Ger.*
La bactérie du charbon, *Fr.*

PART THIRD, SECTION VIII.—THE BACILLUS OF TYPHOID FEVER.

46. *Bacillus typhi abdominalis*.
Bacillus typhosus.
Typhus bacillus (Eberth, Gaffky).

PART THIRD, SECTION IX.—BACTERIA IN DIPHTHERIA.

47. *Bacillus diphtheriæ* (Klebs, Löffler).
48. Pseudo-diphtheritic bacillus (Roux and Yersin).
49. *Bacillus diphtheriæ columbarum* (Löffler).
50. *Bacillus diphtheriæ vitulorum* (Löffler).
51. *Bacillus* of intestinal diphtheria in rabbits (Ribbert).

PART THIRD, SECTION X.—BACTERIA IN INFLUENZA.

52. *Bacillus* of influenza (Pfeiffer, Canon).

PART THIRD, SECTION XI.—BACILLI IN CHRONIC INFECTIOUS DISEASES.

53. *Bacillus tuberculosis* (Koch).
Tubercle bacillus.

54. *Bacillus tuberculosis gallinarum* (Maffucci).
55. *Bacillus lepræ* (Hansen, Neisser).
Leprosy bacillus.
56. *Bacillus mallei* (Löffler and Schütz).
Bacillus of glanders.
Rotzbacillus, *Ger.*
Bacille de la morve, *Fr.*
57. Bacillus of Lustgarten.
Syphilis bacillus (?)
58. Bacillus of rhinoscleroma (?).
59. Bacillus of Koubasoff.
60. Bacillus of Nocard.
Bacille du farcin du bœuf.

PART THIRD, SECTION XII.—BACILLI WHICH PRODUCE SEPTICÆMIA
IN SUSCEPTIBLE ANIMALS.

61. *Bacillus septicæmiæ hæmorrhagicæ* (Hueppe).
Bacillus of fowl cholera.
Microbe du choléra des poules (Pasteur).
Bacillus cholerae gallinarum (Flügge).
Bacillus der Hühnercholera.
Bacillus of rabbit septicæmia.
Bacillus der Kaninchenseptikämie (Koch).
Bacillus cuniculicida (Flügge).
Bacillus der Rinderseuche (Kitt).
Bacillus der Schweineseuche (Löffler and Schütz).
Bacillus der Wildseuche (Hueppe).
Bacillus der Büffelseuche (Oreste-Armanni).
Bacterium of Davaine's septicæmia (?)
62. Bacillus of cholera in ducks (Cornil and Toupet).
63. Bacillus of hog cholera (Salmon and Smith).
Bacillus of swine plague (Billings).
Bacillus of swinepest (Selander).
64. Bacillus of Belfanti and Pascarola.
Impftetanus bacillus (Belfanti and Pascarola).
65. Bacillus of swine plague, Marseilles.
Bacillus der Schweineseuche (Rietsch and Jobert).
Bacillus der Frettenseuche (Eberth and Schimmelbusch).
Bacillus der amerikanischen Rinderseuche (Caneva).
Bacillus of spontaneous rabbit septicæmia (Eberth).
66. *Bacillus septicus agrigenus* (Nicolaiër).
67. Bacillus erysipelatos suis.
Bacillus of hog erysipelas.
Bacillus des Schweinerothlauf (Löffler, Schütz).

- Bacille du rouget du porc (Pasteur).
 Bacillus of mouse septicæmia.
 Bacillus murisepticus (Flügge).
 Bacillus des Mäuseseptikämie (Koch).
 68. Bacillus coprogenes parvus (Bienstock).
 Mäuseseptikämieähnlicher Bacillus (Eisenberg).
 69. Bacillus cavicola (Brieger).
 Brieger's bacillus.
 70. Bacillus cavicola Havaniensis (Sternberg).
 71. Bacillus crassus sputigenus (Kreibohm).
 72. Bacillus pyogenes fœtidus (Passet).
 73. Proteus hominis capsulatus (Bordoni-Uffreduzzi).
 74. Proteus capsulatus septicus (Banti).
 75. Bacillus enteritidis (Gärtner).
 76. Bacillus of grouse disease (Klein).
 77. Bacillus gallinarum (Klein).
 78. Bacillus smaragdinus fœtidus (Reimann).
 79. Bacillus pneumosepticus (Babes).
 80. Bacillus capsulatus (Pfeiffer).
 81. Bacillus hydrophilus fuscus (Sanarelli).
 82. Bacillus tenuis sputigenus (Pansini).
 83. Bacillus of Laser.
 84. Bacillus typhi murium (Löffler).
 85. Bacillus of Cazal and Vaillard.
 86. Bacillus of Babes and Oprescu.
 87. Bacillus of Lucet.
 88. Capsule bacillus of Loeb.

PART THIRD, SECTION XIII.—PATHOGENIC AËROBIC BACILLI NOT
 DESCRIBED IN PREVIOUS SECTIONS.

89. Bacillus coli communis.
 Bacterium coli commune (Escherich).
 Colon bacillus.
 90. Bacillus lactis aërogenes.
 Bacterium lactis aërogenes (Escherich).
 91. Bacillus c of Booker.
 92. Bacillus acidiformans (Sternberg).
 93. Bacillus cuniculicida Havaniensis (Sternberg).
 94. Bacillus leporis lethalis (Sternberg).
 Bacillus of Gibier.
 95. Bacillus pyocyanus (Gessard).
 Bacillus of green pus.
 Microbe du pus bleu.
 Bacillen des grünblauen Eiters.

- Bacterium* *aëruginosum*.
96. *Bacillus* of *Fiocca*.
97. *Proteus vulgaris* (Hauser).
98. *Proteus* of *Karlinsky*.
 Bacillus *murisepticus pleomorphus* (Karlinsky).
99. *Proteus mirabilis* (Hauser).
100. *Proteus Zenkeri* (Hauser).
101. *Proteus septicus* (Babes).
102. *Proteus* *lethalis*.
 Proteus *bei Lungengangrän des Menschen* (Babes).
103. *Bacillus* *A* of *Booker*.
104. *Bacillus endocarditidis griseus* (Weichselbaum).
105. *Bacillus endocarditidis capsulatus* (Weichselbaum).
106. *Bacillus* of *Lesage*.
 Bacillus of green diarrhoea of infants (Lesage).
107. *Bacillus* of *Demme*.
 Bacillus of erythema nodosum (Demme).
108. *Bacillus* *oedematis aërobicus* (Klein).
109. *Bacillus* of *Letzerich*.
 Bacillus of nephritis interstitialis (Letzerich).
110. *Bacillus* of *Schimmelbusch*.
 Bacillus *nomæ* (Schimmelbusch).
111. *Bacillus foetidus ozaenæ* (Hajek).
112. *Bacillus* of *Lumnitzer*.
 Bacillus of putrid bronchitis (Lumnitzer).
113. *Bacillus* of *Tommasoli*.
 Bacillus of sycosis (Tommasoli).
114. *Bacillus* of *Schou*.
 Bacillus of vagus pneumonia (Schou).
115. *Bacillus* *necrophorus* (Löffler).
116. *Bacillus coprogenes foetidus* (Schottelius).
 Darmbacillus of Schottelius.
117. *Bacillus oxytocus perniciosus* (Wyssokowitsch).
118. *Bacillus saprogenes* No. II. (Rosenbach).
119. *Bacillus* of *Afanassiew*.
 Bacillus of whooping cough (Afanassiew).
120. *Pneumobacillus liquefaciens bovis* (Arloing).
121. *Bacillus pseudotuberculosis* (Pfeiffer).
122. *Bacillus gingivæ pyogenes*.
 Bacterium gingivæ pyogenes (Miller).
123. *Bacillus dentalis viridans* (Miller).
124. *Bacillus pulpæ pyogenes* (Miller).
125. *Bacillus septicus keratomalaciæ* (Babes).
126. *Bacillus septicus acuminatus* (Babes).

127. *Bacillus septicus ulceris gangrænosi* (Babes).
128. *Bacillus* of Tricomi.
Bacillus of senile gangrene (Tricomi).
129. *Bacillus albus cadaveris* (Strassmann and Stricker).
130. *Bacillus varicosus conjunctivæ* (Gombert).
131. *Bacillus meningitidis purulentæ* (Neumann and Schäffer).
132. *Bacillus septicus vesicæ* (Clado).
Bacillus of cystitis (Clado).
133. *Bacillus* of Gessner.
Bacterium tholoideum (Gessner).
134. *Bacillus chromo-aromaticus* (Galtier).
135. *Bacillus canalis capsulatus* (Mori).
136. *Bacillus canalis parvus* (Mori).
137. *Bacillus indigenus* (Alvarez).
138. *Bacillus* of Kartulis.
Bacillus of Egyptian ophthalmia (Kartulis).
139. *Bacillus* of Utpadel.
140. *Bacillus alvei* (Cheshire and Cheyne).
Bacillus of foul brood of bees.
141. *Bacillus* of acne contagiosa of horses (Dieckerhoff and Grawitz).
142. *Bacillus* No. I. of Roth.
143. *Bacillus* No. II. of Roth.
144. *Bacillus* of Okada.
145. *Bacillus* of purpura hæmorrhagica of Tizzoni and Giovannini.
146. *Bacillus* of purpura hæmorrhagica of Babes.
147. *Bacillus* of purpura hæmorrhagica of Kolb.
148. *Bacillus heminecrobophilus* (Arloing).

PART THIRD, SECTION XIV.—PATHOGENIC ANAEROBIC BACILLI.

149. *Bacillus tetani* (Nicolai).
Tetanus bacillus.
150. *Bacillus œdematis maligni*.
Bacillus of malignant œdema.
Vibrion septique (Pasteur).
151. *Bacillus cadaveris* (Sternberg).
152. *Bacillus* of symptomatic anthrax.
Bacille du charbon symptomatique (Arloing, Cornevin, and Thomas).
Rauschbrandbacillus.

PART THIRD, SECTION XV.—PATHOGENIC SPIRILLA.

153. *Spirillum Obermeieri*.
Spirochæte Obermeieri.
Spirillum of relapsing fever.

- Recurrentsspirochäte.
154. *Spirillum anserum* (Sakharoff).
 155. *Spirillum cholerae Asiaticæ* (Koch).
Spirillum of Asiatic cholera.
Comma bacillus of Koch.
Bacille-virgule cholérigène.
 156. *Spirillum* of Finkler and Prior.
Vibrio proteus.
 157. *Spirillum tyrogenum*.
Käsespirillen (Deneke).
Cheese spirillum of Deneke.
 158. *Spirillum Metschnikovi*.
Vibrio Metschnikovi (Gameléia).

PART FOURTH, SECTION VIII.—NON-PATHOGENIC MICROCOCCI.

159. *Micrococcus flavus liquefaciens* (Flügge).
160. *Micrococcus flavus desidens* (Flügge).
161. *Micrococcus agilis* (Ali-Cohen).
162. *Micrococcus fuscus* (Maschek).
163. *Diplococcus citreus conglomeratus* (Bumm).
164. *Diplococcus citreus liquefaciens* (Unna).
165. *Diplococcus flavus liquefaciens tardus* (Unna).
166. *Diplococcus fluorescens foetidus* (Klamann).
167. *Diplococcus luteus* (Adametz).
168. *Diplococcus roseus* (Bumm).
169. *Micrococcus cremoides* (Zimmermann).
170. *Micrococcus roseus* (Eisenberg).
171. *Micrococcus aurantiacus* (Cohn).
172. *Micrococcus cerasinus siccus* (List).
173. *Micrococcus versicolor* (Flügge).
174. *Micrococcus* of Dantec.
175. *Micrococcus carneus* (Zimmermann).
176. *Micrococcus cinnabareus* (Flügge).
177. *Micrococcus cereus albus* (Passet).
178. *Micrococcus cereus flavus* (Passet).
179. *Micrococcus citreus*.
Crème-farbiger micrococcus (List).
180. *Micrococcus fervidosus* (Adametz).
181. *Micrococcus flavus tardigratus* (Flügge).
182. *Micrococcus luteus* (Cohn).
183. *Micrococcus violaceus* (Cohn).
184. *Staphylococcus viridis flavescens* (Guttmann).
185. *Micrococcus ochroleucus* (Prove).
186. *Micrococcus acidilactici liquefaciens* (Kreuger).

187. *Micrococcus aërogenes* (Miller).
188. *Micrococcus albus liquefaciens* (Von Besser).
189. *Micrococcus fœtidus* (Klamann).
190. *Micrococcus radiatus* (Flügge).
191. *Diplococcus albicans amplius* (Bumm).
192. *Micrococcus candicans* (Flügge).
193. *Micrococcus candidus* (Cohn).
194. *Micrococcus acidi lactici* (Marpmann).
195. *Micrococcus lactis viscosus* (Conn).
196. *Sphærococcus acidi lactici* (Marpmann).
197. *Micrococcus aquatilis* (Bolton).
198. *Micrococcus concentricus* (Zimmermann).
199. *Micrococcus cumulatus tenuis* (Von Besser).
200. *Micrococcus plumosus* (Bräutigam).
201. *Micrococcus rosettaceus* (Zimmermann).
202. *Micrococcus ureæ* (Pasteur).
203. *Micrococcus ureæ liquefaciens* (Flügge).
204. *Micrococcus viticulosus* (Katz).
205. *Diplococcus albicans tardissimus* (Eisenberg).
Milk-white diplococcus (Bumm).
206. *Diplococcus albicans tardus* (Unna).
207. *Staphylococcus albus liquefaciens*.
White liquefying staphylococcus (Escherich).
208. *Micrococcus ovalis* (Escherich).
209. *Diplococcus coryzæ* (Hajek).
210. *Micrococcus Finlayensis* (Sternberg).
211. *Micrococcus* of Freire.
Cryptococcus xanthogenicus (Freire).
212. *Streptococcus coli gracilis* (Escherich).
213. *Streptococcus acidi lactici* (Grotenfeld).
214. *Streptococcus giganteus urethræ* (Lustgarten).
215. *Streptococcus albus* (Maschek).
216. *Streptococcus vermiformis* (Maschek).
217. *Streptococcus brevis* (Von Lingelsheim).
Streptococcus cadaveris (Sternberg).
218. *Streptococcus Havaniensis* (Sternberg).
219. *Streptococcus liquefaciens* (Sternberg).
220. *Micrococcus tetragenus versatilis* (Sternberg).
221. *Pediococcus albus* (Lindner).
222. *Pediococcus acidi lactici* (Lindner).
223. *Pediococcus cerevisiæ* (Balcke).
224. *Micrococcus tetragenus mobilis ventriculi* (Mendoza).
225. *Micrococcus tetragenus subflavus* (Von Besser).
226. *Sarcina aurantiaca*.

- 227. *Sarcina lutea* (Schröter).
- 228. *Sarcina flava* (De Bary).
- 229. *Sarcina rosea* (Schröter).
- 230. *Sarcina alba* (Eisenberg).
- 231. *Sarcina candida* (Reinke).
- 232. *Sarcina pulmonum* (Hauser).
- 233. *Sarcina ventriculi* (Goodsir).
- 234. *Micrococcus amylovorus* (Burrill).
- 235. *Ascococcus Billrothii* (Cohn).
- 236. *Leuconostoc mesenteroides* (Cienkowski).

PART FOURTH, SECTION IX.—NON-PATHOGENIC BACILLI.

A. Chromogenic, Non-Liquefying Bacilli.

- 237. *Bacterium luteum* (List).
- 238. *Bacillus aurantiacus* (Frankland).
- 239. *Bacillus brunneus* (Adametz).
- 240. *Bacillus aureus* (Adametz).
- 241. *Bacillus flavocoriaceus* (Eisenberg).
Sulphur-yellow bacillus of Adametz.
- 242. *Bacillus berolinensis Indicus* (Clässen).
- 243. *Bacillus constrictus* (Zimmermann).
- 244. *Bacillus fluorescens aureus* (Zimmermann).
- 245. *Bacillus fluorescens longus* (Zimmermann).
- 246. *Bacillus fluorescens tenuis* (Zimmermann).
- 247. *Bacillus fluorescens non-liquefaciens* (Eisenberg).
- 248. *Bacillus fluorescens putidus* (Flügge).
- 249. *Bacillus erythrosporus* (Eidam).
- 250. *Bacillus viridis pallescens* (Frick).
- 251. *Bacillus virescens* (Frick).
- 252. *Bacillus iris* (Frick).
- 253. *Bacillus fuscus* (Zimmermann).
- 254. *Bacillus rubefaciens* (Zimmermann).
- 255. *Bacillus striatus flavus* (Von Besser).
- 256. *Bacillus subflavus* (Zimmermann).
- 257. *Bacillus cyanogenus* (Hueppe).
- 258. *Bacillus fuscus limbatus* (Scheibenzuber).
- 259. *Bacillus latericeus* (Eisenberg).
Ziegelroter bacillus (Adametz).
- 260. *Bacillus spiniferus* (Unna).
- 261. *Bacillus rubescens* (Jordan).
- 262. *Bacillus allii* (Griffiths).

B. Chromogenic, Liquefying Bacilli.

- 263. *Bacillus fulvus* (Zimmermann).
- 264. *Bacillus helvolus* (Zimmermann).
- 265. *Bacillus ochraceus* (Zimmermann).
- 266. *Bacillus plicatilis* (Zimmermann).
- 267. *Bacillus janthinus* (Zopf).
Violet bacillus.
- 268. *Bacillus violaceus* Laurentius (Jordan).
- 269. *Bacillus tremelloides* (Schottelius).
- 270. *Bacillus cuticularis* (Tils).
- 271. Flesh-colored bacillus (Tils).
- 272. *Bacillus arborescens* (Frankland).
- 273. *Bacillus citreus cadaveris* (Strassmann).
- 274. *Bacillus membranaceus amethystinus* (Eisenberg).
- 275. *Ascobacillus citreus* (Unna).
- 276. *Bacillus coeruleus* (Smith).
- 277. *Bacillus fluorescens liquefaciens* (Flügge).
- 278. *Bacillus fluorescens liquefaciens minutissimus* (Unna).
- 279. *Bacillus fluorescens nivalis* (Schmolck).
- 280. *Bacillus lactis erythrogenes* (Hueppe).
- 281. *Bacillus glaucus* (Maschek).
- 282. *Bacillus lividus* (Plagge and Proskauer).
- 283. *Bacillus Indicus* (Koch).
- 284. *Bacillus prodigiosus*.
Micrococcus prodigiosus.
Monas prodigiosa.
- 285. *Bacillus mesentericus ruber*.
Rothen Kartoffelbacillus (Globig).
- 286. *Bacillus pyocyaneus* β (Ernst).
- 287. *Bacillus mycoides roseus* (Scholl).
- 288. *Bacillus rosaceum metalloides* (Dowdeswell).
- 289. *Bacillus viscosus* (Frankland).
- 290. *Bacillus violaceus*.
- 291. *Bacillus sulfureum* (Holschewnikoff).
- 292. *Bacillus rubidus* (Eisenberg).
- 293. *Bacterium termo* of Vignal.
- 294. *Bacillus buccalis minutus*.
Bacillus g of Vignal.
- 295. *Bacillus* of Canestrini.
(Pathogenic for bees.)

C. Non-chromogenic, Non-liquefying Bacilli.

296. *Bacillus ubiquitus* (Jordan).
297. *Bacillus candicans* (Frankland).
298. *Bacillus albus* (Eisenberg).
299. *Bacillus acidi lactici* (Hueppe).
300. *Bacillus limbatus acidi lactici* (Marpmann).
301. *Bacillus lactis pituitosi*.
Bacillus der schleimigen Milch (Löffler).
302. *Bacillus aërogenes* (Miller).
303. *Bacterium aërogenes* (Miller).
304. *Heliobacterium aërogenes* (Miller).
305. *Bacillus aquatilis sulcatus* No. I. (Weichselbaum).
306. *Bacillus aquatilis sulcatus* No. II. (Weichselbaum).
307. *Bacillus aquatilis sulcatus* No. III. (Weichselbaum).
308. *Bacillus aquatilis sulcatus* No. IV. (Weichselbaum).
309. *Bacillus aquatilis sulcatus* No. V. (Weichselbaum).
310. *Bacillus multipediculus* (Flügge).
311. *Bacillus cystiformis* (Clado).
312. *Bacillus hepaticus fortuitus* (Sternberg).
313. *Bacillus intestinalis motilis* (Sternberg).
314. *Bacillus caviæ fortuitus* (Sternberg).
315. *Bacillus coli similis* (Sternberg).
316. *Bacillus filiformis Havaniensis* (Sternberg).
317. *Bacillus Martinez* (Sternberg).
318. *Bacillus epidermidis* (Bizzozero).
319. *Bacillus nodosus parvus* (Lustgarten).
320. *Bacillus hyacinthi septicus* (Heinz).
321. *Bacterium gliscrogenum* (Malerba).
322. *Bacillus ovatus minutissimus* (Unna).
323. Capsule bacilli of Smith.
324. *Bacillus putrificus coli* (Bienstock).
325. *Bacillus subtilis simulans* No. I. (Bienstock).
326. *Bacillus subtilis simulans* No. II. (Bienstock).
327. *Bacillus striatus albus* (Von Besser).
328. *Bacillus stolonatus* (Adametz).
329. *Bacillus ventriculi* (Raczynssky).
330. *Bacterium Zopfi* (Kurth).
331. *Bacterium Zürnianum* (List).
332. *Bacillus* of Colomiatti.
333. *Bacillus scissus* (Frankland).
334. *Bacillus* No. I. of Fulles.
335. *Bacillus* No. II. of Fulles.
336. *Bacillus phosphorescens gelidus* (Forster).

- 337. *Bacillus smaragdino-phosphorescens* (Katz).
- 338. *Bacillus argenteo-phosphorescens* No. I. (Katz).
- 339. *Bacillus argenteo-phosphorescens* No. II. (Katz).
- 340. *Bacillus argenteo-phosphorescens* No. III. (Katz).

D. Non-chromogenic, Liquefying Bacilli.

- 341. *Bacillus cyaneo-phosphorescens* (Katz).
- 342. *Bacillus argenteo-phosphorescens liquefaciens* (Katz).
- 343. *Bacillus phosphorescens Indicus* (Fischer).
- 344. *Bacillus phosphorescens indigenus* (Fischer).
- 345. *Bacillus circulans* (Jordan).
- 346. *Bacillus superficialis* (Jordan).
- 347. *Bacillus reticularis* (Jordan).
- 348. *Bacillus hyalinus* (Jordan).
- 349. *Bacillus cloacæ* (Jordan).
- 350. *Bacillus delicatulus* (Jordan).
- 351. *Bacillus aquatilis* (Frankland).
- 352. *Bacillus diffusus* (Frankland).
- 353. *Bacillus liquidus* (Frankland).
- 354. *Bacillus vermicularis* (Frankland).
- 355. *Bacillus nubilus* (Frankland).
- 356. *Bacillus pestifer* (Frankland).
- 357. *Bacillus filiformis* (Tils).
- 358. *Bacillus devorans* (Zimmermann).
- 359. *Bacillus gracilis* (Zimmermann).
- 360. *Bacillus guttatus* (Zimmermann).
- 361. *Bacillus implexus* (Zimmermann).
- 362. *Bacillus punctatus* (Zimmermann).
- 363. *Bacillus radiatus aquatilis* (Zimmermann).
- 364. *Bacillus vermiculosus* (Zimmermann).
- 365. *Bacillus aërophilus* (Liborius).
- 366. *Bacillus mycoides* (Flügge).
- 367. *Bacillus mesentericus vulgatus* (Flügge).
Kartoffelbacillus.
- 368. *Bacillus mesentericus fuscus* (Flügge).
- 369. *Bacillus megatherium* (De Bary).
- 370. *Bacillus albus putidus* (De Bary).
- 371. *Bacillus brassicæ* (Pommer).
- 372. *Bacillus butyricus* of Hueppe.
- 373. *Bacillus gasoformans* (Eisenberg).
- 374. *Bacillus carabiformis* (Kaczynsky).
- 375. *Bacillus graveolens* (Bordoni-Uffreduzzi).
- 376. *Bacillus carotarum* (A. Koch).

- 377. *Bacillus inflatus* (A. Koch).
- 378. *Bacillus ramosus*.
Wurtzel bacillus.
- 379. *Bacillus subtilis* (Ehrenberg).
- 380. *Bacillus subtilis similis* (Sternberg).
- 381. *Bacillus leptosporus* (L. Klein).
- 382. *Bacillus sessilis* (L. Klein).
- 383. *Bacillus allantoides* (L. Klein).
- 384. *Bacillus* of Scheurlen.
- 385. *Bacillus lactis albus* (Löffler).
- 386. *Bacillus liodermos* (Löffler).
- 387. *Bacillus ulna* (Cohn).
- 388. *Bacillus ulna* of Vignal.
- 389. *Bacillus liquefaciens* (Eisenberg).
- 390. *Bacillus maidis* (Cuboni).
- 391. *Proteus sulfureus* (Lindenborn).
- 392. *Bacillus thermophilus* (Miquel).
- 393. *Bacillus tumescens* (Zopf).
- 394. *Bacillus buccalis maximus* (Miller).
- 395. *Leptothrix buccalis* of Vignal.
- 396. *Bacillus b* of Vignal.
- 397. *Bacillus f* of Vignal.
- 398. *Bacillus buccalis fortuitus*.
Bacillus j of Vignal.
- 399. *Bacillus Havaniensis liquefaciens* (Sternberg).
- 400. *Bacillus liquefaciens communis* (Sternberg).

E. Strictly Anaërobic Bacilli.

- 401. *Bacillus muscoides* (Liborius).
- 402. *Bacillus solidus* (Lüderitz).
- 403. *Bacillus polypiformis* (Liborius).
- 404. *Bacillus butyricus* (Prazmowski).
Bacillus amylobacter.
Clostridium butyricum.
- 405. *Clostridium foetidum* (Liborius).
- 406. *Bacillus liquefaciens magnus* (Lüderitz).
- 407. *Bacillus liquefaciens parvus* (Lüderitz).
- 408. *Bacillus radiatus* (Lüderitz).
- 409. *Bacillus spinosus* (Lüderitz).
- 410. *Bacillus anaërobicus liquefaciens* (Sternberg).

PART FOURTH, SECTION X.—NON-PATHOGENIC SPIRILLA.

- 411. *Spirillum sputigenum* (Miller).
- 412. *Spirillum dentium*.

- Spirochæte denticola*.
 413. *Spirillum plicatile*.
 Spirochæte plicatilis (Ehrenberg).
 414. *Vibrio rugula* (Müller).
 415. *Spirillum volutans* (Ehrenberg).
 416. *Spirillum sanguineum* (Warming).
 Ophidomonas sanguinea.
 417. *Spirillum serpens* (Müller).
 418. *Spirillum undula* (Ehrenberg).
 419. *Spirillum tenue* (Ehrenberg).
 420. *Spirillum linguæ*.
 Vibrio lingualis (Weibel).
 421. *Spirillum nasale*.
 Vibrio nasalis.
 Nasenschleimvibrio (Weibel).
 422. *Spirillum* α of Weibel.
 Vibrio saprophiles α (Weibel).
 423. *Spirillum* β of Weibel.
 Vibrio saprophiles β (Weibel).
 424. *Spirillum* γ of Weibel.
 Vibrio saprophiles γ (Weibel).
 425. *Spirillum aureum*.
 Vibrio aureus (Weibel).
 426. *Spirillum flavescens*.
 Vibrio flavescens (Weibel).
 427. *Spirillum flavum*.
 Vibrio flavus (Weibel).
 428. *Spirillum concentricum* (Kitasato).
 429. *Spirillum rubrum* (Von Esmarch).
 430. *Spirillum* of Smith.
 431. *Spirillum* of Miller.
 Miller's bacillus.

PART FOURTH, SECTION XI.—LEPTOTRICHEÆ AND
 CLADOTRICHEÆ.

432. *Crenothrix Kühniana* (Rabenhorst).
 433. *Beggiatoa alba* (Vauch.).
 434. *Beggiatoa roseo-persicina* (Zopf).
 Clathrocystis roseo-persicina (Cohn).
 Ophidomonas sanguinea (Ehrenberg).
 Bacterium rubescens (Lankester).
 435. *Beggiatoa mirabilis* (Cohn).
 436. *Phragmidiothrix multiseptata* (Engler).

- 437. *Cladothrix dichotoma* (Cohn).
- 438. *Cladothrix Foersteri*.
Streptothrix Foersteri (Cohn).
- 439. *Cladothrix intricata* (Russell).

PART FOURTH, SECTION XII.—ADDITIONAL SPECIES OF BACTERIA,
NOT CLASSIFIED.

- 440. *Nitromonas* of Winogradsky.
- 441. Nitrifying bacillus of Winogradsky.
- 442. *Streptococcus conglomeratus* (Kurth).
- 443. *Bacillus thalassophilus* (Russell).
- 444. *Bacillus granulosus* (Russell).
- 445. *Bacillus limosus* (Russell).
- 446. *Spirillum marinum* (Russell).
- 447. *Bacillus litoralis* (Russell).
- 448. *Bacillus halophilus* (Russell).
- 449. *Bacillus capsulatus mucosus* (Fasching).
- 450. *Bacillus* of potato rot (Kramer).
- 451. *Bacillus vacuolosis* (Sternberg).
- 452. *Bacillus* of Dantec.
- 453. *Bacillus Havaniensis* (Sternberg).
- 454. *Bacillus amylozyma* (Perdrix).
- 455. *Bacillus rubellus* (Okada).
- 456. *Bacterium ureæ* (Jaksch).
- 457. *Sarcina mobilis* (Maurea).
- 458. *Bacillus stoloniferus* (Pohl).
- 459. *Bacillus incanus* (Pohl).
- 460. *Bacillus inunctus* (Pohl).
- 461. *Bacillus flavescens* (Pohl).
- 462. *Bacillus butyricus* of Botkin.
- 463. *Urobacillus Pasteuri* (Miquel).
- 464. *Urobacillus Duclauxi* (Miquel).
- 465. *Urobacillus Freudenreichi* (Miquel).
- 466. *Urobacillus Maddoxi* (Miquel).
- 467. *Urobacillus Schützenbergi* (Miquel).
- 468. *Bacillus* of Bovet.
- 469. *Bacillus Schaffer* (Freudenreich).
- 470. Bacilli of Guillebeau—*a*, *b*, *c*—(Freudenreich).
- 471. *Micrococcus Freudenreichi* (Guillebeau).
- 472. *Bacterium Hessii* (Guillebeau).
- 473. *Bacillus denitrificans* (Giltay and Aberson).
- 474. *Bacillus cyano-fuscus* (Beyerinck).
- 475. *Bacillus pyogenes soli* (Bolton).

- 476. *Micrococcus aquatilis invisibilis* (Vaughan).
- 477. *Bacillus gracilis anaërobiescens* (Vaughan).
- 478. *Bacillus figurans* (Vaughan).
- 479. *Bacillus albus anaërobiescens* (Vaughan).
- 480. *Bacillus invisibilis* (Vaughan).
- 481. *Bacillus venenosus* (Vaughan).
- 482. *Bacillus venenosus brevis* (Vaughan).
- 483. *Bacillus venenosus invisibilis* (Vaughan).
- 484. *Bacillus venenosus liquefaciens* (Vaughan).
- 485. *Bacillus aërogenes capsulatus* (Welch).
- 486. *Bacillus* of Canon and Pielicke.
- 487. *Bacillus sanguinis typhi* (Brannan and Cheesman).
- 488. *Micrococcus agilis citreus* (Menge).
- 489. *Bacillus gracilis cadaveris* (Sternberg).

BACTERIOLOGICAL DIAGNOSIS.

MICROCOCCI.

- I. *Staphylococci*—micrococci, solitary, in pairs, in irregular groups, and occasionally in short chains or in groups of four.
 - A. Grow in nutrient gelatin at the room temperature (20° to 22° C.) and *liquefy* the gelatin.
 - a. Chromogenic ; pigment yellow:
 - Staphylococcus pyogenes aureus* (1).
 - Staphylococcus pyogenes citreus* (3).
 - Micrococcus flavus liquefaciens* (159).
 - Micrococcus citreus liquefaciens* (164).
 - Micrococcus flavus desidens* (160).
 - Micrococcus cremoides* (169).
 - Micrococcus* of Almquist (41).
 - Micrococcus Finlayensis* (210).
 - Staphylococcus salivarius pyogenes* (10).
 - a. Pigment red or pink :
 - Micrococcus fuscus* (162).
 - Micrococcus roseus* (170).
 - Micrococcus agilis* (161).
 - b Non-chromogenic :
 - Staphylococcus pyogenes albus* (2).
 - Staphylococcus pyosepticus* (42).
 - Micrococcus* of Freire (211).
 - Micrococcus albus liquefaciens* (188).
 - Micrococcus* of gangrenous mastitis in sheep (30).
 - Micrococcus ureæ liquefaciens* (203).
 - Micrococcus aërogenes* (187).

Micrococcus radiatus (190).

Micrococcus foetidus (189).

B. Grow in nutrient gelatin at the room temperature (20° to 22° C.), and do not liquefy the gelatin.

a. Chromogenic ; pigment yellow:

Micrococcus versicolor (173).

Micrococcus aurantiacus (171).

Micrococcus cereus flavus (178).

Micrococcus flavus tardigratus (181).

Micrococcus luteus (182).

Micrococcus agilis citreus (488).

a. Pigment greenish-yellow :

Staphylococcus viridis flavescens (184).

a. Pigment violet :

Micrococcus violaceus (183).

a. Pigment red or pink :

Micrococcus carneus (175).

Micrococcus cinnabareus (176).

Micrococcus cerasinus siccus (172).

b. Non-chromogenic :

Micrococcus candicans (192).

Micrococcus cereus albus (177).

Micrococcus concentricus (198).

Micrococcus fervidosus (180).

Micrococcus of bovine pneumonia ? (22).

Micrococcus acidi lactici (194).

Micrococcus aquatilis (197).

Micrococcus cumulatus tenuis (199).

Micrococcus ureæ (202).

Micrococcus of bovine mastitis (21).

Micrococcus salivarius septicus (15).

Micrococcus gingivæ pyogenes (35).

Micrococcus rosettaceus (201).

Micrococcus aquatilis invisibilis (476).

C. Do not grow in nutrient gelatin at the room temperature :

Micrococcus endocarditidis rugatus (29).

Nitromonas of Winogradsky (440).

D. Biological characters not determined :

Micrococcus pyogenes tenuis (4).

Micrococcus of progressive abscess formation in mice (12)

Micrococcus of pyæmia in rabbits (13).

Micrococcus of Forbes (44).

Nosema bombycis (25).

II. Micrococci united in *zoöglæa* masses by an intercellular substance:

- Micrococcus viticulus (204).
- Micrococcus plumosus (200).
- Micrococcus candidus (193).
- Micrococcus amylovorus (234).
- Ascococcus Billrothi (235).
- Leuconostoc mesenteroides (236).

III. Micrococci in pairs—*diplococci*, not forming chains.A. Grow in nutrient gelatin at the room temperature (20° to 22° C.), and *liquefy* the gelatin.

a. Chromogenic ; pigment yellow :

- Diplococcus subflavus (16)—stains by Gram's method.
- Micrococcus botryogenus (19).
- Diplococcus flavus liquefaciens tardus (165).

a. Pigment red :

- Diplococcus roseus (168).

b. Non-chromogenic :

- Diplococcus albicans amplius (191).
- Micrococcus of Heydenreich (26).

B. Grow in nutrient gelatin at the room temperature, and *do not liquefy*; non-chromogenic.

a. Stain by Gram's method :

- Micrococcus of Manfredi (20).
- Micrococcus of trachoma ? (17).

b. Do not stain by Gram's method :

- Diplococcus of pneumonia in horses (32).
- Hæmatococcus bovis (34).
- Diplococcus albicans tardissimus (205).

c. Staining by Gram's method not determined :

- Diplococcus coryzæ (209).

C. Do not grow in nutrient gelatin at the room temperature ; non-chromogenic ; do not stain by Gram's method :

- Diplococcus intercellularis meningitidis (9).
- Micrococcus gonorrhœæ (6).
- Micrococcus of Kirchner (38).
- Micrococcus of Demme (27).

D. Biological characters imperfectly determined :

- Micrococci of Disse and Taguchi (p. 404).

IV. Micrococci which multiply by division in one direction only, forming *diplococci* and *streptococci*.A. Grow in nutrient gelatin at the room temperature (20° to 22° C.), and *liquefy* the gelatin.

- a Chromogenic , pigment yellow :
 - Diplococcus luteus (167).
- a. Pigment green :
 - Diplococcus fluorescens foetidus (166).
- b. Non-chromogenic.
 - 1 Grow on potato :
 - Micrococcus No. II. of Fischel (39).
 - Micrococcus lactis viscosus (195).
 - Streptococcus liquefaciens (219).
 - Streptococcus of Manneberg (28).
 - Streptococcus coli gracilis (212).
 - Micrococcus Freudenreichi (471).
 - Streptococcus albus (215).
 - 2. Does not grow on potato :
 - Streptococcus septicus liquefaciens (37).
- B. Grow in nutrient gelatin at the room temperature, and *do not liquefy*.
 - a. Chromogenic ; pigment yellow :
 - Micrococcus citreus (179).
 - Micrococcus ochroleucus (185).
 - b. Non-chromogenic.
 - 1. Grow on potato :
 - { Streptococcus brevis (217).
 - { Streptococcus cadaveris.
 - Pseudodiplococcus pneumoniae (36).
 - Streptococcus vermiformis (216).
 - 2. Do not grow on potato ; coagulate milk :
 - Streptococcus pyogenes (5).
 - Streptococcus of mastitis in cows (31).
 - 3. Growth on potato not stated :
 - Streptococcus coryzae contagiosa equorum (33).
 - Streptococcus septicus (23).
 - Streptococcus acidi lactici (213).
 - Streptococcus conglomeratus (442).
- C. Do not grow in nutrient gelatin at the room temperature :
 - Micrococcus pneumoniae crouposa (8).
 - Streptococcus of Bonome (40).
 - Streptococcus giganteus urethrae (214).
- D. Biological characters imperfectly known :
 - Streptococcus perniciosus psittacorum (43).
 - Streptococcus bombycis (24).
 - Streptococcus Havaniensis (218).
 - Micrococcus of progressive tissue necrosis in mice (11).

V. Micrococci which multiply by division in two directions, forming *diplococci* and *tetrads*.

A. Grow in nutrient gelatin at the room temperature, and *liquefy* the gelatin.

a. Chromogenic ; pigment yellow :

Diplococcus citreus conglomeratus (163).

Micrococcus tetragenus versatilis (220).

b. Non-chromogenic :

Micrococcus acidi lactici liquefaciens (186).

Pediococcus albus (221).

B. Grow in nutrient gelatin at the room temperature and *do not liquefy*.

Non-chromogenic :

Pediococcus acidi lactici (222).

Micrococcus tetragenus mobilis ventriculi (224).

Pediococcus cerevisiæ (223).

Micrococcus tetragenus (18).

C. Do not grow in nutrient gelatin at the room temperature :

Micrococcus tetragenus subflavus (225).

Micrococcus gonorrhœæ (6).

VI. Micrococci which multiply by division in three directions, forming cubical "packets"—*sarcinæ*.

A. Grow in nutrient gelatin at the room temperature, and *liquefy* the gelatin.

a. Chromogenic ; pigment yellow :

Sarcina aurantiaca (226).

Sarcina lutea (227).

Sarcina flava (228).

a. Pigment red :

Sarcina rosea (229).

Sarcina mobilis (457).

b. Non-chromogenic:

Sarcina alba (230).

Sarcina candida (231).

B. Grow in nutrient gelatin at the room temperature, and *do not liquefy*.

Non-chromogenic :

Sarcina ventriculi (233).

Sarcina pulmonum (232).

BACILLI.

I. *Aërobic bacilli* (many of the bacilli in this group are facultative anaërobics).A. Grow in nutrient gelatin at the room temperature (20° to 22° C.). and *liquefy* the gelatin.

a. Chromogenic ; pigment yellow :

1. Motile ; grow on potato :

Bacillus ochraceus (265).*Bacillus buccalis minutus* (294).*Bacillus plicatilis* (266).*Bacillus citreus cadaveris* (273).*Bacillus fulvus* (263).*Bacillus arborescens* (272).

2. Non-motile or undetermined ; grow on potato :

Bacillus cuticularis (270).*Bacillus hydrophilus fuscus* (81).*Ascobacillus citreus* (275).*Bacillus helvolus* (264).*Bacterium termo* of Vignal (293).*Bacillus lactis erythrogenes* (280).

Does not grow upon potato ; phosphorescent :

Bacillus argenteo-phosphorescens liquefaciens (342).

a. Pigment greenish-yellow or green ; grow on potato.

1. Motile :

Bacillus fluorescens liquefaciens (277).*Bacillus fluorescens liquefaciens minutissimus* (278).*Bacillus fluorescens nivalis* (279).*Bacillus chromo-aromaticus* (134).*Bacillus viscosus* (289).*Bacillus pyocyaneus* (96).*Bacillus pyocyaneus* β (286).

2. Non-motile :

Bacillus smaragdinus foetidus (78).

a. Pigment violet or blue ; grow on potato.

1. Motile :

Bacillus violaceus (290).*Bacillus violaceus Laurentius* (268).*Bacillus janthinus* (267).*Bacillus lividus* (282).*Bacillus cyano-fuscus* (474).

2. Non-motile :

Bacillus membranaceus amethystinus (274).*Bacillus cœruleus* (276).

a. Pigment red, pink, or brown.

1. Motile.

† Form spores :

Bacillus of Canestrini (295).

Bacillus of Dantec (452).

Bacillus mesentericus ruber (285).

* Spore formation not observed :

Bacillus rubidus (292).

Bacillus sulfureum (291).

Bacillus rosaceum metalloides (288).

Flesh-colored bacillus (271).

Bacillus Indicus (283).

2. Non-motile ; spore formation not observed :

Bacillus mycoides roseus (287).

Bacillus glaucus (281).

Bacillus prodigiosus (284).

Bacillus tremelloides (269).

b. Non-chromogenic.

1. Motile.

† Form spores :

Bacillus subtilis (379).

Bacillus subtilis similis (380).

Bacillus of Scheurlen (384).

Bacillus Hessii (472).

Bacillus circulans (345).

Bacillus mycoides (366).

Bacillus mesentericus vulgatus (367).

Bacillus mesentericus fuscus (368).

Bacillus tumescens (393).

Bacillus alvei (140).

Bacillus butyricus, Hueppe (372).

Bacillus liodermos (386).

Bacillus ramosus (378).

Bacillus megatherium (369).

Bacillus of potato rot (450).

Bacillus maidis (390).

Bacillus inflatus (377).

Bacillus gracilis (359).

Bacillus lactis albus (385).

Vibrio rugula (414).

Bacillus limosus (445).

Urobacillus Maddoxi (466).

Bacillus vacuolosis (451).

Urobacillus Pasteuri (463).

Urobacillus Duclauxi (464).

Urobacillus Freudenreichi (465).

* Spore formation not observed.

x. Grow on potato :

Bacillus radiatus aquatilis (363).

Bacillus vermiculosus (364).

Bacillus guttatus (360).

Bacillus pestifer (356).

Bacillus nubilus (355).

Bacillus albus putidus (370).

Bacillus punctatus (362).

Bacillus hyalinus (348).

Bacillus cloacæ (349).

Bacillus liquefaciens (389).

Bacillus liquidus (353).

Bacillus diffusus (352).

Bacillus delicatulus (350).

Bacillus foetidus ozænæ (111).

Bacillus septicus ulceris gangrænosi (127).

Bacillus albus cadaveris (129).

Bacillus leporis lethalis (94).

Bacillus liquefaciens communis (400).

Bacillus reticularis (347).

Proteus vulgaris (97)—polymorphous.

Proteus septicus (101)—polymorphous.

Proteus mirabilis (99)—polymorphous.

Proteus of Karlinsky (98)—polymorphous.

Proteus sulfureus (391)—polymorphous.

Urobacillus Schützenbergi (467).

Bacillus b of Guillebeau (470).

Bacillus figurans (478).

Bacillus venenosus liquefaciens (484).

Bacillus phosphorescens Indicus (343).

Bacillus litoralis (447).

Bacillus halophilus (448).

Bacillus stoloniferus (458).

y. Do not grow on potato :

Bacillus superficialis (346).

Bacillus devorans (358).

Bacillus aquatilis (351).

Bacillus Havaniensis liquefaciens (399).

Bacillus cyaneo-phosphorescens (341).

Bacillus phosphorescens indigenus (344).

z. Growth on potato not determined :

Bacillus gasoformans (373).*Bacillus* of Schou (114).*Bacillus carabiformis* (374).

2. Non-motile.

† Form spores :

Bacillus anthracis (45).*Bacillus brassicæ* (371).*Bacillus carotarum* (376).*Bacillus* of Tricomi (128).*Bacillus vermicularis* (354).*Bacillus* *aërophilus* (365).*Bacillus* of Letzerich (109).*Bacillus implexus* (361).*Bacillus filiformis* (357).*Bacillus granulosus* (444).

* Do not form spores, or undetermined :

Bacillus ulna of Vignal (388).*Leptothrix buccalis* of Vignal (395).*Bacillus f* of Vignal (397).*Bacillus b* of Vignal (396).*Bacillus buccalis fortuitus* (398).*Bacillus varicosus conjunctivæ* (130).*Bacillus pulpæ pyogenes* (124).*Bacillus gingivæ pyogenes* (122).*Bacillus graveolens* (375).*Pneumobacillus liquefaciens bovis* (120).*Bacillus incanus* (459).*Bacillus inunctus* (460).B. Grow in nutrient gelatin at the room temperature, and *do not liquefy*.

a. Chromogenic ; pigment yellow.

1. Motile ; grow on potato ; spore formation not observed :

Bacillus aurantiacus (238).*Bacillus subflavus* (256).*Bacillus constrictus* (243).*Bacillus aureus* (240).*Bacillus fluorescens aureus* (244).*Bacillus heminecrobiophilus* (148).*Bacillus flavescens* (461).

2. Non-motile ; spore formation not observed :

Bacillus spiniferus (260)—grows on potato.*Bacillus* of cholera in ducks (62)—grows on potato.

Bacillus fuscus (253)—grows on potato.

Bac. of Tizzoni and Giovannini (145)—grows on potato.

Bacillus flavocoriaceus (241).

Bacillus luteum (237).

3. Motility not determined ; grows on potato :

Bacillus striatus flavus (255).

a. Pigment yellowish-green or green.

1. Motile ; grow on potato.

† Form spores :

Bacillus of Lesage (106).

Bacillus erythrosporus (249).

* Do not form spores :

Bacillus fluorescens longus (245).

Bacillus fluorescens tenuis (246).

Bacillus virescens (251).

Bacillus fluorescens putidus (248).

Bacillus canalis parvus (136).

Bacillus dentalis viridans (123).

2. Non-motile ; do not form spores ; grow on potato :

Bacillus fluorescens non-liquefaciens (247).

Bacillus iris (252).

a. Pigment violet or blue ; grow on potato.

1. Motile.

† Forms spores (?) :

Bacillus cyanogenus (257).

* Do not form spores, or undetermined.

Bacillus viridis pallescens (250).

Bacillus beroliniensis Indicus (242).

Bacillus cyanogenus Jordaniensis (257).

a. Pigment red, pink, or brown.

1. Motile ; grow on potato ; spore formation not observed :

Bacillus rubescens (261).

Bacillus rubefaciens (254).

Bacillus fuscus limbatus (258).

2. Non-motile.

† Forms spores :

Bacillus brunneus (239).

* Do not form spores :

Bacillus latericeus (259).

Bacillus Havaniensis (453).

b. Non-chromogenic.

1. Motile.

† Form spores :

Bacillus of Afanassiew (119).

- *Bacillus* of Koubasoff (59).
- Bacillus putrificus coli* (324).
- Bacillus septicus vesicæ* (132).
- * Spore formation not observed :
 - x. Grow upon potato :
 - Bacillus endocarditidis griseus* (104).
 - Bacillus meningitidis purulentæ* (131).
 - Bacillus pyogenes fœtidus* (72).
 - Bacillus enteritidis* (75).
 - Bacillus œdematis aërobicus* (108).
 - Bacillus hyacinthis septicus* (320).
 - Bacillus gliscrogenum* (321).
 - Bacillus stolonatus* (328).
 - Bacillus albus* (298).
 - Bacillus aërogenes* (302).
 - Bacterium aërogenes* (303).
 - Bacillus aquatilis sulcatus* No. I. (305).
 - Bacillus aquatilis sulcatus* No. II. (306).
 - Bacillus aquatilis sulcatus* No. III. (307).
 - Bacillus aquatilis sulcatus* No. V. (309).
 - Heliobacterium aërogenes* (304).
 - Bacillus* No. I. of Fulles (334).
 - Proteus lethalis* (102).
 - Proteus Zenkeri* (100).
 - Bacillus* of hog cholera (63).
 - Bacillus* of swine plague, Marseilles (65).
 - Bacillus typhi abdominalis* (46).
 - Bacillus cavicida Havaniensis* (70).
 - Bacillus* No. I. of Roth (142).
 - Bacillus coli communis* (89).
 - Bacillus cuniculicida Havaniensis* (93).
 - Booker's bacilli, *d, e, f, g, h, k, n* (89).
 - Bacillus cavicida* (69).
 - Bacillus* of Bovet (468).
 - Bacillus Schaffer* (469).
 - Bacillus a* of Guillebeau (470).
 - Bacillus c* of Guillebeau (470).
 - Bacillus gracilis anaërobiescens* (477).
 - Bacillus invisibilis* (480).
 - Bacillus venenosus* (481).
 - Bacillus venenosus brevis* (482).
 - Bacillus venenosus invisibilis* (483).
 - y. Do not grow on potato :
 - Bacillus aquatilis sulcatus* No. IV. (308).

} Usually
not motile.

} Motility not
stated.

- Bacillus argenteo-phosphorescens* No. 1 (338).
Bacillus argenteo-phosphorescens No. 3 (340).
- z. Growth on potato undetermined :
Bacterium Zopfii (330).
Bacillus ventriculi (329).
Bacillus of Utpadel (139).
Bacillus cystiformis (311).
2. Non-motile.
- † Form spores :
Bacillus of Colomiatti (332).
Bacillus acidi lactici (Hueppe) (299).
Bacillus subtilis simulans No. I. (325).
Bacillus epidermidis (318).
Bacillus coprogenes foetidus (116).
- * Spore formation not observed.
- x. Grow on potato :
Bacillus diphtheriæ (47).
Bacillus diphtheriæ columbrarum (49).
Bacillus septicæmiæ hæmorrhagicæ (61).
Bacillus of Tommasoli (113).
Bacillus pyogenes soli (475).
Bacillus pneumosepticus (79).
Bacillus acidiformans (93).
Bacillus lactis aërogenes (91).
Bacillus capsulatus mucosus (449).
Bacterium ureæ (456).
Bacillus ubiquitus (296).
Bacillus scissus (333).
Bacillus of Gessner (133).
Bacillus of purpura hæmorrhagica of Kolb (147).
Bacillus of purpura hæmorrhagica of Babes (146).
Bacillus albus anaërobiescens (479).
Bacillus tenuis sputigenus (82).
Bacillus No. II. of Roth (143).
Bacillus of Schimmelbusch (110).
Bacillus crassus sputigenus (71).
Bacillus Zürnianus (331).
Bacillus of Fiocca (96).
Bacillus of intestinal diphtheria in rabbits (51).
Bacillus coli similis (315).
Bacillus limbatus acidi lactici (300).
Bacillus multipediculus (310).
Bacillus No. II. of Fulles (335).
Bacillus candicans (297).

- Bacillus lactis pituitosi* (301).
- Bacillus striatus albus* (327).
- Bacillus* of Belfanti and Pascarola (64).
- Bacillus hepaticus fortuitus* (312).
- Bacillus phosphorescens gelidus* (336).
- Bacillus smaragdino-phosphorescens* (337).

- "Capsule bacilli." {
- Bacillus* of Friedländer (7).
 - Bacillus* of rhinoscleroma (58).
 - Capsule bacilli of Smith (323).
 - Bacillus capsulatus* (80).
 - Bacillus canalis capsulatus* (135).
 - Proteus hominis capsulatus* (73).
 - Proteus capsulatus septicus* (74).

y. Do not grow on potato :

- Bacillus erysipelatos suis* (67).
- Bacillus gallinarum* (77).
- Bacillus* of grouse disease (76).
- Bacillus pseudotuberculosis* (121).
- Bacillus* of Okada (144).
- Bacillus filiformis* Havaniensis (316).
- Bacillus nodosus parvus* (319).
- Bacillus* Martinez (317).
- Bacillus argenteo-phosphorescens* No. II. (339).

z. Growth on potato not determined :

- Bacillus septicus keratomalaciæ* (125).
- Bacillus oxytocus perniciosus* (117).
- Bacillus* of acne contagiosa of horses (141).
- Bacillus endocarditidis capsulatus* (105).
- Pseudo-diphtheritic bacillus (48).

3. Motility not determined :

- Bacillus septicus agrigenus* (66).
- Bacillus ovatus minutissimus* (322).

C. Do not grow in nutrient gelatin at the room temperature, but have been cultivated in other media.

1. Motile :

- Bacillus mallei* (56)—grows on potato.
- Bacillus* of Lumnitz (112)—forms spores.

2. Non-motile :

- Bacillus tuberculosis* (53).
- Bacillus tuberculosis gallinarum* (54).
- Bacillus* of Demme (107).
- Bacillus* of Nocard (60)—grows on potato.
- Bacillus sanguinis typhi* (487).

3. Motility not determined :

- Bacillus necrophorus* (115).

Bacillus of Kartulis (138).

Bacillus septicus acuminatus (126).

Nitrifying bacillus of Winogradsky (441).

Bac. of Canon and Pielicke—measles bacillus ? (486).

D. Growth in nutrient gelatin not determined, but have been cultivated in other media.

a. Chromogenic.

† Pigment green :

Bacillus allii (262).

* Pigment blue :

Bacillus indigogenus (137).

b. Non-chromogenic.

1. Motile.

† Form spores :

Bacillus leptosporus (381).

Bacillus ulna (387).

* Spore formation not observed :

Bacillus allantoides (383).

2. Non-motile.

† Forms spores .

Bacillus sessilis (382).

* Spore formation not determined :

Bacillus coprogenes parvus (68).

II. *Strictly anaërobic bacilli.*

A. Grow in nutrient gelatin at the room temperature, and *liquefy* the gelatin.

1. Motile ; form spores :

Bacillus tetani (149).

Bacillus of symptomatic anthrax (152).

Bacillus oedematis maligni (150).

Bacillus spinosus (409).

Bacillus rubellus (455).

Bacillus butyricus of Botkin (462).

Clostridium foetidum (405).

Bacillus liquefaciens magnus (406).

Bacillus radiatus (408).

Bacillus thalassophilus (443).

2. Non-motile ; form spores :

Bacillus liquefaciens parvus (407).

Bacillus anaërobicus liquefaciens (410).

B. Grow in nutrient gelatin at the room temperature, and *do not liquefy*.

1. Motile ; form spores :

Bacillus polypiformis (403).

Bacillus solidus (402).

Bacillus amylozyma (454).

2. Non-motile ; forms spores :

Bacillus muscoides (401).

3. Non-motile ; does not form spores :

Bacillus aërogenes capsulatus (485).

C. Growth in nutrient gelatin not determined, but have been cultivated in other media.

1. Motile ; forms spores :

Bacillus butyricus (404).

2. Non-motile ; spore formation not observed :

Bacillus cadaveris (151).

III. Have not been cultivated in artificial media :

Bacillus lepræ (cultivation claimed) (54).

Bacillus diphtheriæ vitulorum (50).

Bacillus of Lustgarten (57).

Bacillus buccalis maximus (394).

SPIRILLA.

Aërobic spirilla (many of the species are also facultative anaërobics).

A. Grow in nutrient gelatin at the room temperature, and *liquefy* the gelatin.

1. Motile ; grow upon potato ; spore formation not determined :

Spirillum cholerae Asiaticæ (155).

Spirillum of Finkler and Prior (156).

Spirillum tyrogenum (157).

Spirillum Metschnikovi (158).

Spirillum of Miller (431).

Spirillum marinum (446).

B. Grow in nutrient gelatin, and *do not liquefy* :

a. Chromogenic :

† Pigment yellow ; non-motile :

Spirillum flavum (427).

Spirillum aureum (425).

* Pigment yellowish-green ; non-motile :

Spirillum flavescens (426).

‡ Pigment red ; motile :

Spirillum rubrum (429).

b. Non-chromogenic :

† Motile :

z. Grow on potato :

Spirillum saprophiles α (422).

Spirillum saprophiles β (423).

Spirillum saprophiles γ (424).

Spirillum of Smith (430).

y. Does not grow on potato :

Spirillum concentricum (428).

* Non-motile :

Spirillum nasale (421).

Spirillum linguæ (420).

C. Biological characters not determined.

Pathogenic :

Spirillum Obermeieri (153).

Spirillum anserum (154).

Non-pathogenic, or undetermined :

Spirillum serpens (417).

Spirillum sanguineum (416).

Spirillum dentium (412).

Spirillum tenue (419).

Spirillum undula (418).

Spirillum volutans (415).

Spirillum sputigenum (411).

Spirillum plicatile (413).

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